

Draft Genome Sequences of Two Multidrug-Resistant Acinetobacter baumannii Strains of Sequence Type ST92 and ST96

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The global epidemiology of multidrug-resistant *Acinetobacter baumannii* is dominated by a limited number of clones. Here, we announce the draft genome sequences of two multidrug-resistant *A. baumannii* strains, 1H8 and 4A3, representing the major epidemic clones, sequence type 92 (ST92) and ST96, respectively.

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A cinetobacter baumannii is a major nosocomial pathogen that displays a remarkable ability to acquire mechanisms that confer resistance to virtually all antibiotics (1, 2). Analysis of the population structure of *A. baumannii* has shown that the global emergence of multidrug resistance in this organism is predominated by a number of widely distributed clones (2). In Hong Kong hospitals, the increase in multidrug-resistant *A. baumannii* is mainly attributed to the expansion of two clones, sequence type 92 (ST92) and ST96 (3, 4). ST92 is a pandemic clone with distribution in >30 countries (2), while ST96 is a recently identified clone that has so far been geographically restricted to certain parts of China (2, 4).

In this report, we present the draft genomes of two *A. baumannii* strains, 1H8 (ST92) and 4A3 (ST96), originating from clinical samples of two hospitalized patients in Hong Kong, China (4). The bacterial genomes were sequenced on a GS FLX system (Roche Diagnostics, Basel, Switzerland). A total of 263,241 reads from strain 4A3 (ST96) and 247,690 reads from strain 1H8 (ST92) were generated at approximately 23-fold genome coverage for both strains. *De novo* genome assembly was performed using the Newbler assembler 2.7 (Roche Diagnostics). The contigs were then oriented and ordered into scaffolds using OSLay (5). Gene identification and automatic functional annotation were performed using the RAST (Rapid Annotations using Subsystem Technology) server and Blast2GO (6, 7). The antibiotic resistomes in the genomes were identified using the Antibiotic Resistance Genes Database (8).

The genome assembly of strain 4A3 (ST96) consists of 136 contigs, with a G+C content of 38.9%, a total length of 4,104,065 bp, and 3,916 protein-coding sequences. The genome assembly of strain 1H8 (ST92) consists of 69 contigs, with a G+C content of 39.0%, a total length of 3,993,780 bp, and 3,774 protein-coding sequences. In the genomes of strain 4A3 (ST96) and strain 1H8 (ST92), there were 41 and 49 antimicrobial resistance genes, respectively. These include efflux pumps (ABC, major facilitator superfamily [MFS], small multidrug resistance [SMR], and resistance to a wide range of compounds and genes associated with specific resistance to β -lactams (class A, C, and D β -lactamases), chloramphenicol (*cat, cml*), aminoglycosides (phosphotransferases, adenyl-transferases, nucleotidyltransferases, and dimethyladenosine trans-

ferases), macrolides-lincosamides-streptogramins (*macB*, *mph2*, *msrA*, *vat*, *vgaA*), tetracyclines (*tetB*), trimethoprim (*dhfr*), and sulphonamides (*sul*). Overall, the availability of the present genome sequences facilitates further comparative genomic and bioinformatics analysis in *A. baumannii* populations.

Nucleotide sequence accession numbers. This Whole-Genome Shotgun project has been deposited at NCBI GenBank under the accession no. AOLU00000000 (4A3) and ANNC00000000 (1H8). The genome sequences described in this paper are the first versions, AOLU01000000 (4A3) and ANNC01000000 (1H8).

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REFERENCES

- Ho PL, Cheng VC, Chu CM. 2009. Antibiotic resistance in communityacquired pneumonia caused by *Streptococcus pneumoniae*, methicillinresistant *Staphylococcus aureus*, and *Acinetobacter baumannii*. Chest 136: 1119–1127.
- Karah N, Sundsfjord A, Towner K, Samuelsen Ø. 2012. Insights into the global molecular epidemiology of carbapenem non-susceptible clones of *Acinetobacter baumannii*. Drug Resist. Updat. 15:237–247.
- Bartual SG, Seifert H, Hippler C, Luzon MA, Wisplinghoff H, Rodríguez-Valera F. 2005. Development of a multilocus sequence typing scheme for characterization of clinical isolates of *Acinetobacter baumannii*. J. Clin. Microbiol. 43:4382–4390.
- 4. Ho PL, Ho AY, Chow KH, Lai EL, Ching P, Seto WH. 2010. Epidemiology and clonality of multidrug-resistant *Acinetobacter baumannii* from a healthcare region in Hong Kong. J. Hosp. Infect. 74:358–364.
- Richter DC, Schuster SC, Huson DH. 2007. OSLay: optimal syntenic layout of unfinished assemblies. Bioinformatics 23:1573–1579.
- Conesa A, Götz S, García-Gómez JM, Terol J, Talón M, Robles M. 2005. Blast2GO: a universal tool for annotation, visualization and analysis in functional genomics research. Bioinformatics 21:3674–3676.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: rapid annotations using subsystems technology. BMC Genomics 9:75.
- Liu B, Pop M. 2009. ARDB—antibiotic resistance genes database. Nucleic Acids Res. 37(Database issue):D443–D447. doi:10.1093/nar/gkn656.