

Upregulation of Talin-1 expression associates with advanced pathological features and predicts lymph node metastases and biochemical recurrence of prostate cancer

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Abstract

Talin-1 functions to regulate cell-cell adhesion, and its altered expression was reported to be associated with human carcinogenesis. A total of 280 tissue specimens from prostate cancer (PCa) patients who underwent radical prostatectomy, 75 cases of benign prostatic hyperplasia (BPH) tissue, and 6 cases of normal prostate tissue specimens were collected for construction of tissue microarray and subsequently subjected to immunohistochemical staining of Talin-1 expression.

Talin-1 expression was significantly higher in PCa than both normal and BPH tissues (P < 0.001). Talin-1 expression in PCa tissues was associated with preoperative prostate-specific antigen (PSA) level, Gleason score, tumor stage, lymph node metastasis, positive surgical margin, extracapsular extension and seminal vesicle invasion (all P < 0.05). Logistic regression analysis showed that Talin-1 and Gleason score were independent risk factors for lymph node metastasis of PCa (P < 0.001). Receiver operating characteristic (ROC) curve indicated that Talin-1 expression (AUC = 0.766) had a better accuracy to predict PCa lymph node metastasis than Gleason score (AUC = 0.697), whereas their combination could further enhance the prediction accuracy (AUC = 0.803). Kaplan–Meier curve analysis showed that increased Talin-1 expression was associated with shortened biochemical-free survival of PCa patients after radical prostatectomy (P < 0.001).

These findings suggested that Talin-1 protein was significantly upregulated in PCa tissues compared with that of BPH tissue and Talin-1 expression was an independent predictor for lymph node metastasis and biochemical recurrence of PCa. Further study will investigate the underlying molecular mechanism and the role of Talin-1 in PCa.

Abbreviations: AUC = area under the curve, BFS = biochemical-free survival, BMI = body mass index, BPH = benign prostatic hyperplasia, DAB = 3,3'-diaminobenzidine, DOR = diagnostic odds ratio, EMT = epithelial-mesenchymal transition, H&E = hematoxylin and eosin staining, LN = lymph node, NPV = negative predictive value, PBS = phosphate buffered saline, PCa = prostate cancer, PLND = pelvic lymph node dissection, PPV = positive predictive value, PSA = prostate-specific antigen, PSAD = prostate-specific antigen density, ROC = receiver operating characteristic value.

Keywords: lymph node metastasis, pathological feature, predict, prostate cancer, Talin-1

1. Introduction

Prostate cancer (PCa) is one of the most commonly diagnosed malignancies in men, ranking as the second worldwide incidence rate in male malignancies.^[1] Although most PCa grows and

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progresses slowly, it contributed to more than a quarter million cancer-related death globally as reported in 2008.^[2] The risk factors of PCa are older age, African ethnic, and family history of the disease. For example, approximately 62% of all PCa cases in the United States have been diagnosed in ≥ 65 years old males.^[2] There are several treatment options for PCa, including surgery, radiation therapy, hormone therapy, chemotherapy, or targeted therapy, and treatment selection depends on tumor stages and age of patients.^[3] Radical prostatectomy is one of the most effective therapies of PCa; however, it is debatable whether it needs to combine with pelvic lymph node dissection (PLND),^[4] because most of studies indicated that PLND was only necessary for moderate- and high-risk PCa patients.^[5-7] Like most of other cancers, PCa lymph node metastasis could lead to mortality of the disease; thus, development of biomarkers to predicate tumor metastasis before surgery could help medical oncologists to make an appropriate decision and selection of treatment.^[8,9]

Tumor metastasis is a complex of multiple processes in which cancer cells obtain an ability and capacity to leave the primary tumor site via the lymphatic system and/or the bloodstream and then establish a secondary tumor site in other parts of the body.^[10] Tumor cells undergo the epithelial–mesenchymal

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transition (EMT) to gain mobility and invasion capacity by altering cell adhesion and degradation of extracellular matrix,^[11] thereby leading to cancer metastasis. At the gene levels, a great number of proteins could participate tumor metastasis and inhibition, such as cell adhesion molecules, proteinases, integrins, and EMT-related proteins.^[12] In this study, our research is to focus on Talin-1, a high-molecular-weight cytoskeletal protein. Talin-1 functions to regulate cell-cell adhesion by binding to a variety of adhesion molecules (such as integrins) and actin to induce tumor cytoskeletal remodeling and leading to tumor cell adhesion and migration.^[13,14] In PCa, a previous study demonstrated that Talin-1 could promote PCa cell migration and invasion.^[15] However, another study showed that Talin-1 was an important factor in mediating PCa bone metastasis.^[16] Thus, our present study detected Talin-1 expression in PCa versus benign prostatic hyperplasia (BPH) tissue specimens and then associated Talin-1 expression with clinicopathological characteristics and lymph node metastasis in PCa patients.

2. Materials and methods

2.1. Tissue specimen and data collection

This study was approved by the ethics committee of the First Affiliated Hospital of Fujian Medical University (Fuzhou, Fujian, China). We collected 280 cases of tissue specimens from PCa patients between January 2012 and June 2015 from the First Affiliated Hospital of Fujian Medical University. These patients were diagnosed with PCa according to EAU guidelines^[3] and underwent radical prostatectomy plus PLND without any preoperative adjuvant endocrine therapy or radiotherapy (238 patients received standard PLND with a resection range including obturator foramen and external iliac vein peripheral lymph nodes, whereas 42 patients received extended PLND with a peripheral lymph node fat tissue having an upper bound to the common iliac artery bifurcation, a lower bound to the femoral canal, both sides to the pelvic wall, and a back bound to the obturator nerves, blood vessels, and anterior internal vein).^[17] Moreover, we also collected 75 cases of BPH tissue specimens randomly and 6 cases of normal prostate tissue specimens as a control from our hospital. Clinical data, including age, prostate-specific antigen (PSA), prostate volume, PSA density (PSAD), body mass index (BMI), tumor stage, numbers of lymph node metastasis, positive surgical margin, extracapsular extension, and seminal vesicle invasion, were collected (Table 1). After radical prostatectomy, patients were followed up regularly for physical examination and serum PSA test at the first month after surgery, then every 3 months for 2 years, and semiannually thereafter. As patients with detectable PSA (>0.1 ng/ mL) after surgery usually have persistent cancer, the serum PSA levels of patients >0.1 ng/mL at the first month after surgery were therefore excluded from this study according to previous reports.^[18,19] Patients were diagnosed histologically and did not receive any preoperatively adjuvant endocrine therapy or radiotherapy and patients with uncompleted clinical and pathological data were also excluded from this study. In this study, we excluded 95 cases from further analysis of biochemical recurrence. The time of biochemical recurrence was recorded or to the last follow-up date (on May 2016). Patients were followed up for median 22 months (ranged between 4 and 49 months), and 41 of 185 (14.4%) patients had biochemical disease progression and the median time of biochemical recurrence was 13 months (ranged between 4 and 37 months). We did not calculate the time of overall survival because of the shortness of follow-up time.

Table 1

Clinicopathological parameters of 280 cases of prostate cancer.

Age, y	
Mean \pm SD	68.66 ± 6.75
Range	45–78
BMI, kg/m ²	
Mean \pm SD	25.88 ± 4.38
Range	17.02-33.24
Prostate volume, cm ³	
Mean \pm SD	39.29 ± 9.92
Range	23-56
PSA level, ng/mL	
Mean \pm SD	19.64±13.60
Range	2-62
PSAD, ng/mL·cm ³	
Mean \pm SD	0.54 ± 0.43
Range	0.05-2.25
Percentage of positive biopsies, n (%)	
<50%	176 (62.9)
≥50%	104 (37.1)
Pathological stage (pT), n (%)	
T1	13 (4.6)
T2	129 (46.1)
T3	133 (47.5)
T4	5 (1.8)
Gleason score, n (%)	
2–6	67 (23.9)
7	117 (41.8)
8–10	96 (34.3)
Lymph node metastasis, n (%)	
Yes	126 (45.0)
No	154 (55.0)
Extracapsular extension, n (%)	
Yes	124 (44.3)
No	156 (55.7)
Seminal vesicle invasion, n (%)	
Yes	28 (10.0)
No	252 (90.0)
Positive surgical margin, n (%)	
Yes	36 (12.9)
No	244 (86.8)

BMI = body mass index, PSA=prostate-specific antigen, PSAD=prostate-specific antigen density, SD=standard deviation.

2.2. Construction of tissue microarray

Paraffin blocks from both PCa and BPH were retrieved from Pathology Department and sectioned for hematoxylin and eosin (H&E) staining to confirm diagnosis and identify the representative areas for tissue microarray construction. A tissue microarray maker designed by us was used to generate tissue microarrays using two 2 × 2 mm tissue cores of each case. In the end, we obtained 15 tissue microarrays with 5 × 10 tissue cores of each for both PCa and BPH tissues and then sectioned 4-µm-thick tissue sections for immunohistochemical staining of Talin-1 protein.^[20,21]

2.3. Immunohistochemistry

The sections were conventionally de-waxed and rehydrated and antigen retrieval was performed using a high-pressure cook in 0.1 M citric acid buffer (pH 5.0; Fuzhou Maixin Biotech. Co., Ltd, Fuzhou, China). After that, tissue microarrays were then incubated with 3% H_2O_2 for 10 minutes to block endogenous peroxidase activity and subsequently with 20% normal goat serum at the room temperature for 30 minutes. Next, the tissue

microarray sections were incubated with a rabbit monoclonal anti-Talin-1 antibody (Cell Signaling Technology, Danvers, MA) at a dilution of 1:400 at 4°C overnight. On the next day, the sections were washed briefly with phosphate buffered saline (PBS) 3 times and further incubated with a secondary antibody (DAKO Company, CA) at 37°C for 30 minutes and subsequently with a ChemMate EnVision DetectionKit (DAKO Company, CA). The sections were subjected to color reaction with 3,3'diaminobenzidine (DAB) solution and counterstained with hematoxylin briefly and mounted with a cover slip. Confirmed positive sections were used as positive control and PBS to replace the primary antibody was used as a negative control. The immunostained tissue microarray sections were reviewed and scored under a light microscope (Olympus, Tokyo, Japan) by two pathologists blindly. Any discrepancy was resolved by their rereviewing of the sections. A semi-quantitative method was conducted and the total Talin-1 immunostaining score included staining intensity and proportion of positive cells. Staining intensity (I) was recorded as 0, absent; 1, weak; 2, moderate; 3, strong, whereas the proportion (P) of positive cells was recorded as 0, <5%; 1, 5% to 25%; 2, 26% to 50%; 3, 51% to 75%; and 4, >75%. A score for each histological grade (H-score) was determined as the summary of intensity and proportion Hscore = Σ ($I \times P$) according to a previous study.^[14] The final Talin-1 expression score was calculated using the value of the percent positivity score multiplied by the staining intensity score as "-" (score, 0-1), "+" (score, 2-3), "++" (score, 4-5), and "+++" $(\text{score } \geq 6).^{[22]}$

2.4. Statistical methods

SPSS 19.0 statistical software (SPSS, Chicago, IL) was utilized to perform all statistical analyses. Quantitative data were compared using independent samples t test, Mann–Whitney U test, or

Kruskal–Wallis test. Qualitative data were compared using independent sample χ^2 -square test or Fisher exact test. Lymph node metastasis predictors were analyzed using logistic regression analysis and double-indicator combination was analyzed using logistic regression model. Receiver operating characteristic (ROC) curve was used to evaluate the predictive value of each indicator for PCa lymph node metastasis. Kaplan–Meier plots and the log-rank test were performed to assess the association of Talin-1 expression with biochemical recurrence-free survival (BFS). P < 0.05 was considered statistically significant.

3. Results

3.1. Differential expression of Talin-1 protein in PCa versus normal and BPH tissue specimens

Immunohistochemical data showed that Talin-1 protein was mainly expressed in the cytoplasm of positive cells (Fig. 1) and Talin-1 expression was significantly higher in PCa than in both normal and BPH tissues. Talin-1 expression was significantly higher in poorly differentiated PCa than in both moderately and well-differentiated PCa. Talin-1 expression was significantly higher in LN(+) PCa than in LN(-) PCa (P < 0.05 for all, Tables 1 and 2). However, there was no significant difference in Talin-1 expression between normal and BPH tissue specimens (P > 0.05, Tables 1 and 2).

3.2. Association of Talin-1 expression with clinicopathological data from prostate cancer patients

We then associated the expression of Talin-1 protein with clinicopathological data from PCa patients. We found that high Talin-1 expression was associated with higher PSA levels, Gleason score, tumor stage, lymph node metastasis, positive surgical margin, extracapsular extension, and seminal vesicle

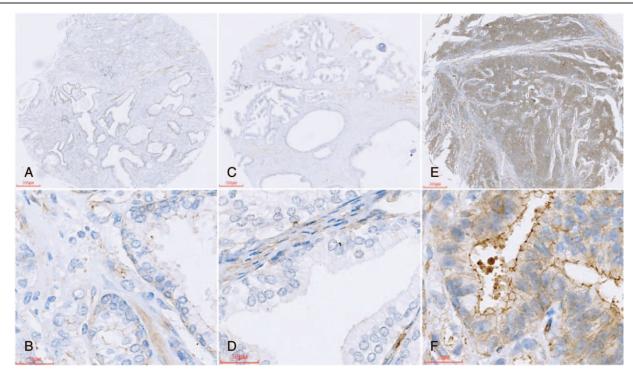


Figure 1. Tissue microarrays containing normal prostate, BPH, and prostate cancer tissues were immunostained with a monoclonal anti-Talin-1 antibody and the data were semiquantitatively analyzed: (A, B) normal prostate; (C,D) BPH; (E,F) prostate cancer. BPH = benign prostatic hyperplasia.

Table 2

Expression of Talin-1 in normal, BPH, and human prostate cancer.

Specimen	n	H-score		<i>P</i> value		
Normal	6	48.3 ± 26.4				
BPH	75	48.2 ± 30.2	0.994*			
Well-differentiated PCa (Gleason score 2-6)	67	92.8 ± 46.8	0.025*			
Moderately differentiated PCa (Gleason score $=$ 7)	117	153.5 ± 46.3	<0.001*	< 0.001 *		
Poorly differentiated PCa (Gleason score 8-10)	96	163.8 ± 41.1	<0.001*	< 0.001 *	0.091 [‡]	
LN () PCa	154	130.2±54.7	< 0.001*			
LN (+) PCa	126	157.5±46.2	< 0.001*	< 0.001§		

BPH = benign prostatic hyperplasia, LN = lymph node, PCa = prostate cancer.

* Compared with normal prostate.

⁺ Compared with well-differentiated PCa.

* Compared with moderately differentiated PCa.

§ Compared with LN (--) PCa.

invasion (P < 0.001; Table 3), whereas Talin-1 expression was not associated with age of patients (P > 0.05; Table 3). Talin-1 expression was more commonly observed in poorly differentiated, high-stage, and lymph node-positive PCa tissue specimens (Fig. 2). Upregulated Talin-1 expression was associated with PCa malignant behaviors and lymph node metastasis.

3.3. Association of clinicopathological factors and Talin-1 expression with pelvic lymph node metastasis of prostate cancer

We then performed subgroup analysis to associate clinicopathological factors and Talin-1 expression with pelvic lymph node metastasis of PCa. The results showed that PCa metastasis to pelvic lymph nodes was associated with Talin-1 expression, higher PSA level, PSAD, Gleason score, tumor grade, positive surgery margin, extracapsular extension, and seminal vesicle invasion (all P < 0.05; Table 4), but not associated with age and BMI of patients, prostate volume, or percentage of positive prostate needle biopsies (Table 4).

Multivariate logistic regression analysis showed that Gleason score and Talin-1 expression were independent risk factors for PCa lymph node metastasis (P < 0.001, Table 5). The ROC curve analysis showed that the area under the curve (AUC) of Talin-1 expression (AUC=0.766) was much higher than that of Gleason scores (AUC=0.699), although their combination could further enhance the accuracy in predicting PCa lymph node metastasis (AUC=0.802) (Fig. 3). Further analysis of their diagnostic

Table 3

Variables		Talin-1 expression				
	Ν	_	+	++	+++	P value
Total, n (%)	280	60 (21.4)	56 (20.0)	91 (32.5)	73 (26.1)	
Age, y						
<70	134 (47.9)	33 (55.0)	23 (41.1)	44 (48.4)	34 (46.6)	0.58
≥70	146 (52.1)	27 (45.0)	33 (58.9)	47 (51.6)	39 (53.4)	
PSA level, ng/mL						
<10	50 (17.9)	32 (53.3)	0 (0.0)	18 (19.8)	0 (0.0)	< 0.001
10-20	148 (52.9)	25 (41.7)	22 (39.3)	73 (80.2)	28 (38.4)	
>20	82 (29.3)	3 (5.0)	34 (60.7)	0 (0.0)	45 (61.6)	
Gleason score						
2–6	67 (23.9)	28 (46.7)	22 (39.3)	8 (8.8)	9 (12.3)	< 0.001
7	117 (41.8)	19 (31.7)	8 (14.3)	54 (59.3)	36 (49.3)	
8–10	96 (34.3)	13 (21.7)	26 (46.4)	29 (31.9)	28 (38.4)	
pT stage						
T1-T2	142 (50.7)	47 (78.3)	20 (35.7)	64 (70.3)	11 (15.1)	< 0.001
T3–T4	138 (49.3)	13 (21.7)	36 (64.3)	27 (29.7)	62 (84.9)	
Lymph node metasta	sis					
Yes	126 (45.0)	5 (8.3)	19 (33.9)	47 (51.6)	55 (75.3)	< 0.001
No	154 (55.0)	55 (91.7)	37 (66.1)	44 (48.4)	18 (24.7)	
Extracapsular extensi	on					
Yes	124 (44.3)	7 (11.7)	28 (50.0)	33 (36.3)	56 (76.7)	< 0.001
No	156 (55.7)	53 (88.3)	28 (50.0)	58 (63.7)	17 (23.3)	
Seminal vesicle invas	ion					
Yes	28 (10.0)	0 (0.0)	2 (3.6)	4 (4.4)	22 (30.1)	< 0.001
No	252 (90.0)	60 (100.0)	54 (96.4)	87 (95.6)	51 (69.9)	
Positive surgical marginal	gin				. ,	
Yes	36 (12.9)	5 (8.3)	4 (7.1)	6 (6.6)	21 (28.8)	< 0.001
No	244 (87.1)	55 (91.7)	52 (92.9)	85 (93.4)	52 (71.2)	

PSA = prostate-specific antigen.

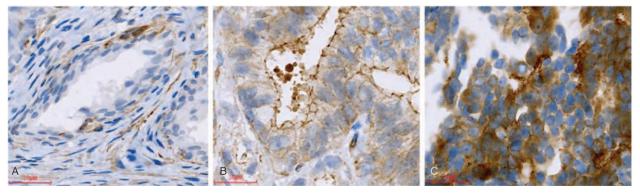


Figure 2. Different expression level of Talin-1 protein in prostate cancer tissues: (A) well differentiated: + (1 × 1 = 1); (B) moderately differentiated: ++ (2 × 2 = 4); (C) poorly differentiated: +++ (3 × 3 = 9).

Table 4

Association of clinicopathological features with lymph node metastasis of prostate cancer.

		Lymph nod	e metastasis		
Variables	Ν	Negative	Positive	P value	
Total, n (%)	280	154 (55.0)	126 (45.0)		
Age, y					
<70	134 (47.9)	77 (50.0)	57 (45.2)	0.427	
≥70	146 (52.1)	77 (50.0)	69 (54.8)		
BMI					
≤25	136 (48.6)	80 (51.9)	56 (44.4)	0.211	
>25	144 (51.4)	74 (48.1)	70 (55.6)		
Prostate volum	e, cm ³				
\leq 35	125 (44.6)	76 (49.4)	49 (38.9)	0.08	
>35	155 (55.4)	78 (50.6)	77 (61.1)		
Percentage of	positive biopsies				
<50%	176 (62.9)	96 (62.3)	80 (63.5)	0.842	
≥50%	104 (37.1)	58 (37.7)	46 (36.5)		
PSA level, ng/r	. ,	· · · ·			
<10	50 (17.9)	31 (20.1)	19 (15.1)	0.006	
10-20	148 (52.9)	90 (58.4)	58 (46.0)		
>20	82 (29.3)	33 (21.4)	49 (38.9)		
PSAD, ng/mL·c			- ()		
< 0.15	30 (10.7)	15 (9.7)	15 (11.9)	0.560	
≥0.15	250 (89.3)	139 (90.3)	111 (88.1)		
Gleason score					
2–6	67 (23.9)	56 (36.4)	11 (8.7)	<0.001	
7	117 (41.8)	65 (42.2)	52 (41.3)		
8–10	96 (34.3)	33 (21.4)	63 (50.0)		
pT stage					
T1-T2	142 (50.7)	91 (59.1)	51 (40.5)	0.002	
T3–T4	138 (49.3)	63 (40.9)	75 (59.5)	01002	
Extracapsular e	()	00 (1010)	10 (0010)		
Yes	124 (44.3)	57 (37.0)	67 (53.2)	0.007	
No	156 (55.7)	97 (63.0)	59 (46.8)	0.001	
Seminal vesicle	. ,	0. (00.0)	00 (1010)		
Yes	28 (10.0)	5 (3.2)	23 (18.3)	<0.001	
No	252 (90.0)	149 (96.8)	103 (81.7)	<0.001	
Positive surgica	. ,	110 (00.0)	100 (0111)		
Yes	36 (12.9)	11 (7.1)	25 (19.8)	0.002	
No	244 (87.1)	143 (92.9)	101 (80.2)	0.002	
Talin-1 express	. ,	140 (02.0)	101 (00.2)		
_	60 (21.4)	55 (35.7)	5 (4.0)	<0.001	
+	56 (20.0)	37 (24.0)	19 (15.1)	<0.001	
++	91 (32.5)	44 (28.6)	47 (37.3)		
+++	73 (26.1)	18 (11.7)	55 (43.7)		
	10 (20.1)	10 (11.7)	00 (40.7)		

BMI = body mass index, PSA = prostate-specific antigen, PSAD = prostate-specific antigen density.

sensitivity, specificity, positive predictive value, negative predictive value, and accuracy showed that combination of Talin-1 expression and Gleason score had the highest accuracy in predicting PCa lymph node metastasis (71.4%), whereas Gleason scores were lowest (64.3%) and Talin-1 expression was moderate (69.2%) (Table 6). Combination of Talin-1 with Gleason score (>7) could help us to predict tumor lymph node metastasis.

3.4. Association of Talin-1 expression with biochemical recurrence-free survival

Kaplan–Meier curve analysis showed that increased Talin-1 expression was associated with shortened BFS of PCa patients after radical prostatectomy (P < 0.001, Fig. 4).

4. Discussion

The 5-year survival rate of PCa patients in the United States could reach to 99% and all patients with PCa may not die from the disease.^[1] It is because different individuals with PCa show very different disease progression. For example, in some moderate/ high-risk cases, there is no disease progression for a long period of time, whereas lymph node metastasis and even bone metastasis may occur in certain low-risk cases in the early stage.^[7,23] To date, the application of PLND to all PCa patients remains controversial and previous studies have suggested that PLND could help medical oncologist to accurately evaluate tumor stage and improve prognosis of patients to certain extent,^[24,25] whereas other studies revealed that PLND may not affect prognosis of PCa patients with postoperatively confirmed pT×N₀ stage, even if such patients had a relatively high PSA

 Table 5

 Multivariable analysis of clinicopathological features for association with lymph node metastasis of prostate cancer.

Variables	OR (95% CI)	P value	
Multivariable analysis			
PSA level (<10 vs10-20 vs >20)	1.003 (0.594-1.693)	0.991	
PSAD (<0.15 vs>0.15)	1.120 (0.604–2.075)	0.480	
Gleason score (2-6 vs7 vs 8-10)	3.466 (2.176-5.520)	< 0.001	
pT stage (T1–T2 vsT3–T4)	1.886 (0.916-3.885)	0.085	
Extracapsular extension	0.583 (0.283-1.203)	0.144	
Seminal vesicle invasion	1.779 (0.511–6.192)	0.365	
Positive surgical margin	2.019 (0.720-5.662)	0.182	
Talin-1 (- /+ /++ /+++)	2.565 (1.786-3.684)	< 0.001	

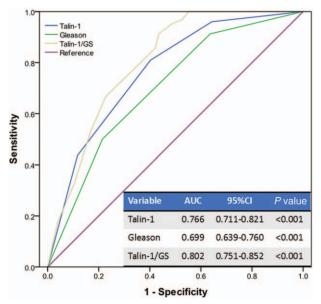


Figure 3. The ROC curves of Talin-1, Gleason score, and their combination in diagnosis of prostate cancer lymph node metastasis. ROC = receiver operating characteristic.

level, pathological grade, or capsule invasion before surgery.^[26,27] Moreover, PLND may also involve in different complications, such as deep venous thrombosis of the lower limbs, lymphatic leakage, pelvic infection, and so on, especially for extended PLND.^[28] Thus, to date, surgeons in the field have suggested that low-risk PCa may not need PLND and that a standard or extended PLND should be performed in the moderate- and high-risk PCa cases.^[29,30] However, it lacks a standard protocol or tumor markers to evaluate or predict the risk of PCa metastasis to the pelvic lymph nodes.

Our present study therefore evaluated Talin-1 expression in 280 PCa versus 75 BPH tissue specimens for association with lymph node metastasis and other clinicopathological data. We found that Talin-1 was highly expressed in PCa tissues compared with BPHs. Talin-1 expression was associated with higher PSA levels, Gleason score, tumor stage and differentiation, lymph node metastasis, positive surgical margin, extracapsular extension, and seminal vesicle invasion, whereas PSA level, PSAD, Gleason score, tumor grade, positive surgery margin, extracapsular extension, and seminal vesicle invasion were all associated with PCa lymph node metastasis. Combination of Talin-1 expression and Gleason score can predicate >70% PCa metastasis to the pelvic lymph node. Further study will identify better biomarkers to predicate lymph node metastasis of PCa.

Indeed, increase in cell proliferation and mobility could promote tumor growth and distant metastasis. The cell migration includes the steps of actin phosphorylation and cytoskeletal remodeling.^[31,32] Talin-1 is a focal adhesion protein to mediate integrin interaction with extracellular matrix and Talin-1 can also bind to a variety of cell–cell adhesion molecules (such as integrin) and actin to induce cell cytoskeleton remodeling; thus, increase in Talin-1 expression could result in promotion of tumor cell adhesion and migration. For example, Lai et al^[33] showed that high Talin-1 expression in tumor tissues resulted in a poor prognosis of oral squamous cell carcinoma in ex vivo, whereas knockdown of Talin-1 expression inhibited proliferation and invasion ability of oral squamous cell carcinoma cells. The data

Table 6

Sensitivity, specificity, PPV, NPV, and accuracy (%) of Talin-1 expression, Gleason score, and their combination in diagnosis of prostate cancer lymph node metastasis.

	Sensitivity (%)	Specificity (%)	PPV (%)	NPV (%)	DOR (%)	Youden's index	Cut-off
Talin-1	81.0	59.7	62.1	79.3	69.2	0.407	+++
Gleason	48.4	77.3	63.6	64.7	64.3	0.257	>7
Talin-1/Gleason	92.1	54.5	62.4	89.4	71.4	0.466	(+++/>7)

CI = confidence interval, DOR = diagnostic odds ratio, NPV = negative predictive value, OR = odd ratio, PPV = positive predictive value, PSA = prostate-specific antigen, PSAD = prostate-specific antigen density.

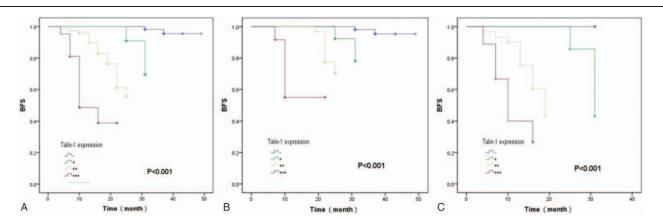


Figure 4. Kaplan–Meier curve analyses of biochemical recurrence-free survival of prostate cancer patients stratified by Talin-1 expression. (A) The total of 185 prostate cancer patients; (B) 134 cases of prostate cancer patients with negative lymph node metastasis; (C) 51 cases of prostate cancer patients with positive lymph node metastasis.

reported by Xing et al^[34] revealed that injection of an anti-Talin-1 antibody or antisense RNA was able to destroy the stress fibers of cellulose cells and inhibit adhesion, extension, and migration of breast cancer cells. In PCa, Zhang et al^[15] showed that Talin-1 was overexpressed in PCa tissue specimens and cell lines, whereas microRNA-124 (miR-124) and Talin-1 impaired cellular adhesion and motility through integrins and the focal adhesion kinase/ Akt pathway. Their data concluded that miR-124 impairs the adhesion, migration, and invasion of PCa cells by downregulation of Talin-1.^[15] Our present data further confirmed these previous studies; however, the underlying molecular mechanisms responsible for Talin-1-asscoaited PCa lymph node metastasis remain to be determined, although a previous study showed that Talin-1 phosphorylation activated β 1 integrin as a novel mechanism to promote PCa bone metastasis.^[16]

Clinically, PCa metastasis is mainly involved in pelvic lymph nodes and contributes to an unfavorable prognosis of PCa patients.^[35] Previous studies have shown that Talin-1 expression associated with lymph node metastasis of oral squamous cell carcinoma and colon cancer.^[33,36] In our present study, we found that Talin-1 expression and other clinicopathological factors, such as higher PSA level, PSAD, Gleason score, tumor grade, positive surgery margin, extracapsular extension, and seminal vesicle invasion, were all associated with PCa metastasis to the pelvic lymph nodes. Thus, we speculate that PCa metastasis is a multiple factor-induced phenomenon and further study is needed to understand the underlying molecular mechanism and therefore, to effectively control disease progression, and provide a means to evaluate and predict PCa metastasis.

Xu et al^[37] demonstrated that expressions of Talin-1 mRNA and protein were significantly upregulated in nasopharyngeal carcinoma tissues; high expression of Talin-1 was associated with poor prognosis of nasopharyngeal carcinoma patients. Lai et al^[33] showed that high Talin-1 expression in tumor tissues is also associated with poor prognosis of oral squamous cell carcinoma. As most patients diagnosed with PCa do have a favorable disease progression-free survival after radical prostatectomy and the follow-up time of our present study was short, we just assessed Talin-1 expression for association with biochemical recurrence of PCa after radical prostatectomy and found that increase in Talin-1 expression was associated with shortened postoperative BFS.

However, our present study does have some limitations. For example, we could include the tumor cell-positive number of lymph nodes versus total numbers of lymph nodes in this study, which could lead to a more accurate determination of association between Talin-1 expression and lymph node metastasis. The reason was simply because surgeons who performed PLND on each patient might not have a consistent technique, leading to inconsistence in the extent of lymph node dissection. Moreover, some patients received a standard PLND, whereas others may have received an extended PLND. In addition, the present study is retrospective and thus was not well controlled. In conclusion, our present data demonstrated that upregulated Talin-1 expression was associated with PCa malignant behaviors and lymph node metastasis. Combination of Talin-1 with Gleason score (>7) could help to predict tumor lymph node metastasis. Talin-1 expression was associated with shortened biochemical-free survival of PCa patients after radical prostatectomy.

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