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Utility of the detection of *Plasmodium* parasites for the diagnosis of malaria in endemic areas

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Abstract

Background: In populations where the prevalence of infection with *Plasmodium* parasites is high, blood tests that identify *Plasmodium* parasites in patients with fever may lead to false positive diagnosis of malaria-disease. We characterised the diminishing value of the parasite detection test as a function of the prevalence of infection.

Methods: We computed the ability of the parasite detection test to identify malaria at various levels of prevalence (0% to 90%), assuming plausible estimates of sensitivity (95% and 85%) and specificity (99% and 95%) for the detection of parasites. In each situation, we computed likelihood ratios of malaria (or absence of malaria) for positive and negative parasite detection tests. Likelihood ratios were classified as clinically useful (≥ 10), intermediate (5–10), or unhelpful (<5).

Results: Likelihood ratios of positive tests were strongly related to the prevalence of infection in the general population: a positive test was unhelpful when the prevalence was 20% or more, and useful only when prevalence was 5% or less. The sensitivity and specificity of the test had little influence on these results. Likelihood ratios of negative tests were clinically useful when prevalence was 70% or less, but only for high levels of sensitivity (95%). If sensitivity was low (85%), the negative test was at best of intermediate utility, and was unhelpful if the prevalence of asymptomatic infection exceeded 30%.

Conclusion: Identification of *Plasmodium* parasites supports a diagnosis of malaria only in areas where the prevalence of *Plasmodium* infection is low. Wherever this prevalence exceeds about 20%, a positive test is clinically unhelpful.

Background

In current practice, the diagnosis of malaria in a patient in whom this disease is suspected rests on the identification of *Plasmodium* parasites in the patient's blood [1]. This diagnostic method is currently recommended by the World Health Organization [2]. Parasite detection tests

include the classical methods of the thick or thin blood smear, and various rapid diagnostic tests [3].

This approach works well in cases of traveller's malaria, and more generally whenever the patient is unlikely to be an asymptomatic carrier of *Plasmodium* parasites. However, in populations where malaria is common, many

people are infected by Plasmodium parasites without being sick. In such situations, using the presence of parasites as a criterion for diagnosing malaria in a febrile patient will lead to over-diagnosis of malaria, unnecessary anti-malarial treatments, and missed diagnoses of other febrile illnesses. While this point has been made by experts previously [1], it is usually framed in general terms, without precise guidance as to when a positive or a negative parasite detection test is clinically useful, and when it is not. In this paper, we examine the influence of the prevalence of asymptomatic parasitic infection on the clinical utility of parasite detection for the diagnosis of malaria in patients with fever.

Methods

We examined various hypotheses regarding test performance and the prevalence of parasitic infection in the general population. A prevalence <20% is sometimes qualified as low, 20–50% as intermediate, and >50% as high.

We distinguish between the ability of a test to detect parasites in a person's blood, i.e., infection, from its ability to identify malaria as clinical disease in a patient with fever. The former is a general characteristic of the test, independent of the prevalence of parasitic infection. The latter pertains to a test used in specific epidemiologic circumstances, and is related to the prevalence of infection (Box 1).

Assessing the utility of a diagnostic test

Classic measures of the performance of a diagnostic test are its sensitivity (Sn) – i.e., the proportion of positive tests among those with the disease – and its specificity (Sp) – i.e., the proportion of negative tests among those without the disease. However, these statistics are not directly applicable in clinical practice, for if we knew who had the disease and who did not, we would not need the test. The clinician is typically interested in the test's positive predictive value – i.e., the proportion of diseased people among those with a positive test, and in its negative predictive value – i.e., the proportion of non-diseased people among those with a negative test. The drawback of these statistics is that they depend strongly on the pre-test probability of disease, which varies from patient to patient.

The statistic that best characterises the clinical utility of a test result is the likelihood ratio (LR) [5-7]. The likelihood ratio is the ratio of probabilities of a given test result under the 2 hypotheses under consideration – i.e., presence versus absence of disease. The likelihood ratio of a positive test is given by $LR+ = Sn/(1 - Sp)$, and the likelihood ratio of a negative test, by $LR- = (1 - Sn)/Sp$. To facilitate comparisons [6,7], likelihood ratios of a negative test are

sometimes shown for the absence of disease, i.e., $LR- = Sp/(1 - Sn)$. This merely inverts the likelihood ratio: a likelihood ratio of 0.1 for the disease becomes 10 for the absence of disease. We have adopted this convention in this paper.

The likelihood ratio allows the computation of the post-test probability of disease. According to the Bayes' theorem, post-test odds of disease are the product of the likelihood ratio and the pre-test odds [5-7]. Because probabilities can be converted into odds ($O = P/(1 - P)$), and odds into probabilities ($P = O/(O + 1)$), the likelihood ratio allows the transformation of a pre-test probability into a post-test probability (Figure 1). For example, if the pre-test probability of disease was 20% (i.e., pre-test odds = 1/4), and the test was positive, with a likelihood ratio of 8, the post-test odds of disease would be 2 (i.e., $8 \times (1/4)$), and the corresponding probability 66.7% (i.e., $2/(2+1)$).

This is how likelihood ratios can be interpreted [6]: a likelihood ratio is clinically useful if it equals 10 or more, of intermediate usefulness between 5 and 10, and clinically unhelpful below 5.

Detection of parasites in blood

To examine the performance of a parasite detection test for determining the presence or absence of Plasmodium parasites in the blood, we assumed a sensitivity of either 85% or 95%, and a specificity of either 95% or 99%. This

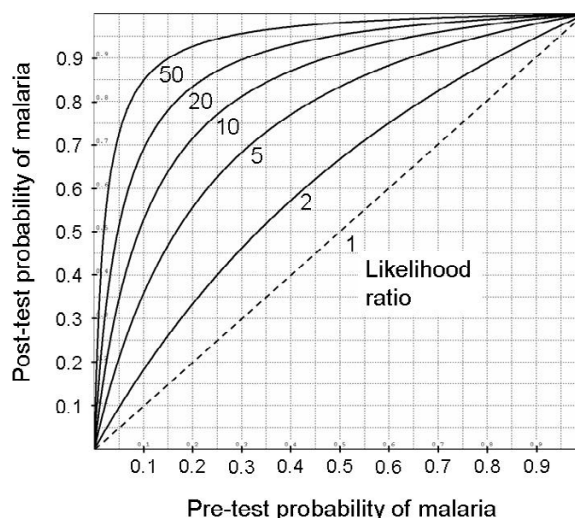


Figure 1 Post-test probability of malaria, as a function of pre-test probability of malaria, for 5 levels of the likelihood ratio of a positive test (2, 5, 10, 20 or 50). An uninformative test (likelihood ratio = 1) is shown as a diagonal dotted line.

means that among 100 persons who harbour parasites, the test will be positive for 85 or 95. The remaining are "false negatives", due to technical problems, or to the parasite being absent from peripheral blood for a portion of its life cycle [4]. Similarly, among 100 persons who do not harbour parasites, the test will be falsely positive for 1 to 5, due to various laboratory errors. These values of sensitivity and specificity can be considered as realistic in a well equipped and expertly managed laboratory.

Diagnosis of malaria

In a clinical context, the purpose of the test is not merely to detect parasites in the blood, but to establish or rule out the diagnosis of malaria in patients in whom the disease is suspected. Three types of patients should be considered (Table 1): a) patients with malaria, all of whom can be assumed to harbour parasites, b) patients with fever from another origin who are also infected – but not ill – with *Plasmodium*, and c) patients with fever from another origin who are not infected with *Plasmodium*. The probability that patients with fever from another origin are also infected with *Plasmodium* equals roughly the prevalence of parasitic infection in the general population. If this prevalence is nil, as is the case in much of the Western world, the performance of the test for diagnosing malaria is the same as for identifying the presence of parasites.

The sensitivity of the parasite detection test for detecting malaria (S_{n_m}) will be the same as its sensitivity for detecting parasites (S_{n_p}). However, the specificity of the test for

detecting malaria (S_{p_m}) will be influenced by the prevalence of infection, because the population under consideration is a mixture of two subgroups: patients who are infected with *Plasmodium* (proportion x), and of those who are not (proportion $1-x$). The specificity of the test for detecting malaria becomes $S_{p_m} = S_{p_p} \cdot (1 - x) + (1 - S_{n_p}) \cdot x$, and its complement, $1 - S_{p_m} = (1 - S_{p_p}) \cdot (1 - x) + S_{n_p} \cdot x$ (Box 1).

Consequently, the likelihood ratio of a positive test for the presence of malaria disease becomes

$$LR_{+m} = \frac{S_{n_p}}{(1 - S_{p_p}) \cdot (1 - x) + S_{n_p} \cdot x}$$

of a negative test for the absence of malaria is

$$LR_{-0m} = \frac{S_{p_p} \cdot (1 - x) + (1 - S_{n_p}) \cdot x}{1 - S_{n_p}}$$

Note that the proportion of *Plasmodium* carriage in the population (denoted by x in Table 1) is distinct from the pre-test probability of malaria in a given patient with fever (denoted by m in Table 1). The former is a characteristic of the population to which the patient belongs. The latter depends on various patient-related factors, such as the intensity and temporal pattern of fever, presence or absence of an enlarged spleen, anemia, pregnancy, age, exposure to mosquito bites, season, likelihood of alternate diseases, etc.

Table 1: Performance of Plasmodium detection tests in endemic populations: a) detection of the presence of parasites, b) diagnosis of a malaria. The parameters considered include the specificity of the test for the detection of parasites in blood (S_{n_p}), the corresponding specificity (S_{p_p}), the prevalence of malaria infection in the general population (x), and the pre-test probability of a malaria attack in a patient with fever (m)

a) Detection of parasites			
	Disease status		
	Malaria	Other febrile illness, parasites present	Other febrile illness, parasites absent
Positive parasite detection test	S_{n_p}	S_{n_p}	$1 - S_{p_p}$
Negative parasite detection test	$1 - S_{n_p}$	$1 - S_{n_p}$	S_{p_p}
Column totals	1	1	1
Prevalence of infection in the general population	-	x	$1 - x$

b) Detection of malaria (disease)		
	Malaria	Other febrile illness
Positive parasite detection test	S_{n_p}	$S_{n_p} \cdot x + (1 - S_{p_p}) \cdot (1 - x)$
Negative parasite detection test	$1 - S_{n_p}$	$(1 - S_{n_p}) \cdot x + S_{p_p} \cdot (1 - x)$
Column totals	1	1
Pre-test probability of malaria	m	$1 - m$

Effect of a higher threshold for a positive test

Thus far we have assumed that the test would perform equally well in patients who suffer from malaria and in those who are infected but not ill with malaria. But because parasite levels are likely higher in the former group, the test may discriminate better if a higher parasite threshold is used to diagnose malaria – say, greater than 1000 parasites per µL instead of greater than 0. Raising the diagnostic threshold will decrease the sensitivity of the test, but less so in patients with malaria ($Sn_{p-malaria}$) than in other patients ($Sn_{p-other}$). At the same time, raising the threshold will increase the specificity of the test to virtually 100%. Under these conditions, the likelihood ratio of a positive test for the presence of malaria becomes:

$$LR_{+m} = \frac{1}{x} \cdot \frac{Sn_{p-malaria}}{Sn_{p-other}}$$

negative test for the absence of malaria becomes

$$LR_{-0m} = \frac{1 - x \cdot Sn_{p-other}}{1 - Sn_{p-malaria}}$$

Analysis of test performance

We considered a range of prevalence of infection in the general population between 0% and 90%, and plausible values of sensitivity (0.85 and 0.95) and specificity (0.95 and 0.99) of the test for the detection of parasites. We show likelihood ratios of malaria for a positive test (Table 2), and of absence of malaria for a negative test (Table 3).

We also explored the effect of raising the diagnostic threshold, assuming that this will lower the sensitivity of the test, but less so in patients with malaria than in patients with an asymptomatic infection (Table 4). The computations were done using macro functions written for SPSS software.

Results

Detection of parasites

The likelihood ratios for the detection of parasites correspond to those of the detection of malaria when the prevalence of asymptomatic infection is assumed to be nil (first result lines in Tables 2 and 3). For a positive test, the likelihood ratio was very high (95 and 85) when specificity was modelled as 0.99, and is still clinically useful (at 19 and 17) when specificity was assumed to be 0.95 (Table 2). In contrast, the likelihood ratio of a negative test depended on the value of sensitivity: it was close to 20 when sensitivity was modelled as 0.95, but of only borderline utility, around 6, when sensitivity was entered as 0.85 (Table 3).

Detection of malaria

Likelihood ratios of malaria for a positive test varied considerably with the prevalence of *Plasmodium* infection (Table 2). When the prevalence was assumed to be 5% or less, the likelihood ratios were about 10 or more, in the clinically useful range. On the other hand, for a prevalence of 20% or more, the likelihood ratio dropped below 5, which would not be considered clinically useful. At low values of prevalence, the specificity of the test influenced

Table 2: Likelihood ratios of malaria when the parasite detection test is positive, according to the prevalence of infection in the general population, and to the sensitivity and specificity of the test for the detection of parasites. Clinically useful tests are in bolded type, tests of intermediate utility are in standard type, clinically unhelpful tests in italics

Prevalence of infection in the general population	Sensitivity = 0.95		Sensitivity = 0.85	
	Spec. = 0.99	Spec. = 0.95	Spec. = 0.99	Spec. = 0.95
0%	95.0	19.0	85.0	17.0
0.1%	86.8	18.7	78.4	16.7
1%	49.0	16.1	46.2	14.7
2%	33.0	14.0	31.7	12.9
5%	16.7	10.0	16.4	9.4
10%	9.1	6.8	9.0	6.5
15%	6.3	5.1	6.2	5.0
20%	4.8	4.1	4.8	4.0
30%	3.2	3.0	3.2	3.0
40%	2.5	2.3	2.5	2.3
50%	2.0	1.9	2.0	1.9
60%	1.7	1.6	1.6	1.6
70%	1.4	1.4	1.4	1.4
80%	1.2	1.2	1.2	1.2
90%	1.1	1.1	1.1	1.1

Table 3: Likelihood ratios of the absence of malaria when the parasite detection test is negative, according to the prevalence of infection in the general population, and to the sensitivity and specificity of the test for the detection of parasites. Clinically useful tests are in bolded type, tests of intermediate utility are in standard type, clinically unhelpful tests in italics

Prevalence of infection in the general population	Sensitivity = 0.95		Sensitivity = 0.85	
	Spec. = 0.99	Spec. = 0.95	Spec. = 0.99	Spec. = 0.95
0%	19.8	19.0	6.6	6.3
0.1%	19.8	19.0	6.6	6.3
1%	19.6	18.8	6.5	6.2
2%	19.4	18.6	6.5	6.2
5%	18.9	18.1	6.3	6.1
10%	17.9	17.2	6.0	5.8
15%	17.0	16.3	5.8	5.5
20%	16.0	15.4	5.5	5.3
30%	14.2	13.6	4.9	4.7
40%	12.3	11.8	4.7	4.2
50%	10.4	10.0	3.8	3.7
60%	8.5	8.2	3.2	3.1
70%	6.6	6.4	2.7	2.6
80%	4.8	4.6	2.1	2.1
90%	2.9	2.8	1.6	1.5

considerably test performance. Whether sensitivity was modelled as 0.85 or as 0.95 was of little import for the likelihood ratios.

The impact of the prevalence of infection on the usefulness of a negative test was also notable (Table 3). The negative test remained clinically useful up to a prevalence of about 50%, as long as the sensitivity was high (0.95). However, even a negative test was not clinically useful

when the prevalence exceeded 70%, or if the sensitivity of the test was assumed to be 0.85.

Impact of a higher threshold for a positive test

Likelihood ratios of presence of malaria for a positive test improved when the diagnostic threshold was increased. If the change of threshold decreased sensitivity to 0.80 in patients with malaria and to 0.60 in patients with asymptomatic infection, a positive test was clinically useful up to

Table 4: Likelihood ratios of positive test for presence of malaria if a high threshold is used to define a positive test. The sensitivity of the test is reduced, but less so for patients with malaria than for other infected individuals, and specificity is considered to be perfect (100%). Clinically useful tests are in bolded type, tests of intermediate utility are in standard type, clinically unhelpful tests in italics

Prevalence of infection in the general population	Sensitivity of test in patients with malaria: 0.80. Sensitivity of test in other infected patients:	
	0.60	0.40
0%	infinite	infinite
0.1%	1333	2000
1%	133	200
2%	67.7	100
5%	26.7	40.0
10%	13.3	20.0
15%	8.9	13.3
20%	6.7	10
30%	4.4	6.7
40%	3.3	5.0
50%	2.7	4.0
60%	2.2	3.3
70%	1.9	2.9
80%	1.7	2.5
90%	1.5	2.2

a prevalence of infection of 10% (Table 4). If the sensitivities were 0.80 and 0.40, the test became useful up to a prevalence of 20%. In contrast, such a change of threshold virtually abolished the utility of a negative test. Even in the best case scenario (prevalence of 0%), the likelihood ratio of a negative test for absence of malaria was 5, and it was less than 5 in all other situations (detailed results not shown).

Discussion

Globally, this analysis confirms that a positive parasite detection test is unhelpful in establishing the diagnosis of malaria in highly endemic areas [1], and defines a range of situations where a negative test is useful in excluding the diagnosis. Even in areas of intermediate prevalence of infection in the general population (20%–50%), a positive test for *Plasmodium* parasites in the blood does not establish the diagnosis of malaria. In such situations, the likelihood ratio is 5 or less, which will push the post-test probability of malaria above 90% only if the pre-test probability was already very high, greater than 70%. In highly endemic areas (>50%), a positive test is virtually meaningless, with likelihood ratios <2. While the general direction of these results is unsurprising, our study provides for the first time a definition of circumstances in which a parasite detection test is or is not useful for the diagnosis of malaria.

Positive parasite detection test

The likelihood ratio of a positive test depends strongly on the prevalence of infection in the general population, and is hardly influenced by the sensitivity or specificity of the test, within the range we considered. In fact, the formula for the likelihood ratio of a positive test ($LR(+)_m$) implies that this ratio cannot exceed the inverse of the prevalence of infection (or $1/x$), even if both sensitivity and specificity for the detection of parasites are a perfect 100%. For example, if half of the population is infected by *Plasmodium* parasites, the likelihood ratio cannot exceed 2. Consequently, if prevalence of infection is high, ruling in malaria cannot be improved much by the development of more sensitive parasite detection tests, such as those based on the polymerase chain reaction assays [9]. Therefore a positive *Plasmodium* detection test has little utility in populations where malaria is most frequent, as in Africa, where most of the burden of mortality and morbidity of malaria occurs [10].

Using a higher parasite density threshold to consider a test as positive may improve the value of a positive test [11]. This is because raising the threshold will reduce the sensitivity among patients who harbour parasites but do not have clinical malaria more than among patients with malaria. However, even this strategy will not extend the usefulness of the positive test beyond a prevalence of

asymptomatic infection greater than 20%. Furthermore, this strategy will invalidate the utility of a negative parasite detection test.

Negative parasite detection test

In contrast, the diagnostic contribution of the *negative* test varied considerably with the sensitivity of the test. When sensitivity was deemed equal to 0.95, the negative test produced high, clinically useful likelihood ratios up to a prevalence of infection of 70%, allowing the exclusion of malaria even in populations where *Plasmodium* infection is highly prevalent. On the other hand, when sensitivity was assumed to be only 0.85, the likelihood ratios of a negative test were lower, and exceeded 5 only up to a prevalence of asymptomatic infection of about 20%. Therefore the sensitivity of the *Plasmodium* detection test is critical for ruling out malaria in highly endemic populations. Ruling out malaria in patients with fever is important, to avoid unnecessary treatment with antimalarial drugs, which is common in highly endemic countries [1], and to provide appropriate etiologic treatment to the patients [12]. In this paper, we do not address the issue of malaria control at the population level, which may or may not include the treatment of asymptomatic carriers.

Good quality laboratory tests to rule out *Plasmodium* infection are therefore important, especially in highly endemic areas. Our estimates assumed rather high values of sensitivity and specificity of the parasite detection test. In precarious field conditions, these test properties may be worse [13,14]. Lower sensitivity would considerably reduce the clinical usefulness of a negative test. Unfortunately, given the insufficient quality of many field laboratories, the practical contribution of a *Plasmodium* detection test to the diagnosis of malaria is often limited [15].

Improving the diagnosis of malaria

Establishing the diagnosis of clinical malaria in endemic areas remains a challenge. As mentioned before, detection of *Plasmodium* infection in a patient presenting with a febrile acute illness is not a sufficient basis for the diagnosis. A useful contribution would be the identification of parameters, whether clinical variables or laboratory measurements, that distinguish patients who suffer from malaria from patients who have another febrile illness and merely harbour *Plasmodium* parasites. If clinical algorithms that yield a high enough pre-test probability of malaria were available, even a positive *Plasmodium* detection test associated with a modest likelihood ratio would help establish the diagnosis.

Several clinical algorithms have been proposed [16-20], but none seems sufficiently accurate as a basis for the diagnosis. A key difficulty in developing good diagnostic algo-

rhythms is the lack of a gold standard for diagnosing malaria. One cannot aim at a target that remains invisible. Development of adequate technologies to diagnose malaria in endemic areas remains an important and difficult public health challenge.

The difficulty in establishing the diagnosis of malaria increases with the prevalence of infection. It is noteworthy that reliance on positive *Plasmodium* detection tests remains justified in patients whose probability of asymptomatic parasitic infection is low, such as among travellers [14]. More generally, the issue of malaria diagnosis explored in this paper illustrates the difficulty in transferring a technological solution that is well suited to the developed world to other contexts.

Conclusion

In populations where the prevalence of *Plasmodium* infection is intermediate or high, a positive *Plasmodium* detection test does not help establish the diagnosis of malaria.

Abbreviations

Sn: Sensitivity

Sp: Specificity

LR: Likelihood ratio

Competing interests

The author(s) declare that they have no competing interests.

Authors' contributions

TVP and AR proposed the study and the study design. All authors discussed the interpretation of the results. TVP developed the statistical models, analysed data, and wrote the first draft. AR and TS performed the literature review and revised the paper.

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References

- Greenwood BM, Bojang K, Whitty CJ, Targett GA: **Malaria**. *Lancet* 2005, **365**:1487-98.
- Anonymous: **New perspectives. Malaria diagnosis**. Report of a Joint WHO/USAID Informal Consultation, 25–27 October 1999 2000 [http://www.who.int/tdr/publications/publications/pdf/malaria_diagnosis.pdf]. Geneva: World Health Organization
- Moody A: **Rapid diagnostic tests for malaria parasites**. *Clin Microbiol Rev* 2002, **15**:66-78.
- Delley V, Bouvier P, Breslow N, Doumbo O, Sagara I, Diakite M, Mauris A, Dolo A, Rougemont A: **What does a single determination of malaria parasite density mean? A longitudinal survey in Mali**. *Trop Med Int Health* 2000, **5**:404-12.
- Grimes DA, Schulz KF: **Refining clinical diagnosis with likelihood ratios**. *Lancet* 2005, **365**:1500-5.
- Jaeschke R, Guyatt GH, Sackett DL: **Users' guides to the medical literature. III. How to use an article about a diagnostic test. B: What are the results and will they help me in caring for**

- my patients? The Evidence-Based Medicine Working Group**. *JAMA* 1994, **271**:703-707.
- Weissler AM: **A perspective on standardizing the predictive power of noninvasive cardiovascular tests by likelihood ratio computation: I. Mathematical principles**. *Mayo Clin Proc* 1999, **74**:1061-71.
- Simel DL, Holleman DR: **Should the definition for the negative likelihood ratio be changed?** *J Clin Epidemiol* 1997, **50**:641-2.
- Hänscheid T, Grobusch MP: **How useful is PCR in the diagnosis of malaria?** *Trends Parasitol* 2004, **18**:395-8.
- Hay SI, Guerra CA, Tatem AJ, Noor AM, Snow RW: **The global distribution and population at risk of malaria: past, present, and future**. *Lancet Infect Dis* 2004, **4**:327-36.
- Boisier P, Jambou R, Raharimalala L, Roux J: **Relationship between parasite density and fever risk in a community exposed to a low level of malaria transmission in Madagascar highlands**. *Am J Trop Med Hyg* 2002, **67**:137-40.
- Reyburn H, Redempta M, Drakeley C, Carnero I, Mwakasungula E, Mwerinde O, Saganda K, Shao J, Kitua A, Olomi R, Greenwood BM, Whitty C: **Overdiagnosis of malaria in patients with severe febrile illness in Tanzania: a prospective study**. *BMJ* 2004, **329**:1212-15.
- Mason DP, Kawamoto F, Lin K, Laoboonchai A, Wongsrichanalai C: **A comparison of two rapid field immunochromatographic tests to expert microscopy in the diagnosis of malaria**. *Acta Trop* 2002, **82**:51-9.
- Marx A, Pewsner D, Egger M, Nuesch R, Bucher HC, Genton B, Hatz C, Jüni P: **Meta-analysis: accuracy of rapid tests for malaria in travelers returning from endemic areas**. *Ann Intern Med* 2005, **142**:836-46.
- Barat L, Chipipa J, Kolczak M, Sukwa T: **Does the availability of blood slide microscopy for malaria at health centers improve the management of persons with fever in Zambia?** *Am J Trop Med Hyg* 1999, **60**:1024-30.
- Chandramohan D, Jaffar S, Greenwood B: **Use of clinical algorithms for diagnosing malaria**. *Trop Med Int Health* 2002, **7**:45-52.
- Rougemont A, Breslow N, Brenner E, Moret AL, Dumbo O, Dolo A, Soula G, Perrin L: **Epidemiological basis for clinical diagnosis of childhood malaria in endemic zone in West Africa**. *Lancet* 1991, **338**:1292-5.
- Olaleye BO, Williams LA, D'Alessandro U, Weber MM, Mulholland K, Okorie C, Langerock P, Bennett S, Greenwood BM: **Clinical predictors of malaria in Gambian children with fever or a history of fever**. *Trans R Soc Trop Med Hyg* 1998, **92**:300-4.
- Bojang KA, Obaro S, Morison LA, Greenwood BM: **A prospective evaluation of a clinical algorithm for the diagnosis of malaria in Gambian children**. *Trop Med Int Health* 2000, **5**:231-6.
- Luxemburger C, Nosten F, Kyle DE, Kiricharoen L, Chongsuphajaisidhi T, White NJ: **Clinical features cannot predict a diagnosis of malaria or differentiate the infecting species in children living in an area of low transmission**. *Trans R Soc Trop Med Hyg* 1998, **92**:45-9.

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