High frequency of antimicrobial resistance and virulence gene in *Shigella* species isolated from pediatric patients in an Iranian Referral Hospital

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Abstract. Background: Shigella is a main cause of gastroenteritis and it is responsible for 5 to 10% of diarrhea through the world. The aims of this study were to assess the antibiotic susceptibility pattern and the presence of 3 common virulence genes (sigA, virF, invE) of Shigella strains isolated from patients with gastroenteritis in Children's Medical Center Hospital, Tehran, Iran. Methods: Over a period of 15 months, all Shigella species collected from the patients with gastroenteritis were entered to the study. Susceptibility testing of all isolates towards different antibiotics was performed using the disk diffusion method and the prevalence of virulence genes was detected by polymerase chain reaction (PCR) technology. Results: Among a total of 183 Shigella strains, 128 Shigella sonnei (70%) and 55 S. flexneri (30%) were isolated. The resistance rate to the antibiotics in S. sonnei strains was higher than S. flexneri. The most sensitive antibiotics for S. flexneri strains were gentamicin (98%), amikacin (85%) and ciprofloxacin (82%), while high resistance rate to trimethoprim-sulfamethoxazole (96%), ampicillin (96%), nalidixic acid (64%) and cefotaxime (60%) was observed. The frequency of *invE*, *virF* and *sigA* gene in S. flexneri strains was 89 %, 93 % and 56 %, respectively; whereas they found in 93 %, 96 %, and 100 % of S. sonnei strains, respectively. SigA gene was identified significantly higher in the S. sonnei strains (100%). There was no significant difference between the presence of virF and invE genes among Shigella strains. Conclusion: The high presence of sigA gene in S. sonnei strains plays an important role in its pathogenesis, and the high frequency of *invE* and *virF* genes showed that this classical pathway regulating the expression of *Shigella* virulence factor genes could play a key role in the pathogenesis of this bacterium. (www.actabiomedica.it)

Key words: Shigella, children, antibiotics resistance, virulence factors

Introduction

Shigellosis continues to be a main public health problem worldwide, mainly in developing countries where it is endemic (1, 2) and it is considered as a prominent global cause of moderate to severe diarrhea in children (3) and adults (4). Children under five years of age face the biggest impact of the disease and 61% of deaths occur in children (5). Unfortunately, poor hygienic circumstances and low quality of water in developing countries enhance the incidence and prevalence of the disease (6). The genus *Shigella* includes four subgroups historically treated as species: *Shigella flexneri*, *Shigella boydii*, *Shigella sonnei*, and *Shigella dysenteriae* (7), among which, *S. flexneri* is the most prevailing species in developing nations followed by *S. sonnei*, whereas *S. boydii* and *S. dysenteriae* are less frequently isolated (8).

Although shigellosis is a self-limiting disease, antibiotics might decrease the duration of illness and consequently reduce the person to person transmission (9). Unfortunately, irregular usage of antimicrobial agents in addition to horizontal gene transfer, have given rise to the increasing resistance of *Shigella spp.* and the growth of multi-drug resistance against common antibiotics worldwide (10, 11). *Shigella* capacity to cause disease is based on genes contained in an invasion plasmid pINV of 220 Kb, such as *ipaH*, *ipaBCD*, *ial*, *sen*, *virA*, *virB* (*invE*), *virF*, *icsA*, *sepA*, and *ipgD* and on chromosomal genes, *ipaH*, *iuc*, *sat*, *sigA*, *pic*, *set1A*, and *set1B* (12, 13).

The aim of this study was to investigate the antibiotic susceptibility pattern and the distributation of three common virulence genes (*sigA*, *virF*, *invE*) of Shigella species isolated from patients with gastroenteritis in Tehran, Iran.

Material and methods

The study was approved by the Ethical Committee from the Tehran University of Medical Sciences, Iran (IR.TUMS.CHMC.REC.1397.008). In our cross-sectional study over a period of 15 months, from September 2018 to February 2020, all *Shigella* species collected from the patients with gastroenteritis were entered to the study. The isolates were identified by standard biochemical tests (14). Susceptibility testing of all *Shigella* isolates towards different antibiotics was performed using the disk diffusion method based on Clinical Laboratory Standard Institute (CLSI) guidelines, 2019 (15). The antimicrobials tested were: nalidixic acid, amikacin, ampicillin, gentamicin, cefotaxime, ciprofloxacin, and trimethoprim sulfamethoxazole.

In order to detect virulence genes, DNA template was obtained as method previously described by Hosseini Nave *et al.* (16). PCR was performed to target the virulence genes (*sigA*, *virF*, *invE*) by using previously reported primers (Table 1).

Amplification was performed in a mixture consisting of 2.5 µl of the PCR buffer (10-times concentrated), 0. 5 µl of Mgcl2 (final concentration 200 µM), 0.5 µl of dNTPs (Fermentas, Vilnius, Lithuania, final concentration 2.5 mM), 0.5 µl of each primer, final concentration of 0.2 mM, 1.5 U of the Tag DNA polymerase (Bioron, Germany), 1 µl of DNA (final concentration 2 ng/µL) and DNase-, RNase-free deionised water (Biomedicals) to a final volume of 25 µl. Cycling conditions were carried out as follows: initial denaturation at 95° C for 7 min, followed by 30 cycles including denaturation for 5 min at 95° C, annealing for 45 s and a single final extension at 72° C for 15 min. The analysis of the amplified products was performed in 1% agarose (Sigma) and DNA bands were visualized by staining with gel red (Biotium), analysed under UV light and photographed using the GEL Doc 2000 documentation system (Bio-Rad).

Statistical Analysis

Data were analyzed using the SPSS version 16.0 software (SPSS Inc., Chicago, IL, United States) and the results were described by frequency (percentage) and mean ± standard deviation (SD). Univariate analysis was performed using the chi-squared test or Fisher's exact test, as appropriate. P-values were based

Gene	Primer sequence (5' –3')	Size of product (bp)	Annealing temperature (° C)	Reference
sigA-forward	CCGACTTCTCACTTTCTCCCG	420	50	(21)
sigA-reverse	CCATCCAGCTGCATAGTGTTTG	430	39	
virF-forward	TCAGGCAATGAAACTTTGAC	(10	ĒG	(21)
virF-reverse	TGGGCTTGATATTCCGATAAGTC	018	30	
invE-forward	CGATAGATGGCGAGAAATTATATCCCG	766	60	(37)
invE-reverse	CGATCAAGAATCCCTAACAGAAGAATCAC	700	00	

Table 1. Primers used in this study

on two-tailed test results, and P < 0.05 were considered statistically significant.

Results

In this study, 183 *Shigella* strains (2.6%) were isolated from a total of 7121 children with gastroenteritis referred to the Children's Medical Center Hospital, Tehran, Iran during the period of 15 months.

The most isolated species were *S. sonnei* with 128 cases (70%) and *S. flexneri* with 55 cases (30%). Among 183 patients whose stool culture was positive for *Shigella* bacteria, 93 patients were boys (51%) with a mean age of 5.7 years old (SD= 3.4 years, the age range of 1 to 16 years old). There was no significant difference between the distribution of *Shigella* bacteria in the children with gastroenteritis by age (p value=0.5) and sex (p value=0.87). The highest rate of *Shigella* isolates was isolated in autumn and from outpatients (n=108, 59%) and among the different wards of the hospital, emergency department had the highest rate of *Shigella* (35%).

The results of antibiotic susceptibility test showed high resistance rate of *Shigella* strains to ampicillin (p value ≥ 0.05) and trimethoprim sulfamethoxazole (p value=0.09). Generally, the resistance rate to cefotaxime, nalidixic acid, ciprofloxacin and ampicillin in *S. sonnei* strains was significantly higher than *S. flexneri* (Table 2).

The prevalence of *invE*, *virF* and *sigA* genes in *S. flexneri* strains was 89%, 93% and 56%, and in *S. sonnei* strains was 93%, 96% and 100%, respectively (Table 2). The *sigA* gene was significantly more detected in *S. sonnei* strains (p value \leq 0.0001). However, no significant difference was observed between *virF* and *invE* gene detection in *Shigella* strains.

Discussion

Gastroenteritis is considered as one of the most vital diseases all around the world and it is more severe and dangerous among children, the elderly, and people who are malnourished or live in poor conditions. *Shigella* is a main cause of gastroenteritis throughout the world and it is responsible for 5 to 10% of diarrhea through the world (17). It can be considered as a major pathogen in developing countries with lower levels of hygiene (17, 18).

In our study, 70% of the strains were *S. sonnei* and 30% of them were confirmed as *S. flexneri*. *S. flexneri* is responsible for most of the shigellosis burden in developing countries worldwide, while *S. sonnei* occurs predominantly in developed countries and in countries shifting from low- to middle-income (19, 20). The reason for this discrepancy is not clear; however, efforts to increase local health have drastically reduced the prevalence of the disease and even changed the distribution pattern of *Shigella* species (21, 22). This pattern change has also been observed in countries such as Brazil (23), South America (24) and China (25), which is similar to the results of studies conducted in our country in the cities of Tehran (22, 26-28), Babol (29) and Abadan (30).

In developing countries where the prevalence of shigellosis is usually reported endemically, evaluating the pattern of antibiotic resistance can be very effective in prescribing appropriate drugs. The resistance rate to the antibiotics studied in our study in *S. sonnei* strains was higher than *S. flexneri*, which was in consistent with previous studies (22, 31). *Shigella* strains showed high sensitivity to aminoglycosides in the present study. The most sensitive antibiotics for *S. flexneri* strains were gentamicin (98%), amikacin (85%) and ciprofloxacin (82%). While high resistance pattern to trimethoprim-sulfamethoxazole (96%),

Table 2. Antibiotic susceptibility pattern and frequency of the virulence genes in Shigella isolates

	Antibiotics						Genes			
					Trimethoprim	Nalidixic				
Bacteria	Amikacin	Gentamycin	Ampicillin	Cefotaxime	Sulfamethoxazole	acid	Ciprofloxacin	invE	sigA	virF
S. sonnei	127 (100%)	122 (96.1%)	6 (4.7%)	6 (4.7%)	0	3 (2.3%)	72 (56.7%)	119 (93%)	128 (100%)	123 (96.1%)
S. flexneri	46 (85.2%)	53 (98.1%)	2 (3.7%)	22 (40%)	2 (3.6%)	20 (36.4%)	40 (81.6%)	49 (89.1%)	31 (56.4%)	51 (92.7%)

ampicillin (96%), nalidixic acid (64%) and cefotaxime (60%) was observed. Several studies around the world have reported increased resistance of *Shigella* species to common antibiotics such as trimethoprim, sulfamethoxazole, and ampicillin (23, 32, 33).

Virulence of Shigella depends on the presence of a large virulence inv plasmid, carrying an operon that encodes the type III-secretion-system (T3SS) responsible for bacterial entry (34, 35). In our study, sigA gene was significantly identified more frequent in S. sonnei strains (p value ≤ 0.0001). The high frequency of sigA gene in S. sonnei strains plays a key role in its pathogenesis, which is consistent with previous studies (7, 16, 36). However, no significant difference was observed between virF and invE genes identification in S. flexneri and sonnei strains. When Shigella growth conditions are suitable for invasion, a transcription cascade begins by activating the *virF* gene to express the AraC-like protein virF, which in turn activates the transcription of the *invE* regulatory gene (36). The high abundance of these genes indicated that this classical regulatory pathway of Shigella virulence gene expression might play a major role in its pathogenesis.

In conclusion, due to the overuse of antibiotics and the consequent increase in drug resistance, some antibiotics should be removed from the list of drugs for the treatment of *Shigella*, which include ampicillin and trimethoprim sulfamethoxazole. The lowest pattern of resistance in the present study was observed to gentamicin, amikacin and ciprofloxacin. The high presence of *sigA*, *invE* and *virF* genes showed that this classical regulatory pathway of *Shigella* virulence factor gene expression can play a major role in the pathogenesis of this bacterium.

Acknowledgments: This study was taken from Dr. Mohammad Ghaffari's postgraduate thesis.

Funding: This project was supported by a grant (grant number: 97-01-88-32526) from Tehran University of Medical Sciences to Dr. Setareh Mamishi.

Conflicts of interest: Each author declares that he or she has no commercial associations (e.g. consultancies, stock ownership, equity interest, patent/licensing arrangement etc.) that might pose a conflict of interest in connection with the submitted article.

Ethical Approval statement: The study was approved by the Ethical Committee from the Tehran University of Medical Sciences, Iran (IR.TUMS.CHMC.REC.1397.008).

Authors' Contribution: SM1 and RHS involved in writing of the manuscript. BP, MGH, and MRA participated in data collection. SM1 and SM2 involved in funding and interpretation of data and supervised the project. BP supervised the project and involved in interpretation of data. All authors discussed the results and contributed to the final manuscript.

References

- 1. Medeiros PHQS, Lima AÂM, Guedes MM, et al. Molecular characterization of virulence and antimicrobial resistance profile of Shigella species isolated from children with moderate to severe diarrhea in northeastern Brazil. Diagn Microbiol Infect Dis 2018;90(3):198-205.
- The HC, Thanh DP, Holt KE, Thomson NR, Baker S. The genomic signatures of Shigella evolution, adaptation and geographical spread. Nat Rev Microbiol 2016;14(4):235-50.
- Bardhan P, Faruque AS, Naheed A, Sack DA. Decrease in shigellosis-related deaths without Shigella spp.-specific interventions, Asia. Emerg Infect Dis 2010;16(11):1718-23.
- Lozano R, Naghavi M, Foreman K, et al. Global and regional mortality from 235 causes of death for 20 age groups in 1990 and 2010: a systematic analysis for the Global Burden of Disease Study 2010. Lancet 2012;380(9859):2095-128.
- Jain PA, Kulkarni RD, Dutta S, et al. Prevalence and antimicrobial profile of *Shigella* isolates in a tertiary care hospital of North Karnataka: A 12-year study. Indian J Med Microbiol 2020;38(1):101-108.
- 6. Eshaghi Zadeh SH, Fahimi H, Fardsanei F, Soltan Dallal MM. Antimicrobial Resistance and Presence of Class 1 Integrons Among Different Serotypes of Salmonella spp. Recovered From Children with Diarrhea in Tehran, Iran. Infect Disord Drug Targets 2020;20(2):160-166.
- 7. da Cruz CB, de Souza MC, Serra PT, Santos I, Balieiro A, Pieri FA, Nogueira PA, Orlandi PP. Virulence factors associated with pediatric shigellosis in Brazilian Amazon. Biomed Res Int 2014;2014:539697.
- Nüesch-Inderbinen M, Heini N, Zurfluh K, Althaus D, Hächler H, Stephan R. Shigella antimicrobial drug resistance mechanisms, 2004–2014. Emerg Infect Dis 2016;22(6):1083.
- Sethuvel DPM, Anandan S, Michael JS, et al. Virulence gene profiles of Shigella species isolated from stool specimens in India: its association with clinical manifestation and antimicrobial resistance. Pathog Glob Health 2019;113(4):173-9.
- Davies J, Davies D. Origins and evolution of antibiotic resistance. Microbiol Mol Biol Rev 2010;74(3):417-33.
- 11. Salimiyan Rizi K, Farsiani H, Sasan MS. High rate of resistance to ceftriaxone and azithromycin among Shigella

spp. isolates at three children's referral hospitals in Northeast Iran. J Infect Chemother 2020;26(9):955-958.

- Bastos FC, Loureiro EC. Antimicrobial resistance of Shigella spp. isolated in the State of Pará, Brazil. Rev Soc Bras Med Trop. 2011;44(5):607-10.
- Gonzales JC, Seribelli AA, Gomes CN, et al. A high number of multidrug-resistant and predominant genetically related cluster of Shigella flexneri strains isolated over 34 years in Brazil. Braz J Microbiol 2020;51(4):1563-1571.
- Mahon CR, Lehman DC, Manuselis G. Textbook of diagnostic microbiology-E-Book: Elsevier Health Sciences 2018;
- <Melvin P. Performance Standards for Antimicrobial Susceptibility Testing 2019.pdf>.
- 16. Hosseini Nave H, Mansouri S, Emaneini M, Moradi M. Distribution of genes encoding virulence factors and molecular analysis of Shigella spp. isolated from patients with diarrhea in Kerman, Iran. Microb Pathog 2016;92:68-71.
- Ahmed AM, Furuta K, Shimomura K, Kasama Y, Shimamoto T. Genetic characterization of multidrug resistance in Shigella spp. from Japan. J Med Microbiol. 2006;55(Pt 12):1685-1691.
- Shakoor S, Zaidi AK, Hasan R. Tropical bacterial gastrointestinal infections. Infect Dis Clin 2012;26(2):437-53.
- Malau E, Ford R, Valcanis M, et al. Antimicrobial sensitivity trends and virulence genes in Shigella spp. from the Oceania region. Infect Genet Evol. 2018;64:52-56
- 20. Taneja N, Mewara A. Shigellosis: epidemiology in India. The Indian journal of medical research. 2016;143(5):565.
- 21. Fan W, Qian H, Shang W, et al. Low distribution of genes encoding virulence factors in Shigella flexneri serotypes 1b clinical isolates from eastern Chinese populations. Gut Pathogens 2017;9(1):76.
- 22. Mahmoudi S, Pourakbari B, Moradzadeh M, et al. Prevalence and antimicrobial susceptibility of Salmonella and Shigella spp. among children with gastroenteritis in an Iranian referral hospital. Microb Pathog 2017;109:45-8.
- 23. Sousa MÂB, Mendes EN, Collares GB, Péret-Filho LA, Penna FJ, Magalhães PP. Shigella in Brazilian children with acute diarrhoea: prevalence, antimicrobial resistance and virulence genes. Memórias do Instituto Oswaldo Cruz 2013;108(1):30-5.
- 24. Orrett FA. Prevalence of Shigella serogroups and their antimicrobial resistance patterns in southern Trinidad. Journal of health, population, and nutrition 2008;26(4):456.
- 25. Qu F, Bao C, Chen S, et al. Genotypes and antimicrobial profiles of Shigella sonnei isolates from diarrheal patients circulating in Beijing between 2002 and 2007. Diagn Microbiol Infect Dis 2012;74(2):166-70.
- 26. MoezArdalan K, Zali MR, Dallal MMS, Hemami MR, Salmanzadeh-Ahrabi S. Prevalence and pattern of antimicrobial resistance of Shigella species among patients with acute diarrhoea in Karaj, Tehran, Iran. J Health Popul Nutr 2003:96-102.
- 27. Pourakbari B, Mamishi S, Mashoori N, et al. Frequency and antimicrobial susceptibility of Shigella species isolated

in Children Medical Center Hospital, Tehran, Iran, 2001-2006. Braz J Infect Dis 2010.

- Mamishi S, Mashoori N, Mahboobi N, Pour Akbari B. Increasing resistance to nalidixic acid in Shigella subgroups in a comparative study between 2001-2003 and 2004-2006. Singapore Med J 2009;50(8):791-3. PMID: 19710978.
- BARARI SKR, AHMADPOUR KM. Prevalence of Shigella species and their antimicrobial resistance patterns at Amirkola Children's Hospital, North of Iran 2007.
- 30. Jomezadeh N, Babamoradi S, Kalantar E, Javaherizadeh H. Isolation and antibiotic susceptibility of Shigella species from stool samples among hospitalized children in Abadan, Iran. Gastroenterol Hepatol Bed Bench 2014;7(4):218-23.
- Mamishi S, Arab ZY, Mahmoudi S, Moradzadeh M, Taghi MHA, Pourakbari B. Antimicrobial-resistance pattern of Shigella species in children: a six-year study in an Iranian referral Hospital. Ann Ig Med Preventiva Comunita 2019;31(4):356-64.
- 32. Aggarwal P, Uppal B, Ghosh R, Krishna Prakash S, Chakravarti A, Rajeshwari K. True Prevalence of Shigellosis in Indian Children with Acute Gastroenteritis: Have We Been Missing the Diagnosis? J Res Health Sci 2016;16(1):11-6.
- 33. Asrat D. Shigella and Salmonella serogroups and their antibiotic susceptibility patterns in Ethiopia. East Mediterr Health J 2008;14(4):760-7.
- Coster TS, Hoge CW, VanDeVerg LL, et al. Vaccination against shigellosis with attenuated Shigella flexneri 2a strain SC602. Infection and immunity 1999;67(7):3437-43.
- 35. Sansonetti PJ. Rupture, invasion and inflammatory destruction of the intestinal barrier by Shigella, making sense of prokaryote-eukaryote cross-talks. FEMS Microbiol Rev 2001;25(1):3-14
- 36. Gu B, Fan W, Qin T, et al. Existence of virulence genes in clinical Shigella sonnei isolates from Jiangsu Province of China: a multicenter study. Ann Transl Med 2019;7(14):305.
- Bonkoungou I, Lienemann T, Martikainen O, et al. Diarrhoeagenic Escherichia coli detected by 16-plex PCR in children with and without diarrhoea in Burkina Faso. Clin Microbiol Infect 2012;18(9):901-6.

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Received: 18 October 2020

Accepted: 13 November 2020

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