

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19

Stochasticity in Dietary Restriction-Mediated Lifespan Outcomes in *Drosophila*

Olivia L. Mosley¹, Joel A. Villa², Advaita Kamalakkannan², Eliyashaib James², Jessica M. Hoffman^{1,*}, Yang Lyu^{2,*}

¹ Department of Biological Sciences, Augusta University, Augusta, GA, USA

² Department of Molecular Biology and Biochemistry, Rutgers, the State University of New Jersey, New Brunswick, NJ, USA

*Co-senior/Co-corresponding authors

20 **Abstract**

21 Dietary restriction (DR) is widely considered to be one of the most potent approaches to extend healthy
22 lifespan across various species, yet it has become increasingly apparent that DR-mediated longevity is
23 influenced by biological and non-biological factors. We propose that current priorities in the field should include
24 understanding the relative contributions of these factors to elucidate the mechanisms underlying the beneficial
25 effects of DR. Our work conducted in two laboratories, represents an attempt to unify DR protocols in
26 *Drosophila* and to investigate the stochastic effects of DR. Across 64 pairs of survival data (DR/ad libitum, or
27 AL), we find that DR does not universally extend lifespan. Specifically, we observed that DR conferred a
28 significant lifespan extension in only 26.7% (17/64) of pairs. Our pooled data show that the overall lifespan
29 difference between DR and AL groups is statistically significant, but the median lifespan increase under DR
30 (7.1%) is small. The effects of DR were overshadowed by stochastic factors and genotype. Future research
31 efforts directed toward gaining a comprehensive understanding of DR-dependent mechanisms should focus on
32 unraveling the interactions between genetic and environmental factors. This is essential for developing
33 personalized healthspan-extending interventions and optimizing dietary recommendations for individual genetic
34 profiles.

35 Introduction

36 Over the past century, the benefits of caloric or dietary restriction (CR or DR) have been extensively studied
37 across organisms ^{1,2}. The concept that reducing food intake without causing malnutrition may promote longevity
38 and health is widely appreciated and generally supported by observations across various species. This field was
39 anchored in early studies of McCay et al. ³, who reported that rats on a calorically restricted diet were longer
40 lived than those fed ad libitum (AL). Since then, the effects of CR/DR have been demonstrated to extend to
41 multiple species including yeast ⁴, invertebrates ^{5,6}, other mammals ⁷, and perhaps even humans ⁸. Remarkably,
42 the underlying biology of CR/DR reveals a complex and conserved molecular machinery, that includes pathways
43 that play a crucial roles in nutrient sensing and DR-mediated outcomes such as the target of rapamycin ⁹ and
44 AMPK-activated protein kinase pathways ¹⁰ (reviewed in ²).

45 While many would argue that DR is the most robust method to extend healthy lifespan known thus far, the
46 complex nature of lifespan modulation under DR has become increasingly evident as genetic factors and other
47 variables have been suggested to play significant roles ¹¹. For instance, grand-offspring of wild-caught mice had
48 no increase in longevity under DR ¹², and less than 50% of 41 recombinant inbred mouse strains subjected to
49 DR exhibited an increase in lifespan ¹³. More recently, Wilson et al. utilized 161 isogenic strains from naturally
50 derived inbred lines of *Drosophila melanogaster*, finding that 29% of these strains did not exhibit DR-induced
51 lifespan extension ¹⁴. These findings underscore the need to further investigate and explore influential variables,
52 including but not limited to genetic background, to enhance our understanding of the relationship between DR
53 and longevity control.

54 In addition to genetic factors associated with response to DR, stochastic events are increasingly recognized as
55 significant contributors to the diversity of aging phenotypes ¹⁵⁻¹⁷. For example, *C. elegans* from an N2 isogenic
56 reference population show varied rates of aging as they approach later life stages ¹⁸, and the *Caenorhabditis*
57 Interventions Testing Program (CITP) has found significant stochastic variation in lifespan across and within
58 laboratories ¹⁹. In flies, stochastic variation has been observed in response to mating status across genetically
59 distinct population ²⁰. Furthermore, recent studies have identified intrinsic noise and variations at the cellular
60 level in aging biomarkers ^{21,22}. Overall, the inclusion and rigorous analysis of stochastic factors in DR studies are
61 critical and currently underexplored, potentially biasing results of DR experiments.

62 Invertebrate models such as *Drosophila* and *C. elegans* have been instrumental in elucidating key factors that
63 contribute to the longevity benefits of DR. These models have primarily explored DR by modulating nutritional
64 concentrations in the food media ²³, not necessarily restricting calories. Therefore, the AL state is better
65 described as a high nutrient state, as both the DR and AL groups have continuous access to food. In *Drosophila*,
66 restrictions of either yeast (a major protein source for flies) or individual amino acids have been extensively used
67 to study DR mechanisms e.g. ^{6,9,24,25}, though these studies have sparked some recent controversies (see recent
68 updates from ²⁶). Notably, the effects of dietary restriction are more consistent when a restricted diet is compared
69 to a nutrient rich diet, rather than to a standard husbandry diet ^{23,27}, though within *Drosophila* there is actually no
70 “standard diet” used consistently across laboratories. This practice in the field presents significant challenges in
71 attributing longevity effects solely to DR, as it has been shown that an enriched diet can lead to desiccation
72 causing increased mortality ²⁷, and overnutrition with a nutrient rich media may lead to obese phenotypes which
73 predictably exhibit a shortened lifespan.

74 We suggest that the subtleties between a restricted diet and a “standard” diet may present challenges in
75 reproducibility due to stochastic variations, and that DR effects may only be biologically relevant when compared
76 to high nutrient, enriched diets. To assess and quantify these variations, we replicated DR experiments that
77 involve multiple cohorts and distinct dietary paradigms, in two geographically distinct laboratories. We find that
78 while genotype emerges as the most significant predictor of lifespan, we recorded considerable variation among
79 cohorts with respect to DR effects, some of which can be attributed to stochastic variation. We conclude that

rigorous understanding of CR/DR outcomes must strongly take genetics and stochastic factors, as well as diet details, into account.

Methods

Drosophila husbandry

Mated male and female flies from four common laboratory strains of *Drosophila melanogaster* were used in each cohort: w^{1118} , Oregon-R (OR), $w^{Dahomey}$, and Canton-S (CS). As an additional control for any potential genetic drift or variations between stocks, the Hoffman lab gifted OR and w^{1118} strains and received the $w^{Dahomey}$ and Canton-S strains from the Lyu lab, so the strains used across labs were genetically identical. After exchange, all new fly strains were acclimated to the laboratory for a period of 6-8 weeks prior to use in experiments. Lab stocks were maintained at 25°C at 65-85% humidity and a diurnal, 12-12 light/dark schedule. All fly stocks were maintained on a cornmeal-based (CT) diet (Table 1).

DR lifespan protocols

Both labs collected time-synchronized eggs for the lifespan assays. In the Hoffman lab, each genotype was placed on fresh CT food, and flies mated and laid eggs for 48-72 hours. After expanding each stock, all adult flies were cleared from the vials and the time-synchronized eggs developed. The Lyu lab used an egg-collecting chamber and grape juice-agar media to gather embryos deposited within a 48-hours period²⁸. For both labs, after 10 days, the new adult flies were transferred onto SY10 (Cohort 1, 3, and 4) or CT (Cohort 2) food and allowed to mate for 48 hours before sexing under light CO₂ anesthesia. The difference in the mating diet introduces variation in early life dietary exposures. The collection process took place over the course of 2-3 days until 300 flies were collected for each genotype and sex with each vial containing 25 flies. The collected flies were randomized onto either a dietary restriction (DR) or ad libitum (AL) media (Table 1). We must note that while we are using the term ad libitum for the higher nutrient treatment due to the ubiquitous use of the term in the aging field, in *Drosophila*, and other invertebrates, this is not a true AL treatment, as all groups have access to their diet 24/7. We varied the diets and mating food in individual cohorts such that cohorts 1-3 used CT/SY10, while cohort 4 used SY5/SY15 as the DR/AL dietary paradigms, respectively. Flies were transferred to fresh media three times a week with deaths recorded at each transfer using D-Life²⁸ and Excel.

Table 1 Ingredients of each of four diets used in the study. Each amount is measured in 1L of water. The nutrient composition is estimated using *Drosophila* Dietary Composition Calculator: <https://brodericklab.com/DDCC.php>.

Ingredient	Experimental Protocol 1		Experimental Protocol 2	
	DR (CT)	AL (SY10)	DR (SY5)	AL (SY15)
Agar	1-2%	1-2%	1-2%	1-2%
Propionic acid (mL)	5	5	5	5
Yeast (g)	25	100	50	150
Sucrose (g)	55	100	100	100
Dextrose (g)	30	0	0	0
Cornmeal (g)	60	0	0	0
Total calories (cal)	628.85	775.70	582.20	969.20
Proteins (g)	17.72	53.03	26.53	79.53
Fat (g)	1.44	0.30	0.30	0.30
Carbohydrates (g)	150.43	151.00	129.50	172.50

108 Climbing and body mass assays

109 At approximately 30 days of age, flies from each group were run through a climbing assay. Briefly, flies were
110 tapped to the bottom of an empty vial and allowed to climb for 10 seconds. At 10 seconds, the number of flies
111 that had climbed at least 5 cm was recorded. Data was collected from cohorts 1-3 in the Hoffman lab and
112 analyzed with all results combined.

113 To determine if flies on low yeast diets were calorically restricted, we weighed flies on each diet to determine if
114 the DR flies weighed less than AL flies. Flies were placed on either a S10Y5 or S10Y15 diet for 30 days prior to
115 weighing. After 30 days, flies were anesthetized on ice, transferred to a 2mL centrifuge tube in groups of 5-10
116 and weighed on a microanalytic balance. Weights were calculated by subtracting the average empty-tube weight
117 per group from the measured weight per sample and adjusting for the number of flies per sample. Both climbing
118 ability and body mass assays were only conducted in the Hoffman lab, as we were looking at general health
119 effects, not reproducibility.

120 Statistical analyses

121 All statistical analyses were completed in program R. Overall, comparisons across labs and variables of interest
122 were determined with Cox proportional hazard models using the “survival” package^{29,30}. Comparisons between
123 individual DR pairs within a lab/genotype/sex/cohort were made with log rank tests. Kaplan-Meier curves were
124 plotted for visualization of the data. Spearman rank correlations were calculated to look at correlations of median
125 longevities across laboratories. Due to the large number of log-rank tests for individual comparisons, we applied
126 a Bonferroni correction with significance set as $p < 0.00078$. Differences in healthspan measures (climbing ability
127 and body mass) were calculated using an ANOVA looking at the effects of sex, genotype, and dietary treatment.

128 We performed Cox regression and model fitting using in-house R script to determine the amount of variance
129 explained by each variable analyzed. We used the `coxph` function from the survival package^{29,30} to fit both full
130 and reduced models. The full model included the covariates lab, sex, cohort, genotype, and diet while the
131 reduced models excluded one covariate at a time to evaluate their individual contributions. The proportional
132 hazards assumption for the Cox regression models was tested using the `cox.zph` function from the survival
133 package. We estimated the Cox-Snell R^2 ³¹ for both full and reduced models. The likelihood of each model was
134 computed using the `logLik` function from the stats package. The contribution of each covariate was estimated
135 using a likelihood-based measure, derived from the differences of log-likelihoods of the full and reduced models:

$$136 \text{Contribution}_{X_j} = \frac{\log \text{Lik}_{full} - \log \text{Lik}_{reduced, X_j}}{\sum_{i \in I} \log \text{Lik}_{full} - \log \text{Lik}_{reduced, X_i}}, \text{ where } i = \{Lab, Sex, Diet, Cohort, Genotype\}$$

139 Results

140 Lab reproducibility

141 To minimize inter-laboratory variability and enhance reproducibility, we utilized the same DR protocols, applied
142 identical experimental procedures, and ordered supplies simultaneously from the same vendors. Detailed
143 approaches are described in the methods section. Our final dataset consisted of 15,935 flies across 64 pairs of
144 DR/AL survival data (128 longevity curves). All raw data can be found in Supplementary Table 1. We used two
145 DR protocols: Protocol 1 utilized the commonly used CT food as the restricted diet and SY10, 10% (w/v)
146 sucrose:yeast as the AL diet, while Protocol 2 controlled for all other ingredients, varying only the concentration
147 of yeast to further test the effects of protein restriction (see Table 1 for detailed ingredients). We ran Protocol 1

three times independently in each lab. We combined data generated from two protocols to estimate overall reproducibility and stochasticity.

Overall, we found reasonable reproducibility in lifespan data from the two labs (Figure 1a). We did find a significant difference in longevity between labs (log-rank $p=0.001$); however the differences in median lifespan are minimal: 53.7 days (95%CI 53-54.1 days) for the Hoffman Lab and 53.1 days (95%CI 52-54.2 days) for the Lyu Lab, a difference of ~1% and driven by our large sample size ($n = 8,475$ for the Hoffman Lab and 7,460 for the Lyu Lab). Across cohorts, there was significant correlation of mean longevity between the labs (Figure 2, Spearman $\rho=0.55$, $p=3.6 \times 10^{-6}$). Together, these results indicate that when applying the same protocols and procedures, laboratories or geographic locations are not major factors influencing lifespan results.

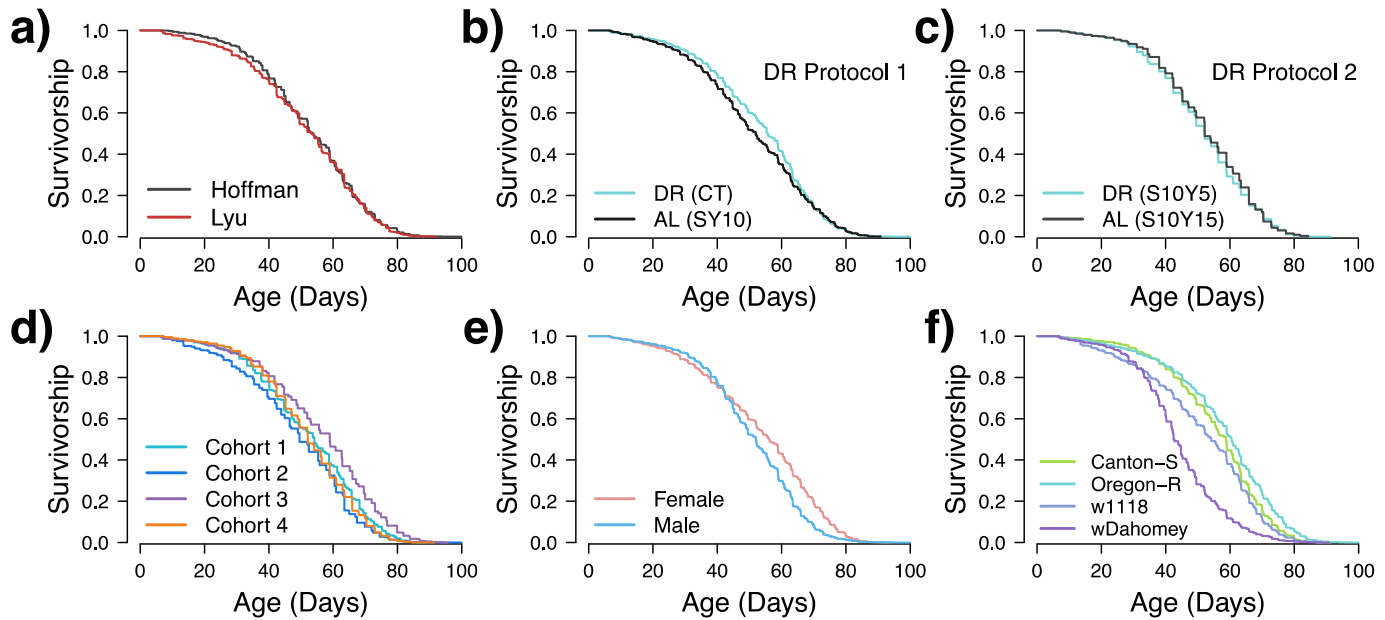


Figure 1. Kaplan–Meier survival analysis of *Drosophila melanogaster*. The survival curves represent the proportion of survivors over time (in days) during the adult stage. Each panel shows the survival curves for a specific factor, illustrating the effects of lab (a), dietary restriction protocol 1 (b), dietary restriction protocol 2 (c), cohorts (d), sex (e), and genotype (f) on the lifespan of the flies. These panels collectively demonstrate how different factors impact the lifespan of the flies.

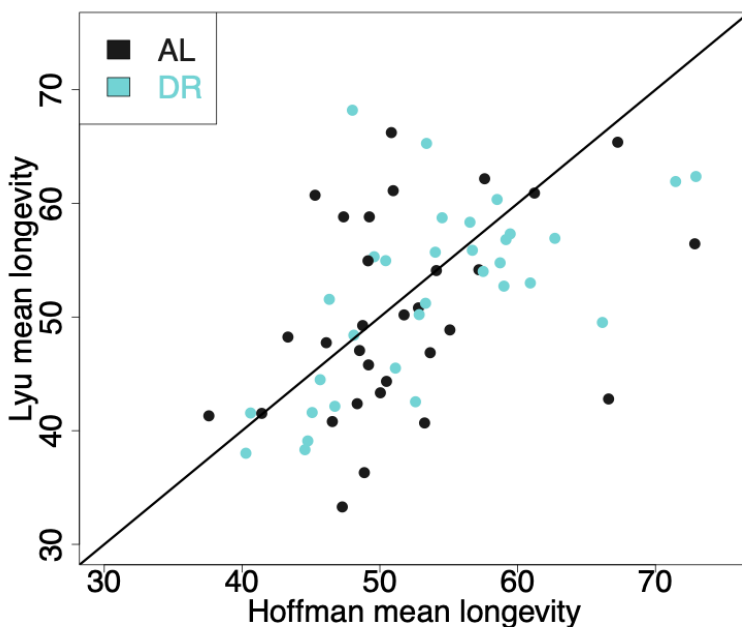


Figure 2. Correlation plot of each set of cohort pairs between the two labs. Each point represents the mean longevity for the Hoffman lab (x-axis) or Lyu lab (y-axis) for each individual treatment, sex, genotype, lab, cohort replicate ($n=64$). Black line is the line of symmetry.

Biological factors and stochasticity together influence lifespan

To understand how each factor influences lifespan, we used Cox regression to estimate the predictive power of each factor in the model fitting (see Methods for detailed calculation and Discussion for limitations). Specifically, we calculated the Cox-Snell R^2 for both the full model and the reduced models to examine the power of each covariate. We found that, compared to the full model, only removing the covariates genotype or cohort resulted in a moderate reduction in Cox-Snell R^2 (Supplementary Table 2), indicating that genotype and cohort are the main factors influencing lifespan in our dataset.

We further estimated the proportion of variance each factor explains using a likelihood-based method, summarized in Table 2. The variability among cohorts (16.35% of total variance) indicates the presence of stochasticity, which explains the results even better than sex (14.59% of total variance), a well-known factor that determines lifespan³². Genotype was the major determinant of variation in our dataset, accounting for 67.97% of the total variance. The variability between labs (3.33% of total variance) is small, consistent with our previous observation. Most surprisingly, dietary treatment, the main focus of this study, accounts for only 0.76% of the total variance. To visualize the differences, we present the average lifespan grouped by each factor in Figure 1, highlighting the negligible differences between labs (Fig. 1a) and dietary conditions (Figs. 1b and 1c), moderate differences in sex (Fig. 1e) and cohort (Fig. 1d), and remarkable differences in genotype (Fig. 1f).

Table 2. We estimated the accounted variance percentages for different factors across two dietary restriction (DR) protocols involving different cohorts, using a likelihood-based method.

Factor	DR Protocol 1&2 (Cohort 1-4) Accounted Variance (%)	DR Protocol 1 (Cohort 1-3) Accounted Variance (%)	DR Protocol 1 w/SY treatment (Cohort 1 & 3) Accounted Variance (%)
Lab	3.33	2.15	0.57
Cohort	16.35	17.01	9.52
Diet	0.76	0.75	0.18
Sex	14.59	12.54	13.4
Genotype	67.97	67.54	76.3

We consider the possibility that different DR protocols might contribute to the stochasticity, even though this is not suggested by data in Figure 1b and 1c. To rule out impact of different DR protocols, we estimated the proportion of variance explained using DR Protocol 1 (Cohorts 1-3), which shows a similar result to the entire dataset, indicating that stochasticity may account for 17.01% of the total variance. We also suspect that different food flies mated on before the lifespan assay (Cohort 1 and 3 versus 2, see Methods for details) may add to the stochasticity. To test this, we estimated the proportion of variance explained with only Cohorts 1 and 3, where the food flies mated on are the same (SY10). Indeed, we observed a decrease in the proportion of contribution by cohort (9.52%), but this number is still much larger than the proportion contributed by lab (0.57%) and diet (0.18%). In summary, our analyses indicate that genetic, sex, and stochastic factors are the predominant determinants of lifespan, with lab and dietary restriction regimen accounting for very little impact on longevity.

DR does not universally extend lifespan

One of the primary objectives of our experiment was to assess the reproducibility and stochastic nature of the longevity effects observed with dietary restriction. Combining two protocols, we found that DR flies were significantly longer-lived than those on high nutrient diets (Log-rank $p=4.7 \times 10^{-7}$), but the difference in median lifespan (7.1%) is rather small. Given the large stochastic effects in our dataset, we asked if the DR effects are

204 reproducible across different replicates. Out of the 64 pairs of DR/AL comparisons, we observed a significant
 205 lifespan effect of dietary restriction in only 17 out of 64 pairs (26.7%, log-rank test, Table 3). Survivorship curves
 206 are shown in Figure S1. Unexpectedly, in five comparisons, the AL group exhibited significantly longer lifespans.
 207 Previous research has consistently indicated that *D. melanogaster* tend to live longer under dietary restriction
 208 (Grandison, Wong et al. 2009; McCracken, Adams et al. 2020). However, our findings can be extrapolated to
 209 suggest that these effects are at least partially attributable to the toxicity of the enriched diet (see Discussion).
 210 The effects of dietary restriction *per se* appear to be minimal and sporadic when compared to what would be
 211 considered a standard diet.
 212

213 **Table 3 Log-rank test results and *P*-values for each AL/DR comparison pair. Bold text indicates *P*-values that pass**
 214 **the Bonferroni correction ($P \leq 0.00078$). The longer-lived group is indicated in brackets for significant comparisons.**

		Hoffman Lab				Lyu Lab			
		Cohort 1	Cohort 2	Cohort 3	Cohort 4	Cohort 1	Cohort 2	Cohort 3	Cohort 4
Canton-S	F	0.07	3 × 10⁻⁵ (AL)	6 × 10⁻¹⁰ (AL)	0.1	0.4	8 × 10⁻¹⁵ (DR)	0.02	6 × 10⁻⁶ (AL)
	M	0.02	0.5	2 × 10⁻⁹ (DR)	0.06	0.1	5 × 10⁻¹⁷ (DR)	0.004	7 × 10⁻⁴ (DR)
Oregon-R	F	1 × 10⁻⁵ (DR)	0.07	0.003	0.8	0.004	0.03	0.01	6 × 10⁻⁵ (DR)
	M	9 × 10⁻⁸ (DR)	0.001	3 × 10⁻¹⁴ (DR)	3 × 10⁻¹⁰ (DR)	0.7	5 × 10⁻⁷ (DR)	0.06	3 × 10⁻⁵ (DR)
w¹¹¹⁸	F	1	0.1	0.7	0.5	0.2	2 × 10⁻⁹ (DR)	0.008	0.8
	M	5 × 10⁻⁹ (DR)	0.03	0.005	3 × 10⁻⁷ (AL)	0.5	1 × 10⁻⁷ (DR)	5 × 10⁻⁴ (DR)	2 × 10⁻⁴ (DR)
w^{Dahomey}	F	1	0.1	1 × 10⁻⁴ (DR)	0.3	1 × 10⁻⁸ (AL)	0.04	0.1	0.6
	M	0.008	0.01	0.2	0.3	0.8	0.6	0.8	0.06

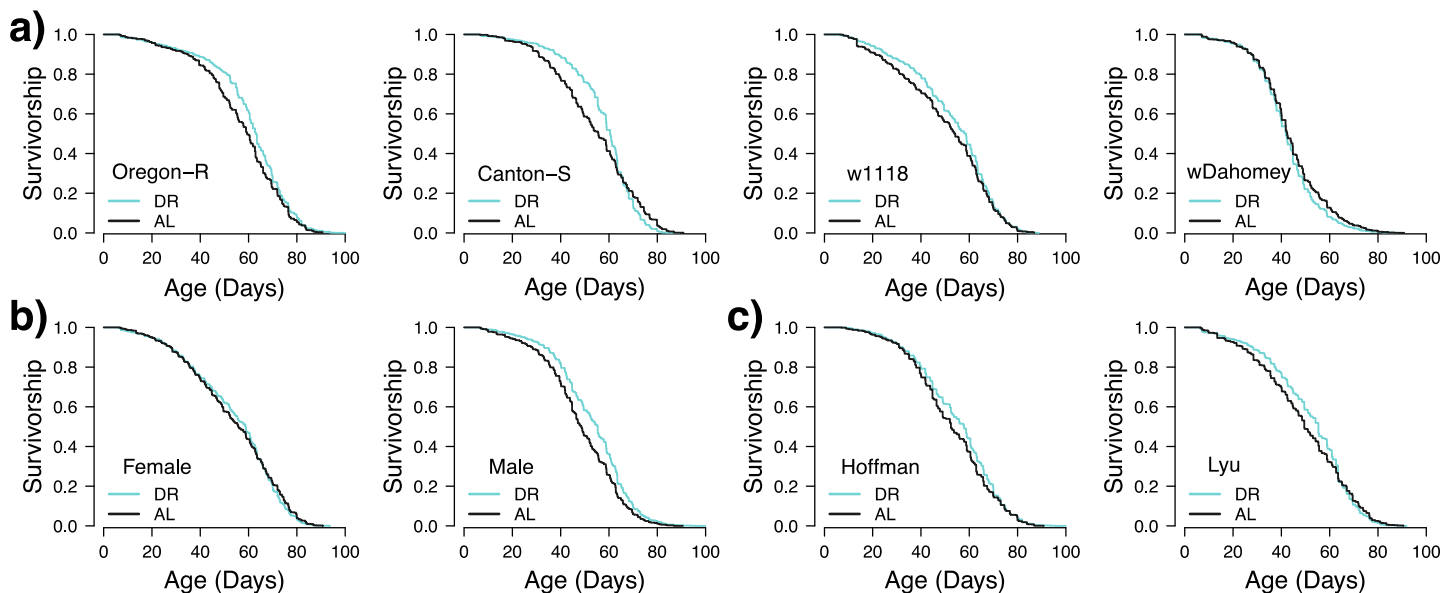
215

216 Within these analyses, certain genotypes were more likely to show an effect of DR, with w^{Dahomey} flies showing
 217 overall no real effect of DR, and Oregon-R flies showing lifespan extension under DR in almost 50% of the
 218 replicates with no increases in the AL groups (Table 3 and Figure 3a). Lastly, we also found significant sex-by-
 219 genotype effects, and in general males were more likely to respond to the different diets (Figure 3b), with the
 220 exception of w^{Dahomey}, in which no male replicates had any significant differences between the DR and AL diets.

221 We also observed variation in the two DR protocols, with Protocol 1 showing more pronounced DR effects
 222 (Figures 1b versus 1c). When examining the two labs separately, the Lyu lab found more DR effects using DR
 223 Protocol 2 (4 out 8 pairs have $P \leq 0.00078$). In addition, looking at the difference between AL-DR median
 224 lifespans, we see a positive trend for similar differences between pairs, though the effect was not significant
 225 (Supplemental Figure 2, Spearman rho=0.313, p=0.081). Given the stochastic effects observed between
 226 different cohorts, it is difficult to determine whether these differences are due to variations between the labs or
 227 stochasticity.

228
229
230
231
232
233

Similar to our longevity results, we found no effects of DR treatment on climbing ability, a marker of healthspan, in middle aged flies (ANOVA $p=0.54$, Supplementary Figure 3), but similar to the longevity results, there were significant effects of genotype (ANOVA $p=1.07 \times 10^{-7}$) and sex (ANOVA $p=0.004$). As expected, our body mass analysis found males were smaller than females (ANOVA $p=1.32 \times 10^{-7}$, Supplemental Figure 4), but there was no effect of DR treatment (ANOVA $p=0.23$) nor genotype on overall body mass (ANOVA $p=0.43$). These combined data suggest there were no effects of our DR protocols on healthspan in the flies.



234

235
236
237

Figure 3. The interaction of DR with sex (a), genotype (b), and lab (c). Kaplan–Meier survival curves represent the proportion of survivors over time (in days) during the adult stage. Only cohorts 1-3 (DR protocol 1) were used for this analysis to control for the protocol.

238

239

240

Discussion

241
242
243
244
245
246
247
248
249
250
251
252
253
254

Although DR is widely acknowledged as one of the most effective pro-longevity interventions across various species, recent studies from several species indicate variable lifespan responses to restricted diets^{13,14,27}. These variations can be attributed to differences in dietary regimes, genetic backgrounds²⁷, laboratory conditions³³, and other stochastic effects. Our collaborative effort, involving the replication of identical sets of experiments between two labs and repeating the same experiments multiple times within each lab, provides a unique opportunity to focus on the stochastic effects of DR. While we observe variations in lifespan between labs, these are not necessarily greater than the variations seen within repeated experiments in the same lab. This finding suggests that with rigorous control of laboratory conditions, inter-laboratory variability can be minimized (less than 4% of the total variance in our dataset), allowing a clearer focus on biological and stochastic effects, and our study strongly suggests that stochastic effects are one of the primary variables influencing lifespan under DR (Table 2). This conclusion is consistent with major findings from the *Caenorhabditis Interventions Testing Program* (CITP)^{19,34,35}, supporting the notion that variability in longevity control might be a universal phenomenon. Therefore, while DR may be a robust method to increase lifespan, there is significant variation in the magnitude and directionality of response.

255
256
257

One of the key reasons we observed significant stochasticity in our results is perhaps the small average lifespan differences between the DR and AL conditions in most of the genotypes, even when the sample size is sufficiently large (Figure 1b and 1c, Table 3). The average lifespan response to varying protein (yeast) concentrations in the

diet typically follows a bell-shaped curve across different genotypes²⁷. A major challenge in designing DR experiments is determining the optimal food formulation that maximizes lifespan under DR conditions, as well as identifying an appropriate standard diet for the high nutrient group. A common misinterpretation of DR effects arises when using an extra high-nutrient diet as the control, often referred to as the AL condition. In such cases, observed lifespan extensions under DR could be misleading, as they may reflect the harmful effects of a high-nutrient diet rather than true benefits of DR (see Discussion in Ref. 27). For example, recent studies suggest a large effect of DR on *Drosophila* lifespan³⁶, but the AL diet was 30% (w/v) Y and S, which is well outside of what is used in standard husbandry. When a standard diet is chosen properly (e.g. 1% compared 10% (w/v) S and Y, as shown in³⁷, the differences between the DR and AL groups tend to be subtle in most genotypes, as observed in our study and reported by others^{23,27,38}. Given this perspective, the lack of significant DR effects, though initially unexpected, becomes less surprising. This subtlety in DR response emphasizes the importance of carefully selecting control diets and highlights the inherent challenges in designing and interpreting DR studies.

Although we did not observe a remarkable lifespan extension with DR, the differences between the DR and AL groups were reasonably repeatable across our labs (Fig. 2). A previous report has analyzed the correlation between the lifespan differences (DR-AL) in their dataset³³ and those by a second study³⁹, reporting a correlation, although not statistically significant. This lack of significant correlation could be influenced by variations in fly husbandry and dietary regimes between the studies³³. Nevertheless, the delta in lifespan between the DR and AL conditions (Δ [DR-AL]) seems relatively consistent, even if the differences are not always significant nor positive (Supplementary Fig. 2). This “rule” suggests that within each genotype, the lifespan response curve to dietary concentration³⁷ is relatively stable.

Our findings underscore the importance of controlled experimental conditions and highlight the inherent challenges in achieving significant lifespan extensions through DR in certain genotypes. However, it is worth noting that the Oregon-R genotype consistently exhibits a DR response in 12 out of 16 trials in our studies ($P < 0.05$), with none showing an increase in the AL group. This suggests that in specific genetic backgrounds, the response to DR may be more predictable and robust. Understanding the genetic bases underlying this robustness is critical for future mechanistic studies, and for translating DR interventions into practical applications in daily life. Interestingly, we found remarkably similar median and maximum lifespans within a genotype across laboratories suggesting strong genetic effects on strain longevity, but not necessarily on strain response to DR. This is similar to our previous work suggesting high genetic correlation across strains within and between labs²⁰. Together, both genotype (G) and the interaction between genotype and diet (G x E) seem to have more significant impact on longevity than diet alone (E). Thus, as has been becoming more and more evident in the aging field, studies of multiple genetic backgrounds are necessary to understand the species level effects of different interventions and environmental conditions.

It may be noteworthy that we found no effect of diet on climbing ability or weight across our treatments, although these data we collected in only one of our labs (Supplementary Fig. 3 and 4). This suggests that first, what we are considering to be AL/DR in flies is not an accurate representation, specifically the AL group, as the DR group did not have a small body mass than the AL group, as would be expected in mammalian CR studies, where CR mice are significantly smaller than those on AL diets^{3,12}. Potentially we need a new way to denote DR studies that refer to the high/low nutrients of the diet but not necessarily the caloric intake of individuals on the diet as is denoted by the name ‘ad libitum’. In addition, as we found minor effects of DR on lifespan, it is not particularly surprising that health was also not affected. This is in line with previous studies showing that the correlation between health- and lifespan also depends on the genetic background^{14,34}. Like our longevity results, we found effects of sex and genotype on the climbing ability (and weight) that completely overshadowed any DR effect. Combined, these results suggest again that DR may have minor effects in *Drosophila* when restricted animals are compared to a ‘standard’ diet, and genetic background effects drive most of the variation in organismal health in fruit flies.

304 **Caveats**

305 DR Protocols - While our results hint toward some of the nuanced conditions that must be considered when
306 interpreting dietary interventions and longevity response in *Drosophila*, our results are not without their
307 limitations. Experimental diets using CT and SY10 foods were selected based on their common use as stock
308 diets in *Drosophila* laboratory husbandry. The addition of cornmeal in the CT food may slow the mechanical
309 ingestion and metabolism and have physiological impacts, though our minor longevity effects seen comparing
310 CT and SY10 suggest these effects are most likely minor. In addition, our SY5 and SY15 diets did not show
311 many DR effects in the Hoffman lab specifically, suggesting our lack of CT/SY10 effects are most likely not due
312 to any intentional differences in the food media. Both labs experienced issues with food quality across the
313 experimental cohorts leading to censoring of flies, usually related to overly wet/sticky food; however, these food
314 issues were random and would have been equally applied to all groups minimizing their overall effects. Still, we
315 cannot rule out a bias in our removal of individual flies from the analysis.

316 Modeling - The assumption of proportional hazards in the Cox Regression model was not met, as indicated by
317 the p-values from the proportional hazards test being less than 0.05 for all covariates except Lab. Given our
318 large sample size, it is challenging to completely avoid violations of this assumption, and even small deviations
319 can look like a violation when they are not biologically meaningful. Perhaps not surprisingly given the rest of our
320 results, the largest deviations from the proportional hazards assumptions were due to the genotype effects. In
321 the future, adjusting the model to include time-dependent covariates may address these violations and improve
322 the accuracy of our results.

323 **Conclusions**

324 Combined, our results find inconsistent DR longevity effects across labs within *Drosophila melanogaster*. As fruit
325 flies are common longevity and dietary intervention models, it is important to note that any observed longevity
326 effects in other studies may be due to stochastic variation within and across labs. We would suggest future
327 studies need to thoughtfully design experiments with appropriate AL diets, and in addition, future studies must
328 carefully interpret data, especially those that apply to minor effects. This caution is also likely relevant to other
329 invertebrate species. Moving forward, one of the priorities perhaps should be focused on mapping the genetic
330 alleles that influence the degree of variation in DR-mediated lifespan changes, as genotype was the largest
331 factor affecting both overall longevity and response to DR. Utilizing existing population genomic resources will
332 be essential in identifying such genetic determinants. Our insights on diet and longevity relative to genetic make-
333 up, food regimen, and stochastic factors, will be crucial for advancing effective approaches for personalized
334 medicine and nutrition, allowing for more tailored and effective longevity interventions.

337 **Acknowledgements**

338 The authors would like to thank members of the Lyu and Hoffman labs for help with fly husbandry. We would
339 also like to thank Monica Driscoll for her valuable comments on the manuscript. This work was funded by
340 R00AG059920 to JMH.

342 **Data availability**

343 Our lifespan data is available in Supplementary Table 1.

345 Competing interests

346 The authors declare no competing interests.

348 Author contributions

349 The paper had the following contributions by each author: conception and experimental design- JMH and YL;
350 data collection- OLM, JV, AK, EJ; data analysis, figure creation, writing first draft- OLM, JMH, YL. All authors
351 edited and approved the final version of the manuscript.

353 Supplemental Figure legends

354 **Supplemental Figure 1. Kaplan-Meier curves of each of 64 pairs of AL/DR experiments.**

355 **Supplemental Figure 2. Difference of AL and DR median lifespan between labs.**

356 **Supplemental Figure 3. Climbing results for 30-day old flies from the Hoffman lab for females (A) and**
357 **males (B).** Each replicate consists of 18 vials of ~20 flies each. Mean climbing values were taken on a per vial
358 average. Cohorts 1-3 were combined for analysis. There were significant effects of sex and genotype with no
359 difference between AL and DR treatments.

360 **Supplemental Figure 4. Body mass results for 30-day old flies on SY5 vs SY15 for females (A) and males**
361 **(B).** Each replicate consisted of ~ 5 measurements of 5 flies each. There were no significant effects of treatment,
362 suggesting that our flies were not calorically restriction on the DR treatment. Females were significantly larger
363 than males as expected, and no genotype effects were seen.

365 References

- 366 1 Fontana, L. & Partridge, L. Promoting health and longevity through diet: from model organisms to humans. *Cell* **161**,
367 106-118 (2015). <https://doi.org/10.1016/j.cell.2015.02.020>
- 368 2 Green, C. L., Lamming, D. W. & Fontana, L. Molecular mechanisms of dietary restriction promoting health and
369 longevity. *Nat Rev Mol Cell Biol* **23**, 56-73 (2022). <https://doi.org/10.1038/s41580-021-00411-4>
- 370 3 McCay, C. M., Crowell, M. F. & Maynard, L. A. The effect of retarded growth upon the length of life span and upon the
371 ultimate body size: one figure. *J Nutr* **10**, 63-79 (1935).
- 372 4 Lin, S. J., Defossez, P. A. & Guarente, L. Requirement of NAD and SIR2 for life-span extension by calorie restriction in
373 *Saccharomyces cerevisiae*. *Science* **289**, 2126-2128 (2000). <https://doi.org/10.1126/science.289.5487.2126>
- 374 5 Hosono, R., Nishimoto, S. & Kuno, S. Alterations of life span in the nematode *Caenorhabditis elegans* under monoxenic
375 culture conditions. *Exp Gerontol* **24**, 251-264 (1989). [https://doi.org/10.1016/0531-5565\(89\)90016-8](https://doi.org/10.1016/0531-5565(89)90016-8)
- 376 6 Chippindale, A. K., Leroi, A. M., Kim, S. B. & Rose, M. R. Phenotypic plasticity and selection in *Drosophila* life-history
377 evolution. I. Nutrition and the cost of reproduction. *J Evol Biol* **6**, 171-193 (1993).
- 378 7 Weindruch, R., Walford, R. L., Fligiel, S. & Guthrie, D. The retardation of aging in mice by dietary restriction: longevity,
379 cancer, immunity and lifetime energy intake. *J Nutr* **116**, 641-654 (1986). <https://doi.org/10.1093/jn/116.4.641>
- 380 8 Waziry, R. *et al.* Effect of long-term caloric restriction on DNA methylation measures of biological aging in healthy adults
381 from the CALERIE trial. *Nat Aging* **3**, 248-257 (2023). <https://doi.org/10.1038/s43587-022-00357-y>
- 382 9 Kapahi, P. *et al.* Regulation of lifespan in *Drosophila* by modulation of genes in the TOR signaling pathway. *Curr Biol* **14**,
383 885-890 (2004). <https://doi.org/10.1016/j.cub.2004.03.059>
- 384 10 Cantó, C. & Auwerx, J. Calorie restriction: is AMPK a key sensor and effector? *Physiology (Bethesda)* **26**, 214-224 (2011).
385 <https://doi.org/10.1152/physiol.00010.2011>
- 386 11 Bylino, O. V., Ogienko, A. A., Batin, M. A., Georgiev, P. G. & Omelina, E. S. Genetic, environmental, and stochastic
387 components of lifespan variability: The *Drosophila* paradigm. *Int J Mol Sci* **25** (2024).
388 <https://doi.org/10.3390/ijms25084482>

- 389 12 Harper, J. M., Leathers, C. W. & Austad, S. N. Does caloric restriction extend life in wild mice? *Aging Cell* **5**, 441-449
390 (2006). <https://doi.org/10.1111/j.1474-9726.2006.00236.x>
- 391 13 Liao, C. Y., Rikke, B. A., Johnson, T. E., Diaz, V. & Nelson, J. F. Genetic variation in the murine lifespan response to
392 dietary restriction: from life extension to life shortening. *Aging Cell* **9**, 92-95 (2010). <https://doi.org/10.1111/j.1474-9726.2009.00533.x>
- 393
394 14 Wilson, K. A. *et al.* GWAS for lifespan and decline in climbing ability in flies upon dietary restriction reveal decima as a
395 mediator of insulin-like peptide production. *Curr Biol* **30**, 2749-2760.e2743 (2020).
396 <https://doi.org/10.1016/j.cub.2020.05.020>
- 397 15 Finch, C. E. & Kirkwood, T. B. *Chance, development, and aging*. (Oxford University Press, USA, 2000).
- 398 16 Martin, G. M. Stochastic modulations of the pace and patterns of ageing: impacts on quasi-stochastic distributions of
399 multiple geriatric pathologies. *Mech Ageing Dev* **133**, 107-111 (2012). <https://doi.org/10.1016/j.mad.2011.09.001>
- 400 17 Finch, C. E. & Haghani, A. Gene-environment interactions and stochastic variations in the gero-exposome. *J Gerontol*
401 *A Biol Sci Med Sci* **76**, 1740-1747 (2021). <https://doi.org/10.1093/gerona/glab045>
- 402 18 Herndon, L. A. *et al.* Stochastic and genetic factors influence tissue-specific decline in ageing *C. elegans*. *Nature* **419**,
403 808-814 (2002). <https://doi.org/10.1038/nature01135>
- 404 19 Lithgow, G. J., Driscoll, M. & Phillips, P. A long journey to reproducible results. *Nature* **548**, 387-388 (2017).
405 <https://doi.org/10.1038/548387a>
- 406 20 Hoffman, J. M., Dudeck, S. K., Patterson, H. K. & Austad, S. N. Sex, mating and repeatability of *Drosophila melanogaster*
407 longevity. *R Soc Open Sci* **8**, 210273 (2021). <https://doi.org/10.1098/rsos.210273>
- 408 21 Mendenhall, A., Driscoll, M. & Brent, R. Using measures of single-cell physiology and physiological state to understand
409 organismic aging. *Aging Cell* **15**, 4-13 (2016). <https://doi.org/10.1111/acer.12424>
- 410 22 Mendenhall, A. R., Martin, G. M., Kaeberlein, M. & Anderson, R. M. Cell-to-cell variation in gene expression and the
411 aging process. *Geroscience* **43**, 181-196 (2021). <https://doi.org/10.1007/s11357-021-00339-9>
- 412 23 Grandison, R. C., Wong, R., Bass, T. M., Partridge, L. & Piper, M. D. Effect of a standardised dietary restriction protocol
413 on multiple laboratory strains of *Drosophila melanogaster*. *PLoS One* **4**, e4067 (2009).
414 <https://doi.org/10.1371/journal.pone.0004067>
- 415 24 Mair, W., Piper, M. D. & Partridge, L. Calories do not explain extension of life span by dietary restriction in *Drosophila*.
416 *PLoS Biol* **3**, e223 (2005). <https://doi.org/10.1371/journal.pbio.0030223>
- 417 25 Juricic, P., Grönke, S. & Partridge, L. Branched-chain amino acids have equivalent effects to other essential amino
418 acids on lifespan and aging-related traits in *Drosophila*. *J Gerontol A Biol Sci Med Sci* **75**, 24-31 (2020).
419 <https://doi.org/10.1093/gerona/glz080>
- 420 26 Austad, S. N., Smith, J. R. & Hoffman, J. M. Amino acid restriction, aging, and longevity: an update. *Front Aging* **5**,
421 1393216 (2024). <https://doi.org/10.3389/fragi.2024.1393216>
- 422 27 McCracken, A. W., Buckle, E. & Simons, M. J. P. The relationship between longevity and diet is genotype dependent and
423 sensitive to desiccation in *Drosophila melanogaster*. *J Exp Biol* **223** (2020). <https://doi.org/10.1242/jeb.230185>
- 424 28 Linford, N. J., Bilgir, C., Ro, J. & Pletcher, S. D. Measurement of lifespan in *Drosophila melanogaster*. *J Vis Exp* (2013).
425 <https://doi.org/10.3791/50068>
- 426 29 Therneau, T. M. *A package for survival analysis in R*, <<https://CRAN.R-project.org/package=surviva>> (2022).
- 427 30 Therneau, T. M. & Grambsch, P. M. *Modeling survival data: Extending the Cox model*. (Springer, 2000).
- 428 31 Cox, D. R. & Snell, E. J. *Analysis of binary data*. (Routledge, 2018).
- 429 32 Austad, S. N. & Fischer, K. E. Sex differences in lifespan. *Cell Metab* **23**, 1022-1033 (2016).
430 <https://doi.org/10.1016/j.cmet.2016.05.019>
- 431 33 Simons, M. J. P. & Dobson, A. J. The importance of reaction norms in dietary restriction and ageing research. *Ageing Res*
432 *Rev* **87**, 101926 (2023). <https://doi.org/10.1016/j.arr.2023.101926>
- 433 34 Banse, S. A. *et al.* The coupling between healthspan and lifespan in *Caenorhabditis* depends on complex interactions
434 between compound intervention and genetic background. *Aging (Albany NY)* **16**, 5829-5855 (2024).
435 <https://doi.org/10.18632/aging.205743>
- 436 35 Onken, B. *et al.* Metformin treatment of diverse *Caenorhabditis* species reveals the importance of genetic background
437 in longevity and healthspan extension outcomes. *Aging Cell* **21**, e13488 (2022). <https://doi.org/10.1111/acer.13488>
- 438 36 Li, M. *et al.* Late-life shift in caloric intake affects fly metabolism and longevity. *Proc Natl Acad Sci U S A* **120**,
439 e2311019120 (2023). <https://doi.org/10.1073/pnas.2311019120>
- 440 37 Partridge, L., Piper, M. D. & Mair, W. Dietary restriction in *Drosophila*. *Mech Ageing Dev* **126**, 938-950 (2005).
441 <https://doi.org/10.1016/j.mad.2005.03.023>
- 442 38 McCracken, A. W., Adams, G., Hartshorne, L., Tatar, M. & Simons, M. J. P. The hidden costs of dietary restriction:
443 Implications for its evolutionary and mechanistic origins. *Sci Adv* **6**, eaay3047 (2020).
444 <https://doi.org/10.1126/sciadv.aay3047>
- 445 39 Jin, K. *et al.* Genetic and metabolomic architecture of variation in diet restriction-mediated lifespan extension in
446 *Drosophila*. *PLoS Genet* **16**, e1008835 (2020). <https://doi.org/10.1371/journal.pgen.1008835>