NEDD4L contributes to ferroptosis and cell growth inhibition in esophageal squamous cell carcinoma by facilitating xCT ubiquitination

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Supplementary results

Supplementary Tables

Table S1. Small interfering RNA sequence for NEDD4L and xCT.

	Sequence (5'-3') forward	Sequence (5'-3') reverse
siNEDD4L#1	CCCUAUACAUUUAAGGACUdTdT	AGUCCUUAAAUGUAUAGGGdTdT
siNEDD4L#2	GGAGAAUUAUGUCCGUGAAdTdT	UUCACGGACAUAAUUCUCCdTdT
sixCT	CCAGGUGGUUUAGAAUAAUTT	AUUAUUCUAAACCACCUGGTT
siControl	UUCUCCGAACGUGUCACGUTT	ACGUGACACGUUCGGAGAATT

Table S2. E3 ligases interact with xCT

E3	E3GENE	SUB	PFAM	GO	NET	MOTIF	SCORE	Confidence
								interaction
Q96PU5	NEDD4L	Q9UPY5	1.00	5.73	1.00	6.61	0.829	High
Q86TM6	SYVN1	Q9UPY5	1.00	1.25	1.00	6.61	0.714	Middle
Q86YT6	MIB1	Q9UPY5	1.00	2.33	1.00	2.80	0.693	Middle
P46934	NEDD4	Q9UPY5	1.00	5.73	1.00	1.00	0.681	Middle
P62879	GNB2	Q9UPY5	1.00	5.73	1.00	1.00	0.681	Middle
P22681	CBL	Q9UPY5	1.00	3.98	1.00	1.00	0.671	Middle
Q9UKV5	AMFR	Q9UPY5	1.00	1.51	1.00	2.80	0.652	Low
P43034	PAFAH1B1	Q9UPY5	1.00	3.77	1.00	1.00	0.640	Low
O14512	SOCS7	Q9UPY5	1.00	2.88	1.29	1.00	0.639	Low
Q9ULV8	CBLC	Q9UPY5	1.00	2.88	1.29	1.00	0.639	Low
Q8TCQ1	MARCH1	Q9UPY5	1.00	1.25	1.00	2.80	0.633	Low
Q8WY64	MYLIP	Q9UPY5	1.00	1.25	1.00	2.80	0.633	Low
Q5T0T0	MARCH8	Q9UPY5	1.00	1.25	1.00	2.80	0.633	Low
P14373	TRIM27	Q9UPY5	1.00	1.51	1.00	2.12	0.624	Low
Q99732	LITAF	Q9UPY5	1.00	2.88	1.00	2.12	0.613	Low
Q9P1Y6	PHRF1	Q9UPY5	1.00	2.88	1.00	2.12	0.613	Low
Q9UNE7	STUB1	Q9UPY5	1.00	2.88	1.00	2.12	0.613	Low
Q8TBB1	LNX1	Q9UPY5	1.00	2.88	1.00	2.12	0.613	Low
Q86Y01	DTX1	Q9UPY5	1.00	2.88	1.00	2.12	0.613	Low
P62873	GNB1	Q9UPY5	1.00	2.88	1.00	2.12	0.613	Low

Supplementary Figures

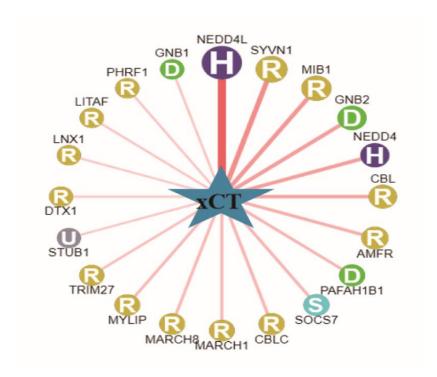


Figure S1. UbiBrowser database showed NEDD4L is the highest confidence to interact with xCT.

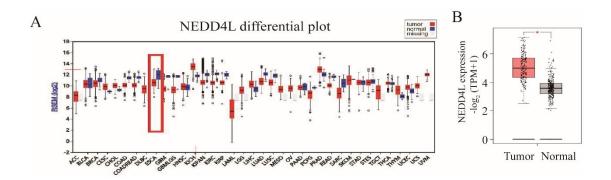


Figure S2. NEDD4L expression in public database. A Analysis of TCGA database showed that NEDD4L mRNA levels were significantly decreased in esophageal cancer compared with normal esophageal tissue. B GEPIA database results indicated that NEDD4L mRNA levels were increased in esophageal cancer (n = 182) compared to normal tissue (n = 286).

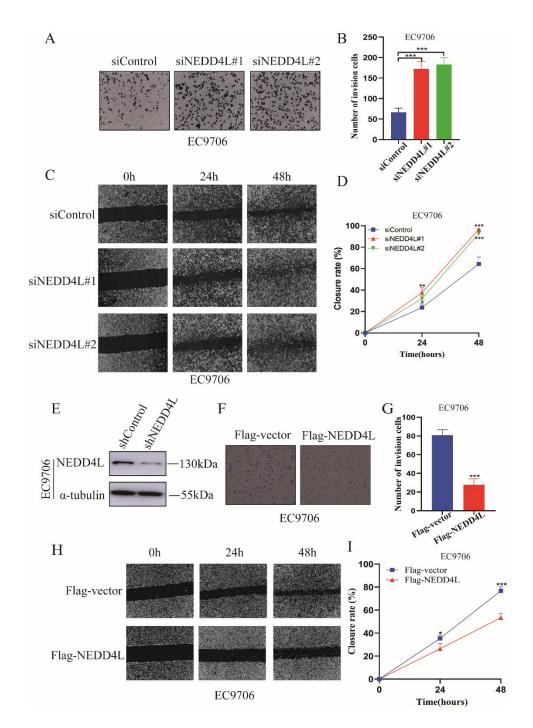


Figure S3. The impact of silencing NEDD4L and overexpressing NEDD4L on invasion capacity and wound closure speed in ESCC cells. A, B silencing NEDD4L promoted cell invasion in EC9706 cells. n = 3 per group. C, D silencing NEDD4L promoted wound closure speed in EC9706 cells. n = 3 per group. E shNEDD4L transfection efficiency in EC9706 cell line. F, G Overexpression NEDD4L suppressed cell invasion in EC9706 cells. n = 3 per group. H, I Overexpression

NEDD4L inhibited wound closure speed in EC9706 cells. n = 3 per group. The data are presented as the mean \pm SD. Statistical significance was determined by one-way ANOVA. *p < 0.05; **p < 0.01; ***p < 0.001.

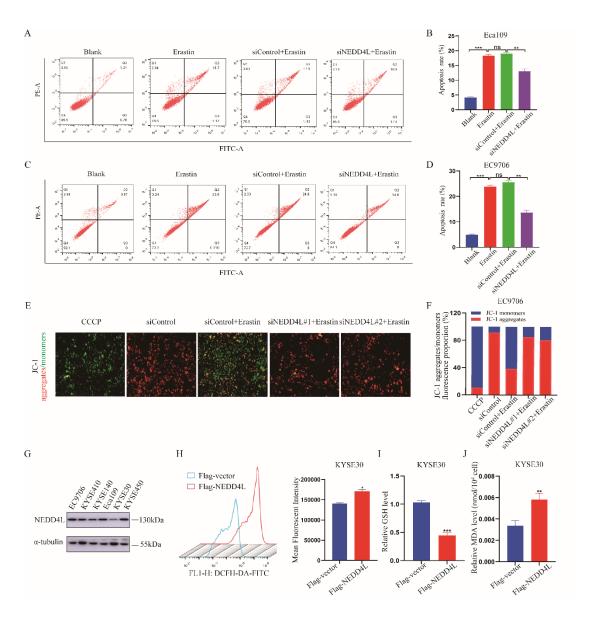


Figure S4. The influence of silencing NEDD4L and overexpressing NEDD4L on ferroptosis. A-D Cell death rate after treated with erastin for 24 h and transfected with siControl/siNEDD4L for 48 h were detected in Eca109 and EC9706 cells. n = 3 per group. **E, F** The effect of silencing NEDD4L on MMP under the treatment of erastin in

EC9706 cells was detected by optical microscopy. **G** NEDD4L expression in EC9706, Eca109, KYSE30, KYSE140, KYSE410 and KYSE450 cells was measured by western blotting. **H** ROS level in KYSE30 cells transfected with Flag-vector or Flag-NEDD4L plasmids for 48 h was detected. n=3 per group. **I** GSH level in KYSE30 cells transfected with Flag-vector or Flag-NEDD4L plasmids for 48 h was detected. n=3 per group. **J** MDA content in KYSE30 cells transfected with Flag-vector or Flag-NEDD4L plasmids for 48 h was detected. n=3 per group. The data are presented as the mean \pm SD. Statistical significance in **B**, **D**, **F** were determined by one-way ANOVA. *p < 0.05; **p < 0.01; ***p < 0.0

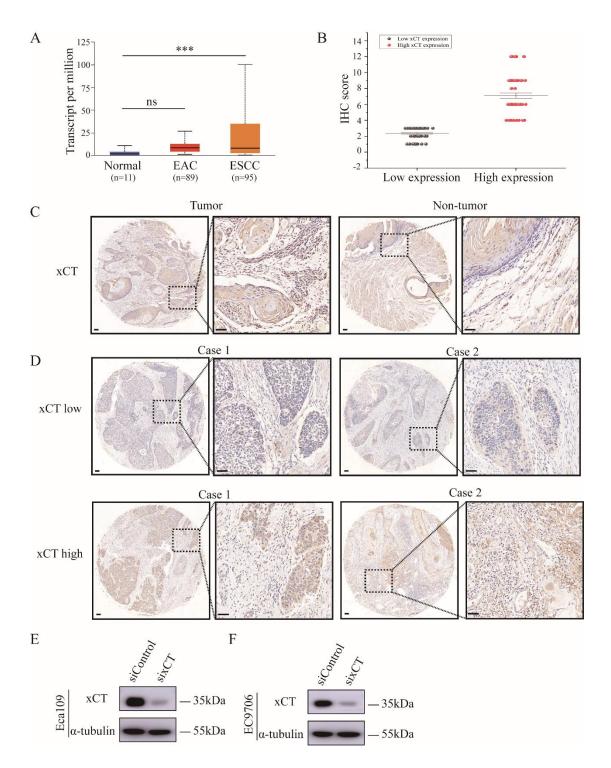


Figure S5. The expression and IHC score of xCT in 142 ESCC cases.

A UALCAN database results indicated that xCT mRNA levels is significantly increased in ESCC (n = 95) compared to normal tissue (n = 11). **B** xCT expression score in 142 cases of ESCC (n = 142). **C** xCT protein expression was increased in ESCC tissues (n = 25) compared with normal tissue (n = 25) as analyzed by IHC. Scale bars, 100 μ m. **D**

Representative images of xCT low expression and high expression analyzed by IHC (n = 142). Scale bars, 100 μ m. **E, F** xCT silencing efficiency in Eca109 and EC9706 cells were evaluated by western blotting.

Supplementary materials and methods

Wound healing assay

EC9706 cells were transfected with 50 nM NEDD4L siRNA or 5 μ g Flag-NEDD4L plasmids for 24 h. Then, cells were transfer into 24-well plates and cultured with 1% FBS. 10 μ L pipette tip was used for scratching when the cell density reached 100%. Wound distance was measured at 0 hour, 24 hours and 48 hours and normalized with the distance at 0 hour. Image J software was used to analyse the recovery speed.

Invasion assay

EC9706 cells were transfected with 50 nM NEDD4L siRNA or 5μg NEDD4L plasmids. To induce invasion, Matrigel (356234, BD) was applied to the upper chamber. Following a 12-hour incubation period, the cells were delicately extracted. After that, cells were fixed by 4% paraformaldehyde and stained by crystal violet. Image J software was used for counting the number of invasive cells.

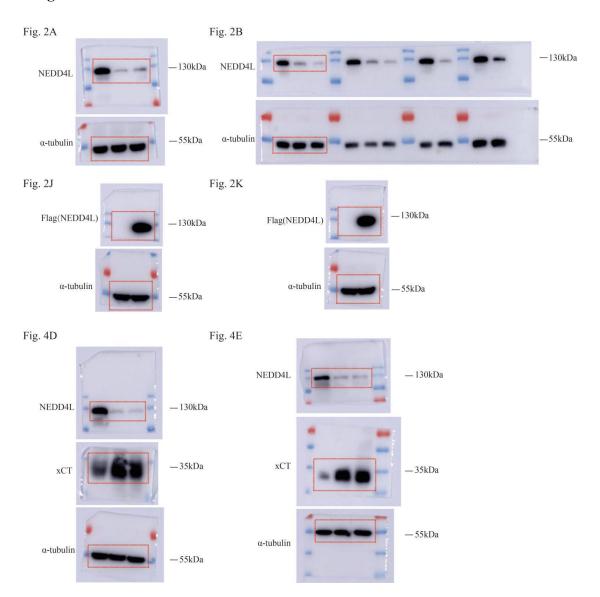
Cell death assay

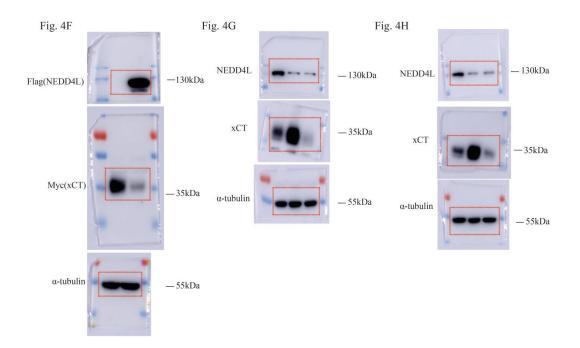
Eca109 and EC9706 cells transfected with siNEDD4L/siControl were treated with erastin (20 μ M) for 24 h. Then, cells were collected and re-suspended with staining buffer containing 5 μ L Annexin V, FITC and 5 μ L PI solution (AD10, DOJINDO). Afterwards, the cells incubated at room temperature in dark for 15 minutes, followed by detection using flow cytometry (Beckman, America).

Mitochondrial membrane potential (MMP) assay

JC-1 kit (C2006, Beyotime) was used to detect MMP. EC9706 cells were transfected with 50 nM siNEDD4L or siControl for 24 h. Subsequently, erastin with final concentration of 20 μM was added into the wells. After 24 h treatment, cells were gently washed with PBS. 1mL of fresh culture medium with JC-1 dyeing working solution (25 μM) were added and mixed well. In addition, cells were treated with carbonyl cyanide-m-chloro phenylhydrazone (CCCP) at a concentration of 10 μM for 20 minutes as a positive control. Afterwards, the cells were incubated in the cell incubator at 37°C in a dark environment for 20 min, followed by being washed twice with 1× JC-1 staining buffer. Then, 2 mL fresh culture medium was added, and fluorescence intensity was observed by a fluorescence microscope.

Original western blot





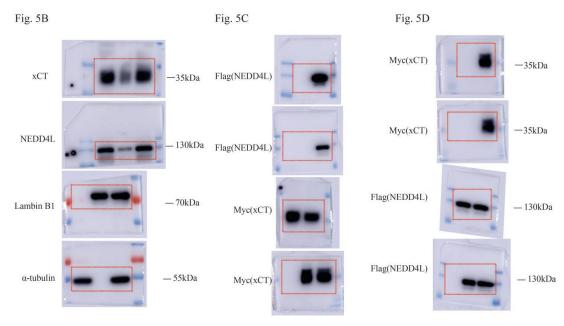
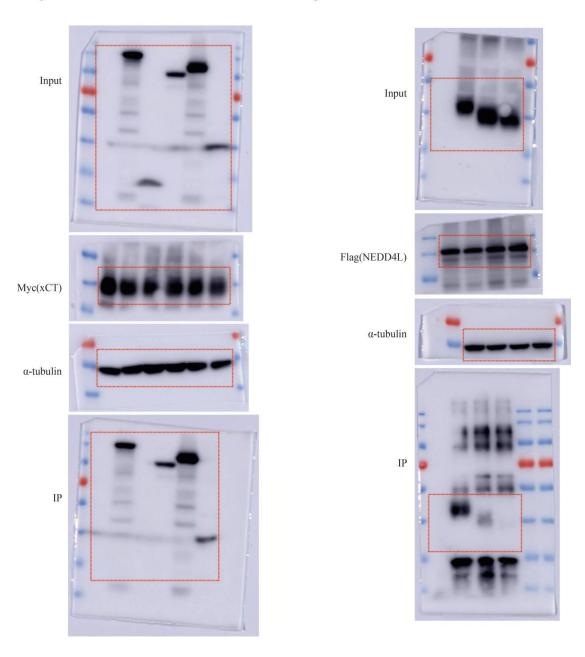
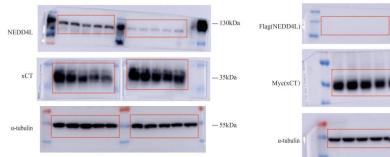
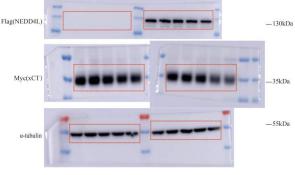
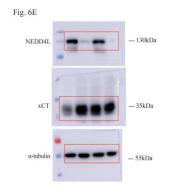


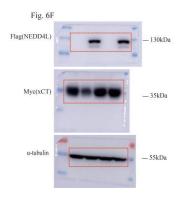
Fig. 5G Fig. 5H

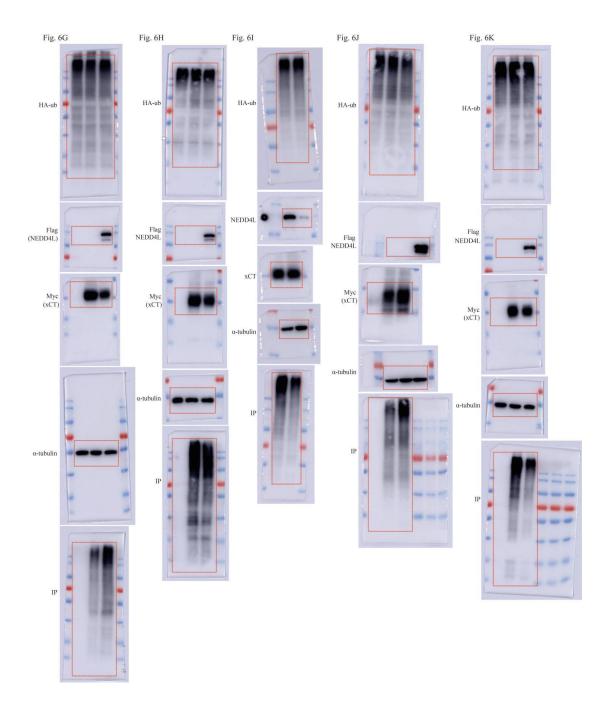












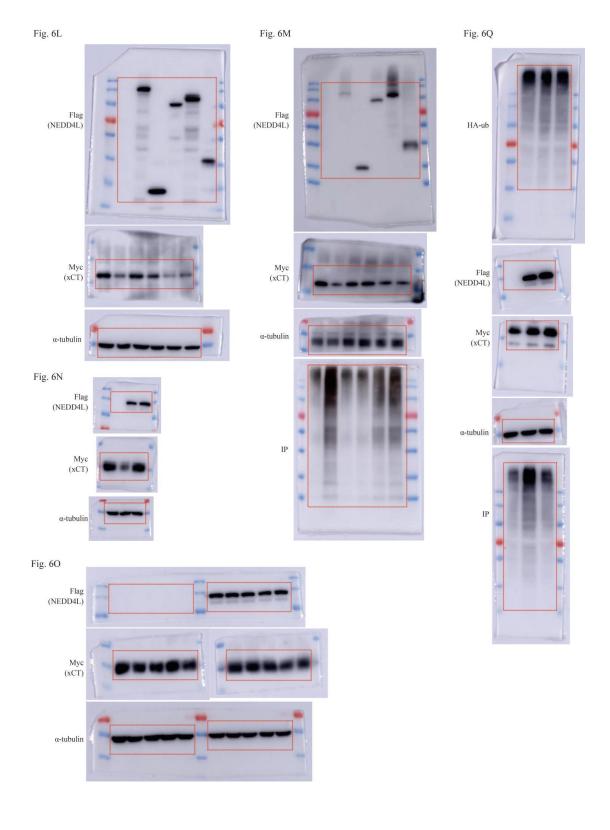


Fig. S4G

