

Intermittent Parathyroid Hormone Accelerates Stress Fracture Healing More Effectively Following Cessation of Bisphosphonate Treatment

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ABSTRACT

Parathyroid hormone (PTH) and bisphosphonates (BPs), including alendronate (ALN), have opposing effects on bone dynamics. The extent to which PTH remains effective in the treatment of stress fracture (SFx) in the presence of an ongoing BP treatment has not been tested. SFx was induced in 150 female Wistar rats, divided into five equal groups ($n = 30$). All rats were pretreated with ALN ($1 \mu\text{g}/\text{kg}^{-1}/\text{day}^{-1}$) for 14 days prior to SFx induction, followed by ALN cessation or continuation for the duration of the experiment; this was combined with daily PTH ($8 \mu\text{g}/100 \text{g}^{-1}/\text{day}^{-1}$) on SFx induction for 14 days, followed by cessation or continuation of ALN after SFx induction or an equivalent vehicle as a control. Ulnas were examined 2 weeks or 6 weeks following SFx. Two toluidine blue and two tartrate-resistant acid phosphatase-stained sections were examined for histomorphometric analysis using Osteomeasure software. There was a significant interaction between the effects of time and treatment type on the woven bone width and apposition rate, as well as an improvement in the woven bone architecture. However, woven bone variables remained unaffected by the cessation or continuation of ALN. Cessation of ALN increased osteoclast number when compared with the ALN-PTH continuation group ($p = 0.006$), and vehicle ($p = 0.024$) after 2 weeks. There was a significant interaction between the effects of time and treatment type on the number of osteoclasts per unit BMU area and length. The number of osteoclasts per unit BMU area and length was significantly greater in ALN cessation groups. It was concluded that intermittent short-duration iPTH treatment effectively increased remodeling of SFx with a concurrent BP treatment, provided that BP was ceased at the time of SFx. Our results could help develop shorter iPTH treatment protocols for the clinical management of SFxs and guide clinical decision-making to cease BP treatment in cases of SFx. © 2020 The Authors. *JBMR Plus* published by Wiley Periodicals LLC. on behalf of American Society for Bone and Mineral Research.

KEY WORDS: BISPHOSPHONATES; STRESS FRACTURE; BONE REMODELING; HEALING; PARATHYROID HORMONE; ULNA

Introduction

Stress fractures (SFxs) account for about 1% to 7% of all athletic injuries.^(1,2) The main difference between SFxs and acute fractures is related to the nature of loading of the bones. An acute fracture typically occurs because of a single maximal loading, whereas SFxs occur because of repetitive submaximal loading.⁽¹⁾ Bisphosphonates (BPs) are stable analogues of pyrophosphatase. They are deposited on bone surfaces within minutes or hours of uptake. The mode of action on the osteoclast is radically different between non-nitrogen-containing BPs (first generation) and nitrogen-containing BPs (second and third generation). Apoptosis is induced by non-nitrogen-containing BPs through the formation of a toxic analogue of adenosine

triphosphate, whereas nitrogen-containing BPs (second and third generation) target the enzyme farnesyl diphosphate synthase needed for posttranslational modification of small guanosine triphosphate- (GTP-) binding proteins required for osteoclastic function.^(3,4) All BPs have an affinity for bone tissue, more specifically to osteoclasts, because the acidic pH of resorption lacunae causes an intracellular uptake of BPs, leading to the internalization of substantial amounts of BP. Regardless of the inhibitory pathway, all BPs depress osteoclast activation and resorption.⁽⁴⁾

BPs are highly effective in the treatment of osteoporosis. Numerous large clinical trials demonstrate their efficacy in reducing bone turnover, increasing BMD, and reducing vertebral and nonvertebral fracture risk in patients with osteoporosis.^(5,6) They

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are incorporated into the bone matrix and have a biological half-life of more than 10 years.⁽⁷⁾ As the duration of antiresorptive treatments increased to 10 years or more, debate arose concerning the risks versus benefits of antiresorptive treatment, and the consequences of “frozen bone.”⁽⁸⁾ One of the side-effects that emerged was a SFx in the proximal femoral diaphysis, which progressed to an atypical femoral fracture (AFF).^(8–13) Recently, a case of fracture in the proximal ulna was also reported after prolonged BP treatment.⁽¹⁴⁾

It is now well-accepted that the pathogenesis of AFF is its initiation as a SFx caused by an accumulation of unrepaired microdamage following suppression of bone turnover by BPs.^(15,16) It is also acknowledged that initiation of the microdamage is facilitated by bone hypermineralization and accumulation of advanced glycation end-products that reduce the toughness of the bone matrix.^(17–20) This hypothesis is supported by the location of the AFF in areas of stress (lateral femur), and the fact that such fractures develop over a long period and are preceded by episodes of prodromal pain.⁽¹⁵⁾ There are a lot of similarities between SFxs and AFFs in terms of etiology^(15,16) and possible management protocols.^(21–23) Therefore, we hypothesized that anabolic PTH treatment would accelerate SFxs, even in the presence of BP treatment.

Many studies confirm the anabolic effect of PTH and its correlation with the timing and duration of treatment.^(24,25) Intermittent administration of PTH results in increased bone formation, and cortical bone volume and width. It also improves bone architecture and mechanical properties in different species.^(26–28) The potential for PTH to treat localized osseous defects and accelerate SFx healing has also been investigated.^(29–31) Conversely, combination therapy of PTH plus BP (alendronate [ALN]) impairs the anabolic function of PTH and its ability to enhance BMD.^(32,33) For example, PTH treatment for 2 months after an antiresorptive agent increased bone formation and the mineralization rate, but it was less than that achieved with PTH treatment alone,^(34–36) though this observation has not been universal.⁽³⁷⁾ So the question remains as to what extent PTH can remain effective in the presence of BP treatment.

We previously showed that BP treatment (risedronate) impaired healing of a SFx by reducing the volume of bone resorbed and replaced during remodeling.⁽³⁸⁾ However, formation of a periosteal callus was not adversely affected. This woven bone reaction acts to return the bone to its original strength 2 weeks after SFx induction.^(39,40) However, there is little analysis available for the combined effects of a BP with PTH. In the ulna SFx model, PTH increased BMC significantly by 7% at 4 weeks and BMD and BMC significantly by 10% and 7% at 8 weeks compared with controls, whereas ALN did not change BMD or BMC.⁽⁴¹⁾ PTH significantly stimulated bone formation by 114% at 2 weeks, increased intracortical resorption area by 23% at 4 weeks, and enhanced the ultimate force of the affected ulnas by 15% at 8 weeks. Similar to Kidd and colleagues,⁽³⁸⁾ ALN significantly suppressed the bone formation rate by 44% compared with the control at 4 weeks.⁽⁴¹⁾ These data suggest that PTH could accelerate SFx remodeling, but the combined effects of PTH with BPs remain unknown.

The ulnar loading model in rats provides an effective approach to examine focal SFx remodeling with a known time course and precise anatomical location.^(22,31,38,42) Patients receiving long-term BP treatment may be at higher risk for development of SFx, and more serious complications including AFF.^(8–13) Although some studies have investigated the effect of combined PTH-BP therapy in osteoporosis, this is not the case for SFx and potential AFF.⁽⁴³⁾ Therefore, the aim of the current experiment was to investigate the efficacy of a daily iPTH treatment for 14 days in acceleration of SFx remodeling and healing

indices in the rat ulna in the presence or absence of a concurrent BP treatment (ALN).

Materials and Methods

The Griffith University Animal Ethics Committee (Nathan, Queensland, Australia) approved the experimental protocols (GU Ref No: MSC/02/13/AEC).

We induced an ulnar SFx in 150 female Wistar rats, 12 weeks of age, weight (300 ± 15 g). Rats were anesthetized with halothane and oxygen for loading. SFx was achieved in a single loading session of the forearm in axial compression at 2 Hz until an increase in displacement of 10% was reached, on average after approximately 8000 cycles or about 60 minutes (range, 4000 to 20,000). This produced a remarkably standardized SFx.^(22,31) Accounting for variability, anesthesia, and recovery, four to five rats were loaded/day. Loading was performed in a loading device using a Linear Voltage Displacement Transducer (LVDT) to monitor displacement in the limb. Because of the natural ulnar curvature, axial compression was converted into bending forces with the lateral cortex in tension and the medial in compression.⁽⁴⁴⁾ Loading involved cyclic compressive loading at 18 to 20 N load and 2-Hz cycle frequency. Loading was stopped at a predetermined point when increased displacement reached 10%. As noted above, this level of stiffness loss consistently produced SFxs. A single injection of opioid analgesia (buprenorphine 0.05 mg/kg s.c.) was used following loading sessions.

Rats were divided into ALN-, combined therapy- (ALN-PTH-), and vehicle- (VEH) treated groups (*n* = 15 each). PTH-treated groups received s.c. injections of human PTH-(1–34) peptide (Sigma-Aldrich, St. Louis, MO, USA) dissolved in 0.9% saline with 1% rat heat-inactivated serum in a final volume of 200 µL with a dose of 8 µg/100 g/day. PTH treatment was started 24 hours after SFx loading and continued daily for 14 days. ALN-treated groups received s.c. injections of ALN in saline carrier, once daily at 1.0 µg/kg/day. The ALN treatment started 14 days before the SFx loading. The ALN dose was adjusted for the metabolic rate of the rat and equivalent to the human clinical dose.⁽²²⁾ VEH groups received an equivalent dose of saline for each ALN injection and an equivalent dose of rat serum for each PTH injection in the corresponding treatment groups. Rats were euthanized at 2- and 6-weeks post-SFx (Table 1).

Table 1. Experimental Design of the Study

Group	14 days (post-SFx)	6 weeks (post-SFx)
Alendronate continuation (ALN ₁)	15	15
Alendronate cessation (ALN ₂)	15	15
Combined treatment with alendronate continuation (ALN-PTH ₁)	15	15
Combined treatment with alendronate cessation (ALN-PTH ₂)	15	15
Vehicle (VEH)	15	15

ALN₁ = Pretreatment with ALN for 14 days before SFx, followed by daily ALN injections up to 6 weeks post-SFx; ALN₂ = pretreatment with ALN for 14 days before SFx, followed by cessation of ALN at time of SFx, no treatment up to 6 weeks; ALN-PTH₁ = pretreatment with ALN for 14 days before SFx, followed by daily ALN injections up to 6 weeks post-SFx + 14 days of PTH treatment post-SFx; ALN-PTH₂ = pretreatment with ALN for 14 days before SFx, followed by cessation of ALN at time of SFx + 14 days of PTH treatment post-SFx; SFx = stress fracture; VEH = control for ALN-PTH groups.

Histomorphometry

Two toluidine blue and two tartrate-resistant acid phosphatase-stained sections were examined for histomorphometric analysis using Osteomeasure software (OsteoMetrics, Decatur, GA, USA). As previously described, histomorphometry was performed at a standard level along the SFx (Fig. 1A), where the microcrack was halfway between the medial cortical margin and the medullary cavity of the bone in transverse section.^(22,31) The area of a BMU was defined as the total area that had been resorbed. Within this BMU, the area filled with new bone formation was defined as the healed area. A distinct cement line around a previously resorbed area of bone defined the healed area. The area that had been resorbed, but not yet filled by new bone formation, was defined as porosity. Morphometric measures included standard variables (Fig. 1):

- 1 Cortical bone area (Ct.Ar [mm^2])
- 2 Cortical bone perimeter (mm)
- 3 Woven bone area (mm^2)
- 4 Woven bone perimeter (mm)
- 5 Woven bone width (Wo.B.Wi [mm])
- 6 Length of SFx (μm)
- 7 Length of remodeling unit along the microcrack (μm)
- 8 Porosity BMU area (μm^2)
- 9 Porosity BMU area perimeter (μm)
- 10 Erosion (unhealed) area (μm^2)
- 11 Erosion (unhealed) perimeter (μm)
- 12 Healing area (μm^2)
- 13 Healing area perimeter (μm)
- 14 Number of osteoclasts
- 15 Osteoclasts surface perimeter (μm)

From the standard variables measured above, the following derived variables were obtained:

- 1 Number of osteoclasts per μm^2 of BMU area (number of osteoclasts / μm^2)
- 2 Number of osteoclasts per μm of BMU length (number of osteoclasts / μm)
- 3 Healing percentage (healing area [μm^2] / porosity BMU area $\times 100$ [μm^2])
- 4 Woven bone apposition rate per day (Wo.B.Wi \times woven bone perimeter / number of days)

In addition, the following derived variables were calculated to correct for variations in the total length of the microcrack.

- 1 Percentage fracture length occupied by bone formation (healing area perimeter [μm] / length of SFx [μm %])
- 2 Healing area per mm^2 of the cortical bone area (healing area [μm^2] / Ct.Ar [mm^2])
- 3 Porosity BMU area per mm^2 of the cortical bone area (porosity BMU area [μm^2] / Ct.Ar [mm^2])
- 4 Percentage fracture length occupied by erosion (erosion unhealed perimeter [μm] / length of SFx [μm %])
- 5 Erosion area per mm^2 of the cortical bone area (erosion unhealed area [μm^2] / Ct.Ar [mm^2])

Statistical analysis

The data collected from this experiment were analyzed using a two-way ANOVA with time and treatment group as the independent variables. In the presence of a significant statistical interaction, the main effects in the original two-way ANOVA were ignored, post hoc pairwise comparisons were performed between individual groups and differences determined using Fisher's least significant difference. In the presence of a nonsignificant statistical interaction, the ANOVA main effect for each independent variable (time or treatment group) was reported independently. Significance was accepted at $p \leq 0.05$.

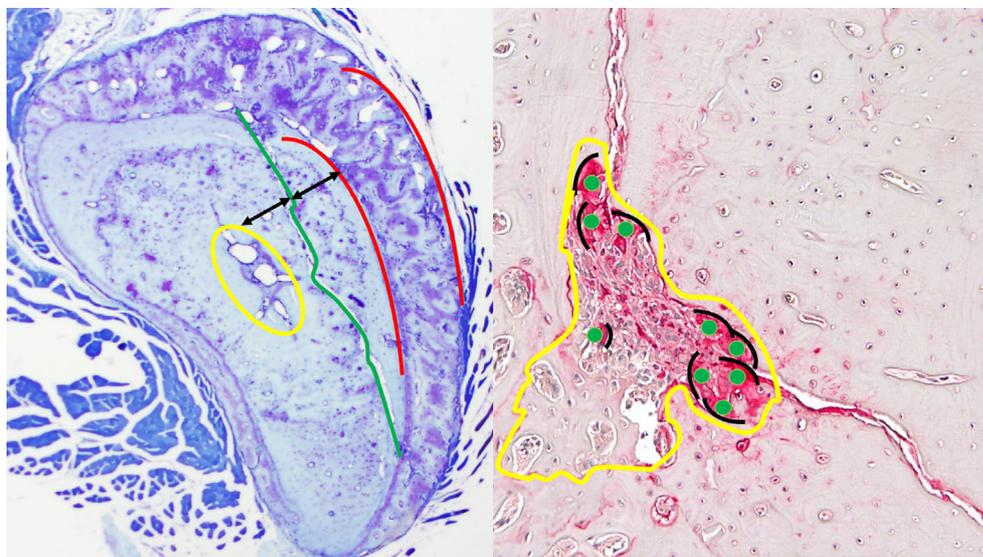


Fig 1. Left panel: Photomicrograph of a transverse section of a rat's ulna showing the standardized position chosen for histomorphometric analysis in this study along the stress fracture. To avoid any bias the slides selected for analysis were chosen at a position where the stress fracture (green) was midway between the outer cortical margins (red) and the inner medullary cavity (yellow; toluidine blue 2X). Right panel: A schematic diagram on a tartrate-resistant acid phosphatase-stained slide showing the boundaries of the BMU (yellow) with osteoclast count (green) and osteoclast perimeter (black; tartrate-resistant acid phosphatase 10X).

Results

There were no significant differences among groups in terms of Ct.Ar and cortical bone perimeter, woven bone area, and the length of SFx (μm).

Woven bone parameters

There were no significant differences in woven bone area between different treatment groups (Fig. 2A). There was a significant interaction between the main effects of time and the type of treatment (ALN₁ versus ALN₂ versus ALN-PTH₁ versus ALN-PTH₂ versus VEH) on Wo.B.Wi ($F = 3.169$; $p = 0.016$). Continuation or cessation of ALN had no effect on Wo.B.Wi 2 weeks post-SFx induction. Wo.B.Wi was significantly higher in the combined ALN-PTH₁ treatment group when compared with the ALN₁ ($p = 0.006$) group after 2 weeks. It was also significantly higher in the combined ALN-PTH₂ treatment group when compared with the ALN₂ ($p < 0.001$) group at the same time frame. After 6 weeks, Wo.B.Wi was significantly greater in the continuous ALN-PTH₁ treatment group when compared with ALN cessation in ALN-PTH₂ treatment ($p = 0.05$) group and the VEH group ($p = 0.016$; Fig. 2B).

There was no significant interaction between the effects of time and treatment type on the woven bone perimeter (Table 2). With regards to the main effect of time on woven bone

perimeter, it was significantly higher at 6 weeks post-SFx induction when compared with 2 weeks ($p = 0.008$; Fig. 2C).

There was a significant interaction between the main effects of time and the type of treatment (ALN₁ versus ALN₂ versus ALN-PTH₁ versus ALN-PTH₂ versus VEH) on the woven bone apposition rate ($F = 4.127$; $p = .003$). The woven bone apposition rate was significantly greater in the combined treatment groups (ALN-PTH₁ and ALN-PTH₂) when compared with the ALN treatment groups (ALN₁ and ALN₂) after 2 weeks of SFx induction ($p = 0.008$ and $p < 0.001$, respectively). Furthermore, the woven bone apposition rate was significantly higher in the ALN-PTH₁ and ALN-PTH₂ groups when compared with the VEH group after 2 weeks of SFx induction ($p = 0.002$ and $p < 0.001$, respectively). Finally, the woven bone apposition rate decreased significantly in all groups after 6 weeks when compared with the second week ($p < 0.001$; Fig. 2D).

Osteoclast parameters

There was no significant interaction between the effects of time and treatment type on the number of osteoclasts or osteoclast perimeters (Table 2).

The number of osteoclasts was significantly higher in the ALN-PTH₂ group when compared with the ALN-PTH₁ ($p = .006$) and VEH groups ($p = .024$; Figs. 2E and 3). The number of osteoclasts and osteoclast perimeters were significantly less after 6 weeks

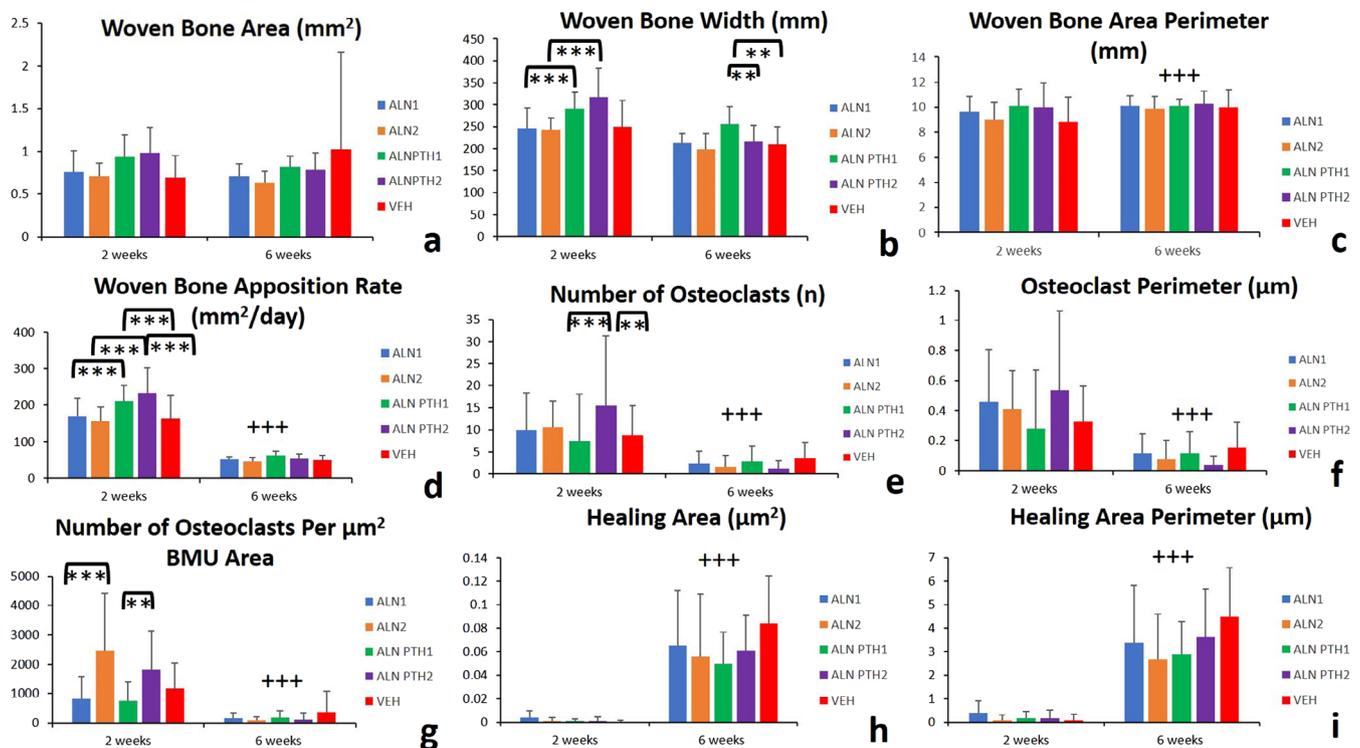


Fig 2. (A–I) Histomorphometric variables of stress fracture woven bone callus, osteoclast, and healing parameters (\pm SD). Combined alendronate-parathyroid hormone (ALN-PTH) treatment was superior to ALN and vehicle treatments in terms of woven bone sculpture and modeling throughout the healing process of a stress fracture. Cessation of ALN results in a significantly less woven bone width, indicating a more rapid progression in healing. Cessation of ALN after 2 weeks resulted in a significantly greater osteoclast number. ALN and combined ALN-PTH treatment modes (cessation and continuation) resulted in less healing area and perimeter after 6 weeks when compared with vehicle. Most variables were significantly influenced by the main effect of time between the 2-week and 6-week time points. Furthermore, there was a significant interaction between the main effects of time and the treatment type on healing perimeter. ** = $p \leq 0.05$; *** = $p \leq 0.01$ (differences between treatment types); +++ = $p \leq 0.01$ (differences compared with the previous time point).

Table 2. A Summary of the Significant Findings From the ALN, ALN-PTH, and VEH Groups Related to the Effect of Time, Treatment as well as the Interaction Between Treatment and Time on Different Variables Using a Linear Model in a Two-Way ANOVA Statistical Analysis

Variable	Effect	F	Significance
Wo.B.Wi (mm)	Treatment	6.403	<0.001
	Time	49.783	<0.001
	Interaction (Treatment * Time)	3.169	0.016
Wo.B.Pm (mm)	Treatment	2.141	0.079
	Time	7.279	0.008
	Interaction (Treatment * Time)	1.004	0.408
SFx.Po.Ar (μm^2)	Treatment	.330	0.858
	Time	31.318	<0.001
	Interaction (Treatment * Time)	.524	0.718
SFx.Po.Pm (μm)	Treatment	.556	0.695
	Time	28.886	<0.001
	Interaction (Treatment * Time)	1.010	0.405
SFx.He.Ar (μm^2)	Treatment	.279	0.891
	Time	149.784	<0.001
	Interaction (Treatment * Time)	1.277	0.282
SFx.He.Pm (μm)	Treatment	.418	.795
	Time	164.804	<0.001
	Interaction (Treatment * Time)	1.662	0.163
SFx.E.Ar (μm^2)	Treatment	.536	.710
	Time	126.722	<0.001
	Interaction (Treatment * Time)	1.841	0.125
SFx.E.Pm (μm)	Treatment	2.237	0.069
	Time	54.302	<0.001
	Interaction (Treatment * Time)	1.450	0.221
N.Oc	Treatment	2.431	0.05
	Time	41.695	<0.001
	Interaction (Treatment * Time)	1.847	0.124
Oc.Pm (μm)	Treatment	2.089	0.086
	Time	42.468	0.001
	Interaction (Treatment * Time)	1.730	0.147
He%	Treatment	.238	0.916
	Time	46.449	<0.001
	Interaction (Treatment * Time)	.838	0.505
Woven bone apposition rate per day (mm^2/day)	Treatment	8.724	<0.001
	Time	425.900	<0.001
	Interaction (Treatment * Time)	4.217	0.003
Number of osteoclasts per μm^2 of BMU area (N.Oc/ μm^2)	Treatment	6.095	<0.001
	Time	46.342	<0.001
	Interaction (Treatment * Time)	4.032	0.005
Number of osteoclasts per μm of BMU length (N.Oc/ μm)	Treatment	3.559	0.009
	Time	38.909	<0.001
	Interaction (Treatment * Time)	2.816	0.028
Healing area per mm^2 of the cortical bone area (SFx.He.Ar [μm^2]/ Ct.Ar [mm^2])	Treatment	.289	0.885
	Time	146.062	<0.001
	Interaction (Treatment * Time)	1.300	0.274
Porosity BMU area per mm^2 of the cortical bone area (SFx.Po.Ar [μm^2]/Ct.Ar [mm^2])	Treatment	.284	0.888
	Time	153.287	<0.001
	Interaction (Treatment * Time)	.721	0.579
Erosion (unhealed) area per mm^2 of the cortical bone area (SFx.E.Ar, μm^2 / Ct.Ar [mm^2])	Treatment	.503	0.733
	Time	130.923	<0.001
	Interaction (Treatment * Time)	1.946	0.107
Percentage fracture length occupied by bone formation (SFx.He.Pm, μm /SFx.Le [μm %])	Treatment	.284	0.888
	Time	153.287	<0.001
	Interaction (Treatment * Time)	.721	0.579
Percentage fracture length occupied by erosion (SFx.E.Pm, μm /SFx.Le [μm %])	Treatment	2.576	0.041
	Time	45.254	<0.001
	Interaction (Treatment * Time)	1.898	0.115

ALN = alendronate; He% = healing percentage; N.Oc = number of osteoclasts; Oc.Pm = osteoclasts surface perimeter; SFx = stress fracture; SFx.E.Ar = erosion (unhealed) area; SFx.E.Pm = erosion (unhealed) perimeter; SFx.He.Ar = healing area; SFx.He.Pm = healing area perimeter; SFx.Le = length of stress fracture; SFx.Po.Ar = porosity BMU area; SFx.Po.Pm = porosity (BMU) area perimeter; VEH = vehicle; Wo.B.Wi = woven bone width; Wo.B.Pm = woven bone perimeter.

when compared with 2 weeks post-SFx induction ($p < 0.001$; Figs. 2E,F, and 3).

There was a significant interaction between the main effects of time and the type of treatment (ALN₁ versus ALN₂ versus ALN-PTH₁ versus ALN-PTH₂ versus VEH) on the number of osteoclasts per unit porosity BMU area ($F = 4.032$; $p = .005$). The number of osteoclasts per unit porosity BMU area was significantly greater after 2 weeks following cessation of ALN (ALN₂), when compared with continuous ALN₁ treatment ($p < .001$). Furthermore, the number of osteoclasts per unit porosity BMU area was significantly greater after 2 weeks of ALN cessation in the ALN-PTH₂ group when compared with when ALN treatment was continued in the ALN-PTH₁ group ($p = 0.03$; Figs. 2G and 3). With regards to the single main effect of time, the number of osteoclasts per μm^2 of BMU area was significantly less after 6 weeks when compared with the second week in the ALN cessation groups (ALN₂ and ALN-PTH₂) when compared with the ALN continuation groups (ALN₁ and ALN-PTH₁; $p < 0.001$; Figs. 2G and 3).

There was a significant interaction between the main effects of time and the type of treatment (ALN₁ versus ALN₂ versus ALN-PTH₁ versus ALN-PTH₂ versus VEH) on the number of osteoclasts per unit BMU length ($F = 2.816$; $p = 0.028$). The number of osteoclasts per unit BMU length was significantly greater 2 weeks after ceasing ALN (ALN-PTH₂ group) when compared with ALN continuation in the ALN-PTH₁ group ($p < 0.001$; Figs. 2G and 3). Furthermore, the number of osteoclasts per μm of BMU length was significantly less after 6 weeks post-SFx induction when compared with the second week in the ALN₁ group

($p = 0.005$), the ALN₂ group ($p = 0.001$), and the ALN-PTH₂ group ($p < 0.001$).

Healing parameters

There was no significant interaction between the effects of time and treatment type on any of the healing parameters (Table 2). With regards to the main effect of time, all healing parameters were significantly greater 6 weeks post-SFx induction when compared with the second week ($p < 0.001$; Fig. 4). This finding includes the healing area (Fig. 2H), healing perimeter (Fig. 2I), healing percentage (Fig. 5A), healing area per mm^2 of the Ct.Ar (Fig. 5B), and the percentage fracture length occupied by bone formation (Fig. 5C).

Erosion (unhealed) parameters

There was no significant interaction between the effects of time and treatment type on any of the erosion parameters (Table 2).

With regards to the main effect of time, all erosion parameters were significantly less after 6 weeks of SFx induction when compared with the second week ($p < 0.001$; Fig. 5D,E,F,G). The percentage fracture length occupied by erosion was significantly less in the ALN-PTH₁ group, when ALN was continued, in comparison with the ALN-PTH₂ group, when the ALN was ceased ($p = 0.006$), and the VEH ($p = 0.01$; Fig. 5F) group.

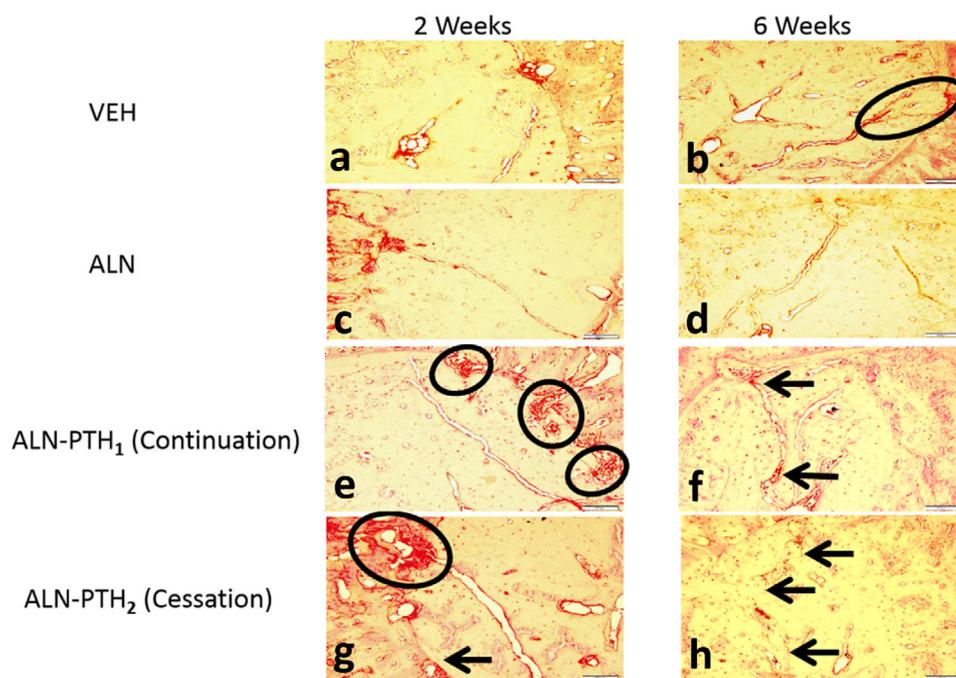


Fig 3. (A–H) Photomicrographs showing osteoclastic activity in different treatment groups and timelines. (A,B) Showing normal osteoclastic activity in vehicle (VEH) groups, (C) suppressed osteoclastic activity in alendronate (ALN) groups, and (G) higher osteoclastic activity in the ALN-PTH₂ group during the resorptive phase of the remodeling cycle after 2 weeks. (E) Continuation of ALN treatment in ALN-PTH₁ resulted in suppression of osteoclastic activity along the stress fracture despite the recruitment and availability of osteoclasts around the woven bone callus. (B,D,F,H) After 6 weeks, osteoclastic activity was significantly less in all groups apart from some osteoclastic trials indicating past resorptive activity. ALN-PTH groups showed evidence of osteoclastic activity only around, but not along the stress fracture because of the progression in healing. (Tartrate-resistant acid phosphatase 10X). Scale bar = 10 μm .

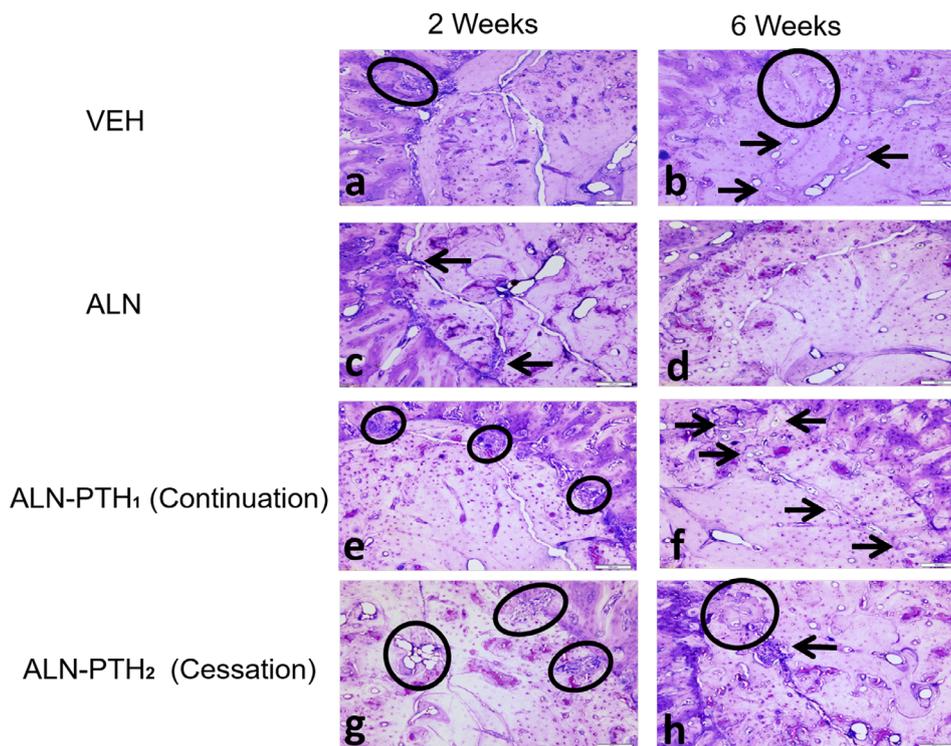


Fig 4. Photomicrographs showing examples of microscopic evaluation of the bone remodeling unit in stress fractures. (A,B) Showing the initiation of remodeling in vehicle (VEH) groups (black arrows and circles). (C) Alendronate (ALN) treatment resulted in the development of smaller BMUs (black arrows). (D) After 6 weeks, healing was completely suppressed in the ALN groups, but continued normally in the VEH groups (B). (E,G) The combined ALN-PTH treatments triggered the development of multiple porosity areas (BMUs) that were significantly larger when ALN treatment was ceased (black circles). (F,H) In the combined ALN-PTH treatment, areas of erosion (black arrows) persisted after 6 weeks along the stress fracture despite the progression of healing in other areas (black circle), especially when ALN treatment was continued (toluidine blue 10X). Scale bar = 10 μ m.

Porosity parameters

There was no significant interaction between the effects of time and treatment type on any of the porosity parameters (Table 2).

There were no significant differences in porosity parameters between the different treatment groups. With regards to the main effect of time, porosity area, perimeter, and the porosity BMU area per mm^2 of the Ct.Ar increased significantly after 6 weeks in all treatment groups when compared with the second week ($p < 0.001$; Fig. 5H and I).

Discussion

Our results can be explained by understanding the mechanism of healing of SFx, which occurs through direct remodeling and starts from the periosteal exit point of the SFx and progresses along the SFx line. BMUs are formed in the first 2 weeks, and osteoclasts play a significant role at this initial stage of healing.⁽²²⁾ This is different from the repair process and healing of a complete fracture, which starts with an internal and external callus formation, with the external callus undergoing endochondral ossification for mineralization. Only after this initial phase of stabilization, bone remodeling starts to replace the lamellar bone.⁽⁴⁵⁾ The rat model used in this study is a standardized and reliable model that has been used by us previously⁽²²⁾; it facilitates the study of SFx remodeling. The time points chosen for

histomorphometric analysis of SFx remodeling were carefully chosen in light of the fact that 2 weeks is the most representative time point for the early (resorptive) phase of the remodeling cycle, whereas 6 weeks is the most representative time point for the later (formation) phase of remodeling.

We also pretreated rats with ALN for 14 days, after which rats were treated with PTH for 14 days in the ongoing presence of ALN or its cessation. Current PTH treatment protocols are 18 to 24 months,⁽⁴⁶⁾ which is approximately 4 weeks in a rat's life span.⁽⁴⁷⁾ We reduced the treatment window of PTH to the shortest possible period through administration of 14 daily injections, which is sufficient to induce stable (plateau) serum PTH levels.⁽⁴⁸⁾ The significant increase in osteoclast number along the SFx was expected at 2 weeks following PTH treatment.^(31,49,50) This may be explained by the PTH induction of monocyte chemoattractant protein-1, which is responsible for differentiation and recruitment of osteoclast precursors in early remodeling phases.^(51,52) The mechanism of the anabolic action of PTH is also supported by other factors, including calcium availability and the calcium sensing receptor.⁽⁵³⁾ The downregulation of sclerostin has also been linked to the anabolic PTH effect, through reduction of Sost expression, which negatively regulates Wnt signaling.^(54,55)

Our study is the first study to investigate the combined effect of intermittent PTH treatment with a concurrent or ceased ALN treatment on the overall remodeling of SFx. There have been inconsistent outcomes with regards to the combined

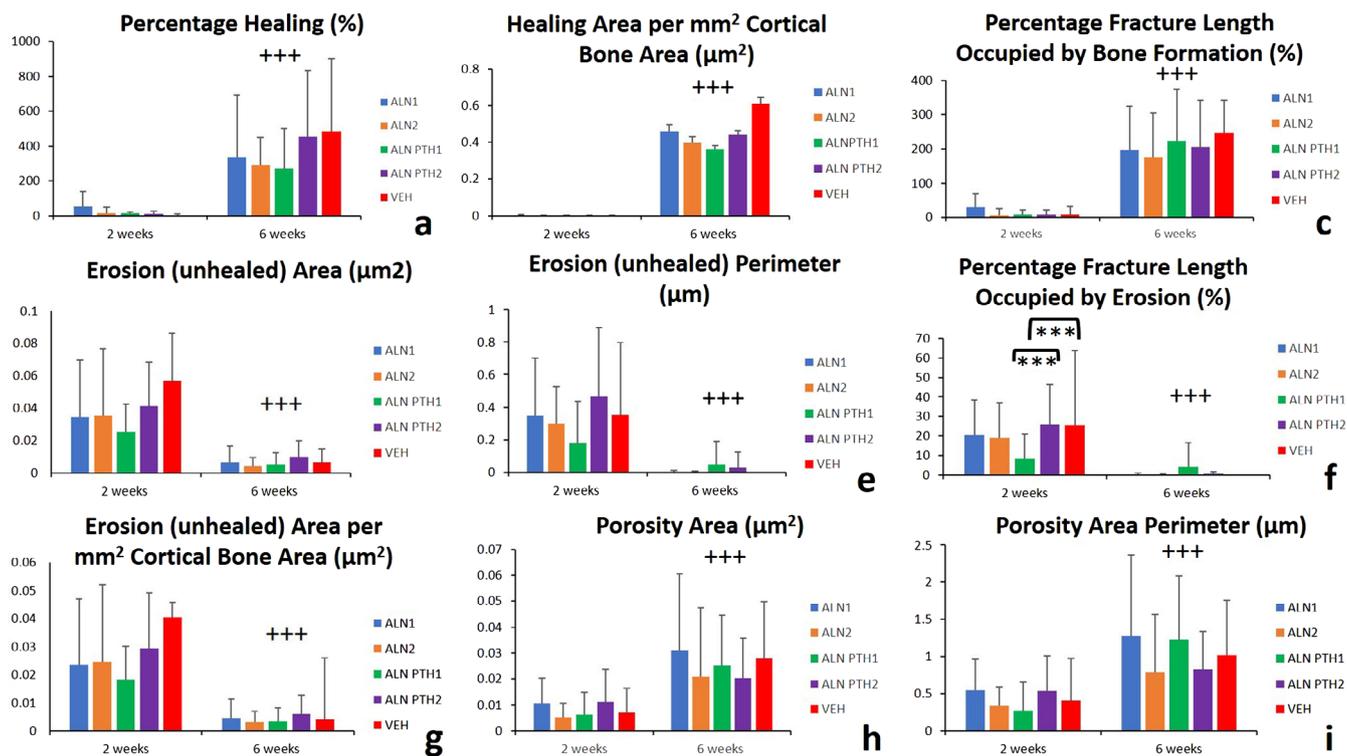


Fig 5. Histomorphometric variables of stress fracture healing (\pm SD). Continuation of alendronate in the combined alendronate-PTH₁ treatment mode resulted in significantly less percentage fracture length occupied by erosion after 2 weeks when compared with the vehicle and cessation of alendronate in the combined alendronate-PTH₂ treatment mode. Healing, porosity variables were significantly greater, whereas erosion variables were significantly less as a result of the main effect of time between the 2-week and 6-week time points. *** = $p \leq 0.01$ (differences between treatment types). ++ = $p \leq 0.01$ (differences compared with the previous time point).

antiresorptive and PTH therapies in the treatment of postmenopausal women with osteoporosis.^(32,33,56–59) For example, greater bone turnover was achieved in women with osteoporosis receiving antiresorptive therapy (ALN) for 18 months after cessation of that therapy and switching to PTH for an additional 18 months.⁽⁵⁶⁾ It was suggested in a previous study that the additive effect of the combined ALN-PTH treatment is attributable to increased bone formation with a combined osteoclast inhibitory action, implying that the anabolic effect of PTH might be independent of resorption.⁽⁶⁰⁾ Furthermore, in cases of combined treatment, decreasing the frequency of an antiresorptive agent administration results in an increase in the overall BMD.⁽⁵⁸⁾ It was also suggested that pretreatment with PTH, followed by an antiresorptive agent like ALN and continuing the combination therapy could improve treatment outcomes.⁽⁵⁹⁾ However, although this could be applied to chronic conditions such as osteoporosis, it is not feasible for SFx, where the timing of injury cannot be predicted beforehand to allow for sufficient time for pretreatment with PTH.

Contrary to our hypothesis, intermittent PTH treatment was more effective in SFx repair when ALN treatment was stopped. This is based on the greater recruitment and availability of osteoclasts that initiate the resorption phase of bone remodeling. Although the percentage healing of the SFx was greater following cessation of ALN, it was not significant at 6 weeks in this experiment. The current clinical treatment protocols for osteoporosis are based on long-term drug administration with chosen periods of cessation called “drug holidays.” The evidence for

the value of the drug holiday periods remains weak, and a careful evaluation for the risks versus benefits of cessation of BPs should be investigated thoroughly before making a decision.⁽⁶¹⁾ The optimal length of a drug holiday has not been established, but existing data suggest 3 to 5 years with ALN, 3 to 6 years with zoledronate, and 1 to 2 years with risedronate. A decision to recommence therapy should then probably be based on regular reassessment of BMD and fracture risk.^(62,63) New alternative drugs with a shorter duration of action like denosumab⁽⁴⁾ could provide a more reversible option as an antiresorptive agent and could potentially be beneficial for patients with SFx to prevent the risk of fractures, or at least allow for a quicker healing of an AFF caused by earlier activation of remodeling. Romosozumab (EVENTY; Amgen, Thousand Oaks, CA, USA), developed as a humanized monoclonal antibody against sclerostin,⁽⁶⁴⁾ could also support the formation phase of remodeling.

BPs are known to have a long residence time in bone, and to also be recycled and return to other resorptive surfaces even after the treatment has ceased.^(3,65) Limited data are available from BP discontinuation clinical trials, but suggest a residual and continuous biological effect of BPs, especially ALN and zoledronic acid for a lengthy period after discontinuation.⁽⁶⁶⁾ It was hypothesized that BP treatment causes tissue brittleness that initiates cracks, increases homogeneity of osteonal and interstitial structures, impairs targeted repair by BMUs, and allows easier accumulation of microdamage at areas of maximum force and mechanical loading.⁽⁶⁷⁾

In clinical studies, healing of nondisplaced AFF improved when BPs were ceased.⁽⁶⁸⁾ There is debate as to whether the improved healing of AFF was because of teriparatide treatment, the discontinuation of BPs, or both factors combined.⁽⁶⁸⁾ Furthermore, a reduction in the incidence of AFF by 70% per year was reported, when BPs were ceased. This was attributed to the osteoclastic bone resorption and repair of microcracks that follows the discontinuation of BP treatment.⁽⁶⁹⁾ iPTH has shown promising results in the treatment of BP-related AFFs.^(43,68,70–73) In one of the case reports, despite successful healing of AFF using teriparatide, the authors questioned that it had a major role in the healing process. It was hypothesized that surgical fixation, vitamin D therapy, calcium, and ALN discontinuation could have played a secondary role in the healing process.^(10,19,71,74) This aligns with the results from the current study where greater remodeling activation was observed following cessation of ALN.

In a recent case report, discontinuation of BPs and treatment with teriparatide resulted in a nearly complete radiographically verified healing of AFF, but unfortunately 12 months after the cessation of BPs, bilateral recurrence of AFF occurred, despite sequential use of teriparatide and denosumab.⁽⁷⁵⁾ This illustrates the difficulty of managing cases of SFx in association with antiresorptive therapy. That is, cessation of antiresorptive agents is favored for healing of SFx, but compromises the structural integrity of bone in osteoporotic patients, whereas continuation of antiresorptive treatment may delay healing and lead to atypical fractures. In the absence of randomized controlled trials on this question, a taskforce of the American Society for Bone and Mineral Research recommends discontinuation of BPs, adequate calcium, and vitamin D, and consideration of teriparatide for those who appear not to heal on conservative therapy.⁽¹⁵⁾ The above-mentioned results highlight the importance of the current findings related to SFx healing and note the variability of treatment outcomes as a limitation that could be related to the different animal model or unique clinical scenario of each individual case, as well as the different treatment protocols adopted in each study.

The higher values for woven bone parameters observed in this study is consistent with others,^(60,76) and dispels the questions concerning efficacy of PTH in the formation of new bone around sites that have been previously suppressed by BP treatment.^(32,33,57) For example, PTH therapy can enable new bone formation and replace the old bone matrix on a previously resorption-suppressed site caused by BP treatment.⁽⁷⁷⁾ Furthermore, PTH when compared with other anabolic agents like strontium ranelate, has a long-term stimulating effect on bone formation and resorption markers in postmenopausal women with osteoporosis previously treated with BPs.⁽⁷⁸⁾

The present study showed that the anabolic PTH action was not affected or inhibited by prior ALN treatment. However, continuation of ALN treatment with a concurrent PTH treatment depressed its anabolic effect, and the increased Wo.B.Wi was consistent with other studies.^(60,76) The efficacy of PTH in the formation of new bone around sites that have been previously suppressed by BP treatment had been questioned.^(32,33,57) Later, it was demonstrated that PTH could enable new bone formation and replace the old bone matrix on a previously resorption-suppressed site following BP treatment.⁽⁷⁷⁾ The increase in healing parameters in the ALN-PTH group following cessation of ALN was greater, but not statistically significant, at 6 weeks. Ma and colleagues⁽⁷⁹⁾ did show that cortical bone formation was lower in ALN-pretreated groups when compared with PTH-only groups. This was explained by the greater intensity of resorption

during remodeling activation that leads in sequence to greater formation. The earlier increase in osteoclastic activity was certainly observed in this study.

Our results show that after 6 weeks, daily PTH injections for 14 days did not elicit the same effect, consistent with activated modeling (formation). This is in alignment with a recent study that showed a higher bone volume with higher daily PTH doses as a result of a higher trabecular number in the healing callus and a denser trabecular bone network, indicating increased bone formation.⁽⁸⁰⁾ This opens the possibilities for investigating the effects of weekly rather than daily PTH injections, which has been investigated previously, but only in the treatment of AFF.⁽⁷³⁾ It also highlights the different effects on bone surfaces, including the woven bone callus, and the remodeling required to heal the SFx line.

Conclusion

It was concluded that cessation of ALN after SFx induction in the combined ALN-PTH treatment was effective in increasing osteoclast parameters. Daily iPTH injections for 14 days improved the woven bone apposition rate and osteoclast parameters. Other woven bone and healing parameters remained unaffected by the cessation or continuation of ALN. It is possible that relatively short periods of iPTH therapy could activate remodeling of SFx following treatment with BPs, as long it is ceased at the time of SFx induction.

Disclosures

The authors declare no conflicts of interest.

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AUTHOR CONTRIBUTIONS

Mahmoud Bakr: Conceptualization; data curation; formal analysis; investigation; methodology; project administration; validation; writing-original draft; writing-review and editing. **Wendy Kelly:** Data curation; investigation; project administration; writing-review and editing. **Athena Brunt:** Data curation; investigation; methodology; project administration; writing-review and editing. **Bradley Paterson:** Data curation; investigation; methodology; project administration. **Helen Massa:** Data curation; investigation; methodology; supervision; validation; writing-review and editing. **Nigel Morrison:** Conceptualization; formal analysis; funding acquisition; investigation; methodology; supervision; validation; writing-review and editing. **Mark**

Forword: Conceptualization; formal analysis; funding acquisition; investigation; methodology; project administration; supervision; validation; writing-review and editing.

References

1. Boden BP, Osbahr DC. High-risk stress fractures: evaluation and treatment. *J Am Acad Orthop Surg.* 2000;8(6):344–53.
2. Gehrmann RM, Renard RL. Current concepts review: stress fractures of the foot. *Foot Ankle Int.* 2006;27(9):750–7.
3. Russell RG, Watts NB, Ebetino FH, Rogers MJ. Mechanisms of action of bisphosphonates: similarities and differences and their potential influence on clinical efficacy. *Osteoporos Int.* 2008;19(6):733–59.
4. Reyes C, Hitz B, Prieto-Alhambra D, Abrahamsen B. Risks and benefits of bisphosphonate therapies. *J Cell Biochem.* 2016;117(1):20–8.
5. Cranney A, Tugwell P, Adachi J, et al. Meta-analysis of risedronate for the treatment of postmenopausal osteoporosis. *Endocr Rev.* 2002a;23(4):517–23.
6. Cranney A, Wells G, Willan A, et al. Meta-analysis of alendronate for the treatment of postmenopausal women. *Endocr Rev.* 2002b;23(4):508–16.
7. Schneider JP. Bisphosphonates and low impact femoral fractures: current evidence on alendronate-fracture risk. *Geriatrics.* 2009;64(1):18–23.
8. Adler RA. Atypical femoral fractures: risks and benefits of long-term treatment of osteoporosis with anti-resorptive therapy. *Eur J Endocrin.* 2018;178(3):R81–7.
9. Watts NB, Diab DL. Long-term use of bisphosphonates in osteoporosis. *J Clin Endocrinol Metab.* 2010;95(4):1555–65.
10. Kim JT, Jeong HJ, Lee SJ, Kim HJ, Yoo JJ. Adjuvant teriparatide therapy for surgical treatment of femoral fractures; does it work? *Hip Pelvis.* 2016;28(3):148–56.
11. Koh A, Guerado E, Giannoudis PV. Atypical femoral fractures related to bisphosphonate treatment. *Bone Joint J.* 2017;99(3):295–302.
12. Lim S-J, Yeo I, Yoon P-W, et al. Incidence, risk factors, and fracture healing of atypical femoral fractures: a multicenter case-control study. *Osteoporos Int.* 2018;29(11):2427–35.
13. Black DM, Abrahamsen B, Bouxsein ML, Einhorn T, Napoli N. Atypical femur fractures: review of epidemiology, relationship to bisphosphonates, prevention, and clinical management. *Endocr Rev.* 2019;40(2):333–68.
14. Oh BK, Heo YM, Yi JW, Kim TG, Lee JS. Atypical fracture of the proximal shaft of the ulna associated with prolonged bisphosphonate therapy. *Clin Orthop Surg.* 2018;10(3):389–92.
15. Shane S, Burr D, Abrahamsen B, et al. Atypical subtrochanteric and diaphyseal femoral fractures: second report of a task force of the American Society for Bone and Mineral Research. *J Bone Miner Res.* 2014;29(1):1–23.
16. Iwata K, Li J, Follet H, Phipps RJ, Burr DB. Bisphosphonates suppress periosteal osteoblast activity independently of resorption in rat femur and tibia. *Bone.* 2006;39(5):1053–8.
17. Vashishth D, Gibson GJ, Khoury JI, Schaffler MB, Kimura J, Fyhrie DP. Influence of nonenzymatic glycation on biomechanical properties of cortical bone. *Bone.* 2001;28(2):195–201.
18. Seeman E, Delmas PD. Reconstructing the skeleton with intermittent parathyroid hormone. *Trends Endocrinol Metab.* 2001;12(7):281–3.
19. Larsen MS, Schmal H. The enigma of atypical femoral fractures: a summary of current knowledge. *EFORT Open Rev.* 2018;3(9):494–500.
20. Lloyd AA, Gludovatz B, Riedel C, et al. Atypical fracture with long-term bisphosphonate therapy is associated with altered cortical composition and reduced fracture resistance. *Proc Natl Acad Sci U S A.* 2017;114(33):8722–7.
21. Schilcher J, Koeppen V, Aspenberg P, Michaelsson K. Risk of atypical femoral fracture during and after bisphosphonate use. *N Engl J Med.* 2014a;371(10):974–6.
22. Kidd LS, Stephens A, Kuliwaba J, Fazzalari N, Forwood MR. Temporal pattern of gene expression and histology of stress fracture healing in the rat ulna-loading model. *Bone.* 2010;46(2):369–78.
23. Schilcher J, Sandberg O, Isaksson H, Aspenberg P. Histology of 8 atypical femoral fractures. *Acta Orthop.* 2014b;85(3):280–6.
24. Dempster DW, Cosman F, Parisien M, Shen V, Lindsay R. Anabolic actions of parathyroid hormone on bone. *Endocr Rev.* 1993;14(6):690–709.
25. Schiller PC, D'Ippolito G, Roos BA, Howard GA. Anabolic or catabolic responses of MC3T3-E1 osteoblastic cells to parathyroid hormone depend on time and duration of treatment. *J Bone Miner Res.* 1999;14(9):1504–12.
26. Burr DB, Hirano T, Turner CH, Hotchkiss C, Brommage R, Hock JM. Intermittently administered human parathyroid hormone(1-34) treatment increases intracortical bone turnover and porosity without reducing bone strength in the humerus of ovariectomized cynomolgus monkeys. *J Bone Miner Res.* 2001;16(1):157–65.
27. Dempster D, Cosman F, Kurland ES, et al. Effects of daily treatment with parathyroid hormone on bone microarchitecture and turnover in patients with osteoporosis: a paired biopsy study. *J Bone Miner Res.* 2001;16(10):1846–53.
28. Seeman E, Duan Y, Fong C, Edmonds J. Fracture site-specific deficits in bone size and volumetric density in men with spine or hip fractures. *J Bone Miner Res.* 2001;16(1):120–7.
29. Andreassen TT, Ejersted C, Oxlund H. Intermittent parathyroid hormone (1–34) treatment increases callus formation and mechanical strength of healing rat fractures. *J Bone Miner Res.* 1999;14(6):960–8.
30. Jahng JS, Kim HW. Effect of intermittent administration of parathyroid hormone on fracture healing in ovariectomized rats. *Orthopedics.* 2002;23(10):1089–94.
31. Bakr MM, Kelly WL, Brunt AR, et al. Single injection of PTH improves osteoclastic parameters of remodeling at a stress fracture site in rats. *J Orthop Res.* 2019;37(5):1172–82.
32. Black DM, Greenspan SL, Ensrud KE, et al. The effects of parathyroid hormone and alendronate alone or in combination in postmenopausal osteoporosis. *N Engl J Med.* 2003;349(13):1207–15.
33. Finkelstein JS, Hayes A, Hunzelman JL, Wyland JJ, Lee H, Neer RM. The effects of parathyroid hormone, alendronate, or both in men with osteoporosis. *N Engl J Med.* 2003;349(13):1216–26.
34. Gasser JA, Kneissel M, Thomsen JS, Mosekilde L. PTH and interactions with bisphosphonates. *J Musculoskelet Neuronal Interact.* 2000;1(1):53–6.
35. Gasser J, Green J. Chronic subcutaneous, but not single intravenous, dosing of rats with bisphosphonates results in reduced anabolic response to PTH. *J Bone Miner Res.* 2006;21(Suppl 1):F386.
36. Ma YL, Bryant HU, Zeng Q, et al. New bone formation with teriparatide [human parathyroid hormone-(1–34)] is not retarded by long-term pretreatment with alendronate, estrogen, or raloxifene in ovariectomized rats. *Endocrinology.* 2003;144(5):2008–15.
37. Boonen S, Marin F, Obermayer-Pietsch B, et al. Effects of previous antiresorptive therapy on the bone mineral density response to two years of teriparatide treatment in postmenopausal women with osteoporosis. *J Clin Endocrinol Metab.* 2008;93(3):852–60.
38. Kidd LJ, Cowling NR, Wu ACK, Kelly WL, Forwood MR. Bisphosphonate treatment delays stress fracture remodeling in the rat ulna. *J Orthop Res.* 2011;29(12):1827–33.
39. Silva MJ, Touhey DC. Bone formation after damaging in vivo fatigue loading results in recovery of whole-bone monotonic strength and increased fatigue life. *J Orthop Res.* 2007;25(2):252–61.
40. Uthgenannt BA, Kramer MH, Hwu JA, Wopenka B, Silva MJ. Skeletal self-repair: stress fracture healing by rapid formation and densification of woven bone. *J Bone Miner Res.* 2007;22(10):1548–56.
41. Sloan AV, Martin JR, Li S, Li J. Parathyroid hormone and bisphosphonate have opposite effects on stress fracture repair. *Bone.* 2010;47(2):235–40.
42. Kidd LJ, Cowling NR, Wu ACK, Kelly WL, Forwood MR. Selective and non-selective cyclooxygenase inhibitors delay stress fracture healing in the rat ulna. *J Orthop Res.* 2013;31(2):235–42.

43. Miller PD, McCarthy EF. Bisphosphonate-associated atypical subtrochanteric femur fractures: paired bone biopsy quantitative histomorphometry before and after teriparatide administration. *Semin Arthritis Rheum.* 2015;44(5):477–82.
44. Mosley JR, March BM, Lynch J, Lanyon LE. Strain magnitude related changes in whole bone architecture in growing rats. *Bone.* 1997;20(3):191–8.
45. Li M, Liang H, Shen Y, Wronski TJ. Parathyroid hormone stimulates cancellous bone formation at skeletal sites regardless of marrow composition in ovariectomized rats. *Bone.* 1999;24(2):95–100.
46. Canalis E, Giustina A, Bilezikian JP. Mechanisms of anabolic therapies for osteoporosis. *N Engl J Med.* 2007;357(9):905–16.
47. Sengupta P. The laboratory rat: relating its age with human's. *Int J Prev Med.* 2013;4(6):624–30.
48. Li X, Liu H, Qin L, et al. Determination of dual effects of parathyroid hormone on skeletal gene expression in vivo by microarray and network analysis. *J Biol Chem.* 2007;282(45):33086–97.
49. Vrahnas C, Pearson TA, Brunt AR, et al. Anabolic action of parathyroid hormone (PTH) does not compromise bone matrix mineral composition or maturation. *Bone.* 2016;93:146–52.
50. Meakin LB, Todd H, Delisser PJ, et al. Parathyroid hormone's enhancement of bones' osteogenic response to loading is affected by ageing in a dose- and time-dependent manner. *Bone.* 2017;98:59–67.
51. Wu AC, Morrison NA, Kelly WL, Forwood MR. MCP-1 expression is specifically regulated during activation of skeletal repair and remodeling. *Calcif Tissue Int.* 2013;92(6):566–75.
52. Morrison NA, Day CJ, Nicholson GC. Dominant negative MCP-1 blocks human osteoclast differentiation. *J Cell Bio.* 2014;115(2):303–12.
53. Al-Dujaili SA, Koh AJ, Dang M, et al. Calcium sensing receptor function supports osteoblast survival and acts as a co-factor in PTH anabolic actions in bone. *J Cell Biochem.* 2016;117(7):1556–67.
54. O'Brien CA, Plotkin LI, Galli C, et al. Control of bone mass and remodeling by PTH receptor signaling in osteocytes. *PLoS One.* 2008;3:e2942.
55. Kramer I, Keller H, Leupin O, Kneissel M. Does osteocytic SOST suppression mediate PTH bone anabolism? *Trends Endocrin Metabol.* 2010;21(4):237–44.
56. Cosman F, Wermers RA, Recknor C, et al. Effects of teriparatide in postmenopausal women with osteoporosis on prior alendronate or raloxifene: differences between stopping and continuing the antiresorptive agent. *J Clin Endocrinol Metab.* 2009;94(10):3772–80.
57. Finkelstein JS, Wyland JJ, Lee H, Neer RM. Effects of teriparatide, alendronate, or both in women with postmenopausal osteoporosis. *J Clin Endocrinol Metab.* 2010;95(4):1838–45.
58. Cosman F, Eriksen EF, Recknor C, et al. Effects of intravenous zoledronic acid plus subcutaneous teriparatide [rhPTH(1–34)] in postmenopausal osteoporosis. *J Bone Miner Res.* 2011;26(3):503–11.
59. Cosman F, Nieves JW, Dempster DW. Treatment sequence matters: anabolic and antiresorptive therapy for osteoporosis. *J Bone Miner Res.* 2017;32(2):198–202.
60. Altman-Singles AR, Tseng W, de Bakker CMJ, et al. A closer look at the immediate trabecula response to combined parathyroid hormone and alendronate treatment. *Bone.* 2014;61:149–57.
61. Ye Y, Mou Y, Bai B, Li L, Chen G-P, Hu S-J. Knockdown of farnesylpyrophosphate synthase prevents angiotensin II-mediated cardiac hypertrophy. *Int J Biochem Cell Biol.* 2010;42(12):2056–64.
62. Anagnostis P, Stevenson JC. Bisphosphonate drug holidays – when, why and for how long? *CLIMACTERIC.* 2015;18(Suppl 2):32–8.
63. Anagnostis P, Paschou SA, Mintzioria G, et al. Drug holidays from bisphosphonates and denosumab in postmenopausal osteoporosis: EMAS position statement. *Maturitas.* 2017;101:23–30.
64. Markham A. Romosozumab: first global approval. *Drugs.* 2019;79(4):471–6.
65. Diab DL, Watts NB. Bisphosphonate drug holiday: who, when and how long. *Ther Adv Musculoskelet Dis.* 2013;5(3):107–11.
66. Black DM, Schwartz AV, Ensrud KE, et al. Effects of continuing or stopping alendronate after 5 years of treatment: the fracture intervention trial long-term extension (FLEX): a randomized trial. *JAMA.* 2006;296(24):2927–38.
67. Ettinger B, Burr D, Ritchie R. Proposed pathogenesis for atypical femoral fractures: lessons from materials research. *Bone.* 2013;55(2):495–500.
68. Watts NB, Aggers D, McCarthy EF, et al. Responses to treatment with teriparatide in patients with atypical femur fractures previously treated with bisphosphonates. *J Bone Miner Res.* 2017;32(5):1027–33.
69. Schilcher J, Koepfen V, Aspenberg P, Michaelsson K. Risk of atypical femoral fracture during and after bisphosphonate use. *Acta Orthopaedica.* 2015;86(1):100–7.
70. Carvalho NN, Voss LA, Almeida MO, Salgado CL, Bandeira F. Atypical femoral fractures during prolonged use of bisphosphonates: short-term responses to strontium ranelate and teriparatide. *J Clin Endocrinol Metab.* 2011;96(9):2675–80.
71. Huang HT, Kang L, Huang PJ, et al. Successful teriparatide treatment of atypical fracture after long-term use of alendronate without surgical procedure in a postmenopausal woman: a case report. *Menopause.* 2012;19(12):1360–3.
72. Chiang CY, Zebaze RM, Ghasem-Zadeh A, Iuliano-Burns S, Hardidge A, Seeman E. Teriparatide improves bone quality and healing of atypical femoral fractures associated with bisphosphonate therapy. *Bone.* 2013;52(1):360–5.
73. Tsuchie H, Miyakoshi N, Iba K, et al. The effects of teriparatide on acceleration of bone healing following atypical femoral fracture: comparison between daily and weekly administration. *Osteoporos Int.* 2018;29(12):2659–65.
74. Gomberg SJ, Wustrack RL, Napoli N, Arnaud CD, Black DM. Teriparatide, vitamin D, and calcium healed bilateral subtrochanteric stress fractures in a postmenopausal woman with a 13-year history of continuous alendronate therapy. *J Clin Endocrinol Metab.* 2011;96(6):1627–32.
75. Ramchand SK, Chiang CY, Zebaze RM, Seeman E. Recurrence of bilateral atypical femoral fractures associated with the sequential use of teriparatide and denosumab: a case report. *Osteoporos Int.* 2016;27(2):821–5.
76. Altman-Singles AR, Jeong Y, Tseng W, et al. Intermittent parathyroid hormone after prolonged alendronate treatment induces substantial new bone formation and increases bone tissue heterogeneity in ovariectomized rats. *J Bone Miner Res.* 2017;32(8):1703–15.
77. Kim SW, Pajevic PD, Selig M, et al. Intermittent parathyroid hormone administration converts quiescent lining cells to active osteoblasts. *J Bone Miner Res.* 2012;27(10):2075–84.
78. De Sousa IO, Diniz ET, Marques TF, Griz L, Coutinho M, Bandeira F. Short-term bone marker responses to teriparatide and strontium ranelate in patients with osteoporosis previously treated with bisphosphonates. *Arq Bras Endocrinol Metab.* 2010;54(2):244–9.
79. Ma YL, Zeng QQ, Chiang AY, et al. Effects of teriparatide on cortical histomorphometric variables in postmenopausal women with or without prior alendronate treatment. *Bone.* 2014;59:139–47.
80. Milstrey A, Wieskoetter B, Hinze D, et al. Dose-dependent effect of parathyroid hormone on fracture healing and bone formation in mice. *J Surg Res.* 2017;220:327–35.