

Physical exercise and liver "fitness": Role of mitochondrial function and epigenetics-related mechanisms in non-alcoholic fatty liver disease



Jelena Stevanović^{1,*}, Jorge Beleza², Pedro Coxito¹, António Ascensão¹, José Magalhães¹

ABSTRACT

Background: Modern lifestyles, especially high-caloric intake and physical inactivity, contribute to the increased prevalence of non-alcoholic fatty liver disease (NAFLD), which becomes a significant health problem worldwide. Lifestyle changes, however, affect not only parental generation, but also their offspring, reinforcing the need for efficient preventive approaches to deal with this disease. This transgenerational influence of phenotypes dependent on parents (particularly maternal) behaviours may open additional research avenues. Despite persistent attempts to design an effective pharmacological therapy against NAFLD, physical activity, as a non-pharmacological approach, emerges as an exciting strategy.

Scope of Review: Here we briefly review the effect of physical exercise on liver mitochondria adaptations in NAFLD, highlighting the importance of mitochondrial metabolism and transgenerational and epigenetic mechanisms in liver diseases.

Major Conclusions: A deeper look into cellular mechanisms sheds a light on possible effects of physical activity in the prevention and treatment of NAFLD through modulation of function and structure of particular organelles, namely mitochondria. Additionally, despite of increasing evidence regarding the contribution of epigenetic mechanisms in the pathogenesis of different diseases, the role of microRNAs, DNA methylation, and histone modification in NAFLD pathogenesis still needs to be elucidated.

© 2019 The Authors. Published by Elsevier GmbH. This is an open access article under the CC BY license (http://creativecommons.org/licenses/by/4.0/).

Keywords NAFLD; Physical exercise; Mitochondrial dysfunction; Epigenetics

1. INTRODUCTION

Sedentary lifestyle together with overconsumption of high-caloric diet has been identified as a major cause of metabolic diseases, including the one defined as non-alcoholic fatty liver disease (NAFLD). Usually described as the hepatic manifestation of metabolic syndrome, NAFLD includes a spectrum of conditions, from hepatic steatosis to non-alcoholic steatohepatitis (NASH) and cirrhosis, characterized by lipid accumulation in liver cells [1—3]. NAFLD is associated with common metabolic disorders, such as, obesity [4], increased cardiovascular risk [5,6], chronic kidney disease [7], insulin resistance (IR) and type 2 diabetes (T2DM) [8], but it has also been associated with obstructive sleep apnea, osteoporosis, psoriasis, periodontitis, hypothyroidism, male sexual dysfunction, polycystic ovarian syndrome, colonorectal cancer, and urolithiasis, proposing NAFLD as a metabolic disease with extra-hepatic manifestations and multi-organ involvement [9,10].

The global prevalence of NAFLD has increased in the last two decades and it is estimated to be about 25% with variations in prevalence in different regions of the world [11,12]. A meta-analytic study of NAFLD global epidemiology showed that the lowest prevalence of NAFLD is observed in Africa, and the highest in Middle East and South America.

Moreover, this same study supports the association of metabolic comorbidities of NAFLD and obesity, T2DM, and metabolic syndrome in the same areas of the globe [11]. The NAFLD prevalence also increases with age, and it is more common in men than in women, in which the prevalence is greatly increased after menopause [13—15]. This gender-specific difference in NAFLD prevalence relies on the possible protective effect of sex hormones, especially estrogen, against the risk of developing NAFLD in women [15].

In this context, it is also important to note that lifestyle may affect not only the parental generation, but also the offspring resistance to develop chronic diseases later in life. Factors to which a mother is exposed during the pregnancy period, such as diet, stress, tobacco and alcohol consumption, or even gestational diabetes mellitus (GDM), may affect fetal development through epigenetic mechanisms that alter fetal phenotype [16,17]. Pregnancy is a period during which a woman's body undergoes multiple metabolic and physiological adaptations necessary to support the demands of pregnancy, as well as to provide physiological environment for fetus development. Changes in maternal lifestyle during pregnancy, especially changes in nutrition and physical activity, may affect those adaptations, and consequently, fetus growth and development [18,19]. Recent data

Received September 30, 2019 • Revision received November 19, 2019 • Accepted November 22, 2019 • Available online 29 November 2019

https://doi.org/10.1016/j.molmet.2019.11.015

¹Laboratory of Metabolism and Exercise (LaMetEx), Research Centre in Physical Activity, Health and Leisure (CIAFEL), Faculty of Sport, University of Porto, Porto, Portugal ²Department of Cell Biology, Physiology & Immunology, Faculty of Biology, University of Barcelona, Barcelona, Spain

^{*}Corresponding author. Laboratory of Metabolism and Exercise (LaMetEx), Research Centre in Physical Activity, Health and Leisure (CIAFEL), Faculty of Sport, University of Porto, Rua Dr. Plácido Costa, 91, 4200-450, Porto, Portugal. E-mail: jela.stevanov@gmail.com (J. Stevanović).

Abbreviations		MiD51 miRNA	mitochondrial dynamics protein of 51 kDa microRNA
Drp1	dynamin-related protein 1	mPTP	mitochondrial permeability transition pores
ER	estrogen receptor	NADH	dihydro-nicotinamide adenine dinucleotide
ERE	estrogen response elements	NAFLD	non-alcoholic fatty liver disease
ET	endurance training	NASH	non-alcoholic steatohepatitis
ETC	electron transport chain	OMM	outer mitochondrial membranes
FFA	free fatty acid	OPA1	optic atrophy 1
Fgf21	fibroblast growth factor-21	OXPHOS	oxidative phosphorylation
GCKR	glucokinase regulatory protein	PGC1α	peroxisome proliferators-activated receptor-gamma
Fis1	human fission factor-1		coactivator 1-alpha
GDM	gestational diabetes mellitus	PNPLA3	patatin-like phospholipase domain-containing 3
HFD	high-fat diet	PPAR	peroxisome proliferators-activated receptor
IMM	inner mitochondrial membranes	ROS	reactive oxygen species
IR	insulin resistance	T2DM	type 2 diabetes mellitus
Lyplal1	lysophospholipase-like-1	TFAM	mitochondrial transcription factor A
MBOAT7	membrane bound 0-acyltransferase domain-containing-7	TG	triglycerides
MFF	mitochondria fission factor	TM6SF2	transmembrane 6 superfamily member-2
Mfn1	mitofusin1	UCP2	uncoupling protein-2
Mfn2	mitofusin2	VLDL	very low-density lipoproteins
MiD49	mitochondrial dynamics protein of 49 kDa	VPA	voluntary physical activity

from animal models show that maternal nutrition during pregnancy and lactation may affect the metabolic phenotype of the offspring, making them more prone to alterations in liver lipid metabolism and hepatic steatosis [20-23].

Therefore, to understand the potential clinical and economic burden of NAFLD in the future, recent studies have shed light on possible therapeutic approaches to address this growing challenge. Still, to design the most effective therapy, it is necessary to recognize the underlying mechanisms in the development and progression of NAFLD. As an active metabolic tissue, the liver has important bioenergetics and xenobiotic detoxification roles, and the disruption of its function may lead to different pathological states [24]. Furthermore, given the pivotal role of mitochondrial network in metabolism and cellular energy production, regulation of ion homeostasis and redox signaling as well as in cellular remodeling and adaptation, dysfunction of liver mitochondria seems to have an important contribution to the development of liver diseases. In fact, liver mitochondrial degeneration has been associated with the hepatocyte death and inflammation, contributing to the degenerative processes. Moreover, mitochondria are recognized as important sources and targets of reactive oxygen species (ROS) associated with liver diseases [1]. They have a central role in cellular lipid metabolism and oxidative stress [25], with mitochondrial network dysfunction being proposed as one of the first events that occur during the development of NAFLD. The inability of mitochondria to coordinate subcellular metabolic processes in the liver results in the impaired regulation of hepatic lipid metabolism and consequent lipid accumulation in hepatocytes [25-27]. Recent data further suggest the important role of endoplasmic reticulum, an important organelle for calcium (Ca²⁺) homeostasis and lipid synthesis, in the pathogenesis of NAFLD. The disruption of endoplasmic reticulum homeostasis activates different signalling pathways related to NAFLD progression. including those involved in inflammation and apoptosis [28-30]. Furthermore, not only that the disturbance of mitochondria and endoplasmic reticulum homeostasis leads to signaling and metabolic changes, but also causes the loss of physical or functional interactions between these two extremely relevant organelles related to cellular metabolism. These tight contact points between mitochondria and

endoplasmic reticulum, known as mitochondria-associated endoplasmic reticulum membranes (MAMs), are vital in the regulation of lipid transport, Ca²⁺ homeostasis, inflammatory signaling and cellular survival [31,32] and thus, the disruption of MAMs integrity is proposed to be involved in hepatic metabolic diseases, including NAFLD [33]. Despite the increased knowledge of underlying cellular mechanisms related to NAFLD, the design of a standard effective therapeutic approach for such complex and multifactorial disease seems to be out of reach so far. Pharmacological interventions and bariatric surgery are proposed in the treatment of NAFLD [34,35]; however, lifestyle modifications, particularly physical exercise, have been suggested as a first-line approach. Being considered one of the most relevant environmental risk factors in NAFLD development, it is no wonder that sedentary behavior is an important target in the NAFLD treatment [1.36]. Physical inactivity has been positively associated with the increased prevalence of a very significant number of chronic diseases and premature mortality. According to the World Health Organization (WHO), it is the 4th leading global risk for mortality, followed by overweight and obesity [37]. Around 30% of people worldwide are physically inactive, with women being slightly more inactive than men [38]. The latest Eurobarometer by the European Commission shows that only 7% of people in the European Union regularly exercise, while 46% of Europeans never exercise, mainly due to lack of time [39,40]. Ironically, whereas physical inactivity rises as a global public health problem, evidence on therapeutic benefits of physical activity on health continue to grow. Regular physical exercise is consensually recognized as an important preventive and/or therapeutic non-pharmacological tool against a range of metabolic, cardiovascular, musculoskeletal, pulmonary, neurological, oncological and psychiatric diseases, [36,41]. Among other mechanisms, physical exercise expresses its beneficial effects against pathophysiological conditions by targeting mitochondrial function, morphology, and bioenergetics [42], which clearly makes it a potent and interesting therapeutic stimulus against, at least, those diseases that are characterized by mitochondrial dysfunction. Our group has already reported the advantageous effects of physical exercise on mitochondriopathies related to cardiac dysfunction induced by doxorubicin (DOX) [43-47] or ischemia-



reperfusion [46,47] and brain dysfunction induced by DOX [48,49], as well as those mitochondrial dysfunction associated with high-fat diet (HFD)-induced adiposopathy [50,51] and NAFLD [52—55]. Therefore, taking into account that NAFLD is a multi-organ disease, lifestyle modifications, including physical activity, should be promoted among patients in order to impact both hepatic and extra-hepatic manifestations of this disease [10]. Furthermore, skeletal muscles act as endocrine-type organs, producing myokines during exercise, also expressing multi-organ effect targeting contractile and non-contractile tissues, including liver, adipose tissue, and brain [36], which proves the exercise as a potent protective mechanism against those multi-organ involving diseases.

2. MECHANISMS OF NAFLD DEVELOPMENT

Since 1980, when an advanced stage of fatty liver diseases, usually thought to be exclusively induced by alcohol overconsumption, was identified in people who do not consume excessive amounts of alcohol, NAFLD has been recognized as a serious metabolic disorder. At that moment, although the cause and the related mechanisms of this condition were still unclear, it was mainly identified in moderately obese patients and/or those with associated diseases such as diabetes [56].

Being considered a complex disease, NAFLD results from the interactions of many environmental and genetic factors. The PNPLA3 (patatin-like phospholipase domain-containing 3), APOB (apolipoprotein B), MBOAT7 (membrane bound 0-acyltransferase domain containing 7), TM6SF2 (transmembrane 6 superfamily member 2), GCKR (glucokinase regulatory protein), and LYPLAL1 (lysophospholipase-like 1) are only some of genetic *loci* recognized as determinants in the development of NAFLD [57], while lifestyle changes, especially overnutrition and physical inactivity, are considered main environmental culprits for NAFLD development.

Being a primary organ of glucose and lipid metabolism, the liver is particularly affected by the excessive accumulation of lipids originated from the diet, lipolysis in adipose tissue, or even de novo lipogenesis in liver itself, which is obviously aggravated by physical inactivity. Excessive accumulation of lipids, in at least 5% of hepatocytes, represents the first recognizable stage of NAFLD - hepatic steatosis. These accumulated lipids, in the form of micro- or macrovesicles, may cause injury to hepatocytes, leading to the advanced stage of NAFLD, known as non-alcoholic steatohepatitis (NASH), characterized also by inflammation and liver fibrosis. Patients with NASH are at higher risk to further develop cirrhosis and/or hepatocellular carcinoma (HCC) [4,58]. Despite current scientific efforts, the mechanisms of NAFLD pathogenesis remain elusive. One of the first proposed hypothesis explaining NAFLD mechanisms was the 'two-hit hypothesis'. According to this suggestion, during the first hit, due to the imbalance in lipid metabolism and transport, the accumulation of lipids in hepatocytes occurs and leads to hepatic steatosis. Accordingly, the first hit makes the liver more susceptible to the second hit, associated with a more progressive hepatic damage. During this following stage, inflammatory cytokines are released, levels of oxidative stress increase, and a number of organelles, including mitochondria and endoplasmic reticulum, undergoes dysfunctional modifications characteristic of NASH [59]. Nevertheless, recent studies have challenged this hypothesis suggesting that multiple other "hits", such as IR, adipokines release, nutrition (overconsumption and caloric intake), alteration in intestinal microbiota composition and activity, increased gut permeability, and (epi) genetic constraints may be involved in the pathogenesis of NAFLD and its progression from hepatic steatosis to NASH [60,61].

This 'multiple hit hypothesis' has also been referred to explain the development and progression of NAFLD in children and adolescents, the so-called pediatric NAFLD [62], with in utero environment suggested as the 'first hit' in the development of pediatric NAFLD [63]. The rising global prevalence of pediatric NAFLD [64,65] urges for understanding the onset mechanisms and improvement of current preventive and therapeutic approaches. Data suggest that children with family history of hepatic steatosis are at higher risk to develop this disease [66]; however unhealthy maternal lifestyles and IR during pregnancy, particularly increased in women with GDM, may also cause hepatic lipid excess in fetus. These conditions create a cytotoxic environment in the early fetal life, which "opens the door" for the development and progression of metabolic disorders [63], namely major alterations in the glucose and lipid metabolism. With advancing pregnancy, fasting glucose levels decrease while glucose production in liver increases. subsequently leading to increased fasting insulin levels and IR. Consequently, circulating glucose and free fatty acid levels increase in mothers to provide enough nutrients for fetal development [67]. However, in women who already have family history of diabetes, who are obese and/or have hyperglycemia, these metabolic alterations during pregnancy are even more exacerbated, being the levels of IR even higher and the risk to develop GDM increased. Although the impaired glucose tolerance and IR, characteristic for this type of diabetes, usually disappear after delivery, GDM carries an increased risk for the development of different metabolic disorders in both mothers and children [68,69]. In fact, as fetal metabolism cannot buffer the excess of lipids originated from maternal nutrition and transmitted through placenta [70], exposure to maternal GDM increases the susceptibility to NAFLD in children [71,72]. Moreover, despite the higher prevalence of NAFLD in men than in women in younger to middle age. a few clinical studies have shown that women with prior GDM are more likely to develop NAFLD in middle age [73-75].

3. MITOCHONDRIAL NETWORK FAILURE IN NAFLD

3.1. Lipid metabolism in mitochondria

Being one of the high-metabolic-rate tissues, the liver plays an important role in the metabolism of carbohydrates, proteins and lipids. However, unlike adipose tissue, the liver is not specialized in lipid storage. The pool of free fatty acids (FFAs) is maintained, on the one hand, by the balance between the lipid intake from the diet, lipolysis in adipose tissue, and de novo lipogenesis in the liver and, on the other hand, by the removal of liver lipids through secretion into the circulation, β -oxidation [76], or autophagy [77]. In physiological conditions, FFAs enter liver mitochondria where FFAs undergo β-oxidation or remain in the cytoplasm where they are esterified into triglycerides (TGs), which can be secreted into the circulation as very low-density lipoproteins (VLDLs). The increase of FFAs due to excess caloric intake and decreased energy expenditure associated to sedentary behavior leads to the TGs accumulation as lipid droplets in hepatocytes [25], which is characteristic for NAFLD [52,54]. Additionally, the availability of FFAs is also regulated by a selective form of autophagy, also known as "lipophagy", which appears to have an important function in the regulation of hepatic lipid stores [77]. Excessive lipid storage, however, may inhibit autophagy, leading to a vicious cycle in which decreased autophagy increases lipid accumulation, which further impairs autophagy and promotes over-excessive lipid accumulation [78]. Indeed, decreased autophagy and associated endoplasmic reticulum stress were reported in murine genetic and dietary models of obesity [78,79], as well as in NAFLD-diagnosed patients, dietary NAFLD murine model and in vitro human hepatic cells [80]. The finding that autophagy is involved in breakdown of lipid droplets in liver cells suggests that autophagy impairments may have a critical role in the pathogenesis of NAFLD [78], which supports the role of physical activity as an important tool in the prevention of NAFLD through autophagy promotion [81,82].

Until recently, lipid droplets were considered simple lipid storage vesicle. However, their complex dynamic nature and functions contributed to their distinction as intracellular organelles. Lipid droplets have a neutral lipid core (TG and esters) coated by a phospholipid monolayer and specific scaffolding proteins re-localized from the endoplasmic reticulum or cytosol to their surface. The synthesis of neutral lipid begins within the endoplasmic reticulum membrane bilayer, which further deforms and lipid droplet buds forms an initial lipid droplet. These initial lipid droplets are relatively small (less than 1-100 μm) with their size and number differing among cell types [83,84]. Apart from their role in lipid storage and metabolism, lipid droplets are also involved in numerous cellular events, such as fatty acids trafficking, protein degradation, modulation of nuclear function, virus assembly and pathogen infections, and response to ER stress [83,85]. Due to these various roles of lipid droplets, aberrations in their biology, as well as, lipolysis and autophagy as main catabolic pathways of lipid droplets, are associated to many pathological conditions [86]. Moreover, by isolating lipids, lipid droplets protect cells from excessive lipid accumulation, which could eventually affect membrane integrity and lead to lipotoxicity. This way excessive fatty acids are relatively inert and harmless. However, due to imbalances in processes that maintain FFAs pool in the liver, a non-physiological accumulation of lipid results in changes not only of the number, but also the size of lipid droplets in liver cells. The hallmark of NAFLD is the formation and accumulation of small (microvesicular steatosis) or large lipid droplets (macrovesicular steatosis) in liver cells. In both cases, lipid droplets are considerably larger than those lipid droplets present in liver cells in physiological conditions [84]. The increased size of lipid droplets in pathological conditions arises from several processes: a) TG synthesis in lipid droplets, or b) fusion processes — ripening (diffusion of contents of one lipid droplet into another one) and coalescence (a true fusion of lipid droplets) [84,85]. Besides, some lipid droplets scaffolding proteins, like perilipins, might be involved in physical and metabolic interaction between lipid droplets and mitochondria, and thus in control of local fatty acid flux [87], suggesting that altered interaction between lipid droplets and mitochondria via lipid droplets scaffolding proteins might be present in pathological conditions.

3.2. Mitochondrial bioenergetics

Mitochondria, which occupy almost 20% of hepatocytes area, markedly contribute to the hepatocyte metabolism, especially to β -oxidation and oxidative phosphorylation [88]. Therefore, the lipid accumulation in hepatocytes is only a superficial part of NAFLD pathology. Given their key role in energy production, pH regulation, calcium (Ca²⁺) and redox homeostasis, liver mitochondria are essential sensors of cell toxicity. Mitochondria control lipid metabolism in liver: however, as in pathological conditions the liver takes over the role of lipid storage, mitochondrial dysfunction is the logical consequence. In fact, lipid accumulation in hepatocytes is known to disrupt liver mitochondrial bioenergetics, which works in a way to maintain cellular homeostasis through multiple pathways [89]. In pathological conditions, increased FFAs accumulation favors an increase of β-oxidation and subsequent generation of NADH, which serves as co-enzyme in the mitochondrial electron transport chain (ETC). However, this increased electron flux through the ETC combined with cytochrome c (Cyt c) release due, at least in part, to tumour necrosis factor- α (TNF- α) membrane

permeabilization promotes an ETC impairment and an over-reduction of the respiratory chain. In these disturbed metabolic conditions, mitochondrial-related 'electron leakage' results in the increased ROS generation, which are able to interact with different macromolecules and affect the normal structure and function of cells and their organelles, including mitochondria themselves [1]. In addition, mitochondrial DNA is more susceptible to DNA damage by ROS than nuclear DNA as it lacks histones, and has only a limited repair capacity [90]. These enhanced oxidative stress conditions trigger the production of inflammatory cytokines, causing inflammation and fibrogenic response, and all of these, together with already extant lipid droplets in hepatocytes, lies in the heart of NASH [91—93].

During the initial phase of NAFLD development, liver mitochondrial bioenergetics alters and undergoes 'remodeling' in order to adapt to nutrients overload and protect against NAFLD progression. In this first stage, mitochondrial respiration increases, but as the disease progresses it becomes inefficient due to mitochondrial electron leakage, leading to the increased ROS production [89,94]. Among other factors [95], the failure of such an important mitochondrial adaptive mechanism is the trigger for progression from simple steatosis to NASH [89,94,96]. These alterations will depend on the progression of the disease and the ability of liver to adapt and possibly store lipids. Still, under chronic conditions, compensatory mechanisms of the liver fail to adapt, and these metabolic pathways are constrained, resulting in lipotoxicity and consequent inflammation and development of NASH [89]. Lipotoxicity-induced liver inflammation has been associated with adipose tissue remodeling and obesity-prompted NAFLD [97]. Impaired metabolic compensation in adipose tissue results in compromised energy homeostasis, and subsequently to obesity and NAFLD progression [98].

In the prolonged state of over-nutrition and excess accumulation of lipids in the liver, mitochondrial bioenergetics collapse and ATP synthesis rates significantly begin to decrease. Indeed, dysfunction of mitochondrial ETC and correspondent decrease in ATP synthesis rates are documented in NAFLD patients and animal models [76,94,99]. Ironically, the flux of TCA cycle in the liver remains elevated, probably in response to the high-energy demands in these conditions, which supplies an inefficient mitochondrial ETC with reducing equivalents and provides conditions for a vicious-cycle of increased ROS production [99].

3.3. Mitochondrial dynamics

Taken together, the above-referred rationale highlights that, although the mechanisms of NAFLD are still not fully clarified, it is consensually accepted that the structural and functional alterations of the mitochondrial network significantly contribute to the pathogenesis of NAFLD [100]. Being morphologically dynamic organelles, mitochondria change shape through fusion and fission-related mechanisms depending on the available energy and supply as well as the levels of stress [101]. Processes of mitochondrial fusion, fission and biogenesis are tightly controlled in response to metabolic stressors. Mitochondrial fission considers the division of one mitochondrion into two mitochondria and it is mediated by the interaction of cytoplasmic mitochondrial fission protein dynamin-like/related protein 1 (DLP1/Drp1) and other mitochondrial fission proteins, such as human fission factor-1 (Fis1), mitochondrial fission factor (MFF), and mitochondrial dynamics proteins of 49 kDa (MiD49) and 51 kDa (MiD51) [102]. Mitochondria fusion is characterized by the union of two mitochondria into one mitochondrion. Mitofusin 1 (Mfn1) and mitofusin 2 (Mfn2) mediate fusion of the outer mitochondrial membranes while optic atrophy 1 (OPA1) fuses the inner mitochondrial membranes and enable



mitochondrial fusion. In normal conditions damaged and/or dysfunctional mitochondria are degraded through mitophagy in order to prevent a release of "toxic" signals from mitochondria into the cytosol. Partly damaged mitochondrion undergoes mitochondrial fission where it is divided into two smaller mitochondria — a healthy one and a damaged one. The damaged mitochondria are further degraded by mitophagy, while healthy mitochondria may eventually fuse contributing to a healthy mitochondrial pool [103].

Mitochondrial dynamics plays an important role in the bioenergetics adaptation to cellular metabolic demands. With alterations in nutrient availability, mitochondria will undergo fusion or fission in order to maintain mitochondrial energy production and bioenergetics [104]. In a rich-nutrient environment, mitochondria fragment to avoid energy waste, by decreasing bioenergetic efficiency and increasing mitochondrial uncoupling, which will eventually lead to the increased nutrient storage [104,105]. During high levels of cellular stress or when damaged, mitochondrial network undergo fission to separate damaged mitochondria from a healthy cellular content and maintain cellular and mitochondrial bioenergetic homeostasis [101]. DLP1/Drp1 mediates mitochondrial fission and some studies show increase in protein expression in animals models of NAFLD [106-108]. In starvation, mitochondria tend to fuse in order to maintain ATP production and, thus, increase bioenergetics efficiency. Mitochondrial elongation during starvation protects cells from autophagy and cell death [104,109]. In the presence of functionally damaged mitochondria, mitochondrial quality control mechanisms are activated and apart from potentially mitophagy-related outcomes, mitochondria may fuse to share RNAs or proteins and compensate for the functional defects. The reduced signaling for mitochondrial fusion detected in NAFLD animal models underlines its potential role in the pathology of NAFLD [106,107]. When mitophagy pathways in hepatocytes are altered, damaged mitochondria accumulate, leading eventually to the release of mitochondrial damage signals into the cytosol resulting in inflammatory cytokines expression and inflammasomes activation, and eventually development of steatohepatitis [103]. Additionally, mitochondrial dynamics is also associated with excessive ROS production and enhanced oxidative damage, therefore prompting mitochondria as a target of oxidative stress levels in metabolic diseases [108,110].

3.4. Mitochondrial permeability transition pore

Once mitochondrial functioning starts failing, their role in ${\rm Ca^{2+}}$ homeostasis also becomes endangered. ${\rm Ca^{2+}}$ is recognized as an important factor in the regulation of mitochondrial function. Adequate levels of cytosolic Ca²⁺ are referred as important mediators in the activation of some of the enzymes involved in oxidative phosphorylation. Furthermore, data suggest that extramitochondrial Ca²⁺ is also involved in supplying oxidative phosphorylation with substrates on demand [111,112]. However, in pathological conditions, disrupted Ca²⁺ homeostasis activates mitochondrial buffering ability to uptake it and leads to the overaccumulation of Ca²⁺ in mitochondria, which further affects mitochondrial redox homeostasis and ATP production. Moreover, Ca²⁺-related involvement in the activation of complex-like pores - mitochondrial permeability transition pores (mPTP), induces the loss of inner mitochondrial membrane permeability. As a consequence, the induction of mPTP stimulates both mitochondrial dysfunction and the release of proapoptotic proteins from mitochondria to the cytosol resulting in the activation of apoptotic-related signaling [113], and eventually of cellular quality control mechanisms, such as autophagy [114,115]. Accordingly, the activation of the mPTP is considered an important factor in the pathogenesis of fatty liver diseases, with susceptibility to mPTP opening significantly increased in animal models of NAFLD [53,116,117]. Together with other typical metabolic disturbances observed in NAFLD conditions, namely the ${\rm Ca}^{2+}$ overload and enhanced oxidative stress [116], it seems that high fat and glucose fluctuation may also trigger mPTP, leading to the unregulated flux of molecules into the mitochondrial matrix and, eventually, to mitochondrial swelling, which jeopardizes transmembrane potential and induces the collapse of mitochondrial bioenergetics [118].

3.5. Estrogen signaling and mitochondrial function

Another interesting issue associating NAFLD to impaired mitochondrial function are low levels of estrogen [119]. In fact, estrogen signaling, mediated by estrogen receptor α (ER α), seems to affect mitochondrial function by modulating ATP production, mitochondrial membrane potential, mitochondrial biogenesis, and Ca²⁺ signaling. ERs appear to translocate and to be transitionally present in mitochondria, where they can bind to an estrogen response elements (ERE)-like element of the mitochondrial genome and induce the transcription of mitochondrial DNA genes [120]. The protective effect of estrogen against the risk of developing NAFLD in women has already been proposed [15], while the significance of $ER\alpha$ in insulin signaling has been noted. Data suggest that ERa knockout mice express impaired glucose tolerance and IR, downregulation of peroxisome proliferators-activated receptors (PPAR), PPAR α and PPAR Δ , and uncoupling protein-2 (UCP2) expression in skeletal muscle as well as elevated inflammatory signaling in liver [121]. Considering that ER α can be co-activated by peroxisome proliferators-activated receptor-gamma coactivator 1-α (PGC1α), the loss of estrogen signaling, present in postmenopausal women, is possibly linked to reduced PGC1 α and associated with liver fat and NAFLD progression, which may explain, at least in part, gender-specific difference in NAFLD development and the increased risk for NAFLD in women after menopause. Furthermore, the loss of estrogen signaling can also exacerbate diet-induced NASH in a rodent model, which highlights the impact of this hormonal-mediated effect in the progression of this pathological condition [122]. Ultimately, an epigenetics-mediated regulation may also explain some of the possible mechanisms that associate estrogen with protective effects against NAFLD risk in women. In a study performed by Zhang at al. [123] and devoted to better understand the role of estrogen signaling in liver of male and female mice diagnosed with NAFLD, in vivo and in vitro experiments showed that the upregulation of miR-125b might explain, at least in part, the protective mechanism associated to estrogen. The miR-125b, activated by estrogen through ERα, targets fatty acid synthase and possibly alter lipid metabolism in a way that protects against NAFLD [123].

4. PHYSICAL (IN) ACTIVITY-RELATED NAFLD

Being NAFLD one of the most prevalent liver diseases in western countries, the questions of developing effective preventive and/or therapeutic countermeasures against the disease emerge. A design of effective pharmacological treatments was the first starting point, usually including drugs targeting associated metabolic disorders, such as diabetes and IR or pharmacological strategies used in the prevention of mitochondrial degeneration, which is also present in NAFLD. Some of these strategies also included mitochondrial-targeted molecules, antioxidants administration, and anti-obesity drugs; however, all of them showed low levels of effective therapeutic efficiency. Therefore, the US Food and Drug Administration (FDA) has not approved any of them in the treatment for this disease. So far, weight reduction and lifestyle modifications (dietary changes and physical activity) are still

the only ones proposed as effective interventions for the treatment of NAFLD [124,125].

4.1. Physical exercise to prevent NAFLD

Despite the fact that the knowledge regarding the effects of physical exercise on non-contractile tissues is emerging, a research in the area of NAFLD treatment with exercise is still scarce. However, a number of studies with NAFLD patients evidence the importance of lifestyle changes in dealing against this disease [126-128]. Several clinical studies suggest a positive association between sedentary time and the prevalence of NAFLD. Using subjective (questionnaires) or objective (motion sensors) methods to assess physical activity, several studies report that patients with NAFLD spend more of their daily time in sedentary activities than healthy people. In these NAFLD patients, body mass index and weight are positively associated with sedentary lifestyle. Thus, it is not a surprise that the number of steps per day, total daily energy expenditure, and metabolic equivalences (METs) are lower in NAFLD patients than in healthy subjects. Furthermore, these studies showed that higher-intensity exercise is associated with lower prevalence of NAFLD [129-132]. Therefore, increased physical activity seems to be able to prevent the development and progression of NAFLD through different pathways, including the increase of energy expenditure by muscle contractions and through the stimulation of glucose uptake from the circulation into the muscles [131]. Skeletal muscle is the primary organ of disposal of glucose, as the response to insulin. Therefore, the loss of muscle mass, might be involved in the development of IR and IR-related metabolic disorders, including NAFLD. On the other hand, sarcopenia, an aging-related loss of muscle mass, can also occur earlier in life due to lifestyle changes, particularly unhealthy diet and physical inactivity, which are also in the background of NAFLD development [133]. Thus it seems that sarcopenia and NAFLD share similar risk factors and underlying mechanisms. By improving energy balance and insulin sensitivity, physical activity can contribute to the reduction of hepatic lipid accumulation. Better understanding of liver-muscle interaction could improve the management of NAFLD [134].

Taken together, data suggest that increased physical activity is an effective protective non-pharmacological strategy against several metabolic disorders and a therapeutic approach for patients diagnosed with NAFLD. Indeed, regular exercise has been shown to improve liver histological and metabolic features in NAFLD patients. Studies in both NAFLD patients [126-128] and animal models of NAFLD [52,54,55,135] indicate that physical activity is effective in reducing lipid content in hepatocytes and enhancing insulin sensitivity. Although the underlying mechanisms of the protective effects perpetrated by physical activity still remain elusive, some of them may rely on the inter-organ pathways established between liver, skeletal muscles, and adipose tissue. At a glance, skeletal muscles are the main disposal sites for the glucose in post-prandial state. Therefore, by increasing the level of glucose uptake by the muscles and the activity of glycogen synthase in myocytes, the peripheral insulin sensitivity is improved [136]. Also, regular physical activity reduces visceral adiposity in NAFLD patients [126,137], which contributes to lower the influx of fatty acids from visceral adipose tissue to the liver and prevent possible liver lipid accumulation [126].

4.2. Physical exercise during pregnancy

As previously referred, GDM enhances the risk of metabolic impairments in the offspring, but also, an increased likelihood for the development of NAFLD in mothers with GDM sooner than it is expected in non-pregnant women. Considering this transgenerational burden, strategies that mitigate the development of GDM need to be seriously implemented in order to protect both mother and fetus from this deleterious condition, being physical exercise a non-pharmacological approach that is becoming more and more exciting. Taking into consideration that in uncomplicated pregnancy, there are no impediments to engage in exercise programs and it is considered safe and beneficial during pregnancy, the American College of Obstetricians and Gynecologists (ACOG) recommends that healthy women or even athletes, after the consultation and clinical evaluation by obstetricians/ gynecologists, continue to regularly exercise, although some adjustments should be made on the exercise programs and training routines due to the anatomical and physiological changes associated to pregnancy. In fact, non-athlete pregnant women are encouraged to perform at least 30 min of moderate intensity physical exercise every day or, at least, most days of the week [138]. In women diagnosed with GDM. moderate- or high-intensity aerobic or resistance exercise, at least three times per week, lowers fasting blood glucose levels and glycated hemoglobin, and also improves glycemic control [139]. Moreover, it is known that physical exercise in pregnancy can improve physical fitness, cardiovascular function, muscle mass, weight management, sleep quality, and psychologic well-being, as well as, reduce the risk of developing GDM [140]. Therefore, it is expected that exercise may reduce the risk of maternal, fetal, and neonatal complications in GDM that could aggravate hepatic glucose and lipid metabolism and prompt development of NAFLD in both generations.

A deeper look into cellular mechanisms sheds a light on possible effects of physical activity in the prevention and treatment of NAFLD through modulation of function and structure of particular organelles. namely mitochondria.

5. PHYSICAL EXERCISE AND LIVER MITOCHONDRIAL **ADAPTATIONS**

Depending on the exercise type, intensity and duration, the predominance of substrates utilization may vary during exercise. Generically, the main substrates during low-to moderate-intensity exercise are FFAs and glucose, while with the increase of intensity, glucose becomes a more prominent fuel source, which challenges the energy homeostasis of muscles in several circumstances [42]. Considering the importance of mitochondria in cell metabolism, the effects of exercise in mitochondria are intensively studied. Although it is expected that exercise-related adaptations mainly affect muscle mitochondria, liver mitochondria are also a target during exercise. Given the role of liver in maintaining glucose and lipid metabolism, liver mitochondria are essential for producing energy for these energy-demanding processes. Therefore, as a stimulus for liver mitochondria to produce ATP to maintain homeostasis of those metabolic processes, exercise per se also activates systemic pathways that indirectly modulate liver mitochondria bioenergetics, function, and structure [42,141]. In fact, during exercise-related contractions, skeletal-muscle fibers can produce a number of musclederived molecules, the so-called myokines that are secreted into the bloodstream and target different non-contractile tissues, such as liver, brain, and adipose tissue. Apart from many other potential pathways, the myokine interaction with tissue-specific receptors on those tissues, modulates molecules that ultimately may activate PGC-1a, resulting in an improved regulation of mitochondrial biogenesis [126-128]. Still, the mechanisms are rather obscure. Given the known deleterious consequences of distinct pathological conditions on mitochondrial bioenergetics and dynamics, including in the liver, and the health-related relevance of maintaining an adequate mitochondrial network function and structure for cell functioning, the question whether and how exercise



can positively modulate liver mitochondria, and thereby counteract adverse impact of malnutrition and sedentary behavior in the development and progress of NAFLD arises.

So far, there are no consensual suggestions regarding how physical exercise can impact liver mitochondrial signaling pathways in the animal models. Data from several research groups confirm the positive relevance of active lifestyles in the adaptation of liver mitochondria harmful consequences of NAFLD 55,82,135,142,143]. In fact, two models of exercise (voluntary physical activity (VPA) performed in free-running wheels and endurance training (ET) in the treadmill) in animal models were able to mitigate impairments in mitochondrial bioenergetics and dynamics present in a NASH model induced by a HFD. Both exercise modalities were able to positively modulate liver mitochondria bioenergetics function, reflected by the beneficial changes in the respiratory control ratio [54]. The preservation of liver mitochondrial function can, at least in part, be explained by improvements of mitochondrial membrane structured in the exercise groups. The ratio between phosphatidylcholine and phosphatidylethanolamine, as a marker of membrane integrity, was reduced in sedentary HFD group, suggesting an impaired membrane integrity that was reverted by physical exercise. Nevertheless, cardiolipin, which plays a main role in mitochondrial bioenergetics, decreased in both sedentary and exercise HFD groups [52]. Alterations in the mitochondrial membrane permeability and the induction of mPTP lead to the consequent release of proapoptic molecules and, ultimately, the progression of both the apoptotic and autophagic signaling [114,115]. These results highlight the relevance of active lifestyle strategies to reduce the susceptibility of liver mitochondria to permeability transition pore opening and to positively modulate apoptotic and autophagic signaling in animal models of NAFLD [53]. Even tough, it is important to note that, in some circumstances related with the severity of the deleterious consequences, physical exercise may also stimulates autophagy in various tissues, including liver [82,144], suggesting that one of the mechanisms through which exercise could induce its beneficial effects is the activation of the autophagic signalling [81].

Regarding the positive modulation of physical exercise on liver mitochondrial network remodeling, a study designed by Santos-Alves et al. [145] comparing two chronic exercise modalities (VPA and ET) during 12 weeks in rat model, showed that both models of exercise were able to positively alter the expression of proteins associated with mitochondrial biogenesis and dynamics and increase liver autophagy signaling, thus inducing adaptive remodeling of liver mitochondrial network. On the other hand, another study showed that 5 weeks of endurance treadmill training per se was not a sufficiently strong physiological stimulus to significantly alter liver mitochondrial function or protein expression of OXPHOS subunits. Still, endurance training was able to counteract the deleterious impact induced by salicylate and benefit liver mitochondrial function in the context of toxicity [146]. In contrast, an elegant study designed by Fletcher et al. [142] suggests that 4 weeks of endurance training increases OXPHOS activity mediated by complex I-related substrates although without significantly modulating liver mitochondrial biogenesis markers. In accordance, other studies suggest changes in the activity of liver mitochondrial ETC complexes in response to exercise, particularly of complexes I, IV, and V, followed by increase in mitochondrial glutathione content, and without increased ROS production [147]. Likewise, 6 weeks of swimming exercise positively modulated oxidative stress in rats' liver mitochondria by increasing the activity of the antioxidant system and concomitantly decreasing the levels of oxidative stress [148]. Taking together, the increased antioxidant defense and decreased oxidative stress observed in the referred studies suggest that physical exercise is effective in managing ROS production during exercise and in preserving liver mitochondria redox homeostasis [147,148]. In Otsuka Long-Evans Tokushima Fatty (OLETF) rats, it was shown that, even though both aerobic exercise and treatment with metformin, usually used for the treatment of type 2 diabetes, were effective in modulating NAFLD phenotype regarding fatty acid metabolism and liver mitochondrial function, exercise training per se was even more effective, while the combination of drug treatment and a lifestyle change had only a slight synergistic effect [149]. Even though the general perspective highlights a positive impact of physical exercise in liver mitochondrial function and dynamics, some of the discrepancies among these studies may be related to the differences in animal species and strains used and in the features of the distinct exercise models, namely the distinct type, duration, and intensities of exercise programs [24].

6. EPIGENETIC PERSPECTIVE OF NAFLD AND LIFESTYLE MODIFICATIONS

Although the precise underlying mechanisms linking lifestyle changes and development of NAFLD are not completely understood, epigenetic regulation of genes associated with NAFLD development and progression recently came to the fore [150]. Expression of microRNAs (miRNAs) as well as DNA methylation and histone modification by altering DNA accessibility, regulate the activity of genes involved in, among others, lipid metabolism, mitochondrial dysfunction, oxidative stress, and inflammation. In fact, studies on liver biopsy samples from patients diagnosed with NAFLD confirmed the alterations in methylation of genes related to lipid metabolism and mitochondrial biogenesis. Furthermore, the observed differences in DNA methylation status may help to distinguish patients with mild and advanced stages of NAFLD and can be useful to predict progression of NAFLD [151,152]. Despite of increasing evidence regarding the contribution of epigenetic mechanisms in the pathogenesis of different diseases, the role of miRNAs. DNA methylation, and histone modification in NAFLD pathogenesis is still not clear. Global screening of circulating miRNA expressed in liver under NAFLD conditions showed that the upregulation of miR-122, mir-34a, miR-15b, and miR-16 in circulation were positively associated with NAFLD-related evidence. Increased circulatory levels of miR-122, miR-192, and miR-375 could be proposed as clinical markers of NAFLD progression from hepatic steatosis to NASH. Among these miRNAs, it seems that increased circulatory miR-122, the most abundant miRNA in the liver with an important role in lipid metabolism, has a high impact in NAFLD pathogenesis [153]. Furthermore, NASH was associated with decreased liver expression of miR-122 in NAFLD patients [154]. Similarly, the liver expression of miR-122 was downregulated, together with miR-27 and miR-451, while miR-200a, miR-200b, and mir-429, were upregulated in the liver of a diet-induced rat model of NAFLD [155]. This inverse correlation between circulatory and liver levels of miRNAs may explain their influence in the pathophysiology of NAFLD. Liver lipid-responsive miRNAs miR-34a, which targets one of the members of sirtuin family of NAD-dependent histone deacetylases, is also shown to be upregulated in NASH [154] apparently affecting histone modifications in liver. In addition, histone modifications and DNA methylation regulate gene activity and transcription. Among several transcription factors involved in de novo lipogenesis in liver and contributing to liver steatosis, PPAR- γ [156] and its coactivator PGC-1 α drew attention in epigenetics research associated with NAFLD and its progression [156-159]. The involvement of PGC-1 α relies in mitochondrial

biogenesis, β-oxidation, and lipid transport [158,160] and it's reduction in liver is associated with NAFLD progression in murine model of NAFLD [161]. It is also an important factor for expression of mitochondrial transcription factor A (TFAM) [160], which plays an important role in replication and gene expression of mitochondrial DNA [143]. Hypermethylation of PPARGC1A, but not of Tfam promoter, was detected in liver biopsy specimens from NAFLD patients [157]. Given a potency of epigenetics in diseases' pathophysiology, new emphasis is being placed on the influence of epigenetic modifications at the mitochondrial level. miRNAs detected in liver mitochondria may act as sensors of mitochondrial environment leading to the modulation of mitochondrial function and cell signaling [162,163]. Even though, miRNAs regulation is usually related to the cytosol, it seems that mitochondria have a unique pool of miRNAs, independent of total miRNA pool in the cell. The expression of some of these mitochondriaspecific miRNAs was increased in liver mitochondria of diabetic mice, such as miR-202-5p, miR-134, miR-155, as well as, miR-494, which affects Tfam expression and thus has a role in regulation of mitochondrial DNA [163]. Mitochondrial miRNAs are involved in regulation of transcription, gene expression, gene silencing, cell cycle and cell division, chromatin modification, and in utero embryonic development [164]. Additionally, methylation of mitochondrial DNA has been proposed as a biomarker in disease diagnosis, considering that this epigenetic mechanism may be involved in the pathophysiology of some diseases. However, due to the specificity and structural organization of mitochondrial DNA, which lacks in histones, the methylation of mitochondrial DNA may also be a very specific mechanism and further investigation is necessary. So far, some studies have been conducted in areas of drug treatment, oxidative stress, aging, and neurodegenerative diseases [165]. Still, in liver mitochondria of NAFLD patients, methylation of mitochondria encoded NADH dehydrogenase 6 was higher in NASH patients when compared to hepatic steatosis patients

The emerging epigenetic research seems to explain possible underlying mechanisms in NAFLD pathogenesis, but also provides new surprising perspectives on how epigenetic mechanisms play a role in offspring programming. In fact, maternal nutrition has been associated with epigenetic alterations in key genes involved in lipid metabolism and accumulation, such as Ppara, Ppargc, and Fgf21, suggesting that altered DNA methylation patterns in fetal development due to maternal lifestyles may influence offspring health later in life [22,23]. In rodent models, offspring of mothers fed with HFD had altered gene expression, inhibition of hepatic cell cycle and associated DNA methylation [167,168]. In utero exposure to maternal HFD affected the genes related to liver dysfunction, immune response, and inflammation, and led to DNA hypermethylation of the genes. Furthermore, these epigenetic alterations, some of them persistent through the life, were associated with prolonged gene expression alterations in the offspring liver, possibly enabling the development of metabolic dysfunctions later in life [169]. Moreover, maternal GDM impacts the epigenome of their offspring, mainly through metabolic pathways affecting fetal growth and development, suggesting that DNA methylation is one of the mechanisms mediating fetal programming. The exposure to GDM in early fetal life is associated with DNA methylation alterations in the genes involved in metabolism, endocrine function, and insulin signaling [170,171].

[166], suggesting a role of epigenetic alterations not only on nuclear

DNA, but also on mitochondrial DNA in the development and pro-

aression of NAFLD.

Ultimately, considering the above-mentioned relevance of epigenetic regulation in the development of NAFLD, epigenetic-related analysis regarding the impact of physical exercise in liver mitochondrial function and dynamics will be, for sure, a future emerging and growing field of study. Some studies propose DNA methylation status as a marker of progression of NAFLD [151,152], however, this field remains quite open for research regarding NAFLD treatment through changes in lifestyle. Presently, the modulation of the epigenome by changes in lifestyle seems to be able to revert the adverse effect of malnutrition and sedentary lifestyle. Zhou et al. [172] showed that fast food diet increased liver epigenome susceptibility to develop metabolic diseases. On the other hand, physical exercise was able to protect against this fast food diet-induced DNA hypermethylation [172]. Still, most of the studies address the modulating effect of exercise on skeletal muscles epigenome rather than in the liver. For instance, acute exercise was able to decrease promotors methylation of genes involved in mitochondrial biogenesis, PGC-1 α , PPAR- γ , and TFAM in soleus muscles, which suggest that DNA hypomethylation is an early event in contraction-induced gene activation for structural and metabolic adaptations in exercised skeletal muscles [173]. Furthermore, circulating miRNAs can be altered as a physiological response to physical exercise. For instance, in muscle and heart, mir-486 may affect glucose metabolism, while mir-499 is involved in mitochondrial metabolism and apoptosis through Drp1-related signaling. The better understanding of the effect of physical activity on specific circulating miRNAs could justify the use of exercise as a therapeutic strategy [174]. In a HFD-induced animal model of NAFLD, exercise was able to reduce hepatic overexpression of mir-212, which appeared to be involved in lipogenesis and development of NAFLD by targeting FGF-21 [175]. In the study involving HFD-induced obesity in rats, hypoxic training reduced hepatic levels of mir-378, involved in the regulation of lipid metabolism and production of TGs, and, subsequently, increased energy expenditure, β -oxidation in rat liver, and obesity-related morphological and biochemical features. The study also suggests that the probable mechanism is through muscle utilization of fatty acids produced in the liver [176].

In this regard, it is important to note that not only nuclear DNA, but also mitochondrial DNA may undergo epigenetic modifications that are associated with the development of several diseases. In fact, mitochondrial DNA can also be methylated, which attracts attention in the field of disease treatment and prevention. In NASH patients, the methylation level of NADH dehydrogenase 6 encoded by mitochondrial genome was inversely correlated to the level of physical activity [166]. Still, data are scarce and mitochondrial epigenetics and its association to disease risks are still to be elucidated. Therefore, the clarification of the impact of exercise-related epigenetic mechanisms on cellular processes may contribute to this emerging field and promote active lifestyles.

7. CONCLUDING REMARKS

The prevalence of NAFLD and related metabolic disorders (metabolic syndrome, insulin resistance, obesity) is increasing worldwide. Despite numerous pharmacological therapeutic approaches used to counteract multi-organ manifestation of NAFLD, which appear as highly ineffective, physical exercise emerges as a beneficial non-pharmacological preventive and/or therapeutic strategy against this disease. Physical exercise has already been shown as beneficial against some pathological features characteristic for metabolic liver dysfunction, and especially mitochondrial dysfunction. Additionally, considering that the lifestyle changes of parental generation may influence metabolic status of their offspring, widely known benefits of physical exercise, particularly during pregnancy period, should be taken into account. The use of physical activity during pregnancy as a preventive tool against



developing NAFLD in mothers and even against transmitting their impaired metabolic burden to offspring is challenging, being that the mechanisms behind the supposed resistant phenotype are so far not studied. Bearing in mind the potency of epigenetic mechanisms in the development of NAFLD and the transmission of those metabolic impairments to future generation, better understanding of epigeneticrelated impact of physical exercise on liver mitochondrial function will be a determining factor in the design of beneficial exercise programs in the future. The further mechanistic look of how physical activity interacts with hepatic mitochondrial "fitness", particularly via epigenetic mechanisms, may provide necessary clues for proposing active lifestyles during pregnancy as the important preventive strategy to reduce the risk of liver injury through generations. Additionally, a better understanding of muscle-liver axis is expected to delineate mechanisms by which exercise offers benefits in the prevention and treatment of liver diseases.

ACKNOWLEDGEMENT

This work was supported by the EU's Horizon 2020 Research and Innovation program under the Marie Skłodowska-Curie Actions (No.722619, FOIE GRAS; No.734719, mtFOIE GRAS) and by Portuguese Foundation for Science and Technology (FCT) to the Research Center in Physical Activity, Health and Leisure (FCT/UID/DTP/00617/2019; POCI-01-0145-FEDER-016690-PTDC/DTP-DES/7087/2014; POCI-01-0145-FEDER-016657—PTDC/DTP-DES/1082/2014) and to JB (SFRH/BD/129645/2017).

CONFLICT OF INTERESTS

None declared

REFERENCES

- [1] Goncalves, I.O., Oliveira, P.J., Ascensao, A., Magalhaes, J., 2013. Exercise as a therapeutic tool to prevent mitochondrial degeneration in nonalcoholic steatohepatitis. European Journal of Clinical Investigation 43(11):1184—1194.
- [2] Schuppan, D., Schattenberg, J.M., 2013. Non-alcoholic steatohepatitis: pathogenesis and novel therapeutic approaches. Journal of Gastroenterology and Hepatology 28(Suppl. 1):68—76.
- [3] Nseir, W., Hellou, E., Assy, N., 2014. Role of diet and lifestyle changes in nonalcoholic fatty liver disease. World Journal of Gastroenterology 20(28): 9338—9344.
- [4] Dietrich, P., Hellerbrand, C., 2014. Non-alcoholic fatty liver disease, obesity and the metabolic syndrome. Best Practice & Research Clinical Gastroenterology 28(4):637—653.
- [5] Liu, H., Lu, H.Y., 2014. Nonalcoholic fatty liver disease and cardiovascular disease. World Journal of Gastroenterology 20(26):8407—8415.
- [6] Athyros, V.G., Tziomalos, K., Katsiki, N., Doumas, M., Karagiannis, A., Mikhailidis, D.P., 2015. Cardiovascular risk across the histological spectrum and the clinical manifestations of non-alcoholic fatty liver disease: an update. World Journal of Gastroenterology 21(22):6820—6834.
- [7] Musso, G., Gambino, R., Tabibian, J.H., Ekstedt, M., Kechagias, S., Hamaguchi, M., et al., 2014. Association of non-alcoholic fatty liver disease with chronic kidney disease: a systematic review and meta-analysis. PLoS Medicine 11(7):e1001680.
- [8] Birkenfeld, A.L., Shulman, G.I., 2014. Nonalcoholic fatty liver disease, hepatic insulin resistance, and type 2 diabetes. Hepatology 59(2):713—723.
- [9] Rosato, V., Masarone, M., Dallio, M., Federico, A., Aglitti, A., Persico, M., 2019. NAFLD and extra-hepatic comorbidities: current evidence on a multiorgan metabolic syndrome. International Journal of Environmental Research and Public Health 16(18).

- [10] VanWagner, L.B., Rinella, M.E., 2016. Extrahepatic manifestations of nonalcoholic fatty liver disease. Current Hepatitis Reports 15(2):75—85.
- [11] Younossi, Z.M., Koenig, A.B., Abdelatif, D., Fazel, Y., Henry, L., Wymer, M., 2016. Global epidemiology of nonalcoholic fatty liver disease-Meta-analytic assessment of prevalence, incidence, and outcomes. Hepatology 64(1): 73–84.
- [12] Rinella, M., Charlton, M., 2016. The globalization of nonalcoholic fatty liver disease: prevalence and impact on world health. Hepatology 64(1): 19—22.
- [13] Williams, C.D., Stengel, J., Asike, M.I., Torres, D.M., Shaw, J., Contreras, M., et al., 2011. Prevalence of nonalcoholic fatty liver disease and nonalcoholic steatohepatitis among a largely middle-aged population utilizing ultrasound and liver biopsy: a prospective study. Gastroenterology 140(1): 124–131.
- [14] Lazo, M., Hernaez, R., Eberhardt, M.S., Bonekamp, S., Kamel, I., Guallar, E., et al., 2013. Prevalence of nonalcoholic fatty liver disease in the United States: the third national health and nutrition examination survey, 1988-1994. American Journal of Epidemiology 178(1):38—45.
- [15] Ballestri, S., Nascimbeni, F., Baldelli, E., Marrazzo, A., Romagnoli, D., Lonardo, A., 2017. NAFLD as a sexual dimorphic disease: role of gender and reproductive status in the development and progression of nonalcoholic fatty liver disease and inherent cardiovascular risk. Advances in Therapy 34(6): 1291—1326.
- [16] Barua, S., Junaid, M.A., 2015. Lifestyle, pregnancy and epigenetic effects. Epigenomics 7(1):85—102.
- [17] Lynch, C., Chan, C.S., Drake, A.J., 2017. Early life programming and the risk of non-alcoholic fatty liver disease. Journal of Developmental Origins of Health and Disease 8(3):263—272.
- [18] King, J.C., 2000. Physiology of pregnancy and nutrient metabolism. American Journal of Clinical Nutrition 71(5 Suppl.):1218s—1225s.
- [19] Zeng, Z., Liu, F., Li, S., 2017. Metabolic adaptations in pregnancy: a review. Annals of Nutrition & Metabolism 70(1):59—65.
- [20] Bayol, S.A., Simbi, B.H., Fowkes, R.C., Stickland, N.C., 2010. A maternal "junk food" diet in pregnancy and lactation promotes nonalcoholic Fatty liver disease in rat offspring. Endocrinology 151(4):1451—1461.
- [21] Oben, J.A., Mouralidarane, A., Samuelsson, A.M., Matthews, P.J., Morgan, M.L., McKee, C., et al., 2010. Maternal obesity during pregnancy and lactation programs the development of offspring non-alcoholic fatty liver disease in mice. Journal of Hepatology 52(6):913—920.
- [22] Pruis, M.G., Lendvai, A., Bloks, V.W., Zwier, M.V., Baller, J.F., de Bruin, A., et al., 2014. Maternal western diet primes non-alcoholic fatty liver disease in adult mouse offspring. Acta Physiologica 210(1):215—227.
- [23] Wankhade, U.D., Zhong, Y., Kang, P., Alfaro, M., Chintapalli, S.V., Thakali, K.M., et al., 2017. Enhanced offspring predisposition to steatohepatitis with maternal high-fat diet is associated with epigenetic and microbiome alterations. PLoS One 12(4):e0175675.
- [24] Ascensao, A., Martins, M.J., Santos-Alves, E., Goncalves, I.O., Portincasa, P., Oliveira, P.J., et al., 2013. Modulation of hepatic redox status and mitochondrial metabolism by exercise: therapeutic strategy for liver diseases. Mitochondrion 13(6):862—870.
- [25] Gusdon, A.M., Song, K.X., Qu, S., 2014. Nonalcoholic Fatty liver disease: pathogenesis and therapeutics from a mitochondria-centric perspective. Oxidative Medicine and Cellular Longevity 2014:637027.
- [26] Nassir, F., Ibdah, J.A., 2014. Role of mitochondria in nonalcoholic fatty liver disease. International Journal of Molecular Sciences 15(5):8713—8742.
- [27] Fu, A., Shi, X., Zhang, H., Fu, B., 2017. Mitotherapy for fatty liver by intravenous administration of exogenous mitochondria in male mice. Frontiers in Pharmacology 8:241.
- [28] Zhang, X.Q., Xu, C.F., Yu, C.H., Chen, W.X., Li, Y.M., 2014. Role of endoplasmic reticulum stress in the pathogenesis of nonalcoholic fatty liver disease. World Journal of Gastroenterology 20(7):1768—1776.

- [29] Ashraf, N.U., Sheikh, T.A., 2015. Endoplasmic reticulum stress and Oxidative stress in the pathogenesis of Non-alcoholic fatty liver disease. Free Radical Research 49(12):1405—1418.
- [30] Passos, E., Ascensao, A., Martins, M.J., Magalhaes, J., 2015. Endoplasmic reticulum stress response in non-alcoholic steatohepatitis: the possible role of physical exercise. Metabolism 64(7):780—792.
- [31] Vance, J.E., 2014. MAM (mitochondria-associated membranes) in mammalian cells: lipids and beyond. Biochimica et Biophysica Acta 1841(4):595—609.
- [32] Hayashi, T., Rizzuto, R., Hajnoczky, G., Su, T.P., 2009. MAM: more than just a housekeeper. Trends in Cell Biology 19(2):81—88.
- [33] Rieusset, J., 2017. Endoplasmic reticulum-mitochondria calcium signaling in hepatic metabolic diseases. Biochimica et Biophysica Acta (BBA) - Molecular Cell Research 1864(6):865—876.
- [34] Thoma, C., Day, C.P., Trenell, M.I., 2012. Lifestyle interventions for the treatment of non-alcoholic fatty liver disease in adults: a systematic review. Journal of Hepatology 56(1):255—266.
- [35] Dyson, J., Day, C., 2014. Treatment of non-alcoholic fatty liver disease. Digestive Diseases 32(5):597—604.
- [36] Fiuza-Luces, C., Garatachea, N., Berger, N.A., Lucia, A., 2013. Exercise is the real polypill. Physiology 28(5):330—358.
- [37] World Health Organization, 2009. Global health risks: mortality and burden of disease attributable to selected major risks. Geneva: World Health Organization.
- [38] Hallal, P.C., Andersen, L.B., Bull, F.C., Guthold, R., Haskell, W., Ekelund, U., 2012. Global physical activity levels: surveillance progress, pitfalls, and prospects. Lancet 380(9838):247—257.
- [39] European Commission, 2018. Special Eurobarometer 472. Sport and physical activity
- [40] Commission, E., 2014. Special Eurobarometer 412. Sport and physical activity. European Commission, Directorate-General for Education and Culture and co.
- [41] Pedersen, B.K., Saltin, B., 2015. Exercise as medicine evidence for prescribing exercise as therapy in 26 different chronic diseases. Scandinavian Journal of Medicine & Science in Sports 25(Suppl. 3):1—72.
- [42] Beleza, J., Rizo-Roca, D., Ascensão, A., Magalhães, J., 2018. Targeting mitochondria with sweat: improving mitochondrial function with physical activity. Mitochondrial biology and experimental therapeutics. Springer. p. 379—406.
- [43] Ascensao, A., Magalhaes, J., Soares, J., Ferreira, R., Neuparth, M., Marques, F., et al., 2005. Endurance training attenuates doxorubicin-induced cardiac oxidative damage in mice. International Journal of Cardiology 100(3):451—460.
- [44] Ascensao, A., Magalhaes, J., Soares, J.M., Ferreira, R., Neuparth, M.J., Marques, F., et al., 2005. Moderate endurance training prevents doxorubicininduced in vivo mitochondriopathy and reduces the development of cardiac apoptosis. American Journal of Physiology - Heart and Circulatory Physiology 289(2):H722—H731.
- [45] Ascensao, A., Lumini-Oliveira, J., Machado, N.G., Ferreira, R.M., Goncalves, I.O., Moreira, A.C., et al., 2011. Acute exercise protects against calcium-induced cardiac mitochondrial permeability transition pore opening in doxorubicin-treated rats. Clinical Science 120(1):37—49.
- [46] Ascensao, A., Ferreira, R., Oliveira, P.J., Magalhaes, J., 2006. Effects of endurance training and acute Doxorubicin treatment on rat heart mitochondrial alterations induced by in vitro anoxia-reoxygenation. Cardiovascular Toxicology 6(3—4):159—172.
- [47] Ascensao, A., Magalhaes, J., Soares, J.M., Ferreira, R., Neuparth, M.J., Marques, F., et al., 2006. Endurance training limits the functional alterations of rat heart mitochondria submitted to in vitro anoxia-reoxygenation. International Journal of Cardiology 109(2):169—178.
- [48] Marques-Aleixo, I., Santos-Alves, E., Balca, M.M., Rizo-Roca, D., Moreira, P.I., Oliveira, P.J., et al., 2015. Physical exercise improves brain

- cortex and cerebellum mitochondrial bioenergetics and alters apoptotic, dynamic and auto(mito)phagy markers. Neuroscience 301:480-495.
- [49] Marques-Aleixo, I., Santos-Alves, E., Balca, M.M., Moreira, P.I., Oliveira, P.J., Magalhaes, J., et al., 2016. Physical exercise mitigates doxorubicin-induced brain cortex and cerebellum mitochondrial alterations and cellular quality control signaling. Mitochondrion 26:43—57.
- [50] Rocha-Rodrigues, S., Rodriguez, A., Becerril, S., Ramirez, B., Goncalves, I.O., Beleza, J., et al., 2017. Physical exercise remodels visceral adipose tissue and mitochondrial lipid metabolism in rats fed a high-fat diet. Clinical and Experimental Pharmacology and Physiology 44(3):386—394.
- [51] Rocha-Rodrigues, S., Rodriguez, A., Gouveia, A.M., Goncalves, I.O., Becerril, S., Ramirez, B., et al., 2016. Effects of physical exercise on myokines expression and brown adipose-like phenotype modulation in rats fed a high-fat diet. Life Sciences 165:100—108.
- [52] Goncalves, I.O., Maciel, E., Passos, E., Torrella, J.R., Rizo, D., Viscor, G., et al., 2014. Exercise alters liver mitochondria phospholipidomic profile and mitochondrial activity in non-alcoholic steatohepatitis. The International Journal of Biochemistry & Cell Biology 54:163—173.
- [53] Goncalves, I.O., Passos, E., Diogo, C.V., Rocha-Rodrigues, S., Santos-Alves, E., Oliveira, P.J., et al., 2016. Exercise mitigates mitochondrial permeability transition pore and quality control mechanisms alterations in nonalcoholic steatohepatitis. Applied Physiology Nutrition and Metabolism 41(3):298—306.
- [54] Goncalves, I.O., Passos, E., Rocha-Rodrigues, S., Diogo, C.V., Torrella, J.R., Rizo, D., et al., 2014. Physical exercise prevents and mitigates non-alcoholic steatohepatitis-induced liver mitochondrial structural and bioenergetics impairments. Mitochondrion 15:40—51.
- [55] Goncalves, I.O., Passos, E., Rocha-Rodrigues, S., Torrella, J.R., Rizo, D., Santos-Alves, E., et al., 2015. Physical exercise antagonizes clinical and anatomical features characterizing Lieber-DeCarli diet-induced obesity and related metabolic disorders. Clinical Nutrition 34(2):241—247.
- [56] Ludwig, J., Viggiano, T.R., McGill, D.B., Oh, B.J., 1980. Nonalcoholic steatohepatitis: mayo Clinic experiences with a hitherto unnamed disease. Mayo Clinic Proceedings 55(7):434—438.
- [57] Del Campo, J.A., Gallego-Duran, R., Gallego, P., Grande, L., 2018. Genetic and epigenetic regulation in nonalcoholic fatty liver disease (NAFLD). International Journal of Molecular Sciences 19(3).
- [58] Drew, L., 2017. Fighting the fatty liver. Nature 550(7675):S102-S103.
- [59] Rowell, R.J., Anstee, Q.M., 2015. An overview of the genetics, mechanisms and management of NAFLD and ALD. Clinical Medicine 15(Suppl. 6):s77—s82.
- [60] Takaki, A., Kawai, D., Yamamoto, K., 2013. Multiple hits, including oxidative stress, as pathogenesis and treatment target in non-alcoholic steatohepatitis (NASH). International Journal of Molecular Sciences 14(10):20704—20728.
- [61] Buzzetti, E., Pinzani, M., Tsochatzis, E.A., 2016. The multiple-hit pathogenesis of non-alcoholic fatty liver disease (NAFLD). Metabolism 65(8):1038—1048.
- [62] Clemente, M.G., Mandato, C., Poeta, M., Vajro, P., 2016. Pediatric non-alcoholic fatty liver disease: Recent solutions, unresolved issues, and future research directions. World Journal of Gastroenterology 22(36):8078–8093.
- [63] Stewart, M.S., Heerwagen, M.J., Friedman, J.E., 2013. Developmental programming of pediatric nonalcoholic fatty liver disease: redefining the first hit". Clinical Obstetrics and Gynecology 56(3):577—590.
- [64] Anderson, E.L., Howe, L.D., Jones, H.E., Higgins, J.P., Lawlor, D.A., Fraser, A., 2015. The prevalence of non-alcoholic fatty liver disease in children and adolescents: a systematic review and meta-analysis. PLoS One 10(10):e0140908.
- [65] Vos, M.B., Abrams, S.H., Barlow, S.E., Caprio, S., Daniels, S.R., Kohli, R., et al., 2017. NASPGHAN clinical practice guideline for the diagnosis and treatment of nonalcoholic fatty liver disease in children: recommendations from the Expert Committee on NAFLD (ECON) and the North American Society of Pediatric Gastroenterology. Hepatology and Nutrition (NASPGHAN) 64(2):319.



- [66] Long, M.T., Gurary, E.B., Massaro, J.M., Ma, J., Hoffmann, U., Chung, R.T., et al., 2019. Parental non-alcoholic fatty liver disease increases risk of non-alcoholic fatty liver disease in offspring. Liver International 39(4):740—747.
- [67] Lain, K.Y., Catalano, P.M., 2007. Metabolic changes in pregnancy. Clinical Obstetrics and Gynecology 50(4):938—948.
- [68] American Diabetes Association, 2004. Gestational diabetes mellitus. Diabetes Care 27(Suppl. 1):S88—S90.
- [69] Coustan, D.R., 2013. Gestational diabetes mellitus. Clinical Chemistry 59(9):1310—1321.
- [70] Brumbaugh, D.E., Friedman, J.E., 2014. Developmental origins of nonalcoholic fatty liver disease. Pediatric Research 75(1—2):140—147.
- [71] Patel, S., Lawlor, D.A., Callaway, M., Macdonald-Wallis, C., Sattar, N., Fraser, A., 2016. Association of maternal diabetes/glycosuria and prepregnancy body mass index with offspring indicators of non-alcoholic fatty liver disease. BMC Pediatrics 16:47.
- [72] Brumbaugh, D.E., Tearse, P., Cree-Green, M., Fenton, L.Z., Brown, M., Scherzinger, A., et al., 2013. Intrahepatic fat is increased in the neonatal offspring of obese women with gestational diabetes. The Journal of Pediatrics 162(5):930—936 e931.
- [73] Forbes, S., Taylor-Robinson, S.D., Patel, N., Allan, P., Walker, B.R., Johnston, D.G., 2011. Increased prevalence of non-alcoholic fatty liver disease in European women with a history of gestational diabetes. Diabetologia 54(3):641—647.
- [74] Ajmera, V.H., Gunderson, E.P., VanWagner, L.B., Lewis, C.E., Carr, J.J., Terrault, N.A., 2016. Gestational diabetes mellitus is strongly associated with non-alcoholic fatty liver disease. American Journal of Gastroenterology 111(5):658—664.
- [75] Foghsgaard, S., Andreasen, C., Vedtofte, L., Andersen, E.S., Bahne, E., Strandberg, C., et al., 2017. Nonalcoholic fatty liver disease is prevalent in women with prior gestational diabetes mellitus and independently associated with insulin resistance and waist circumference. Diabetes Care 40(1):109—116.
- [76] Nassir, F., Rector, R.S., Hammoud, G.M., Ibdah, J.A., 2015. Pathogenesis and prevention of hepatic steatosis. Gastroenterology and Hepatology 11(3):167—175.
- [77] Martinez-Lopez, N., Singh, R., 2015. Autophagy and lipid droplets in the liver. Annual Review of Nutrition 35:215—237.
- [78] Singh, R., Kaushik, S., Wang, Y., Xiang, Y., Novak, I., Komatsu, M., et al., 2009. Autophagy regulates lipid metabolism. Nature 458(7242):1131—1135.
- [79] Yang, L., Li, P., Fu, S., Calay, E.S., Hotamisligil, G.S., 2010. Defective hepatic autophagy in obesity promotes ER stress and causes insulin resistance. Cell Metabolism 11(6):467–478.
- [80] Gonzalez-Rodriguez, A., Mayoral, R., Agra, N., Valdecantos, M.P., Pardo, V., Miquilena-Colina, M.E., et al., 2014. Impaired autophagic flux is associated with increased endoplasmic reticulum stress during the development of NAFLD. Cell Death & Disease 5:e1179.
- [81] Dasarathy, S., 2017. Are exercise benefits in nonalcoholic fatty liver disease due to increased autophagy? Exercise and Sport Sciences Reviews 45(3): 125.
- [82] Rosa-Caldwell, M.E., Lee, D.E., Brown, J.L., Brown, L.A., Perry Jr., R.A., Greene, E.S., et al., 2017. Moderate physical activity promotes basal hepatic autophagy in diet-induced obese mice. Applied Physiology Nutrition and Metabolism 42(2):148—156.
- [83] Welte, M.A., 2015. Expanding roles for lipid droplets. Current Biology 25(11): R470—R481.
- [84] Gluchowski, N.L., Becuwe, M., Walther, T.C., Farese Jr., R.V., 2017. Lipid droplets and liver disease: from basic biology to clinical implications. Nature Reviews Gastroenterology & Hepatology 14(6):343—355.
- [85] Wilfling, F., Haas, J.T., Walther, T.C., Farese Jr., R.V., 2014. Lipid droplet biogenesis. Current Opinion in Cell Biology 29:39—45.
- [86] Onal, G., Kutlu, O., Gozuacik, D., Dokmeci Emre, S., 2017. Lipid droplets in health and disease. Lipids in Health and Disease 16(1):128.

- [87] Wang, H., Sreenivasan, U., Hu, H., Saladino, A., Polster, B.M., Lund, L.M., et al., 2011. Perilipin 5, a lipid droplet-associated protein, provides physical and metabolic linkage to mitochondria. The Journal of Lipid Research 52(12): 2159—2168.
- [88] Wei, Y., Rector, R.S., Thyfault, J.P., Ibdah, J.A., 2008. Nonalcoholic fatty liver disease and mitochondrial dysfunction. World Journal of Gastroenterology 14(2):193—199.
- [89] Patterson, R.E., Kalavalapalli, S., Williams, C.M., Nautiyal, M., Mathew, J.T., Martinez, J., et al., 2016. Lipotoxicity in steatohepatitis occurs despite an increase in tricarboxylic acid cycle activity. American Journal of Physiology. Endocrinology and Metabolism 310(7):E484—E494.
- [90] Kauppila, J.H., Stewart, J.B., 2015. Mitochondrial DNA: radically free of freeradical driven mutations. Biochimica et Biophysica Acta 1847(11):1354— 1361.
- [91] Leung, T.M., Nieto, N., 2013. CYP2E1 and oxidant stress in alcoholic and non-alcoholic fatty liver disease. Journal of Hepatology 58(2):395—398.
- [92] Liang, W., Menke, A.L., Driessen, A., Koek, G.H., Lindeman, J.H., Stoop, R., et al., 2014. Establishment of a general NAFLD scoring system for rodent models and comparison to human liver pathology. PLoS One 9(12):e115922.
- [93] Satapati, S., Kucejova, B., Duarte, J.A., Fletcher, J.A., Reynolds, L., Sunny, N.E., et al., 2015. Mitochondrial metabolism mediates oxidative stress and inflammation in fatty liver. Journal of Clinical Investigation 125(12): 4447—4462.
- [94] Koliaki, C., Szendroedi, J., Kaul, K., Jelenik, T., Nowotny, P., Jankowiak, F., et al., 2015. Adaptation of hepatic mitochondrial function in humans with nonalcoholic fatty liver is lost in steatohepatitis. Cell Metabolism 21(5):739—746.
- [95] Pierantonelli, I., Svegliati-Baroni, G., 2019. Nonalcoholic fatty liver disease: basic pathogenetic mechanisms in the progression from NAFLD to NASH. Transplantation 103(1):e1—e13.
- [96] Grattagliano, I., Montezinho, L.P., Oliveira, P.J., Fruhbeck, G., Gomez-Ambrosi, J., Montecucco, F., et al., 2019. Targeting mitochondria to oppose the progression of nonalcoholic fatty liver disease. Biochemical Pharmacology 160:34—45.
- [97] Ishtiaq, S.M., Rashid, H., Hussain, Z., Arshad, M.I., Khan, J.A., 2019. Adiponectin and PPAR: a setup for intricate crosstalk between obesity and non-alcoholic fatty liver disease. Reviews in Endocrine & Metabolic Disorders 20(3):253—261.
- [98] Taketani, H., Nishikawa, T., Nakajima, H., Kodo, K., Sugimoto, S., Aoi, W., et al., 2019. Aging-associated impairment in metabolic compensation by subcutaneous adipose tissue promotes diet-induced fatty liver disease in mice. Diabetes, Metabolic Syndrome and Obesity 12:1473—1492.
- [99] Satapati, S., Sunny, N.E., Kucejova, B., Fu, X., He, T.T., Mendez-Lucas, A., et al., 2012. Elevated TCA cycle function in the pathology of diet-induced hepatic insulin resistance and fatty liver. The Journal of Lipid Research 53(6):1080—1092
- [100] Paradies, G., Paradies, V., Ruggiero, F.M., Petrosillo, G., 2014. Oxidative stress, cardiolipin and mitochondrial dysfunction in nonalcoholic fatty liver disease. World Journal of Gastroenterology 20(39):14205—14218.
- [101] Nasrallah, C.M., Horvath, T.L., 2014. Mitochondrial dynamics in the central regulation of metabolism. Nature Reviews Endocrinology 10(11): 650—658.
- [102] Hall, A.R., Burke, N., Dongworth, R.K., Hausenloy, D.J., 2014. Mitochondrial fusion and fission proteins: novel therapeutic targets for combating cardiovascular disease. British Journal of Pharmacology 171(8):1890—1906.
- [103] Mansouri, A., Gattolliat, C.H., Asselah, T., 2018. Mitochondrial dysfunction and signaling in chronic liver diseases. Gastroenterology 155(3):629—647.
- [104] Schrepfer, E., Scorrano, L., 2016. Mitofusins, from mitochondria to metabolism. Molecular Cell 61(5):683—694.
- [105] Jheng, H.F., Tsai, P.J., Guo, S.M., Kuo, L.H., Chang, C.S., Su, I.J., et al., 2012. Mitochondrial fission contributes to mitochondrial dysfunction and

- insulin resistance in skeletal muscle. Molecular and Cellular Biology 32(2): 309—319
- [106] Du, J., Zhang, X., Han, J., Man, K., Zhang, Y., Chu, E.S., et al., 2017. Proinflammatory CXCR3 impairs mitochondrial function in experimental nonalcoholic steatohepatitis. Theranostics 7(17):4192—4203.
- [107] Xu, J., Cao, K., Li, Y., Zou, X., Chen, C., Szeto, I.M., et al., 2014. Bitter gourd inhibits the development of obesity-associated fatty liver in C57BL/6 mice fed a high-fat diet. Journal of Nutrition 144(4):475—483.
- [108] Galloway, C.A., Lee, H., Brookes, P.S., Yoon, Y., 2014. Decreasing mitochondrial fission alleviates hepatic steatosis in a murine model of nonalcoholic fatty liver disease. American Journal of Physiology - Gastrointestinal and Liver Physiology 307(6):G632—G641.
- [109] Gomes, L.C., Di Benedetto, G., Scorrano, L., 2011. During autophagy mitochondria elongate, are spared from degradation and sustain cell viability. Nature Cell Biology 13(5):589—598.
- [110] Yu, T., Sheu, S.S., Robotham, J.L., Yoon, Y., 2008. Mitochondrial fission mediates high glucose-induced cell death through elevated production of reactive oxygen species. Cardiovascular Research 79(2):341—351.
- [111] Gellerich, F.N., Gizatullina, Z., Trumbeckaite, S., Nguyen, H.P., Pallas, T., Arandarcikaite, O., et al., 2010. The regulation of OXPHOS by extramitochondrial calcium. Biochimica et Biophysica Acta 1797(6—7):1018— 1027.
- [112] Williams, G.S., Boyman, L., Chikando, A.C., Khairallah, R.J., Lederer, W.J., 2013. Mitochondrial calcium uptake. Proceedings of the National Academy of Sciences of the U S A 110(26):10479—10486.
- [113] Figueira, T.R., Barros, M.H., Camargo, A.A., Castilho, R.F., Ferreira, J.C., Kowaltowski, A.J., et al., 2013. Mitochondria as a source of reactive oxygen and nitrogen species: from molecular mechanisms to human health. Antioxidants and Redox Signaling 18(16):2029—2074.
- [114] Lemasters, J.J., 2007. Modulation of mitochondrial membrane permeability in pathogenesis, autophagy and control of metabolism. Journal of Gastroenterology and Hepatology 22(Suppl. 1):S31—S37.
- [115] Rasola, A., Bernardi, P., 2011. Mitochondrial permeability transition in Ca(2+)-dependent apoptosis and necrosis. Cell Calcium 50(3):222-233.
- [116] Teodoro, J.S., Rolo, A.P., Duarte, F.V., Simoes, A.M., Palmeira, C.M., 2008. Differential alterations in mitochondrial function induced by a cholinedeficient diet: understanding fatty liver disease progression. Mitochondrion 8(5-6):367-376.
- [117] Navarro, C.D.C., Figueira, T.R., Francisco, A., Dal'Bo, G.A., Ronchi, J.A., Rovani, J.C., et al., 2017. Redox imbalance due to the loss of mitochondrial NAD(P)-transhydrogenase markedly aggravates high fat diet-induced fatty liver disease in mice. Free Radical Biology and Medicine 113:190—202.
- [118] Yin, X., Zheng, F., Pan, Q., Zhang, S., Yu, D., Xu, Z., et al., 2015. Glucose fluctuation increased hepatocyte apoptosis under lipotoxicity and the involvement of mitochondrial permeability transition opening. Journal of Molecular Endocrinology 55(3):169—181.
- [119] Mooga, V.P., White, C.R., Giordano-Mooga, S., 2018. Estrogen and mitochondrial function in disease. Mitochondrial diseases. IntechOpen.
- [120] Gupte, A.A., Pownall, H.J., Hamilton, D.J., 2015. Estrogen: an emerging regulator of insulin action and mitochondrial function. Journal of Diabetes Research 2015:916585.
- [121] Ribas, V., Nguyen, M.T., Henstridge, D.C., Nguyen, A.K., Beaven, S.W., Watt, M.J., et al., 2010. Impaired oxidative metabolism and inflammation are associated with insulin resistance in ERalpha-deficient mice. American Journal of Physiology. Endocrinology and Metabolism 298(2): E304—E319.
- [122] Besse-Patin, A., Leveille, M., Oropeza, D., Nguyen, B.N., Prat, A., Estall, J.L., 2017. Estrogen signals through peroxisome proliferator-activated receptorgamma coactivator 1alpha to reduce oxidative damage associated with dietinduced fatty liver disease. Gastroenterology 152(1):243—256.

- [123] Zhang, Z.C., Liu, Y., Xiao, L.L., Li, S.F., Jiang, J.H., Zhao, Y., et al., 2015. Upregulation of miR-125b by estrogen protects against non-alcoholic fatty liver in female mice. Journal of Hepatology 63(6):1466—1475.
- [124] LaBrecque, D.R., Abbas, Z., Anania, F., Ferenci, P., Khan, A.G., Goh, K.L., et al., 2014. World Gastroenterology Organisation global guidelines: nonal-coholic fatty liver disease and nonalcoholic steatohepatitis. Journal of Clinical Gastroenterology 48(6):467—473.
- [125] Tsochatzis, E.A., Papatheodoridis, G.V., 2011. Is there any progress in the treatment of non-alcoholic fatty liver disease? World Journal of Gastrointestinal Pharmacology and Therapeutics 2(1):1—5.
- [126] Houghton, D., Thoma, C., Hallsworth, K., Cassidy, S., Hardy, T., Burt, A.D., et al., 2017. Exercise reduces liver lipids and visceral adiposity in patients with nonalcoholic steatohepatitis in a randomized controlled trial. Clinical Gastroenterology and Hepatology 15(1):96—102 e103.
- [127] Shojaee-Moradie, F., Cuthbertson, D.J., Barrett, M., Jackson, N.C., Herring, R., Thomas, E.L., et al., 2016. Exercise training reduces liver fat and increases rates of VLDL clearance but not VLDL production in NAFLD. Journal of Clinical Endocrinology & Metabolism 101(11):4219—4228.
- [128] Sullivan, S., Kirk, E.P., Mittendorfer, B., Patterson, B.W., Klein, S., 2012. Randomized trial of exercise effect on intrahepatic triglyceride content and lipid kinetics in nonalcoholic fatty liver disease. Hepatology 55(6):1738— 1745.
- [129] Zelber-Sagi, S., Nitzan-Kaluski, D., Goldsmith, R., Webb, M., Zvibel, I., Goldiner, I., et al., 2008. Role of leisure-time physical activity in nonalcoholic fatty liver disease: a population-based study. Hepatology 48(6):1791—1798.
- [130] Kwak, M.S., Kim, D., Chung, G.E., Kim, W., Kim, Y.J., Yoon, J.H., 2015. Role of physical activity in nonalcoholic fatty liver disease in terms of visceral obesity and insulin resistance. Liver International 35(3):944—952.
- [131] Hallsworth, K., Thoma, C., Moore, S., Ploetz, T., Anstee, Q.M., Taylor, R., et al., 2015. Non-alcoholic fatty liver disease is associated with higher levels of objectively measured sedentary behaviour and lower levels of physical activity than matched healthy controls. Frontline Gastroenterology 6(1):44—51.
- [132] Gerber, L., Otgonsuren, M., Mishra, A., Escheik, C., Birerdinc, A., Stepanova, M., et al., 2012. Non-alcoholic fatty liver disease (NAFLD) is associated with low level of physical activity: a population-based study. Alimentary Pharmacology and Therapeutics 36(8):772—781.
- [133] Kim, J.A., Choi, K.M., 2019. Sarcopenia and fatty liver disease. Hepatology International 13(6):674—687.
- [134] Kim, H.Y., Choi, J.Y., Park, Y.M., 2019. Relative skeletal muscle mass and non-alcoholic fatty liver disease: from association to causation. Hepatobiliary Surgery and Nutrition 8(5):509—511.
- [135] Rector, R.S., Uptergrove, G.M., Morris, E.M., Borengasser, S.J., Laughlin, M.H., Booth, F.W., et al., 2011. Daily exercise vs. caloric restriction for prevention of nonalcoholic fatty liver disease in the OLETF rat model. American Journal of Physiology - Gastrointestinal and Liver Physiology 300(5):G874—G883.
- [136] Roberts, C.K., Little, J.P., Thyfault, J.P., 2013. Modification of insulin sensitivity and glycemic control by activity and exercise. Medicine & Science in Sports & Exercise 45(10):1868—1877.
- [137] Zhang, H.J., He, J., Pan, L.L., Ma, Z.M., Han, C.K., Chen, C.S., et al., 2016. Effects of moderate and vigorous exercise on nonalcoholic fatty liver disease: a randomized clinical trial. JAMA Internal Medicine 176(8):1074—1082.
- [138] ACOG Committee Opinion No. 650, 2015. Physical activity and exercise during pregnancy and the postpartum period. Obstetrics & Gynecology 126(6):e135—e142.
- [139] Harrison, A.L., Shields, N., Taylor, N.F., Frawley, H.C., 2016. Exercise improves glycaemic control in women diagnosed with gestational diabetes mellitus: a systematic review. Journal of Physiotherapy 62(4):188—196.
- [140] Prather, H., Spitznagle, T., Hunt, D., 2012. Benefits of exercise during pregnancy. PM sR 4(11):845—850 quiz 850.



- [141] Fletcher, J.A., Linden, M.A., Sheldon, R.D., Meers, G.M., Morris, E.M., Butterfield, A., et al., 2016. Fibroblast growth factor 21 and exercise-induced hepatic mitochondrial adaptations. American Journal of Physiology -Gastrointestinal and Liver Physiology 310(10):G832—G843.
- [142] Fletcher, J.A., Meers, G.M., Linden, M.A., Kearney, M.L., Morris, E.M., Thyfault, J.P., et al., 2014. Impact of various exercise modalities on hepatic mitochondrial function. Medicine & Science in Sports & Exercise 46(6): 1089—1097
- [143] Lezi, E., Lu, J., Burns, J.M., Swerdlow, R.H., 2013. Effect of exercise on mouse liver and brain bioenergetic infrastructures. Experimental Physiology 98(1):207—219.
- [144] He, C., Bassik, M.C., Moresi, V., Sun, K., Wei, Y., Zou, Z., et al., 2012. Exercise-induced BCL2-regulated autophagy is required for muscle glucose homeostasis. Nature 481(7382):511—515.
- [145] Santos-Alves, E., Marques-Aleixo, I., Rizo-Roca, D., Torrella, J.R., Oliveira, P.J., Magalhaes, J., et al., 2015. Exercise modulates liver cellular and mitochondrial proteins related to quality control signaling. Life Sciences 135:124—130.
- [146] Ascensao, A., Goncalves, I.O., Lumini-Oliveira, J., Marques-Aleixo, I., Dos Passos, E., Rocha-Rodrigues, S., et al., 2012. Endurance training and chronic intermittent hypoxia modulate in vitro salicylate-induced hepatic mitochondrial dysfunction. Mitochondrion 12(6):607—616.
- [147] Sun, L., Shen, W., Liu, Z., Guan, S., Liu, J., Ding, S., 2010. Endurance exercise causes mitochondrial and oxidative stress in rat liver: effects of a combination of mitochondrial targeting nutrients. Life Sciences 86(1-2):39-44.
- [148] Lima, F.D., Stamm, D.N., Della-Pace, I.D., Dobrachinski, F., de Carvalho, N.R., Royes, L.F., et al., 2013. Swimming training induces liver mitochondrial adaptations to oxidative stress in rats submitted to repeated exhaustive swimming bouts. PLoS One 8(2):e55668.
- [149] Linden, M.A., Fletcher, J.A., Morris, E.M., Meers, G.M., Kearney, M.L., Crissey, J.M., et al., 2014. Combining metformin and aerobic exercise training in the treatment of type 2 diabetes and NAFLD in OLETF rats. American Journal of Physiology. Endocrinology and Metabolism 306(3): E300—E310.
- [150] Lee, J.H., Friso, S., Choi, S.W., 2014. Epigenetic mechanisms underlying the link between non-alcoholic fatty liver diseases and nutrition. Nutrients 6(8): 3303—3325.
- [151] Murphy, S.K., Yang, H., Moylan, C.A., Pang, H., Dellinger, A., Abdelmalek, M.F., et al., 2013. Relationship between methylome and transcriptome in patients with nonalcoholic fatty liver disease. Gastroenterology 145(5):1076—1087.
- [152] Zeybel, M., Hardy, T., Robinson, S.M., Fox, C., Anstee, Q.M., Ness, T., et al., 2015. Differential DNA methylation of genes involved in fibrosis progression in non-alcoholic fatty liver disease and alcoholic liver disease. Clinical Epigenetics 7:25
- [153] Pirola, C.J., Fernandez Gianotti, T., Castano, G.O., Mallardi, P., San Martino, J., Mora Gonzalez Lopez Ledesma, M., et al., 2015. Circulating microRNA signature in non-alcoholic fatty liver disease: from serum non-coding RNAs to liver histology and disease pathogenesis. Gut 64(5):800—812.
- [154] Cheung, O., Puri, P., Eicken, C., Contos, M.J., Mirshahi, F., Maher, J.W., et al., 2008. Nonalcoholic steatohepatitis is associated with altered hepatic MicroRNA expression. Hepatology 48(6):1810—1820.
- [155] Alisi, A., Da Sacco, L., Bruscalupi, G., Piemonte, F., Panera, N., De Vito, R., et al., 2011. Mirnome analysis reveals novel molecular determinants in the pathogenesis of diet-induced nonalcoholic fatty liver disease. Laboratory Investigation 91(2):283—293.
- [156] Cheung, O., Sanyal, A.J., 2008. Abnormalities of lipid metabolism in nonalcoholic fatty liver disease. Seminars in Liver Disease 28(4):351—359.
- [157] Sookoian, S., Rosselli, M.S., Gemma, C., Burgueno, A.L., Fernandez Gianotti, T., Castano, G.O., et al., 2010. Epigenetic regulation of insulin resistance in nonalcoholic fatty liver disease: impact of liver methylation of

- the peroxisome proliferator-activated receptor gamma coactivator 1alpha promoter. Hepatology 52(6):1992—2000.
- [158] Schmidt, S.F., Mandrup, S., 2011. Gene program-specific regulation of PGC-1{alpha} activity. Genes & Development 25(14):1453—1458.
- [159] Ahrens, M., Ammerpohl, O., von Schonfels, W., Kolarova, J., Bens, S., Itzel, T., et al., 2013. DNA methylation analysis in nonalcoholic fatty liver disease suggests distinct disease-specific and remodeling signatures after bariatric surgery. Cell Metabolism 18(2):296—302.
- [160] Aharoni-Simon, M., Hann-Obercyger, M., Pen, S., Madar, Z., Tirosh, O., 2011. Fatty liver is associated with impaired activity of PPARgammacoactivator 1alpha (PGC1alpha) and mitochondrial biogenesis in mice. Laboratory Investigation 91(7):1018—1028.
- [161] Estall, J.L., Kahn, M., Cooper, M.P., Fisher, F.M., Wu, M.K., Laznik, D., et al., 2009. Sensitivity of lipid metabolism and insulin signaling to genetic alterations in hepatic peroxisome proliferator-activated receptor-gamma coactivator-1alpha expression. Diabetes 58(7):1499—1508.
- [162] Kren, B.T., Wong, P.Y., Sarver, A., Zhang, X., Zeng, Y., Steer, C.J., 2009. MicroRNAs identified in highly purified liver-derived mitochondria may play a role in apoptosis. RNA Biology 6(1):65—72.
- [163] Bian, Z., Li, L.M., Tang, R., Hou, D.X., Chen, X., Zhang, C.Y., et al., 2010. Identification of mouse liver mitochondria-associated miRNAs and their potential biological functions. Cell Research 20(9):1076—1078.
- [164] Sripada, L., Tomar, D., Prajapati, P., Singh, R., Singh, A.K., Singh, R., 2012. Systematic analysis of small RNAs associated with human mitochondria by deep sequencing: detailed analysis of mitochondrial associated miRNA. PLoS One 7(9):e44873.
- [165] Iacobazzi, V., Castegna, A., Infantino, V., Andria, G., 2013. Mitochondrial DNA methylation as a next-generation biomarker and diagnostic tool. Molecular Genetics and Metabolism 110(1-2):25-34.
- [166] Pirola, C.J., Gianotti, T.F., Burgueno, A.L., Rey-Funes, M., Loidl, C.F., Mallardi, P., et al., 2013. Epigenetic modification of liver mitochondrial DNA is associated with histological severity of nonalcoholic fatty liver disease. Gut 62(9):1356—1363.
- [167] Dudley, K.J., Sloboda, D.M., Connor, K.L., Beltrand, J., Vickers, M.H., 2011. Offspring of mothers fed a high fat diet display hepatic cell cycle inhibition and associated changes in gene expression and DNA methylation. PLoS One 6(7):e21662.
- [168] Keleher, M.R., Zaidi, R., Shah, S., Oakley, M.E., Pavlatos, C., El Idrissi, S., et al., 2018. Maternal high-fat diet associated with altered gene expression, DNA methylation, and obesity risk in mouse offspring. PLoS One 13(2): e0192606.
- [169] Seki, Y., Suzuki, M., Guo, X., Glenn, A.S., Vuguin, P.M., Fiallo, A., et al., 2017. In utero exposure to a high-fat diet programs hepatic hypermethylation and gene dysregulation and development of metabolic syndrome in male mice. Endocrinology 158(9):2860—2872.
- [170] Petropoulos, S., Guillemin, C., Ergaz, Z., Dimov, S., Suderman, M., Weinstein-Fudim, L., et al., 2015. Gestational diabetes alters offspring DNA methylation profiles in human and rat: identification of key pathways involved in endocrine system disorders, insulin signaling, diabetes signaling, and ILK signaling. Endocrinology 156(6):2222—2238.
- [171] Ruchat, S.M., Houde, A.A., Voisin, G., St-Pierre, J., Perron, P., Baillargeon, J.P., et al., 2013. Gestational diabetes mellitus epigenetically affects genes predominantly involved in metabolic diseases. Epigenetics 8(9):935—943.
- [172] Zhou, D., Hlady, R.A., Schafer, M.J., White, T.A., Liu, C., Choi, J.H., et al., 2017. High fat diet and exercise lead to a disrupted and pathogenic DNA methylome in mouse liver. Epigenetics 12(1):55—69.
- [173] Barres, R., Yan, J., Egan, B., Treebak, J.T., Rasmussen, M., Fritz, T., et al., 2012. Acute exercise remodels promoter methylation in human skeletal muscle. Cell Metabolism 15(3):405—411.

Review

- [174] Xu, T., Liu, Q., Yao, J., Dai, Y., Wang, H., Xiao, J., 2015. Circulating microRNAs in response to exercise. Scandinavian Journal of Medicine & Science in Sports 25(2):e149-e154.
- [175] Xiao, J., Bei, Y., Liu, J., Dimitrova-Shumkovska, J., Kuang, D., Zhou, Q., et al., 2016. miR-212 downregulation contributes to the protective effect of
- exercise against non-alcoholic fatty liver via targeting FGF-21. Journal of Cellular and Molecular Medicine 20(2):204-216.
- [176] Lu, Y.L., Jing, W., Feng, L.S., Zhang, L., Xu, J.F., You, T.J., et al., 2014. Effects of hypoxic exercise training on microRNA expression and lipid metabolism in obese rat livers. Journal of Zhejiang University - Science B 15(9):820-829.