

REVIEW ARTICLE OPEN



The role of protein lactylation in brain health and disease: current advances and future directions

Mingrui Han^{1,2,5}, Wenfeng He^{1,5™}, Wengen Zhu 10 and Linjuan Guo^{4™}

© The Author(s) 2025

Lactate, the end product of glycolysis, plays a crucial role in cellular signaling and metabolism. The discovery of lactylation, a novel post-translational modification, has uncovered the role of lactate in regulating diseases, especially in the brain. Lactylation connects genetic encoding with protein function, thereby influencing key biological processes. Increasing evidence supports lactate-mediated lactylation as a critical modulator in neurological disorders. This review offers an overview of lactate metabolism and lactylation, highlighting recent advances in understanding the regulatory enzymes of lactylation and their role in the central nervous system. We investigate the impact of lactylation on brain dysfunctions, including neurodegenerative diseases, cerebrovascular disorders, neuroinflammation, brain tumors, and psychiatric conditions. Moreover, we highlight the therapeutic potential of targeting lactylation in treating brain disorders and outline key research gaps and future directions needed to advance this promising field.

Cell Death Discovery (2025)11:213; https://doi.org/10.1038/s41420-025-02408-w

FACTS

- Lactate, a carboxylic acid, plays a significant role in cellular signaling and brain metabolism.
- Lactylation, is an emerging post-translational modification that adds lactyl groups to both histone and non-histone proteins, participating in a variety of pathophysiological processes
- Lactylation not only alters gene expression levels but also impacts biological functions by modifying the physicochemical properties of proteins.
- Studies indicate that targeting lactylation modifications is an effective strategy for treating central nervous system diseases.

OPEN QUESTIONS

- Research on the regulatory enzymes of lactylation modifications in the central nervous system is currently limited.
 Are there specific enzymes that mediate lactylation in the brain?
- What are the potential molecular mechanisms linking lactylation to central nervous system diseases, and how can we utilize these biological processes to guide the development of new therapeutic strategies?
- What role does lactylation play in the pathophysiological processes of the central nervous system? A protector or a disruptor?

INTRODUCTION

Brain diseases, such as neurodegenerative disorders, cerebrovascular conditions, and psychiatric illnesses, are major health challenges that are increasingly associated with metabolic dysfunctions [1]. Among these, the role of lactate has emerged as a key area of interest because of its role in cellular signaling and brain metabolism. Although traditionally viewed as a metabolic waste product linked to hypoxic conditions, the significance of lactate in various biological processes is now being reevaluated [2].

Lactylation, a post-translational modification (PTM) identified in 2019, introduces a novel mechanism by which lactate can modulate protein function and gene expression [3]. This discovery has opened new avenues for understanding how metabolic changes impact brain health and diseases. The regulation of lactylation involves specific enzymes, termed writers, erasers, and readers, which control the addition and removal of lactyl groups and thereby influence key cellular functions [4]. While the role of lactate in brain metabolism is well-studied, the impact of lactylation is still under-researched, particularly in relation to brain disorders.

Although prior studies have discussed the roles of lactylation in conditions such as gliomas [5], neurogenesis [6], and neurodegenerative diseases [6, 7], they often suffer from incomplete coverage of regulatory enzymes and outdated references. To bridge these gaps, this review offers an up-to-date overview of lactylation research in brain health by including the latest published studies. Furthermore, we systematically categorize lactylation regulatory enzymes for the first time to investigate

Received: 6 November 2024 Revised: 19 February 2025 Accepted: 18 March 2025

Published online: 30 April 2025

¹Department of Medical Genetics, The Second Affiliated Hospital of Nanchang University, Nanchang, Jiangxi, China. ²Queen Mary school, medical department, Nanchang University, Nanchang, Jiangxi, China. ³Department of Cardiology, The First Affiliated Hospital of Sun Yat-Sen University, Guangzhou, China. ⁴Department of Cardiology, Jiangxi Provincial People's Hospital, The First Affiliated Hospital of Nanchang Medical College, Nanchang, Jiangxi, China. ⁵These authors contributed equally: Mingrui Han, Wenfeng He. [™]email: fengfeng_982366@126.com; 18170044224@qq.com; 727456372@qq.com

their role in lactylation across various neurological diseases. We have summarized the pathophysiological processes of neurodegenerative diseases, cerebrovascular disorders, neuroinflammation, brain tumors, and psychiatric conditions, along with the mechanisms of lactylation modifications' involvement. In conclusion, we determine that targeting lactylation possesses substantial therapeutic potential and significantly contributes to mitigating metabolic dysfunctions in neurological diseases.

LACTATE METABOLISM AND ITS ROLE IN THE BRAIN

Under normal physiological conditions, glucose is converted to pyruvate in the cytoplasm and subsequently enters the tricarboxylic acid (TCA) cycle in the mitochondria, producing substantial energy, oxygen, and water for cellular activities. In hypoxic or other adverse environments, respiration proceeds anaerobically, converting pyruvate from glucose to lactate by lactate dehydrogenase (LDH) via anaerobic glycolysis [8]. At rest, the brain is responsible for 13% of the body's lactate production and 8% of its utilization. During exercise, the brain's lactate uptake increases significantly, consuming about 11% of the body's lactate, underscoring the critical role of lactate metabolism in brain function and systemic homeostasis [9]. Even under aerobic conditions, cancer cells often prefer anaerobic glycolysis, known as the Warburg effect [8, 10], which produces energy more rapidly than oxidative phosphorylation (OXPHOS) and satisfies the increased demand for glycolytic intermediates, supporting rapid tumor growth and leading to poor clinical outcomes [8, 11, 12].

Lactate is cleared either by oxidizing to pyruvate for re-entry into the TCA cycle or by converting back to glucose in the liver and skeletal muscle via the Cori cycle [13, 14]. Lactate exists primarily in two enantiomeric forms: L-lactate and D-lactate [8]. L-lactate is produced directly by glycolysis, whereas D-lactate formation in eukaryotes occurs mainly through two pathways: the glyoxalase enzyme system and gut microbiota [13]. In the glyoxalase pathway, a branch of glycolysis, glucose is converted to methylglyoxal and subsequently processed by glyoxalase 1 (GLO1) and GLO2 into D-lactate and glutathione. Meanwhile, gut microbes, such as lactic acid bacteria, also convert glucose into D-lactate, which enters the bloodstream [13].

L-lactate, the predominant form in the human body and brain, is involved in various biological processes, such as regulating energy production and metabolic pathways [13, 14]. Lactate enters cells through monocarboxylate transporters (MCTs) on the cell membrane or via GPR81 receptor-mediated signaling pathways [14]. MCTs are extensively expressed throughout the brain [8, 14]. The intercellular lactate shuttle describes the transfer of lactate from glycolytic cells to surrounding cells. In the nervous system, the most extensively studied is the astrocyte-neuron lactate shuttle (ANLS) [7]. Astrocytes take up glucose from the bloodstream, store it as glycogen, and produce lactate via rapid glycolysis in response to heightened neuronal activity. This lactate is then transported via MCTs to the extracellular space, where neurons uptake it through MCTs, converting it to pyruvate and acetyl-CoA for ATP production via OXPHOS, which supports synaptic formation and neurotransmitter release [7, 15]. This evidence challenges the conventional view of lactate merely as a metabolic waste product. In the brain, lactate's metabolic ecology and its spatial and temporal distribution are closely tied to cerebrovascular blood flow, neuronal excitation and neuroplasticity [16]. A significant outcome of the inflammatory pathology linked to cytotoxicity and hypoxia within the tumor microenvironment is an increase in local tissue lactate levels [17]. Recent research underscores the vital roles of lactate in enhancing memory impairments, alleviating neurological dysfunctions, and fostering neuronal repair and regeneration [8].

LACTYLATION DEVELOPMENT AND ITS MECHANISMS Development of lysine lactylation

PTMs alter the biochemical properties of specific amino acid residues in proteins, which changes their functions and regulates gene expression. The concept of enhanced lactate production was first described a century ago by the Warburg effect, which revealed that tumor cells rely on anaerobic glycolysis even in the presence of oxygen, providing them with numerous advantages. Subsequently, extensive research on lactate has led to the recognition that lactate is no longer considered a metabolic waste product, but rather plays a crucial role in various biological processes such as cellular metabolism and signaling [4]. In 2019, Professor Zhao Yingming's team made a groundbreaking discovery, identifying lactyl-CoA-mediated histone L-lysine lactylation (Kla) in human breast cancer MCF-7 cells and its involvement in gene expression regulation. This modification was confirmed through immunoblotting and isotope labeling experiments, demonstrating that it originates from intracellular glycolysis [3].

Lactylation modifications exist in three distinct stereochemical forms: lysine I-lactylation (KI-la), ε-(carboxyethyl)-lysine (Kce), and dlactyl-lysine (Kd-la). Kl-la, the predominant form of lactylation mediated by lactyl-CoA, is tightly regulated by glucose concentration and glycolytic levels [18]. This novel modification type is present in various cellular compartments, and its extensive diversity in protein substrates is evidenced by its dual targeting of nuclear histones and cytoplasmic non-histones. Since 2019, histone lactylation has been extensively studied. Changes in chromatin structure affect gene expression, which in turn plays a decisive role in cellular function and fate. It has been confirmed to play a critical role in the development of various diseases, including cancer, neuropsychiatric disorders, and cardiovascular diseases [19]. For instance, H4 lysine 12 lactylation (H4K12la) induces smooth muscle cell senescence by promoting the expression of aging-related genes, thereby accelerating atherosclerosis [20]. As research progresses, non-histone targets have also been gradually identified, which exhibit a pronounced preference for enzymes involved in metabolic pathways [21]. Non-histone lactylation can influence the spatial localization, conformation, and other characteristics of target proteins, thereby regulating the stability of protein-protein interactions and ultimately altering their functional state. By constructing L-lactate sodium derivatives, numerous potential non-histone sites have been identified [22], further affirming their significance beyond histones. Notably, recent studies have for the first time revealed that guanosine triphosphate (GTP)-specific SCS (GTPSCS) and acetyl-CoA synthetase 2 (ACSS2) are responsible for synthesizing lactyl-CoA. Both of them participate in histone lactylation, with the assistance of P300 and KAT2A, respectively, playing a significant role in the development of gliomas [23, 24]. With this discovery, the synthesis mechanism of lactylation modification precursors has been elucidated, marking a new milestone in lactylation research. These studies position lactylation as a crucial player in the pathophysiological processes of various diseases, significantly expanding our understanding of the role of lactate in disease contexts.

Enzyme-mediated lysine lactylation

The precise execution of enzyme-mediated lactylation requires the assistance of specialized tools. Writers, erasers, and readers are three pivotal regulatory enzymes in this process, managing the addition of lactylation modifications, the elimination of lactyl groups, and their spatial distribution. L-lactate, the substrate for lactylation, is supplied through two pathways: one is the conversion of extracellular glucose molecules into pyruvate through glycolysis, which is then catalyzed by LDH to form L-lactate; the other method involves direct transport of L-lactate into the cell via MCT. GTPSCS-mediated conversion of L-lactate to lactyl-CoA is considered a prerequisite for catalysis, as the writers cannot directly add the lactyl group from lactate to lysine residues [8, 23]. Recently, the discovery of new catalytic mechanisms of

alanyl-tRNA synthetase 1 (AARS1) has shown that L-lactate can be converted into lactyl-AMP, which can also be utilized by writers, indicating that they are not entirely reliant on lactyl-CoA [25, 26]. Once the catalytic substrates are ready, writers, erasers, and readers are responsible for regulating the subsequent lactylation modification process (Supplemental Table 1).

Writers. Writers are responsible for adding lactylation modifications to both histone and non-histone proteins. Researchers have identified numerous enzymes that can act as lactylation architects. Most enzymes belong to three main classical histone acetyltransferase (ACT) family (HAT or KAT): p300/CREB-binding protein (CBP) family involving p300 and CBP, MYST family including MOF, HBO1, and Tip60 and Gcn5-related N-acetylase (GNAT) family involving GCN-5 and Yiac [27]. Other enzymes, such as AARS and alphatubulin ACT1 (ATAT1) have also been found to play a role. Current research indicates that the efficiency of most writers in adding lactyl groups depends on the intracellular levels of lactate synthesis and the ability of MCT to transport lactate into cells, using lactyl-CoA as the substrate. However, AARS is an exception, which relies on lactate-AMP as an intermediate to directly transfer lactyl group to the lysine residues of target proteins with the assistance of ATP [25, 26]

As the most well-studied writer, p300-catalyzed lactylation mainly promotes gene transcription [28-39] and maintains normal protein structure and functions [40-45]. In the central nervous system (CNS), P300 and its homologous protein CBP participate in the physiological processes of various diseases by mediating histone and non-histone lactylation. For example, pre-treatment with a small molecule inhibitor of P300 significantly reduced the level of lactylation in a mouse model of ischemic stroke, helping to maintain neuronal activity and alleviate ischemic brain injury [46]. In microglia, lactylation modifications act as a double-edged sword in their impact on abnormal brain pathological conditions. Retinal microglial cells undergo P300-mediated lactylation of YY1, activating microglial cells by promoting the transcription of certain inflammatory genes [38]. In addition, the promoting role of P300/CBP and its associated factors in the aging process of microglia in Alzheimer's disease (AD) has also been observed. However, in certain cases, this epigenetic modification can induce the transition of microglia to the M2 repair phenotype when lactate levels are increased [47]. For glioblastoma multiforme (GBM), the P300-mediated writing process profoundly influences disease progression and treatment strategies. P300-mediated histone lactylation has been observed to be predominantly localized at transcription start sites, significantly driving the malignancy of the U87MG GBM cell line and promoting immunosuppression in the GBM microenvironment. The enhanced efficacy of GBM immunotherapy is attributed to the elimination of histone lactylation by P300 inhibitors [48]. Recent groundbreaking findings have identified GTPSCS as lactyl-CoA synthetase in the nucleus of glioma cells, promoting glioma progression through P300-mediated histone lactylation [23]. Thus, P300/CBP-mediated protein lactylation plays a crucial role in various brain diseases and brain cell activities.

Overall, the mechanisms by which P300 mediates lactylation modifications require further investigation and refinement. The process of gene expression regulation involves a complex regulatory network, including chromatin assembly and the transmission of genetic information, with histone chaperones being one of the key components [49]. Initially recognized in the cardiovascular field, ASF1A has been identified as a cofactor that assists P300 in catalyzing H3K18 lactylation [39]. This indicates that the P300-mediated lactylation process is unlikely to be isolated, and its regulatory factors warrant further study. In terms of disease treatment, regulating lactylation levels in pathological states presents a potential strategy. However, it is crucial to consider how to balance lactate concentration with the catalytic effects of

P300. Among the determinants of the final lactylation modification levels, the "substrate" appears to be more critical than the "tool". In conditions such as pancreatic ductal adenocarcinoma and heart failure, the effects caused by changes in lactate levels cannot be reversed merely by manipulating P300 [33, 41].

Erasers. The "eraser" refers to enzymes that remove lactivation modifications on lysine residues. Histone deacetylases (HDAC), common in mammals [33, 50], are de-lactylation enzymes that effectively remove Kl-la and Kd-la modifications. Mammalian deacetylases are divided into two major families: zinc-dependent HDAC 1-11, also known as Class I, II, and IV HDACs, and NADdependent sirtuins (SIRT) 1-7, categorized as Class III HDACs [27]. Initially, HDACs are generally believed to preferentially target histones, whereas sirtuins exhibit a notable preference for nonhistone components in the cytoplasm or within the nucleus, exhibiting complementary functions [51, 52]. Nonetheless, sirtuins also play a role in the removal of histone lactylation at the cellular level. The potent de-lactylation activity of HDAC1-3, observed in both cellular and in vitro purified proteins, was first identified by Yruela et al., particularly regarding Kd-la [53]. While HDACs may more commonly erase Kl-la modifications, the removal of Kl-la compared to Kd-la might involve specific mechanisms and depend on the cross-linking of different substances within the nucleus [51].

The regulation of erasers is of great significance for the development and function of CNS. Inhibition of HDAC1-3 has been shown to modulate neuronal identity during murine neurodevelopment, activating transcription. H3K9 crotonylation (H3K9cr), H3K9 acetylation (H3K9ac) and H3K18la dynamically change in the genome during embryonic development, working together with other PTM modifications to determine chromatin accessibility and neuronal fate [54]. Additionally, the de-lactylation of H4K8 by SIRT2 has been shown to suppress neuroglioma cells [55]. HDACs and SIRTs, by removing lactylation modifications, regulate the levels of lactylation in the CNS together with the writers. The balance between these enzymes plays a crucial role in gene expression and protein stability in normal brain cells.

Readers. Readers are defined as enzymes that regulate and recognize lactylation modifications, with research in this area being largely unexplored. Current research on readers primarily focuses on their role in tumor progression, such as in colorectal cancer [31] and cervical cancer [56]. In the nervous system, the function of readers has also been elucidated. Evidence suggests that H4K8la levels of astrocytes in subarachnoid hemorrhage (SAH) are modulated by Bromodomain-containing protein 4 (BRD4). Silencing of BRD4 profoundly affects H4K8la levels, with immunoprecipitation experiments directly demonstrating their interaction [57]. Given BRD4's capacity to engage with enhancers and histone acetylation marks, forming intricate interactions to exert robust regulatory effects on gene transcription, BRD4 may be a potential reader in this context [58, 59]. Readers, in collaboration with writers and erasers, collectively influence the dynamic balance of lactylation modifications in cells, playing a significant role in the regulation of cellular activities. Further studies are required to fill the gaps in this field.

Non-enzymatic lysine lactylation

The non-enzymatic lactylation process involves the interaction of lactoylglutathione (LGSH) with lysine residues. The spontaneous incorporation of lactoylLys was initially validated by Gaffney et al. and this process is meticulously regulated by LGSH and GLO2 [60]. Compared to Kl-la, Kd-la is more prone to spontaneous lactyl group addition mediated by LGSH [11]. This is further supported by Zhang et al., who demonstrated that impairing the GLO pathway specifically disrupts the production of Kce and Kd-la, while the generation of lactyl-CoA appears to be positively correlated with Kl-la [18] (Fig. 1).

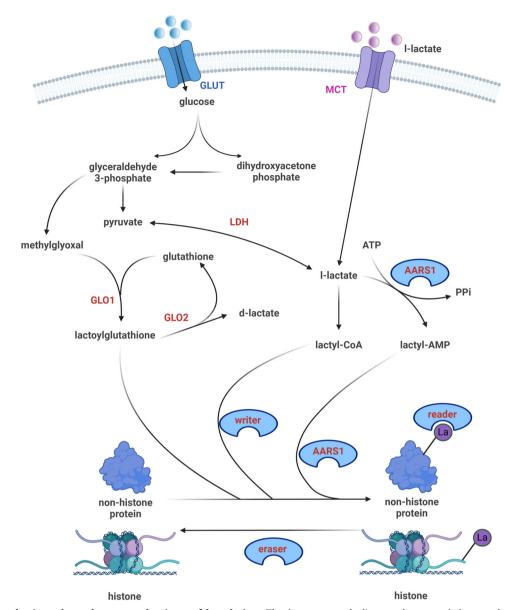


Fig. 1 Lactate synthesis and regulatory mechanisms of lactylation. The lactate metabolism pathway and the regulatory mechanisms of lactylation are illustrated as shown. Lactylation modification targets include both histones and non-histones, and it is accomplished through two pathways: enzyme-catalyzed and non-enzyme-catalyzed reactions. The first pathway: L-lactate which is transported into the cell through MCT or synthesized from pyruvate is the primary substrate for enzyme-catalyzed lactylation. The catalytic process of lactylation by the writer requires the synthesis of lactyl-CoA, while AARS1 uses lactyl-AMP as an intermediate. The balance of lactylation modification is ultimately regulated by erasers and readers, with the former removing lactylation marks and the latter influencing the addition of lactyl groups. The second pathway: non-enzyme-catalyzed lactylation refers to the spontaneous interaction between lactoylglutathione and lysine residues, mainly regulated by GLO1 and GLO2. The substrate, lactoylglutathione, is derived from a specific metabolic pathway from glycolysis.

Methodology in lactylation measurement and identification

The study of lactylation modifications involves various technical approaches and detection methods to thoroughly elucidate the complex biochemical metabolic pathways and regulatory mechanisms. Mass spectrometry (MS), which chemically cleaves proteins into peptide fragments for characteristic peak analysis, is one of the first high-sensitivity classical methods employed. In 2019, Zhang et al. conducted high-performance liquid chromatography (HPLC)–tandem mass spectrometric (MS/MS) analysis on peptides derived from histones in MCF-7 cells and surprisingly discovered a mass shift identical to that induced by the addition of a lactyl group to lysine residues [3]. The advantage of this method lies not only in its ability to confirm the presence of lysine modifications but also in its capacity to identify specific modification sites.

In addition to MS, the use of anti-lactylation antibodies represents another well-established detection method. Generally, antibodies are categorized into those that target all lactylation modifications, such as Pan-Kla, and those that target specific sites, such as anti-H3K18la. By specifically recognizing lactylation sites and combining this with immunofluorescence staining, it is possible to detect lactylation modifications on target proteins and is available to precise localization within tissues. When investigating the role of lactylation modifications in the tumorigenesis of ocular melanoma, researchers utilized Pan-Kla antibodies and DAPI to perform immunofluorescence staining on ocular melanoma tissues and normal melanocyte tissues. This approach clearly demonstrated an elevated level of lactylation in the ocular melanoma tissues [29].

Proteomics and bioinformatics analyses are often used in conjunction to identify candidate genes and explore the functions of lactylation in different cell types and tissues. By performing enrichment analysis on metabolic pathways and signaling pathways related to lactylation modifications in relevant databases, researchers can infer the effects of this PTM on target proteins. For instance, when Yao et al. aimed to investigate the critical role of lactylation in cerebral ischemia-reperfusion injury (CIRI), they first identified upregulated and downregulated lactylation sites among various proteins through proteomics. Subsequently, they explored the biological pathways involving differentially expressed proteins in the database, ultimately elucidating the potential impacts of lactylation modifications [61].

In the future, with the advancement of artificial intelligence (AI), the accuracy, speed, and scalability of identifying PTMs will continue to improve. Al algorithms, particularly machine learning models, can analyze large datasets to identify patterns and associations that traditional methods may struggle to detect. This capability will facilitate the more efficient discovery of lactylation sites, enhance our understanding of their functional roles in various diseases, and drive the development of new therapeutic strategies.

PROTEIN LACTYLATION IN BRAIN HEALTH AND DISEASES Neurodegenerative diseases

AD is characterized by pathological features including amyloid-\(\beta \) (Aβ) accumulation, tau protein aggregation, neuroinflammation, and age-related neuronal degeneration [62]. Compromised glucose metabolism is a key feature of AD, typically appearing as the second pathological change after AB deposition [63, 64]. This metabolic impairment in neurons directly disrupts OXPHOS, resulting in elevated lactate levels and increased lactylation [65]. Astrocytes, through the ANLS, are pivotal in sustaining the normal physiological functioning of neurons [15]. The dysregulation of glycolysis in these cells has been identified as a significant initiator of neuronal impairments in AD [66]. Additionally, microglial and astrocytic inflammation can damage neurons, induce oxidative stress, and worsen AD progression [67]. Recent studies have established a close link between amyloid protein, which is associated with metabolic reprogramming and the acute inflammation of microglia, and the activation of glycolysis, which in turn directly stimulates microglial activation [68]. A metabolic shift from OXPHOS to glycolysis raises tissue lactate levels, thereby establishing a foundation for lactylation in AD. Lactylation-related genes such as ARID5B, SESN1, and XPA have been identified as pivotal regulators in AD pathology [69]. Recently, lactylation has garnered research interest for its potential link to AD.

Pan and colleagues [70] discovered that in the 5XFAD transgenic mouse model of AD, lactate-mediated H4K12la expression in the hippocampus shows a marked preference for microglia surrounding Aß plaques. This process further leads to lactate accumulation through enrichment in the transcriptional start site of the glycolysisrelated gene pyruvate kinase M2 (PKM2), highlighting the glycolysis/ H4K12la/PKM2 feedback loop's significant role in AD progression. The periodontal pathogen Porphyromonas gingivalis can express small RNAs (msRNAs) similar to microRNAs to modulate host gene expression. Macrophages transfected with msRNA P.G 45033 exhibit enhanced glycolysis levels increased histone Kla levels, and accumulation of AB in periodontal tissue that may cross the bloodbrain barrier and enter the brain [71]. In addition, the reduced activity of a key TCA cycle enzyme, isocitrate dehydrogenase 3β (IDH3β), can cause lactate buildup and histone lactylation at sites like H4K12, H4K8, and H3K18. The transcription factor PAX6, a downstream target of IDH3β, negatively controls IDH3β expression, creating a positive feedback loop with IDH3β, lactate, lactylation, and PAX6 [65].

Glucose metabolism disruptions occur not only in pathology but also with aging, marked by metabolic abnormalities and bioenergetic dysregulation [72]. Thus, aging is closely linked to the onset of AD. Cognitive impairment in AD patients is closely associated with age-related genomic damage and mitochondrial dysfunction [73]. Recent research has extensively explored the link between lactylation and aging, beyond neurodegeneration, in conditions like intervertebral disc degeneration [74] and skin aging [75]. In AD research, Wei et al. have created mouse models like the aging FAD4T, AD-specific APP/PS1, and mature microglial Dox-BV2, showing that H3K18la enhances NFkB signaling by upregulating the expression of Rela (p65) and NFκB1 (p50) [76]. This process ultimately triggers aging-related events, including the secretion of senescence-associated secretory phenotype (SASP) factors like IL-6 and IL-8 by aging cells, which are strongly correlated with histopathological features [77]. Their findings underscore the critical role of the lactate/H3K18la/NFkB/IL-6 and IL-8 pathways in both aging and AD [76]. In addition, pan-Kla has been found to regulate Kmt2a gene transcription in the hippocampal tissue, potentially disrupting synaptic plasticity and affecting cognitive functions in mice, such as spatial exploration and memory, by altering synaptic protein expression [78]. Longterm intervention with lactate and high-intensity interval training (HIIT) in elderly mice showed an increase in the expression of several angiogenesis-associated pathway proteins in the hippocampus, enhancing mitochondrial activity and metabolism-related substances. This further illustrates the beneficial long-term effects of lactate and lactylation on enhancing brain neuroplasticity and slowing neurodegeneration [79]. It is noteworthy that the hippocampus, the brain region most closely associated with AD, exhibits AB accumulation in nearly all AD mouse models. The aforementioned studies further underscore its critical role in susceptibility to aging and AD progression, serving as a key memory controller [80, 81].

Microglia, key regulators of neuroinflammation, have been demonstrated to modulate AD progression through lactylation modifications. Notably, tau protein lactylation at K677 initiates ferroautophagy and ferroptosis through the MAPK pathway, leading to microglial activation and neuroinflammation [82]. Microglial autophagy, mediated by the lysosomal system, effectively clears Aβ. An imbalance in Aβ generation and clearance is a pathogenic factor in AD [83]. EPB41L4A-AS1 is a metabolismrelated long non-coding RNA (lncRNA), that is thought to be closely associated with aging and AD progression due to its deficiency causing ATP synthesis defects [84]. In the EPB41L4A-AS1-specific knockout astrocyte cell line U251, $A\beta_{1-42}$ addition led to significant impairment in $A\beta_{1-42}$ clearance and expression of autophagy-related genes (ARGs). It is hypothesized that EPB41L4A-AS1 modulates the transcription of specific ARGs by regulating GCN5L2 lactylation enzyme activity, thus affecting Aß removal [85].

Promising therapeutic strategies can be provided from multiple perspectives. Considering the upstream and downstream associated proteins, inhibiting PKM2 might reduce abnormal microglial activation and inflammation, thereby improving regional neuronal health and enhancing spatial learning and memory in mice [70]. Similarly, upregulating IDH3B or downregulating PAX6 could prevent phosphorylated tau accumulation, restoring synaptic protein expression and ameliorating pathological symptoms in 5xFAD mice [65]. In addition, the role of physical exercise in brain function recovery and neuronal repair should not be overlooked [47, 79]. It is important to note that the role of lactylation in Alzheimer's disease has not been fully elucidated, meaning this specific modification does not necessarily promote disease progression. Occasionally, lactylation can act as a protective response to neuroinflammation. In this process, exercise-induced lactate accumulation can trigger the "lactate clock," promoting the transition of microglia to a reparative phenotype [47]. Further research is required to investigate the targeted cell types and specific stages of the disease process in which lactylation plays a role in AD (Fig. 2).

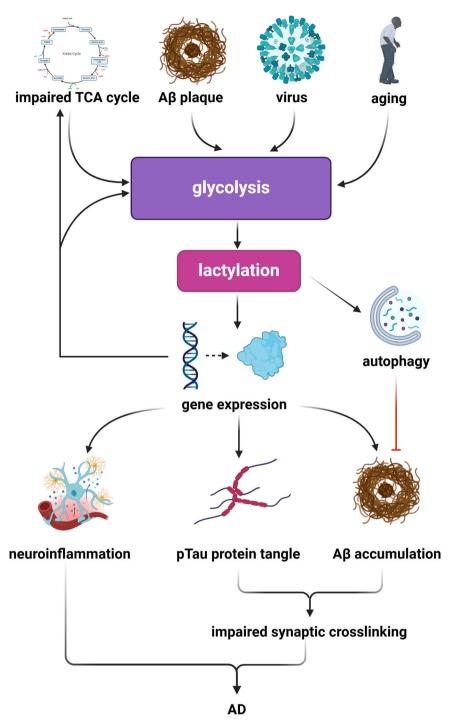


Fig. 2 The regulatory role of lactylation in AD pathology. In AD, the enhancement of glycolytic metabolism is associated with various factors, such as dysregulation of glucose metabolism, accumulation of Aβ plaques, viral infections, and natural aging. Increased lactate production further regulates gene expression through lactylation modification, which can exacerbate neuroinflammation, pTau protein tangles, and Aβ plaque deposition. These pathological changes can impair synaptic connectivity and ultimately accelerate the progression of AD. It is important to note that, in certain instances, lactylation can promote the transcription of autophagy-related genes, playing a beneficial role in the clearance of Aβ plaque accumulation.

In addition to AD, lactate metabolism dysfunction and lactylation modifications are also associated with other neurodegenerative diseases. Parkinson's disease (PD) is primarily characterized by motor symptoms, with the loss of dopaminergic neurons being a key feature [86]. Due to mitochondrial dysfunction and a lack of energy supply, cellular metabolism gradually shifts toward glycolysis, leading to lactate accumulation and further dopaminergic neuron damage [87]. Recent studies have

shown that enhanced glycolytic turnover promotes microglial activation through H3K9 lactylation, which in turn triggers neuroinflammation and accelerates the progression of PD. The use of the glycolysis inhibitor 2-DG has been shown to alleviate the pathological condition in mice by reducing lactate accumulation, suggesting that targeting glucose metabolism may be a potential strategy for treating PD [88]. Multiple sclerosis (MS) is a chronic, immune-related neurodegenerative disease [89].

Increased aerobic glycolysis has been observed in newly diagnosed MS, and the extent of this increase correlates positively with the severity of the disease [90]. Studies have shown that patients with relapsing-remitting MS exhibit elevated lactate levels in their cerebrospinal fluid and monitoring this biomarker may become a powerful tool for early identification and prognosis assessment of the disease [91]. Although no research has yet established a direct link between lactylation modifications and MS, this area holds significant exploratory potential.

Overall, there is a significant and undeniable connection between lactylation modifications and neurodegenerative disorders. However, it is essential to recognize that AD, as a classic neurodegenerative condition, has a complex and multifaceted pathogenesis. A significant limitation of current studies on lactylation is their predominant reliance on mouse models that represent only specific pathological features, which do not fully reflect the actual conditions of patients' brains. A new mouse model has been developed that simultaneously characterizes AB accumulation and tau protein aggregation [92]; however, a significant challenge remains in improving these models to better reflect real-life conditions. Moreover, the study of the localization of lactylation modifications in relation to disease pathology is extremely important. For example, Pan et al. reported elevated levels of Pan-Kla and H4K12la in the cortex and hippocampus of 5XFAD mice, showing significant co-localization with microglia [70]. In contrast, another study found that H3K18la levels were only elevated in the hippocampus [76]. Thus, it is essential to focus on the differences among various tissues. Furthermore, current studies indicate that lactylation can have both positive and inhibitory effects on the pathology of AD. We speculate that this phenomenon may be related to temporal factors and cell types. One possible explanation is that microglia possess specific mechanisms to sense intracellular lactate concentrations. The initial metabolic shifts in microglia tend to trigger neuroinflammation and phagocytic suppression [65, 76]. When abnormalities in the local microenvironment and further increases in lactate levels exceed a certain threshold, microglia may undergo a compensatory shift toward a repair phenotype through lactylation. Another possibility is that the involvement of astrocytes contributes to the recovery of cognitive function during physical activity. The elevated levels of glycolysis in astrocytes can enhance the neuronal energy supply via ANLS, thereby alleviating energy deficits and facilitating functional recovery.

Cerebrovascular disorders

Cerebrovascular disorders (CVD) constitute a spectrum of disorders marked by pathological alterations in the cerebral vasculature. These changes result in functional impairments due to changes in hemodynamic forces, the structural integrity of the vascular walls, and other contributing factors [93]. Stroke, as the most common form of CVD, encompasses ischemic stroke (IS), hemorrhagic stroke, and the less frequently occurring SAH [94]. The IS is the leading killer affecting human health in CVD, characterized by reduced tissue oxygen availability, a shift in metabolic pathways towards glycolysis, and increased lactate concentration [95]. Additionally, elevated serum lactate is also an important marker for intracerebral hemorrhage (ICH) [96]. Beyond the optimal treatment timeframe for IS, the resumption of blood flow and oxygen delivery can precipitate additional tissue damage, a phenomenon termed CIRI. CIRI is closely associated with perturbations in reactive oxygen species (ROS), which stem from inflammatory reactions, excessive calcium levels, and mitochondrial impairment [97]. This injury manifests in various cellular damages, encompassing apoptosis [98], ferroptosis [99] and pyroptosis [100]. Lactate has traditionally been considered a parallel indicator of neuronal dysfunction. However, recent studies have found that moderate supplementation of lactate after ischemic injury and reperfusion is more effective than glucose in improving neurological outcomes [101]. Lactate has also been shown to stimulate angiogenesis and facilitate tissue repair subsequent to ICH [102]. Lactylation, which occurs in response to heightened lactate concentrations, is intricately connected with the pathogenesis and progression of CVD.

In ischemic CVD, there is an inseparable link between damage caused by IS and inflammation in the central nervous system. Within the brain, vascular dysfunction, pathological alterations in the cellular environment, and microcirculatory hypoxia frequently act as instigators of neuroinflammation, often interconnecting in a complex web [103-105]. The neuroinflammatory damage they induce is a key factor in the progression of IS. Timely termination of the inflammatory activation of microglia and astrocytes is a crucial prerequisite for tissue recovery and the improvement of neurological function [106, 107]. As important components of the innate immune system, cyclic GMP-AMP synthase (cGAS) and its downstream signaling protein stimulator of interferon genes (STING) are adept at swiftly detecting foreign danger signals in cells and initiating an inflammatory response [108]. In the brain, they are considered key mediators in regulating the inflammatory activity of microglia during the pathological process of IS [109]. Infants suffering from hypoxic-ischemic encephalopathy (HIE) demonstrate an accelerated rate of glycolytic turnover and a marked increase in cGAS expression. The lactylation modification at the K162 site of cGAS is a critical determinant in the polarization of microglia towards the M1 phenotype [110]. A groundbreaking study has unveiled that lactylation of the Lys131 residue at the N-terminus of cGAS by AARS2 in the wake of viral infection leads to a loss of DNA recognition capacity. This mechanism could underlie the immune suppression associated with elevated tissue lactate levels [111]. Similarly, A1 astrocyte activation, along with lactatemediated lactylation, has been noted to occur in the early stages of IS. Proteins that have undergone lactylation are inclined to engage in the modulation of neuronal activity, which can exacerbate neuronal damage and lead to impaired brain function [46].

Neuroinflammation is a critical event following cerebral ischemia, with extensive inflammatory responses leading to cell damage after reperfusion. In this process, the role of lactylation is not only evident in mediating neuroinflammation but also instrumental in regulating various cellular death pathways. These modifications exert their influence by modulating the protein stability and altering the expression levels of specific mediators. High mobility group box 1 (HMGB1) serves as a key nuclear protein, representing a damage-associated molecular symbol when exists in extracellular environment. Its expression is regulated by various PTMs and has been shown to be closely related to pyroptosis [112]. Yao et al. found that in the CIRI model, increased LDHA-mediated H3K18 lactylation modification promotes the transcription of HMGB1. Specific knockdown of LDHA using SiRNA significantly reduces intracellular HMGB1 levels and prevents pyroptosis [113]. Nuclear receptor coactivator 4 (NCOA4) acts as a crucial receptor mediating the transport of ferritin cargo to the lysosome, making it an essential regulator of ferritinophagy [114]. The stability of the NCOA4 protein can be reinforced by lactylation at the K450 residue, which in turn triggers ferritinophagy and leads to ischemic stroke-related neurological dysfunction [115]. The cytoskeletal protein Lymphocyte cytosolic protein 1 (LCP1), which is integral to cell movement, has been observed to be upregulated in rats subjected to CIRI [116]. Recent studies have further supported this view and additionally identified lactylation modification of LCP1. Knocking down LCP1 or reducing LCP1 lactylation levels through glycolysis inhibition can rescue cell viability and prevent cell apoptosis [117]. Furthermore, a comparative analysis of calcium signaling pathway proteins regulated by Pan-Kla between CIRI and control groups in rats has revealed significant disparities. Notably, lactylation of the solute carrier family 25 members, Slc25a4 and Slc25a5, was exclusive to the CIRI group, while lactylation of voltage-dependent anion channel protein 1 (Vdac1) was detected solely in the control group [61]. These findings advocate for the hypothesis that calcium homeostasis in the brain, particularly under hypoxic conditions, is modulated by lactylation modifications, which in turn, have a substantial impact on the pathological mechanisms of CIRI.

ICH and SAH are another two common types of CVD, differentiated by the location of the bleeding. ICH is a type of hemorrhagic cerebrovascular disease characterized by localized bleeding in the brain due to non-traumatic rupture of blood vessels, closely linked to the process of neuronal ferroptosis [118]. The level of METTL3 lactylation is significantly increased along with its upregulation in the ICH cell model, resulting in reduced cell vitality. Lowering METTL3 decreases the m6A modification level of transferrin receptor (TFRC) and impairs its expression, effectively reducing intracellular iron concentration and preventing cellular ferroptosis [119]. SAH, a form of CVD, has been linked to lactylation modification, in addition to its occurrence in brain parenchyma. During the initial phase of SAH injury, astrocytes exhibit increased levels of lactate and Pan-Kla, coincident with a rise in BRD4 protein levels. In the C8D1A mouse astrocyte cell line. knockdown of BRD4 promotes the generation of the A1 subtype by obstructing lactylation modifications of H4K18. This leads to the release of numerous inflammatory factors and toxic substances into the surrounding microenvironment, threatening neuronal survival and resulting in impaired motor skills and behavioral changes observed in mouse behavioral tests [57].

Cerebral aneurysm (CA), a distinct variety of CVD, is closely associated with inflammatory processes triggered by oxidative stress within the cerebrovascular cellular milieu [120]. Under oxidative stress conditions, such as those induced by H2O2 treatment in vascular endothelial cells (VECs), the expression of LDHA is markedly diminished. Nevertheless, the cytoprotective effect of LDHA overexpression can be counteracted by vascular endothelial growth factor A (VEGFA). When vessels are in an oxidative stress state, the lactylation modification of LDHA induced by local hypoxia represents timely self-regulation and adaptation to the harsh environment. In CA, LDHA and VEGFA closely interact and coordinate with each other, influencing the pathophysiology of VECs under the backdrop of enhanced glycolysis, and providing new insights for management [121].

A variety of therapeutic strategies have been crafted to target lactylation modification pathways, offering promising avenues for treatment. A traditional approach involves the direct inhibition of lactylated proteins to achieve therapeutic effects, such as the inhibition of LCP1 and cGAS [110, 117]. From a metabolic standpoint, directly inhibiting LDHA or employing glycolysis inhibitors can curtail lactate production at its source, thereby mitigating CIRI and aiding in the restoration of cellular function [113, 117]. An alternative strategy involves modulating the activity of lactylation enzymes, such as the downregulation of the reader enzyme BRD4, to suppress lactylation processes. This approach can improve the motor and exploratory capabilities of mice with SAH, thereby enhancing neurological function [57].

Recognized as vital nutritional supporters and glycogen reservoirs for neurons, astrocytes play a pivotal role in maintaining neuronal health [122]. The phenomenon of mitochondrial transfer between neurons and astrocytes, as observed by Ni et al., is seen as one of the potential hopes that we can take advantage of for neuronal survival during ischemic brain injury [122]. Recently, Zhou et al. found that knockdown of low-density lipoprotein receptor-related protein 1 (LRP1) enhances cellular glucose uptake and glycolytic capability by increasing the density of glucose transporter type 1 (GLUT1) on the astrocyte membrane. This knockdown also induces lactylation at the K73 site in ADP-ribosylation factor 1 (ARF1), which affects vesicle assembly and induces mitochondrial loading abnormalities, thereby blocking the release of mitochondria from astrocytes, ultimately

exacerbating the extent of brain infarction in mice. However, the innovative replacement of lysine at the K73 position in ARF1 can inhibit lactylation, foster the transfer of mitochondria from astrocytes to neurons, and effectively reduce the infarct area in mice [123]. It is important to note that, compared to astrocytes, the regulation of the multifunctional transmembrane receptor LRP1 on the surface of neurons has the opposite effect on glucose uptake capacity [124]. Therefore, it is necessary to explore the cellular differences in manipulating specific cell receptors and metabolic pathways.

In the realm of Traditional Chinese Medicine, ischemic CVD is often managed with Buyang Huanwu decoction (BHD), which has demonstrated favorable clinical efficacy [125]. Song et al. further validated the efficacy of BHD in rescuing the infarct area and blood-brain barrier permeability with the use of a rat model. They revealed that BHD diminishes lactylation modifications, including Pan-Kla and H3K18la, by restricting lactate synthesis, which leads to the normalization of apoptotic protease activating factor-1 (Apaf-1) transcription levels and ultimately promotes cellular survival [126].

The localized ischemic and hypoxic environment in brain tissue, along with the release of cytotoxic substances and the accumulation of inflammatory mediators due to tissue damage in CVD, creates favorable conditions for lactate accumulation, thereby supporting the rationale for lactylation research. Current studies involve various cell types, including microglia, astrocytes, and brain endothelial cells. Among these, the complex mechanisms of phenotypic transformation of astrocytes and microglia in CVD represent an important direction for future research. Although Wang et al. found that targeting cGAS and inhibiting lactylation modifications can promote the transition of microglia from the M1 phenotype to a repair phenotype [110], the addition of lactate has no effect on the expression of A2 astrocytes and only weakens M1 polarization process [57]. The complexity of this regulation requires further investigation for clarification. Furthermore, the pathogenic pathways mediated by lactylation in CVD are highly diverse, encompassing various forms such as apoptosis, necroptosis, ferroptosis, and inflammation. Therefore, the interconnections between different mechanisms should be emphasized, as this will enhance our understanding of metabolic changes and tissue damage in CVD (Fig. 3).

Neuroinflammation

The dynamic activities and physiological shifts within microglia and astrocytes play a pivotal role in the initiation and escalation of neuroinflammation. While acute inflammation serves a beneficial function in tissue repair and the clearance of harmful elements following injury, its chronic manifestation is often implicated in the onset of neurodegenerative alterations [127]. The activation of microglia is typically bifurcated into two distinct phenotypes: the M1 phenotype, which is linked to pro-inflammatory activities and neurotoxic responses, and the M2 phenotype, which is engaged in tissue repair and neuroprotective processes. Astrocytes, in a similar vein, are classified into A1 and A2 subtypes, which bear functional resemblances to the M1 and M2 microglial phenotypes, respectively [128]. The transition of cellular metabolism towards glycolysis is closely related to the inflammatory activation of microglia, which can trigger neuroinflammation and neuronal cytotoxicity [129]. Under conditions of oxygen-glucose deprivation (OGD), astrocytes have also been observed to undergo abnormal activation, participating in mediating neuronal dysfunction through the secretion of inflammatory components [130]. Therefore, lactate and lactylation-related subtype transformation of astrocytes and microglia may play a crucial role in the onset and progression of neuroinflammation.

The aberrant inflammatory state triggered by cGAS lactylation-induced M1 polarization of microglia has been discussed above. The transcription factor NF-κB, integral to neuroinflammation, is

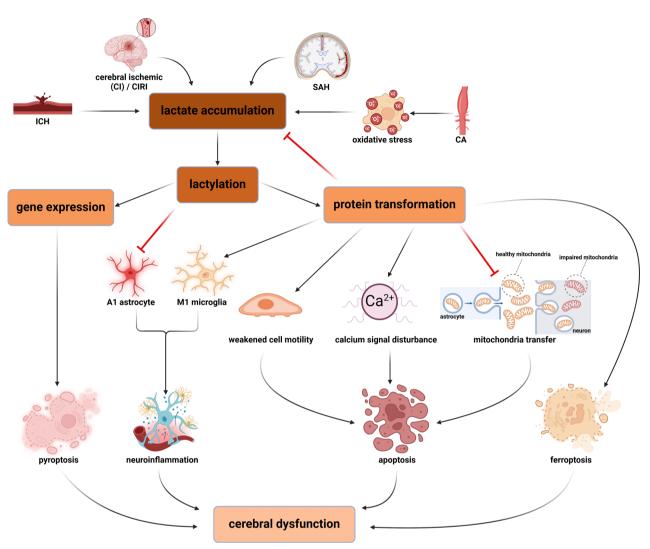


Fig. 3 The regulatory role of lactylation in CVD pathology. The local tissue hypoxic damage caused by CI, ICH, and SAH, along with oxidative stress from CA, plays a critical role in the elevation of lactate levels. Lactate-mediated lactylation induces various cell death pathways and neuroinflammation through the regulation of gene expression and protein stability, ultimately leading to brain dysfunction. First, the initiation of pyroptosis is primarily attributed to the increased expression of the HMGB1 gene. The neuroinflammatory state involves the inflammatory activation of microglia and astrocytes, and lactylation modification can regulate the polarization of these two cell types to control neuroinflammation in the brain. Additionally, the onset of apoptosis is influenced by multiple factors. From the perspective of the cell itself, protein lactylation weakens cell motility and disrupts calcium signaling, ultimately triggering apoptosis. From the perspective of intercellular interactions, lactylation can disrupt mitochondrial transport between astrocytes and neurons, leading to neuronal death. Ferroptosis occurs due to the elevation of intracellular iron levels induced by protein lactylation.

intricately linked with the cGAS signaling pathway [131]. Recent studies have revealed its significant role in lactate-mediated inflammatory activation of microglia. In BV2 microglial cell lines subjected to lipopolysaccharide (LPS)-induced neuroinflammation, both lactylation-mediated inhibition of p53 activity and direct knockdown of p53 protein can disrupt the control of the NF-кВ inflammatory signaling pathway, ultimately leading to abnormal expression of pro-inflammatory factors and M1 polarization of microglia [132]. In high-altitude cerebral edema (HACE) which is characterized by inflammation and hypoxia, proteins in the nucleosome remodeling and histone deacetylase (NuRD) complex of microglial is lactylated, which upregulates the expression of various pro-inflammatory factors in M1-type microglia and enhances NF-kB pathway activity, thereby exacerbating LPS-induced neuroinflammatory conditions [133]. For experimental autoimmune uveitis, abnormal activation of retinal microglia activated by YY1 lactylation also provides strong evidence that the subtype transitions of microglia are closely regulated by this PTM [38].

The inflammatory transitions of astrocytes exhibit complexity in tissue distribution and are regulated in a temporal and spatial dimension [134]. Emerging research on the lactylation of astrocytes suggests that this novel PTM is more inclined to direct cellular processes towards beneficial outcomes, rather than being a driver of inflammation. Given the close collaboration between astrocytes and neurons in energy metabolism [135], it is plausible that lactylation, triggered by the accumulation of lactate, may play a supportive role in bolstering neuronal energy metabolism and be a crucial factor in maintaining the metabolic equilibrium necessary for the proper functioning of neural networks.

What is worth mentioning is that the role of lactate in neuroinflammation mediated by microglia and astrocytes is dual-faceted, not acting merely as an initiator. Various studies have shown that lactylation can play a role in tissue repair and protection, showing potential as a therapeutic target. For example, under conditions of OGD, the local concentration of lactate increases, which triggers astrocyte responses by upregulating

N-Myc downstream-regulated gene 2 (NDRG2) to alleviate the inflammatory state [136]. In the SAH mouse model, BRD4 can impede the generation of A1 subtype astrocytes by inducing Pan-Kla and H4K1la, thereby providing protection for neuronal survival [57]. For microglia, after spinal cord injury (SCI), the concentration of lactate increases, triggering H4K12 lactylation, which is involved in the repair and functional recovery of spinal cord injury [137]. Exercise treatment inhibited the overall expression of microglia in the hippocampus and cortex of the mice while upregulating M2 reparative microglia expression. Exogenous supplementation of lactate resulted in a comprehensive enhancement of the lactate/H3Kla/M2 phenotype conversion axis, indicating that lactylation acts as a protective mechanism, facilitating timely self-adjustment in response to inflammatory environments [47].

Based on existing research, we hypothesize that lactylation modifications have a time-specific impact on the activation state of microglia, similar to the lactate concentration sensing effect observed in AD. In the progression of neuroinflammation-related AD, the subtype transition of microglia exhibits a biphasic pattern [138]. During the acute phase of the disease, the body requires an inflammatory response to eliminate cellular damage, and now lactylation primarily supports the activation of pro-inflammatory microglia, inducing the first peak. Over time, the intrinsic regulatory mechanisms within microglia trigger protective responses that promote the transition to the M2 phenotype. The emergence of the second peak is related to the limitations of the M2 protective mechanisms induced by lactylation and the excessive deterioration of the disease state, as studies have shown that this protective response does not completely counteract the M1-induced inflammatory microenvironmentthis transition begins with a delay and is relatively weak [47].

Brain tumors

Gliomas are prevalent within the central nervous system, with GBM being particularly aggressive. GBM, which typically occurs in adults and the elderly [139], is notorious for its resistance to treatment and its grim prognosis, reflected in a 5-year survival rate of ~10%, and a <1% chance of surviving a decade post-diagnosis [140, 141]. The NF-kB pathway is widely expressed in various cell types within the central nervous system, supporting neuronal function [142]. However, this pathway can become detrimental when overactivated in GBM, promoting tumorigenesis and contributing to resistance to therapeutic interventions [143]. The high metabolic demands of malignant gliomas create a hypoxic tumor microenvironment (TME), triggering a shift in tissue metabolism towards glycolysis, which ultimately leads to elevated lactate levels [144]. To accommodate the tumor's growth, there is an upregulation in the expression of angiogenic factors like vascular endothelial growth factor (VEGF), leading to the abnormal proliferation and expansion of tumor vasculature, a hallmark of high-grade gliomas [145]. Vasculogenic mimicry (VM) represents an innovative paradigm in tumor vascularization, diverging from the conventional angiogenesis that is endothelial cell-dependent. VM constitutes an alternative microcirculatory network, where tumor cells themselves contribute to the formation of blood vessels, playing a crucial role in various malignant diseases [146, 147]. The VEGFR2 present in situ within GBM is of significant importance in both angiogenesis and the construction and dynamic remodeling of VM [148]. In contrast to GBM, neuroblastoma (NB), which originates from the sympathetic nervous system, has a lower malignancy level and primarily affects children [149].

Recently, lactylation modifications have been observed to play an important role in tumor-associated signaling pathways in glioma. Wang et al. identified the lncRNA LINC01127 as a factor that exerts detrimental effects which is linked to lactylation modifications and LPS produced by the gut microbiota in colorectal cancer patients [150]. Subsequent reports have identified LINC01127 in the LN229 GBM cell line and in patient-

derived HG7 cells. In the GBM microenvironment, increased lactate levels associated with the Warburg effect induced lactylation of histone H3, regulating LINC01127 transcriptional activity. LINC01127, in collaboration with Mitogen-activated protein kinase kinase kinase kinase 4 (MAP4K4), activates the JNK pathway to enhance the communication with the NF-κB pathway, culminating in the augmentation of the migratory capabilities and the self-renewal rate of glioma stem cells (GSCs) [151]. GSCs, as a highly specialized population of stem cells, play a crucial role in the progression and recurrence of GBM due to their ability to self-renew and continuously generate new tumor cells [152].

Aberrant activation of tumor signaling pathways leads to uncontrolled tumor proliferation, and the resultant unregulated cell growth induces the formation of new blood vessels locally to meet metabolic demands. Krüppel-like factors (KLFs) are pivotal in governing the biomechanical properties and homeostasis of the vascular wall, with their activity often modulated by a spectrum of PTMs [153]. Recent research has demonstrated that the encoding of P4-135aa peptide by pseudogene MAPK6P4 is responsible for the phosphorylation of the S238 site in KLF15 which targets LDHA to increase its expression. The ensuing lactylation modifications of VE-cadherin and VEGFR2 are closely linked to LDHA levels, which in turn exacerbate the progression of GBM [154].

In the context of high-grade NB, a defining characteristic is the overexpression of the oncogene MYCN [155]. Additionally, heightened levels of Hexokinase-2 (HK2) and LDHA have been observed [156]. Recent studies suggest that elevated HK3 expression in NB cells can intensify lactate metabolism, thereby fostering lactylation modifications that are mediated by intracellular lactate. This metabolic shift subsequently modulates the expression of CXCL14 (C-X-C Motif Chemokine Ligand 14), a protein implicated in the regulation of macrophage homeostasis, contributing to the polarization of M0-type tumor-associated macrophages (TAMs) towards the M2 phenotype, which is associated with enhanced motility and proliferative capacity of NB cells [157].

The pronounced malignancy of gliomas, particularly GBM, poses a significant therapeutic challenge worldwide. A strategic approach to treating GBM involves exploiting the nuances of DNA repair mechanisms to create a situation where the repair capacity is overwhelmed by the rate of damage, thereby enhancing the efficacy of therapies [158]. Recent research has shed light on the connection between homologous recombination (HR) repair and protein lactylation modifications, highlighting their impact on tumor progression and the development of therapeutic resistance [159, 160]. Yue et al. indicated that H3K9la binds to the transcription start site of LUC7L2, enhancing its transcriptional levels. LUC7L2 plays a role in the selective splicing that preserves the termination codon of MLH1, a key component of the mismatch repair (MMR) system. The impairment of MMR is a notable contributor to resistance against the chemotherapeutic agent Temozolomide (TMZ). Stiripentol, an LDH inhibitor, has been shown to significantly boost the effectiveness of TMZ through a mechanism that involves modulating lactylation [161]. Furthermore, Aldehyde dehydrogenase 1A3 (ALDH1A3) was observed to promote the tetramerization of PKM2, which facilitates lactate-induced lactylation of X-ray cross-complementing protein 1 (XRCC1) at the K247 site, directly leading to a diminished response of tumor cells to TMZ and radiation therapy. The small molecule compound D34-919, which targets the ALDH1A3-PKM2 interaction, has demonstrated clinically significant outcomes. This therapeutic strategy has been validated across various platforms, including in vitro studies, mouse models, and GBM organoids derived from patient tumor samples [162].

It is crucial to highlight that the resistance of GBM to treatments is not solely attributed to its enhanced DNA repair mechanisms. TMZ functions by methylating adenine and guanine at specific sites within the DNA sequence, resulting in the formation of three methylated products: N7-MeG, N3-MeA, and O6-meG. MMR system futilely attempts to remove thymine, which pairs optimally with O6-meG, yet fails to address the O6-meG lesion itself, leading to DNA strand breaks [163]. The efficacy of TMZ is intricately linked to the proper functioning of the MMR system, which explains why the downregulation of MLH1 can result in resistance to TMZ [161]. However, studies by Li et al. have demonstrated that heightened DNA repair activity mediated by XRCC1 is associated with resistance to TMZ as well as other radiotherapy and chemotherapy treatments. This association arises because XRCC1 plays a pivotal role in the base excision repair (BER) pathway, rather than in MMR [164]. BER enzymes are primarily targeted at removing N7-MeG and N3-MeA lesions. subsequently repairing the DNA through the collaborative action of DNA polymerase and DNA ligase, thus restoring the DNA strand to its original, non-damaged state [165]. In conclusion, when confronting drug resistance related to DNA repair, it is essential to focus on the specific DNA repair pathways implicated and the mechanisms by which therapeutic agents operate. Additionally, it is crucial to account for the interactions and synergies between various repair pathways, as the resolution of DNA damage in tumor cells often engages multiple mechanisms.

Although advancements have been made in targeting DNA repair mechanisms to treat GBM through chemotherapy, such conventional treatments also involving surgical interventions and radiotherapy have yet to yield consistently satisfactory outcomes [166]. Immunotherapy, which does not require drugs to traverse the blood-brain barrier, has emerged as an exceptionally promising treatment modality in recent years [167]. In GBM, a high proportion of indifferent immune cells and immunosuppressive non-lymphoid cells impair the immune system's ability to combat tumor growth and invasion. The significance of Chimeric Antigen Receptor-T (CAR-T) cell therapy lies in its ability to enhance and equip the patient's own T cells ex vivo to more effectively target tumor-associated antigens, thus rejuvenating weakened immune environments [168]. Sun et al. found that H3K18 lactylation positively regulates the transcription of CD39, CD73, and chemokine (C-C motif) receptor 8 (CCR8) to increase the proportion of Tregs, dampening immune activity. The application of oxamate by inhibiting lactate production can convert tumor-infiltrating lymphocyte and CAR-T cell phenotypes to a more potent state, maximizing tumor-killing effects and improving survival rates in GBM mouse models when combined with CAR-T [169]. TAMs, which include both microglia and monocyte-derived macrophages (MDMs), constitute the predominant immune cell population in GBM [170]. It has been reported that as GBM progresses to later stages of malignancy, MDMs increasingly replace microglia, becoming the dominant cell type and mediating immune suppression associated with the M2 phenotype. This shift is largely attributed to the expression of GLUT1 on the MDM cell surface, as MDMs lacking GLUT1 exhibit significantly diminished immune suppressive capabilities. Notably, lactylation related to GLUT-1 accumulates in the promoter region of the immunosuppressive factor interleukin-10 (IL-10), thereby significantly enhancing IL-10 transcription [48]. Moreover, lactatemediated H3K18 lactylation positively regulates the transcription of tumor necrosis factor superfamily member 9 (TNFSF9), further linking lactate metabolism with M2 polarization events [171].

Developing effective therapeutic strategies that target the immune ecology in gliomas warrants further exploration. In the longitudinal immune landscape, gliomas are characterized in their early stages by proliferative and pro-inflammatory microglia [172]. Investigating the lactylation modifications of microglia during glioma progression should be a focus, as this can help rapidly control disease progression and effectively improve prognosis before deterioration occurs. Up to now, the choice of the specific treatment can be based on a lactylation scoring model, which reflected the intercellular activation and communication of

immune cells within the TME, providing a new approach for assessing the disease status of glioma patients and guiding treatment strategies [173]. Furthermore, non-coding RNAs have been shown to be involved in the progression of gliomas [174]. Among these, both lncRNAs and pseudogenes have been found to be closely associated with lactylation in gliomas, while the unexplored domains of microRNAs and circRNAs are promising and intriguing (Fig. 4).

Neurodevelopmental diseases

The dynamics of energy metabolism and histone modification are pivotal during neurodevelopment. Maturation from neural stem cells (NSCs) to fully differentiated neurons is characterized by a metabolic shift from predominant glycolysis to OXPHOS, which is instrumental in guiding the developmental trajectory [175]. During the posterior embryonic axial development in mice and chicks, a glycolytic gradient is observed, which modulates the expression of various signaling pathways, thereby regulating developmental processes [176]. This spatiotemporal heterogeneity in glycolysis indicates that fluctuations in lactate levels during the early stages of neurodevelopment may exert significant regulatory effects. Furthermore, neural crest cells (NCCs), which are poised to undergo epithelial-mesenchymal transition, also demonstrate heightened glycolytic activity and lactate accumulation. These metabolic changes are crucial for NCCs' migration and the determination of their cellular identities [177]. Recently, the impact of lactatemediated lactylation in this process has also been unveiled.

In terms of the overall regulation of neurodevelopment, Dai et al. demonstrated that the levels of H3K18 lactylation gradually decrease in P19 embryonic carcinoma (EC) cells as neurogenesis and differentiation progress. Despite H3K18la being predominantly found in transcriptionally inert regions like introns, the optimal functioning of transcriptional start sites necessitates a complex interplay of various PTMs, among which lactylation is a key player [54].

Within the nascent phases of neurodevelopment, NCCs ramp up their production of lactate through LDHA/B, steering their metabolism toward glycolysis. This metabolic shift promotes lactylation of certain nuclear genes related to cell adhesion and migration, such as SOX10, thereby enhancing the migratory capacity of NCCs [178]. The migration of NCCs plays an indispensable role in the development of midfacial structures, and the precise control of tissue growth and fusion is paramount to prevent significant developmental anomalies [179]. In an innovative approach, a mouse model was engineered to express bone morphogenetic protein (BMP) receptors ACVR1 constitutively in the neural crest region, thereby enhancing BMP/Smad signaling. The intensification of the BMP pathway was observed to downregulate the expression of pivotal glycolytic genes in a p53dependent fashion, consequently inhibiting Pan-Kla and H3K18la. As a result, the expression of epigenetic downstream targets, notably Pdgfra—a gene essential for facial development—was impeded, leading to severe defects in midline facial development in the majority of the mice [180].

As neurodevelopment progresses, the intricate regulatory network of histone PTMs becomes increasingly vital in sculpting neuronal formation. Genes that exhibit increased enrichment of H3K9cr and H3K18la are predominantly involved in neuronal differentiation and maturation. These modifications, in conjunction with H3K9ac, orchestrate the intricate processes governing neuronal cell fate [54]. Beyond the realm of neurons, the development and functionality of microglia and astrocytes are also influenced by the metabolic landscape, which is associated with defects in the glycolytic pathway mediated by the loss of BTB and CNC homology 1 (Bach1) [181]. The role of Bach1 in glycolytic enzymes and lactate metabolism has been well documented [182]. Bach1 induces the accumulation of lactate-mediated H4K12 lactylation at the promoter region of LRRC15, targeting CD248/GP130/JAK-STAT3 axis to influence the development of astrocytes [181].

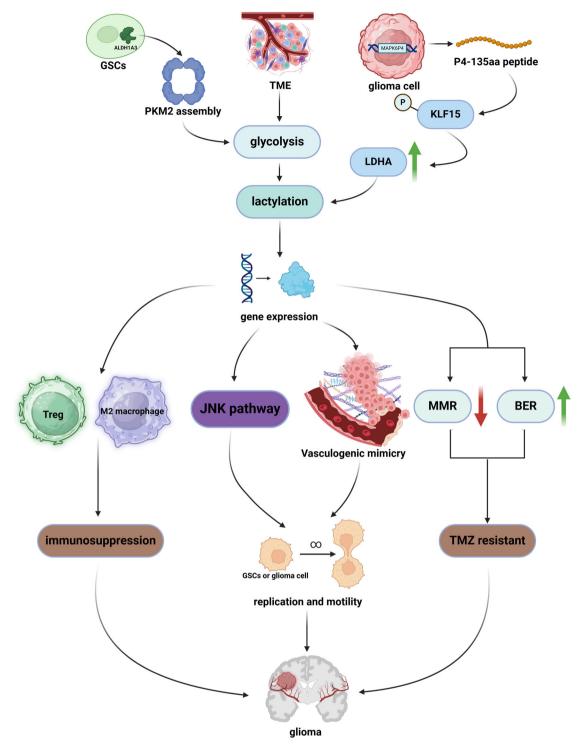


Fig. 4 The regulatory role of lactylation in glioma pathology. The metabolic characteristics of the glioma microenvironment, along with changes in the activity of metabolic enzymes induced by abnormal gene expression, are key factors contributing to the increased lactylation modification in gliomas. Lactylation-mediated regulation of gene expression promotes glioma progression through various pathways. Firstly, enhanced interconnection of JNK and NF-κB pathways promotes the tumor cells' inherent proliferative capacity, while abnormal vascular proliferation provides the necessary external conditions for the unlimited growth of tumor cells. Together, these factors synergistically enhance the cell's motility and invasiveness. Additionally, lactate accumulation in advanced gliomas leads to an increase in the proportion of Treg cells and M2 macrophages in the tumor microenvironment, which mediates immune suppression and hinders the effectiveness of immunotherapy. Lactylation modification also contributes to tumor cell resistance to radiotherapy and chemotherapy by regulating DNA repair processes.

Mental disorders

Anxiety is acknowledged as the most widespread condition among mental disorders, often occurring alongside depression [183]. Schizophrenia (SCZ), another prevalent mental disorder, has

an incidence rate of \sim 0.5% in China [184]. The hallmark of epilepsy is the imbalance in brain energy metabolism. During seizures, the excessive neuronal excitability is closely linked to the energy support provided by astrocytes to neurons, known as the ANLS

[185]. Research has established a strong correlation between lactate levels and both the precision of synaptic networks and neuronal excitability [186]. Enhanced glycolysis or alterations in brain energy metabolism, marked by lactate accumulation, have been detected in major depressive disorder [187], anxiety disorders [188], SCZ [189], and epilepsy [190]. Recently, chronic stress and increased neuronal excitability have been identified as pivotal factors that stimulate lactylation in various neural cells of the brain, especially in the medial prefrontal cortex (mPFC), a region considered to be closely related to anxiety and depression-like behaviors in mice [191]. This represents the first comprehensive systemic analysis of lactylation modifications in the brain, providing a solid foundation for further exploration into the role of lactylation in the spectrum of psychiatric disorders.

Recent studies have uncovered that regular physical activity can modulate lactate levels in the mPFC of mice, concurrently augmenting lactylation at the K885 residue of synaptosomeassociated protein 91 (SNAP91). This specific lactylation event is crucial for maintaining the activity of neuronal networks and for stabilizing synaptic structures, highlighting its essential role as a mediator in the anxiolytic effects of exercise [192]. These findings, in conjunction with prior studies [193], corroborate the mPFC's centrality in the development of anxiety and depression, underscoring the necessity for a deeper exploration into the regulatory dynamics of lactylation modifications within this region. Moreover, in mouse and cellular models of SCZ constructed by MK801, an accelerated glycolytic metabolism accompanied by high lactate accumulation has been observed, resulting in increased Pan-Kla, H3K9la, and H3K18la levels. Impairments in neuronal survival mediated by HMGB1 are believed to be closely associated with the glycolytic pathway, as evidenced by the mitigation of these adverse effects through treatment with the glycolysis inhibitor 2-DG [194]. Furthermore, 2-DG-mediated inhibition of glycolysis effectively reverses various electrophysiological alterations induced by epilepsy, suggesting that targeting lactate metabolism pathways could be a promising approach for the development of novel antiepileptic drugs [195] (Supplemental Table 2).

FUTURE PERSPECTIVES

Lactate, a key participant in anaerobic metabolism, has increasingly become the focus of research, particularly regarding its bioavailability and production in various pathological and physiological states. The phenomenon of lactylation represents a burgeoning field, indicating a more advanced role of lactate in metabolic pathways associated with disease. Based on existing research, the accumulation of lactate triggered by various biological processes is an important prerequisite for the occurrence of lactylation modifications. However, it remains unclear whether lactylation is merely a natural consequence of elevated lactate levels or if it requires specific regulatory factors for precise modulation. Future research needs to continue elucidating the specific mechanisms involved, as this is crucial for guiding us toward a deeper understanding and the development of more effective treatment strategies.

It is noteworthy that the role of LDH in lactylation should be reevaluated. Immunoprecipitation has revealed that LDHA can directly bind to proteins and participate in lactylation modifications.. Silencing LDHA results in a significant reduction in the lactylation levels of its target proteins, with no observed effect on LDHA's role in metabolism. This suggests that LDHA may not function as a metabolic enzyme within the conventional context of glycolytic pathways, but rather plays a direct regulatory role in the addition of lactylation modifications [154]. Additionally, despite the strong correlation between LDH activity and adverse prognosis as well as malignancy in NB cells, sometimes the downregulation of LDHA and LDHB does not impact cellular glycolytic activity or lactate concentration [196]. This observation suggests a potentially different mechanism underlying the

suppressive effects of LDH inhibition in NB, it may extend beyond its recognized metabolic functions in tumorigenesis.

Here, we offer a review of the latest research developments concerning lactylation in the context of brain diseases. Overall, research into the regulatory role of lactylation in brain dysfunction remains largely confined to pathogenic pathways, with limited exploration of drug targets and therapeutic prospects. Although some studies have found that targeting lactylation-related regulatory axes can improve the behavioral activities or physiological states of mice, these are few in number and require broader research support. Notably, the efficacy of the small molecule compound D34-919 has been validated in organoid models of gliomas [162]. Organoids, due to their capacity to replicate the original tumor characteristics and immune microenvironment, are recognized as powerful tools for studying cancer immunotherapy [197]. In future investigations into the regulation of lactylation in brain tumors, incorporating organoid models should be considered.

Furthermore, given that lactylation regulatory enzymes often exhibit ACT properties, the therapeutic prospects for targeting these enzymes remain concerning. Research has already revealed a competitive relationship between acetylation and lactylation modifications at the H3K18 site, and inhibition of erasers has not yielded the anticipated restoration of lactylation levels [50]. Cui and colleagues have also expressed similar concerns, positing that p300 is not only one of the most crucial ACTs but also possesses a multitude of other cellular functions [28]. The competition between different protein acylations, particularly lactylation and acetylation, is a key consideration when exploring treatment strategies for CNS diseases. Although this competitive phenomenon has not yet been observed in the brain, it may be a significant reason for the poor efficacy and side effects of many therapeutic approaches, warranting further investigation. Specifically targeting lactylation without affecting other acylations is crucial for achieving therapeutic effects and avoiding unintended side effects. A more comprehensive understanding of the relationship between lactylation and other PTMs, as well as the regulatory strategies for protein lactylation, will aid in developing new therapeutic ideas and expanding the treatment potential of lactylation modifications in CNS diseases.

CONCLUSION

In conclusion, this review demonstrates that lactate and lactylation modifications, due to their involvement in critical brain metabolic pathways and their regulation of various gene transcription and protein functions, play a crucial regulatory role in the context of brain diseases. Writers, erasers, and readers contribute to maintaining the dynamic balance of enzyme-catalyzed lactylation modifications, which holds significant implications for the progression of brain dysfunction.

REFERENCES

- 1. Bonvento G, Bolanos JP. Astrocyte-neuron metabolic cooperation shapes brain activity. Cell Metab. 2021;33:1546–64.
- Ferguson BS, Rogatzki MJ, Goodwin ML, Kane DA, Rightmire Z, Gladden LB. Lactate metabolism: historical context, prior misinterpretations, and current understanding. Eur J Appl Physiol. 2018;118:691–728.
- 3. Zhang D, Tang Z, Huang H, Zhou G, Cui C, Weng Y, et al. Metabolic regulation of gene expression by histone lactylation. Nature. 2019;574:575–80.
- 4. Li X, Yang Y, Zhang B, Lin X, Fu X, An Y, et al. Lactate metabolism in human health and disease. Signal Transduct Target Ther. 2022;7:305.
- Liu X, Zhou Y, Wang H. The role of lactate-induced protein lactylation in gliomas: implications for preclinical research and the development of new treatments. Front Pharm. 2024;15:1383274.
- 6. Zhao A, Xu W, Han R, Wei J, Yu Q, Wang M, et al. Role of histone modifications in neurogenesis and neurodegenerative disease development. Ageing Res Rev. 2024-08:102324
- 7. Yang C, Pan RY, Guan F, Yuan Z. Lactate metabolism in neurodegenerative diseases. Neural Regen Res. 2024;19:69–74.

- Li R, Yang Y, Wang H, Zhang T, Duan F, Wu K, et al. Lactate and lactylation in the brain: current progress and perspectives. Cell Mol Neurobiol. 2023;43:2541–55.
- van Hall G. Lactate kinetics in human tissues at rest and during exercise. Acta Physiol (Oxf). 2010;199:499–508.
- Warburg O, Wind F, Negelein E. The metabolism of tumors in the body. J Gen Physiol. 1927:8:519–30.
- 11. Liberti MV, Locasale JW. The Warburg effect: how does it benefit cancer cells? Trends Biochem Sci. 2016;41:211–8.
- Vander HM, Cantley LC, Thompson CB. Understanding the Warburg effect: the metabolic requirements of cell proliferation. Science. 2009;324:1029–33.
- Zhu W, Guo S, Sun J, Zhao Y, Liu C. Lactate and lactylation in cardiovascular diseases: current progress and future perspectives. Metabolism. 2024;158:155957.
- Yao S, Chai H, Tao T, Zhang L, Yang X, Li X, et al. Role of lactate and lactate metabolism in liver diseases (review). Int J Mol Med. 2024;54:59.
- Pellerin L, Magistretti PJ. Glutamate uptake into astrocytes stimulates aerobic glycolysis: a mechanism coupling neuronal activity to glucose utilization. Proc Natl Acad Sci USA. 1994;91:10625–9.
- Magistretti PJ, Allaman I. Lactate in the brain: from metabolic end-product to signalling molecule. Nat Rev Neurosci. 2018;19:235–49.
- Certo M, Tsai CH, Pucino V, Ho PC, Mauro C. Lactate modulation of immune responses in inflammatory versus tumour microenvironments. Nat Rev Immunol. 2021;21:151–61.
- 18. Zhang D, Gao J, Zhu Z, Mao Q, Xu Z, Singh PK, et al. Lysine I-lactylation is the dominant lactylation isomer induced by glycolysis. Nat Chem Biol. 2025;21:91–9.
- Xu K, Zhang K, Wang Y, Gu Y. Comprehensive review of histone lactylation: structure, function, and therapeutic targets. Biochem Pharm. 2024;225:116331.
- Li X, Chen M, Chen X, He X, Li X, Wei H, et al. TRAP1 drives smooth muscle cell senescence and promotes atherosclerosis via HDAC3-primed histone H4 lysine 12 lactylation. Eur Heart J. 2024;45:4219–35.
- Yang Z, Yan C, Ma J, Peng P, Ren X, Cai S, et al. Lactylome analysis suggests lactylation-dependent mechanisms of metabolic adaptation in hepatocellular carcinoma. Nat Metab. 2023;5:61–79.
- Sun Y, Chen Y, Peng T. A bioorthogonal chemical reporter for the detection and identification of protein lactylation. Chem Sci. 2022;13:6019–27.
- Liu R, Ren X, Park YE, Feng H, Sheng X, Song X, et al. Nuclear GTPSCS functions as a lactyl-CoA synthetase to promote histone lactylation and gliomagenesis. Cell Metab. 2025;37:377–94.e9.
- Zhu R, Ye X, Lu X, Xiao L, Yuan M, Zhao H, et al. ACSS2 acts as a lactyl-CoA synthetase and couples KAT2A to function as a lactyltransferase for histone lactylation and tumor immune evasion. Cell Metab. 2025;37:361–76.e7.
- Ju J, Zhang H, Lin M, Yan Z, An L, Cao Z, et al. The alanyl-tRNA synthetase AARS1 moonlights as a lactyltransferase to promote yap signaling in gastric cancer. J Clin Investig. 2024;134:e174587.
- Zong Z, Xie F, Wang S, Wu X, Zhang Z, Yang B, et al. Alanyl-tRNA synthetase, AARS1, is a lactate sensor and lactyltransferase that lactylates p53 and contributes to tumorigenesis. Cell. 2024;187:2375–92.
- Zhao S, Zhang X, Li H. Beyond histone acetylation-writing and erasing histone acylations. Curr Opin Struct Biol. 2018;53:169–77.
- Cui H, Xie N, Banerjee S, Ge J, Jiang D, Dey T, et al. Lung myofibroblasts promote macrophage profibrotic activity through lactate-induced histone lactylation. Am J Respir Cell Mol Biol. 2021;64:115–25.
- Yu J, Chai P, Xie M, Ge S, Ruan J, Fan X, et al. Histone lactylation drives oncogenesis by facilitating m(6)a reader protein YTHDF2 expression in ocular melanoma. Genome Biol. 2021;22:85.
- Wang N, Wang W, Wang X, Mang G, Chen J, Yan X, et al. Histone lactylation boosts reparative gene activation post-myocardial infarction. Circ Res. 2022;131:893–908.
- Chen B, Deng Y, Hong Y, Fan L, Zhai X, Hu H, et al. Metabolic recoding of NSUN2-mediated m(5)c modification promotes the progression of colorectal cancer via the NSUN2/YBX1/m(5)c-ENO1 positive feedback loop. Adv Sci (Weinh). 2024;11:e2309840.
- 32. Hu X, Huang X, Yang Y, Sun Y, Zhao Y, Zhang Z, et al. Dux activates metabolism-lactylation-met network during early IPSC reprogramming with BRG1 as the histone lactylation reader. Nucleic Acids Res. 2024;52:5529–48.
- Li F, Si W, Xia L, Yin D, Wei T, Tao M, et al. Positive feedback regulation between glycolysis and histone lactylation drives oncogenesis in pancreatic ductal adenocarcinoma. Mol Cancer. 2024;23:90.
- Dai W, Wu G, Liu K, Chen Q, Tao J, Liu H, et al. Lactate promotes myogenesis via activating H3K9 lactylation-dependent up-regulation of Neu2 expression. J Cachexia Sarcopenia Muscle. 2023;14:2851–65.
- 35. Zhang Y, Zhang X. Virus-induced histone lactylation promotes virus infection in crustacean. Adv Sci (Weinh). 2024;11:e2401017.
- Xu GE, Yu P, Hu Y, Wan W, Shen K, Cui X, et al. Exercise training decreases lactylation and prevents myocardial ischemia-reperfusion injury by inhibiting YTHDF2. Basic Res Cardiol. 2024;119:651–71.

- Wang X, Fan W, Li N, Ma Y, Yao M, Wang G, et al. YY1 lactylation in microglia promotes angiogenesis through transcription activation-mediated upregulation of FGF2. Genome Biol. 2023;24:87.
- Huang J, Wang X, Li N, Fan W, Li X, Zhou Q, et al. YY1 lactylation aggravates autoimmune uveitis by enhancing microglial functions via inflammatory genes. Adv Sci (Weinh). 2024:11:e2308031.
- Dong M, Zhang Y, Chen M, Tan Y, Min J, He X, et al. Asf1a-dependent p300mediated Histone H3 lysine 18 lactylation promotes atherosclerosis by regulating endmt. Acta Pharm Sin B. 2024;14:3027–48.
- Wu D, Spencer CB, Ortoga L, Zhang H, Miao C. Histone lactylation-regulated METTL3 promotes ferroptosis via m6a-modification on ACSL4 in sepsisassociated lung injury. Redox Biol. 2024;74:103194.
- Zhang N, Zhang Y, Xu J, Wang P, Wu B, Lu S, et al. Alpha-myosin heavy chain lactylation maintains sarcomeric structure and function and alleviates the development of heart failure. Cell Res. 2023;33:679–98.
- Huang H, Wang S, Xia H, Zhao X, Chen K, Jin G, et al. Lactate enhances NMNAT1 lactylation to sustain nuclear NAD(+) salvage pathway and promote survival of pancreatic adenocarcinoma cells under glucose-deprived conditions. Cancer Lett. 2024:588:216806.
- Yang L, Niu K, Wang J, Shen W, Jiang R, Liu L, et al. Nucleolin lactylation contributes to intrahepatic cholangiocarcinoma pathogenesis via RNA splicing regulation of madd. J Hepatol. 2024;81:651–66.
- Wang YH, Gao P, Wang YQ, Xu LZ, Zeng KW, Tu PF. Small-molecule targeting PKM2 provides a molecular basis of lactylation-dependent fibroblast-like synoviocytes proliferation inhibition against rheumatoid arthritis. Eur J Pharm. 2024:972:176551.
- 45. Yang K, Fan M, Wang X, Xu J, Wang Y, Tu F, et al. Lactate promotes macrophage HMGB1 lactylation, acetylation, and exosomal release in polymicrobial sepsis. Cell Death Differ. 2022;29:133–46.
- Xiong XY, Pan XR, Luo XX, Wang YF, Zhang XX, Yang SH, et al. Astrocyte-derived lactate aggravates brain injury of ischemic stroke in mice by promoting the formation of protein lactvlation. Theranostics. 2024:14:4297–317.
- 47. Han H, Zhao Y, Du J, Wang S, Yang X, Li W, et al. Exercise improves cognitive dysfunction and neuroinflammation in mice through Histone H3 lactylation in microglia. Immun Ageing. 2023;20:63.
- De Leo A, Ugolini A, Yu X, Scirocchi F, Scocozza D, Peixoto B, et al. Glucosedriven histone lactylation promotes the immunosuppressive activity of monocyte-derived macrophages in glioblastoma. Immunity. 2024;57:1105–23.
- 49. Hammond CM, Stromme CB, Huang H, Patel DJ, Groth A. Histone chaperone networks shaping chromatin function. Nat Rev Mol Cell Biol. 2017;18:141–58.
- Rho H, Terry AR, Chronis C, Hay N. Hexokinase 2-mediated gene expression via histone lactylation is required for hepatic stellate cell activation and liver fibrosis. Cell Metab. 2023;35:1406–23.
- 51. Moreno-Yruela C, Baek M, Monda F, Olsen CA. Chiral posttranslational modification to lysine epsilon-amino groups. Acc Chem Res. 2022;55:1456–66.
- Sun Y, Chen Y, Xu Y, Zhang Y, Lu M, Li M, et al. Genetic encoding of epsilon-n-lactyllysine for detecting delactylase activity in living cells. Chem Commun (Camb). 2022;58:8544–7.
- Moreno-Yruela C, Zhang D, Wei W, Baek M, Liu W, Gao J, et al. Class I histone deacetylases (HDAC1-3) are histone lysine delactylases. Sci Adv. 2022:8:eabi6696.
- Dai SK, Liu PP, Li X, Jiao LF, Teng ZQ, Liu CM. Dynamic profiling and functional interpretation of histone lysine crotonylation and lactylation during neural development. Development. 2022;149:dev200049.
- Zu H, Li C, Dai C, Pan Y, Ding C, Sun H, et al. SIRT2 functions as a histone delactylase and inhibits the proliferation and migration of neuroblastoma cells. Cell Discov. 2022;8:54.
- Zhai G, Niu Z, Jiang Z, Zhao F, Wang S, Chen C, et al. DPF2 reads histone lactylation to drive transcription and tumorigenesis. Proc Natl Acad Sci USA. 2024;121:e1873471175.
- Zhang F, Zhou J, Lu P, Zhang X, Yang L, Wu J, et al. Lactylation of histone by brd4 regulates astrocyte polarization after experimental subarachnoid hemorrhage. J Neuroinflamm. 2024;21:186.
- Mochizuki K, Ishiyama S, Hariya N, Goda T. Regulation of carbohydrateresponsive metabolic genes by histone acetylation and the acetylated histone reader BRD4 in the gene body region. Front Mol Biosci. 2021;8:682696.
- 59. Qian H, Zhu M, Tan X, Zhang Y, Liu X, Yang L. Super-enhancers and the super-enhancer reader BRD4: tumorigenic factors and therapeutic targets. Cell Death Discov. 2023;9:470.
- Gaffney DO, Jennings EQ, Anderson CC, Marentette JO, Shi T, Schou OA, et al. Non-enzymatic lysine lactoylation of glycolytic enzymes. Cell Chem Biol. 2020;27:206–13.
- Yao Y, Bade R, Li G, Zhang A, Zhao H, Fan L, et al. Global-scale profiling of differential expressed lysine-lactylated proteins in the cerebral endothelium of cerebral ischemia-reperfusion injury rats. Cell Mol Neurobiol. 2023;43:1989–2004.

- Scheltens P, De Strooper B, Kivipelto M, Holstege H, Chetelat G, Teunissen CE, et al. Alzheimer's disease. Lancet. 2021;397:1577–90.
- 63. Butterfield DA, Halliwell B. Oxidative stress, dysfunctional glucose metabolism and Alzheimer disease. Nat Rev Neurosci. 2019;20:148–60.
- Chen Z, Zhong C. Decoding Alzheimer's disease from perturbed cerebral glucose metabolism: implications for diagnostic and therapeutic strategies. Prog Neurobiol. 2013;108:21–43.
- 65. Wang X, Liu Q, Yu HT, Xie JZ, Zhao JN, Fang ZT, et al. A positive feedback inhibition of isocitrate dehydrogenase 3beta on paired-box gene 6 promotes Alzheimer-like pathology. Signal Transduct Target Ther. 2024;9:105.
- Minhas PS, Jones JR, Latif-Hernandez A, Sugiura Y, Durairaj AS, Wang Q, et al. Restoring hippocampal glucose metabolism rescues cognition across Alzheimer's disease pathologies. Science. 2024;385:eabm6131.
- 67. Twarowski B, Herbet M. Inflammatory processes in Alzheimer's disease-pathomechanism, diagnosis and treatment: a review. Int J Mol Sci. 2023;24:6518.
- Baik SH, Kang S, Lee W, Choi H, Chung S, Kim JI, et al. A breakdown in metabolic reprogramming causes microglia dysfunction in Alzheimer's disease. Cell Metab. 2019:30:493–507.
- Guo S, Zhu W, Bian Y, Li Z, Zheng H, Li W, et al. Developing diagnostic biomarkers for Alzheimer's disease based on histone lactylation-related gene. Heliyon. 2024;10:e37807.
- Pan RY, He L, Zhang J, Liu X, Liao Y, Gao J, et al. Positive feedback regulation of microglial glucose metabolism by histone h4 lysine 12 lactylation in Alzheimer's disease. Cell Metab. 2022;34:634–48.
- 71. Zhang Y, Sun Y, Hu Y, Zheng S, Shao H, Lin L, et al. Porphyromonas gingivalis msRNA p.g_45033 induces amyloid-beta production by enhancing glycolysis and histone lactylation in macrophages. Int Immunopharmacol. 2023;121:110468.
- Yang S, Park JH, Lu HC. Axonal energy metabolism, and the effects in aging and neurodegenerative diseases. Mol Neurodegener. 2023;18:49.
- 73. Hou Y, Dan X, Babbar M, Wei Y, Hasselbalch SG, Croteau DL, et al. Ageing as a risk factor for neurodegenerative disease. Nat Rev Neurol. 2019;15:565–81.
- Zhang Y, Huang Z, Han W, Wu J, Li S, Qin T, et al. Glutamine suppresses senescence and promotes autophagy through glycolysis inhibition-mediated AMPKalpha lactylation in intervertebral disc degeneration. Commun Biol. 2024;7:325.
- Zou Y, Cao M, Tao L, Wu S, Zhou H, Zhang Y, et al. Lactate triggers KAT8mediated LTBP1 lactylation at lysine 752 to promote skin rejuvenation by inducing collagen synthesis in fibroblasts. Int J Biol Macromol. 2024;277:134482.
- 76. Wei L, Yang X, Wang J, Wang Z, Wang Q, Ding Y, et al. H3K18 lactylation of senescent microglia potentiates brain aging and Alzheimer's disease through the NFkappaB signaling pathway. J Neuroinflamm. 2023;20:208.
- 77. Han X, Zhang T, Liu H, Mi Y, Gou X. Astrocyte senescence and Alzheimer's disease: a review. Front Aging Neurosci. 2020;12:148.
- 78. Wu Y, Hu H, Liu W, Zhao Y, Xie F, Sun Z, et al. Hippocampal lactate-infusion enhances spatial memory correlated with monocarboxylate transporter 2 and lactylation. Brain Sci. 2024;14:327.
- Lei Z, Mozaffaritabar S, Kawamura T, Koike A, Kolonics A, Keringer J, et al. The effects of long-term lactate and high-intensity interval training (HIIT) on brain neuroplasticity of aged mice. Heliyon. 2024;10:e24421.
- Belfiore R, Rodin A, Ferreira E, Velazquez R, Branca C, Caccamo A, et al. Temporal and regional progression of Alzheimer's disease-like pathology in 3xTg-AD mice. Aging Cell. 2019;18:e12873.
- 81. Jaroudi W, Garami J, Garrido S, Hornberger M, Keri S, Moustafa AA. Factors underlying cognitive decline in old age and Alzheimer's disease: the role of the hippocampus. Rev Neurosci. 2017;28:705–14.
- 82. An X, He J, Xie P, Li C, Xia M, Guo D, et al. The effect of tau K677 lactylation on ferritinophagy and ferroptosis in Alzheimer's disease. Free Radic Biol Med. 2024;224:685–706.
- Cho MH, Cho K, Kang HJ, Jeon EY, Kim HS, Kwon HJ, et al. Autophagy in microglia degrades extracellular beta-amyloid fibrils and regulates the NLRP3 inflammasome. Autophagy. 2014;10:1761–75.
- 84. Yang T, Wang Y, Liao W, Zhang S, Wang S, Xu N, et al. Down-regulation of EPB41L4A-AS1 mediated the brain aging and neurodegenerative diseases via damaging synthesis of NAD(+) and ATP. Cell Biosci. 2021;11:192.
- Wang Z, Wang R, Niu L, Zhou X, Han J, Li K. EPB41L4A-AS1 is required to maintain basal autophagy to modulates abeta clearance. NPJ Aging. 2024;10:24.
- 86. Kalia LV, Lang AE. Parkinson's disease. Lancet. 2015;386:896–912.
- 87. Li J, Chen L, Qin Q, Wang D, Zhao J, Gao H, et al. Upregulated Hexokinase 2 expression induces the apoptosis of dopaminergic neurons by promoting lactate production in Parkinson's disease. Neurobiol Dis. 2022;163:105605.
- Qin Q, Wang D, Qu Y, Li J, An K, Mao Z, et al. Enhanced glycolysis-derived lactate promotes microglial activation in Parkinson's disease via histone lactylation. NPJ Parkinsons Dis. 2025;11:3.
- 89. Dobson R, Giovannoni G. Multiple sclerosis—a review. Eur J Neurol. 2019;26:27-40.

- Brier MR, Judge B, Ying C, Salter A, An H, Patel A, et al. Increased white matter aerobic glycolysis in multiple sclerosis. Ann Neurol. 2025;97:766–78.
- Albanese M, Zagaglia S, Landi D, Boffa L, Nicoletti CG, Marciani MG, et al. Cerebrospinal fluid lactate is associated with multiple sclerosis disease progression. J Neuroinflamm. 2016:13:36.
- Zampar S, Wirths O. Characterization of a mouse model of Alzheimer's disease expressing Aβ4-42 and human mutant tau. Int J Mol Sci. 2021;22:5191.
- Yan F, Yan S, Wang J, Cui Y, Chen F, Fang F, et al. Association between triglyceride glucose index and risk of cerebrovascular disease: systematic review and meta-analysis. Cardiovasc Diabetol. 2022;21:226.
- 94. Delapaz RL. Cerebrovascular disease. AJNR Am J Neuroradiol. 2007;28:1197–9.
- Shin TH, Lee DY, Basith S, Manavalan B, Paik MJ, Rybinnik I, et al. Metabolome changes in cerebral ischemia. Cells. 2020;9:1630.
- Bender M, Haferkorn K, Nagl J, Uhl E, Stein M. Serum lactate as serum biomarker for cardiopulmonary parameters within the first 24 h after a spontaneous intracerebral hemorrhage. Diagnostics (Basel). 2022;12:2414.
- Shen L, Gan Q, Yang Y, Reis C, Zhang Z, Xu S, et al. Mitophagy in cerebral ischemia and ischemia/reperfusion injury. Front Aging Neurosci. 2021:13:687246.
- Gong L, Tang Y, An R, Lin M, Chen L, Du J. Rtn1-c mediates cerebral ischemia/ reperfusion injury via er stress and mitochondria-associated apoptosis pathways. Cell Death Dis. 2017;8:e3080.
- 99. Tuo QZ, Liu Y, Xiang Z, Yan HF, Zou T, Shu Y, et al. Thrombin induces acsl4dependent ferroptosis during cerebral ischemia/reperfusion. Signal Transduct Target Ther. 2022;7:59.
- 100. Li J, Xu P, Hong Y, Xie Y, Peng M, Sun R, et al. Lipocalin-2-mediated astrocyte pyroptosis promotes neuroinflammatory injury via nlrp3 inflammasome activation in cerebral ischemia/reperfusion injury. J Neuroinflamm. 2023;20:148.
- Berthet C, Lei H, Thevenet J, Gruetter R, Magistretti PJ, Hirt L. Neuroprotective role of lactate after cerebral ischemia. J Cereb Blood Flow Metab. 2009;29:1780–9.
- Zhou J, Liu T, Guo H, Cui H, Li P, Feng D, et al. Lactate potentiates angiogenesis and neurogenesis in experimental intracerebral hemorrhage. Exp Mol Med. 2018:50:1–12.
- Tian Z, Ji X, Liu J. Neuroinflammation in vascular cognitive impairment and dementia: current evidence, advances, and prospects. Int J Mol Sci. 2022;23:6224.
- Moretti R, Caruso P. Small vessel disease: ancient description, novel biomarkers. Int J Mol Sci. 2022;23:3508.
- 105. Leng F, Edison P. Neuroinflammation and microglial activation in Alzheimer disease: where do we go from here? Nat Rev Neurol. 2021;17:157–72.
- Cai W, Hu M, Li C, Wu R, Lu D, Xie C, et al. FOXP3+ macrophage represses acute ischemic stroke-induced neural inflammation. Autophagy. 2023;19:1144–63.
- Liu M, Xu Z, Wang L, Zhang L, Liu Y, Cao J, et al. Cottonseed oil alleviates ischemic stroke injury by inhibiting the inflammatory activation of microglia and astrocyte. J Neuroinflamm. 2020;17:270.
- Ablasser A, Chen ZJ. Cgas in action: expanding roles in immunity and inflammation. Science. 2019;363:eaat8657.
- 109. Jiang GL, Yang XL, Zhou HJ, Long J, Liu B, Zhang LM, et al. Cgas knockdown promotes microglial m2 polarization to alleviate neuroinflammation by inhibiting cGAS-STING signaling pathway in cerebral ischemic stroke. Brain Res Bull. 2021;171:183–95.
- Wang L, Cai Z, Gu Q, Xu C. Cgas deficiency regulates the phenotypic polarization and glycolysis of microglia through lactylation in hypoxic-ischemic encephalopathy cell model. Biochem Genet. 2024;62:3961–76.
- 111. Li H, Liu C, Li R, Zhou L, Ran Y, Yang Q, et al. AARS1 and AARS2 sense L-lactate to regulate cGAS as global lysine lactyltransferases. Nature. 2024;634:1229–37.
- 112. Chen R, Kang R, Tang D. The mechanism of HMGB1 secretion and release. Exp Mol Med. 2022;54:91–102.
- Yao X, Li C. Lactate dehydrogenase a mediated histone lactylation induced the pyroptosis through targeting HMGB1. Metab Brain Dis. 2023;38:1543–53.
- 114. Mancias JD, Wang X, Gygi SP, Harper JW, Kimmelman AC. Quantitative proteomics identifies NCOA4 as the cargo receptor mediating ferritinophagy. Nature. 2014;509:105–9.
- 115. He X, Wang Z, Ge Q, Sun S, Li R, Wang B. Lactylation of nuclear receptor coactivator 4 promotes ferritinophagy and glycolysis of neuronal cells after cerebral ischemic injury. Neuroreport. 2024;35:895–903.
- 116. Wen M, Jin Y, Zhang H, Sun X, Kuai Y, Tan W. Proteomic analysis of rat cerebral cortex in the subacute to long-term phases of focal cerebral ischemiareperfusion injury. J Proteome Res. 2019;18:3099–118.
- Zhang W, Xu L, Yu Z, Zhang M, Liu J, Zhou J. Inhibition of the glycolysis prevents the cerebral infarction progression through decreasing the lactylation levels of LCP1. Mol Biotechnol. 2023;65:1336–45.
- Xiao Z, Shen D, Lan T, Wei C, Wu W, Sun Q, et al. Reduction of lactoferrin aggravates neuronal ferroptosis after intracerebral hemorrhagic stroke in hyperglycemic mice. Redox Biol. 2022;50:102256.

- Zhang L, Wang X, Che W, Zhou S, Feng Y. METTL3 silenced inhibited the ferroptosis development via regulating the TFRC levels in the intracerebral hemorrhage progression. Brain Res. 2023;1811:148373.
- Starke RM, Chalouhi N, Ali MS, Jabbour PM, Tjoumakaris SI, Gonzalez LF, et al. The role of oxidative stress in cerebral aneurysm formation and rupture. Curr Neurovasc Res. 2013:10:247–55.
- 121. Wu J, Lu L, Dai B, Yu A. Unraveling the role of LDHA and VEGFA in oxidative stress: a pathway to therapeutic interventions in cerebral aneurysms. Biomol Biomed. 2025;25:360–74.
- Guo H, Fan Z, Wang S, Ma L, Wang J, Yu D, et al. Astrocytic A1/A2 paradigm participates in glycogen mobilization mediated neuroprotection on reperfusion injury after ischemic stroke. J Neuroinflamm. 2021:18:230.
- Zhou J, Zhang L, Peng J, Zhang X, Zhang F, Wu Y, et al. Astrocytic LRP1 enables mitochondria transfer to neurons and mitigates brain ischemic stroke by suppressing ARF1 lactylation. Cell Metab. 2024;36:2054–68.
- Liu CC, Hu J, Tsai CW, Yue M, Melrose HL, Kanekiyo T, et al. Neuronal LRP1 regulates glucose metabolism and insulin signaling in the brain. J Neurosci. 2015;35:5851–9.
- Lee YS, Woo SC, Kim SY, Park JY. Understanding the multi-herbal composition of Buyang Huanwu decoction: a review for better clinical use. J Ethnopharmacol. 2020:255:112765.
- Song C, Fang X, Fang N, Hu F. Buyang Huanwu decoction suppresses ischemic stroke by suppressing glycolysis and cell apoptosis in rat brain microvascular endothelial cells. Brain Res Bull. 2024;215:111032.
- Singh D. Astrocytic and microglial cells as the modulators of neuroinflammation in Alzheimer's disease. J Neuroinflamm. 2022;19:206.
- Kwon HS, Koh SH. Neuroinflammation in neurodegenerative disorders: the roles of microglia and astrocytes. Transl Neurodegener. 2020;9:42.
- Cheng J, Zhang R, Xu Z, Ke Y, Sun R, Yang H, et al. Early glycolytic reprogramming controls microglial inflammatory activation. J Neuroinflamm. 2021;18:129.
- Yang XL, Wang X, Shao L, Jiang GT, Min JW, Mei XY, et al. TRPV1 mediates astrocyte activation and interleukin-1beta release induced by hypoxic ischemia (HI). J Neuroinflamm. 2019;16:114.
- 131. Shao J, Meng Y, Yuan K, Wu Q, Zhu S, Li Y, et al. Ru.521 mitigates subarachnoid hemorrhage-induced brain injury via regulating microglial polarization and neuroinflammation mediated by the cGAS/STING/NF-kappaB pathway. Cell Commun Signal. 2023;21:264.
- Fei X, Chen L, Gao J, Jiang X, Sun W, Cheng X, et al. P53 lysine-lactylated modification contributes to lipopolysaccharide-induced proinflammatory activation in BV2 cell under hypoxic conditions. Neurochem Int. 2024;178:105794.
- 133. Jiang X, Gao J, Fei X, Geng Y, Yue X, Shi Z, et al. Global profiling of protein lactylation in microglia in experimental high-altitude cerebral edema. Cell Commun Signal. 2024;22:374.
- Hasel P, Rose I, Sadick JS, Kim RD, Liddelow SA. Neuroinflammatory astrocyte subtypes in the mouse brain. Nat Neurosci. 2021;24:1475–87.
- Belanger M, Allaman I, Magistretti PJ. Brain energy metabolism: focus on astrocyte-neuron metabolic cooperation. Cell Metab. 2011;14:724–38.
- 136. Xu J, Ji T, Li G, Zhang H, Zheng Y, Li M, et al. Lactate attenuates astrocytic inflammation by inhibiting ubiquitination and degradation of NDRG2 under oxygen-glucose deprivation conditions. J Neuroinflamm. 2022;19:314.
- 137. Hu X, Huang J, Li Z, Li J, Ouyang F, Chen Z, et al. Lactate promotes microglial scar formation and facilitates locomotor function recovery by enhancing histone h4 lysine 12 lactylation after spinal cord injury. J Neuroinflamm. 2024;21:193.
- 138. Fan Z, Brooks DJ, Okello A, Edison P. An early and late peak in microglial activation in Alzheimer's disease trajectory. Brain. 2017;140:792–803.
- Gusyatiner O, Hegi ME. Glioma epigenetics: from subclassification to novel treatment options. Semin Cancer Biol. 2018;51:50–8.
- Tykocki T, Eltayeb M. Ten-year survival in glioblastoma. A systematic review. J Clin Neurosci. 2018:54:7–13.
- Ostrom QT, Price M, Neff C, Cioffi G, Waite KA, Kruchko C, et al. Cbtrus statistical report: primary brain and other central nervous system tumors diagnosed in the United States in 2015–2019. Neuro Oncol. 2022;24:v1–95.
- Dresselhaus EC, Meffert MK. Cellular specificity of NF-kappaB function in the nervous system. Front Immunol. 2019;10:1043.
- 143. Khabibov M, Garifullin A, Boumber Y, Khaddour K, Fernandez M, Khamitov F, et al. Signaling pathways and therapeutic approaches in glioblastoma multiforme (review). Int J Oncol. 2022;60:69.
- 144. Reuss AM, Groos D, Buchfelder M, Savaskan N. The acidic brain-glycolytic switch in the microenvironment of malignant glioma. Int J Mol Sci. 2021;22:5518.
- 145. van Tellingen O, Yetkin-Arik B, de Gooijer MC, Wesseling P, Wurdinger T, de Vries HE. Overcoming the blood-brain tumor barrier for effective glioblastoma treatment. Drug Resist Updat. 2015;19:1–12.
- Wei X, Chen Y, Jiang X, Peng M, Liu Y, Mo Y, et al. Mechanisms of vasculogenic mimicry in hypoxic tumor microenvironments. Mol Cancer. 2021;20:7.

- 147. Luo Q, Wang J, Zhao W, Peng Z, Liu X, Li B, et al. Vasculogenic mimicry in carcinogenesis and clinical applications. J Hematol Oncol. 2020;13:19.
- 148. Plate KH, Scholz A, Dumont DJ. Tumor angiogenesis and anti-angiogenic therapy in malignant gliomas revisited. Acta Neuropathol. 2012;124:763–75.
- 149. Qiu B, Matthay KK. Advancing therapy for neuroblastoma. Nat Rev Clin Oncol. 2022:19:515–33.
- Wang J, Liu Z, Xu Y, Wang Y, Wang F, Zhang Q, et al. Enterobacterial LPSinducible LINC00152 is regulated by histone lactylation and promotes cancer cells invasion and migration. Front Cell Infect Microbiol. 2022;12:913815.
- 151. Li L, Li Z, Meng X, Wang X, Song D, Liu Y, et al. Histone lactylation-derived LINC01127 promotes the self-renewal of glioblastoma stem cells via the cisregulating the MAP4K4 to activate JNK pathway. Cancer Lett. 2023;579:216467.
- 152. Valor LM, Hervas-Corpion I. The epigenetics of glioma stem cells: a brief overview. Front Oncol. 2020;10:602378.
- Fan Y, Lu H, Liang W, Hu W, Zhang J, Chen YE. Kruppel-like factors and vascular wall homeostasis. J Mol Cell Biol. 2017;9:352–63.
- Zhang M, Zhao Y, Liu X, Ruan X, Wang P, Liu L, et al. Pseudogene MAPK6P4encoded functional peptide promotes glioblastoma vasculogenic mimicry development. Commun Biol. 2023;6:1059.
- Matthay KK, Maris JM, Schleiermacher G, Nakagawara A, Mackall CL, Diller L, et al. Neuroblastoma. Nat Rev Dis Prim. 2016;2:16078.
- Qing G, Skuli N, Mayes PA, Pawel B, Martinez D, Maris JM, et al. Combinatorial regulation of neuroblastoma tumor progression by N-MYc and hypoxia inducible factor HIF-1alpha. Cancer Res. 2010;70:10351–61.
- 157. Wu X, Mi T, Jin L, Ren C, Wang J, Zhang Z, et al. Dual roles of HK3 in regulating the network between tumor cells and tumor-associated macrophages in neuroblastoma. Cancer Immunol Immunother. 2024;73:122.
- Elmore KB, Schaff LR. DNA repair mechanisms and therapeutic targets in glioma. Curr Oncol Rep. 2021;23:87.
- Chen Y, Wu J, Zhai L, Zhang T, Yin H, Gao H, et al. Metabolic regulation of homologous recombination repair by Mre11 lactylation. Cell. 2024;187:294–311.
- Chen H, Li Y, Li H, Chen X, Fu H, Mao D, et al. NBS1 lactylation is required for efficient DNA repair and chemotherapy resistance. Nature. 2024;631:663–9.
- Yue Q, Wang Z, Shen Y, Lan Y, Zhong X, Luo X, et al. Histone H3K9 lactylation confers temozolomide resistance in glioblastoma via LUC7L2-mediated MLH1 intron retention. Adv Sci (Weinh). 2024;11:e2309290.
- 162. Li G, Wang D, Zhai Y, Pan C, Zhang J, Wang C, et al. Glycometabolic reprogramming-induced XRCC1 lactylation confers therapeutic resistance in ALDH1A3-overexpressing glioblastoma. Cell Metab. 2024;36:1696–710.
- Choi S, Yu Y, Grimmer MR, Wahl M, Chang SM, Costello JF. Temozolomideassociated hypermutation in gliomas. Neuro Oncol. 2018;20:1300–9.
- 164. Mutamba JT, Svilar D, Prasongtanakij S, Wang XH, Lin YC, Dedon PC, et al. XRCC1 and base excision repair balance in response to nitric oxide. DNA Repair (Amst). 2011:10:1282–93.
- 165. Tomar MS, Kumar A, Srivastava C, Shrivastava A. Elucidating the mechanisms of temozolomide resistance in gliomas and the strategies to overcome the resistance. Biochim Biophys Acta Rev Cancer. 2021;1876:188616.
- Nabors LB, Portnow J, Ammirati M, Baehring J, Brem H, Butowski N, et al. NCCN guidelines insights: central nervous system cancers, version 1.2017. J Natl Compr Canc Netw. 2017;15:1331–45.
- 167. Xu S, Tang L, Li X, Fan F, Liu Z. Immunotherapy for glioma: current management and future application. Cancer Lett. 2020;476:1–12.
- 168. Thomas BC, Staudt DE, Douglas AM, Monje M, Vitanza NA, Dun MD. CAR T cell therapies for diffuse midline glioma. Trends Cancer. 2023;9:791–804.
- 169. Sun T, Liu B, Li Y, Wu J, Cao Y, Yang S, et al. Oxamate enhances the efficacy of CAR-T therapy against glioblastoma via suppressing ectonucleotidases and CCR8 lactylation. J Exp Clin Cancer Res. 2023;42:253.
- 170. Khan F, Pang L, Dunterman M, Lesniak MS, Heimberger AB, Chen P. Macrophages and microglia in glioblastoma: heterogeneity, plasticity, and therapy. J Clin Investig. 2023;133:e163446.
- 171. Li M, Sun P, Tu B, Deng G, Li D, He W. Hypoxia conduces the glioma progression by inducing M2 macrophage polarization via elevating TNFSF9 level in a histone-lactylation-dependent manner. Am J Physiol Cell Physiol. 2024;327:C487–504.
- 172. Yeo AT, Rawal S, Delcuze B, Christofides A, Atayde A, Strauss L, et al. Single-cell RNA sequencing reveals evolution of immune landscape during glioblastoma progression. Nat Immunol. 2022;23:971–84.
- 173. Lu X, Zhou Z, Qiu P, Xin T. Integrated single-cell and bulk RNA-sequencing data reveal molecular subtypes based on lactylation-related genes and prognosis and therapeutic response in glioma. Heliyon. 2024;10:e30726.
- 174. Goenka A, Tiek DM, Song X, Iglesia RP, Lu M, Hu B, et al. The role of non-coding RNAs in glioma. Biomedicines. 2022;10:2031.
- 175. Khacho M, Harris R, Slack RS. Mitochondria as central regulators of neural stem cell fate and cognitive function. Nat Rev Neurosci. 2019;20:34–48.

- 176. Oginuma M, Moncuquet P, Xiong F, Karoly E, Chal J, Guevorkian K, et al. A gradient of glycolytic activity coordinates FGF and Wnt signaling during elongation of the body axis in amniote embryos. Dev Cell. 2017;40:342–53.
- Bhattacharya D, Azambuja AP, Simoes-Costa M. Metabolic reprogramming promotes neural crest migration via YAP/TEAD signaling. Dev Cell. 2020:53:199–211.
- 178. Merkuri F, Rothstein M, Simoes-Costa M. Histone lactylation couples cellular metabolism with developmental gene regulatory networks. Nat Commun. 2024:15:90
- 179. Suzuki A, Sangani DR, Ansari A, Iwata J. Molecular mechanisms of midfacial developmental defects. Dev Dyn. 2016;245:276–93.
- Yang J, Zhu L, Pan H, Ueharu H, Toda M, Yang Q, et al. A BMP-controlled metabolic/epigenetic signaling cascade directs midfacial morphogenesis. J Clin Investiq. 2024;134:e165787.
- 181. Wang Y, Wang W, Su L, Ji F, Zhang M, Xie Y, et al. BACH1 changes microglial metabolism and affects astrogenesis during mouse brain development. Dev Cell. 2024;59:108–24.
- 182. Wiel C, Le Gal K, Ibrahim MX, Jahangir CA, Kashif M, Yao H, et al. BACH1 stabilization by antioxidants stimulates lung cancer metastasis. Cell. 2019:178:330–45.
- 183. Penninx BW, Pine DS, Holmes EA, Reif A. Anxiety disorders. Lancet. 2021;397:914–27.
- 184. Long J, Huang G, Liang W, Liang B, Chen Q, Xie J, et al. The prevalence of schizophrenia in mainland China: evidence from epidemiological surveys. Acta Psychiatr Scand. 2014:130:244–56.
- Boison D, Steinhauser C. Epilepsy and astrocyte energy metabolism. Glia. 2018:66:1235–43.
- Monsorno K, Ginggen K, Ivanov A, Buckinx A, Lalive AL, Tchenio A, et al. Loss of microglial MCT4 leads to defective synaptic pruning and anxiety-like behavior in mice. Nat Commun. 2023;14:5749.
- Ernst J, Hock A, Henning A, Seifritz E, Boeker H, Grimm S. Increased pregenual anterior cingulate glucose and lactate concentrations in major depressive disorder. Mol Psychiatry. 2017;22:113–9.
- 188. Ham BJ, Sung Y, Kim N, Kim SJ, Kim JE, Kim DJ, et al. Decreased GABA levels in anterior cingulate and basal ganglia in medicated subjects with panic disorder: a proton magnetic resonance spectroscopy (1H-MRS) study. Prog Neuropsychopharmacol Biol Psychiatry. 2007;31:403–11.
- 189. Dogan AE, Yuksel C, Du F, Chouinard VA, Ongur D. Brain lactate and ph in schizophrenia and bipolar disorder: a systematic review of findings from magnetic resonance studies. Neuropsychopharmacol. 2018;43:1681–90.
- Angamo EA, ul Haq R, Rösner J, Gabriel S, Gerevich Z, Heinemann U, et al. Contribution of intrinsic lactate to maintenance of seizure activity in neocortical slices from patients with temporal lobe epilepsy and in rat entorhinal cortex. Int J Mol Sci. 2017;18:1835.
- 191. Hagihara H, Shoji H, Otabi H, Toyoda A, Katoh K, Namihira M, et al. Protein lactylation induced by neural excitation. Cell Rep. 2021;37:109820.
- 192. Yan L, Wang Y, Hu H, Yang D, Wang W, Luo Z, et al. Physical exercise mediates cortical synaptic protein lactylation to improve stress resilience. Cell Metab. 2024;36:2104–17.
- 193. Wang R, Liu ZH, Bi N, Gu X, Huang C, Zhou R, et al. Dysfunction of the medial prefrontal cortex contributes to bpa-induced depression- and anxiety-like behavior in mice. Ecotoxicol Environ Saf. 2023;259:115034.
- 194. Xie J, Hong S, Zhang X, Li Y, Xie R. Inhibition of glycolysis prevents behavioural changes in mice with MK801-induced SCZ model by alleviating lactate accumulation and lactylation. Brain Res. 2023;1812:148409.
- 195. Khatibi VA, Salimi M, Rahdar M, Rezaei M, Nazari M, Dehghan S, et al. Glycolysis inhibition partially resets epilepsy-induced alterations in the dorsal

- hippocampus-basolateral amygdala circuit involved in anxiety-like behavior. Sci Rep. 2023;13:6520.
- 196. Dorneburg C, Fischer M, Barth T, Mueller-Klieser W, Hero B, Gecht J, et al. Ldha in neuroblastoma is associated with poor outcome and its depletion decreases neuroblastoma growth independent of aerobic glycolysis. Clin Cancer Res. 2018;24:5772–83
- 197. Neal JT, Li X, Zhu J, Giangarra V, Grzeskowiak CL, Ju J, et al. Organoid modeling of the tumor immune microenvironment. Cell. 2018;175:1972–88.

AUTHOR CONTRIBUTIONS

WGZ and MRH conceptualized and developed the design of the article, interpreted relevant literature, and wrote the original manuscript. WFH and LJG conducted the review and editing steps of the manuscript. WGZ was responsible for funding acquisition and validation. All authors have read and approved the final submission.

FUNDING

This work was supported by the National Natural Science Foundation of China (82370383), Guangdong Natural Science Foundation (2024A1515013289), Jiangxi Provincial Health Commission Science and Technology Plan Project (202310008), and Jiangxi Natural Science Foundation (20232BAB206055).

COMPETING INTERESTS

The authors declare no competing interests.

ADDITIONAL INFORMATION

Supplementary information The online version contains supplementary material available at https://doi.org/10.1038/s41420-025-02408-w.

Correspondence and requests for materials should be addressed to Wenfeng He, Wengen Zhu or Linjuan Guo.

Reprints and permission information is available at http://www.nature.com/reprints

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing,

adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

© The Author(s) 2025