

RESEARCH

Open Access

# New clinical and molecular insights on Barth syndrome

Lorenzo Ferri<sup>1</sup>, Maria Alice Donati<sup>2</sup>, Silvia Funghini<sup>3</sup>, Sabrina Malvagia<sup>3</sup>, Serena Catarzi<sup>1</sup>, Licia Lugli<sup>4</sup>, Luca Ragni<sup>5</sup>, Enrico Bertini<sup>6</sup>, Frédéric M Vaz<sup>7</sup>, David N Cooper<sup>8</sup>, Renzo Guerrini<sup>1,9</sup> and Amelia Morrone<sup>1\*</sup>

## Abstract

**Background:** Barth syndrome (BS) is an X-linked infantile-onset cardioskeletal disease characterized by cardiomyopathy, hypotonia, growth delay, neutropenia and 3-methylglutaconic aciduria. It is caused by mutations in the *TAZ* gene encoding tafazzin, a protein involved in the metabolism of cardiolipin, a mitochondrial-specific phospholipid involved in mitochondrial energy production.

**Methods:** Clinical, biochemical and molecular characterization of a group of six male patients suspected of having BS. Three patients presented early with severe metabolic decompensation including respiratory distress, oxygen desaturation and cardiomyopathy and died within the first year of life. The remaining three patients had cardiomyopathy, hypotonia and growth delay and are still alive. Cardiomyopathy was detected during pregnancy through a routine check-up in one patient. All patients exhibited 3-methylglutaconic aciduria and neutropenia, when tested and five of them also had lactic acidosis.

**Results:** We confirmed the diagnosis of BS with sequence analysis of the *TAZ* gene, and found five new mutations, c.641A>G p.His214Arg, c.284dupG (p.Thr96Aspfs\*37), c.678\_691del14 (p.Tyr227Trpfs\*79), g.8009\_16445del8437 and g.[9777\_9814del38; 9911-?\_14402del] and the known nonsense mutation c.367C>T (p.Arg123Term). The two gross rearrangements ablated *TAZ* exons 6 to 11 and probably originated by non-allelic homologous recombination and by Serial Replication Slippage (SRS), respectively. The identification of the breakpoints boundaries of the gross deletions allowed the direct detection of heterozygosity in carrier females.

**Conclusions:** Lactic acidosis associated with 3-methylglutaconic aciduria is highly suggestive of BS, whilst the severity of the metabolic decompensation at disease onset should be considered for prognostic purposes. Mutation analysis of the *TAZ* gene is necessary for confirming the clinical and biochemical diagnosis in probands in order to identify heterozygous carriers and supporting prenatal diagnosis and genetic counseling.

**Keywords:** Barth syndrome, *TAZ* gene mutation, *In utero* cardiomyopathy, Metabolic decompensation, Lactic acidosis, 3-methylglutaconic aciduria, Gross deletions, Metabolic cardiomyopathy

## Background

The X-linked Barth syndrome (BS; OMIM #302060) is a cardioskeletal myopathy that manifests in early infancy with cardiomyopathy, hypotonia, growth delay, and neutropenia [1,2]. Barth syndrome is caused by mutations in the tafazzin (*TAZ*) gene on Xq28 and is generally associated with 3-methylglutaconic aciduria [3]. It also represents one of the neuromuscular

disorders most frequently associated with left ventricular non-compaction (LVNC) [4].

Heart failure is the main cause of death in infancy followed by sepsis due to neutropenia [5], which causes some variability in the clinical course of BS [6]. Until ten years ago, mortality peaked within the first 4 years of life but over the last decade has shifted to around puberty. Improved survival is probably due to prompt diagnosis together with early and aggressive treatment of infections and cardiomyopathy [7].

BS is caused by alterations to the mitochondrial metabolism of cardiolipin (CL), a dimeric phosphoglycerolipid

\* Correspondence: a.morrone@meyer.it

<sup>1</sup>Department of Neurosciences, Psychology, Pharmacology and Child Health, University of Florence and Paediatric Neurology Unit and Laboratories, Meyer Children's Hospital, Viale Pieraccini n. 24, Florence 50139, Italy  
Full list of author information is available at the end of the article

present predominantly in mitochondrial membranes [8,9]. CL is involved in the mitochondrial electron transport chain, playing an essential role in cellular energy metabolism, mitochondrial dynamics and the inception of apoptotic pathways [10-13]. Recently, CL has been shown to be critical for the correct organization of ATP synthase assembly [10]. CL deficiency has been detected in several tissues and cell types from BS patients [9,14,15] and it has been shown to affect both the organization and morphology of the mitochondrial inner membrane [10].

Tafazzins are phospholipid-lysophospholipid transacylases which promote structural uniformity and molecular symmetry among the cardiolipins [16,17]. Inhibition of the tafazzin pathway leads to ultrastructural and functional alteration of the mitochondria [16,18,19]. Studies carried out on a BS zebrafish model suggest that the expression of tafazzins is both tissue-specific and age-dependent, and plays an essential role in cardiac development and function [20]. A rat tafazzin knockdown model exhibits hypertrophy ventricular myocytes at birth [21]. Recently, two murine models of tafazzin deficiency were produced by inducible short hairpin shRNA-mediated *TAZ* knockdown [22,23]. Both models showed decrease of tetralineoleyl cardiolipin and accumulation of monolysocardiolipins in cardiac and skeletal muscle, associated with mitochondrial abnormalities and cardiac and skeletal muscle impairment, similar to BS [22,23].

Several *TAZ* transcript variants encoding different tafazzin isoforms have been described [16,24,25], but the function of several of them is still unknown. Four distinct tafazzin isoforms, *TAZ-FL* (encoding full-length tafazzin, GenBank:NP\_000107.1), *TAZ-Δ5* (encoding tafazzin lacking exon 5, GenBank:NP\_851828.1), *TAZ-Δ7* (encoding tafazzin lacking exon 7, GenBank:NP\_851829.1) and *TAZ-Δ5Delta;7* (encoding tafazzin lacking exons 5 and 7, GenBank:NP\_851830.1) have been reported [26,27]. All four proteins are localized to the mitochondria if expressed in HeLa cells [28], but only two of them, *TAZ-FL* and *TAZ-Δ5*, have transacylase activity [16,28] with different acyl chain substrate specificity [16].

Mutations in the *TAZ* gene cause tafazzin deficiency and sequence analysis of this gene is necessary to confirm the clinical and biochemical diagnosis of BS [24,29]. The human *TAZ* gene contains 11 short exons and 10 variably long introns. At present, 105 different *TAZ* gene mutations have been reported [30,31], 94 of them associated with BS [30]. However, no correlation between the genotype and either cardiac phenotype or disease severity has been reported in BS [32]. A database of human *TAZ* gene mutations and other variants is available on-line from the Barth Syndrome Foundation

[33]. Here we report five new *TAZ* gene mutations in six unrelated BS patients, including two new gross gene rearrangements.

## Methods

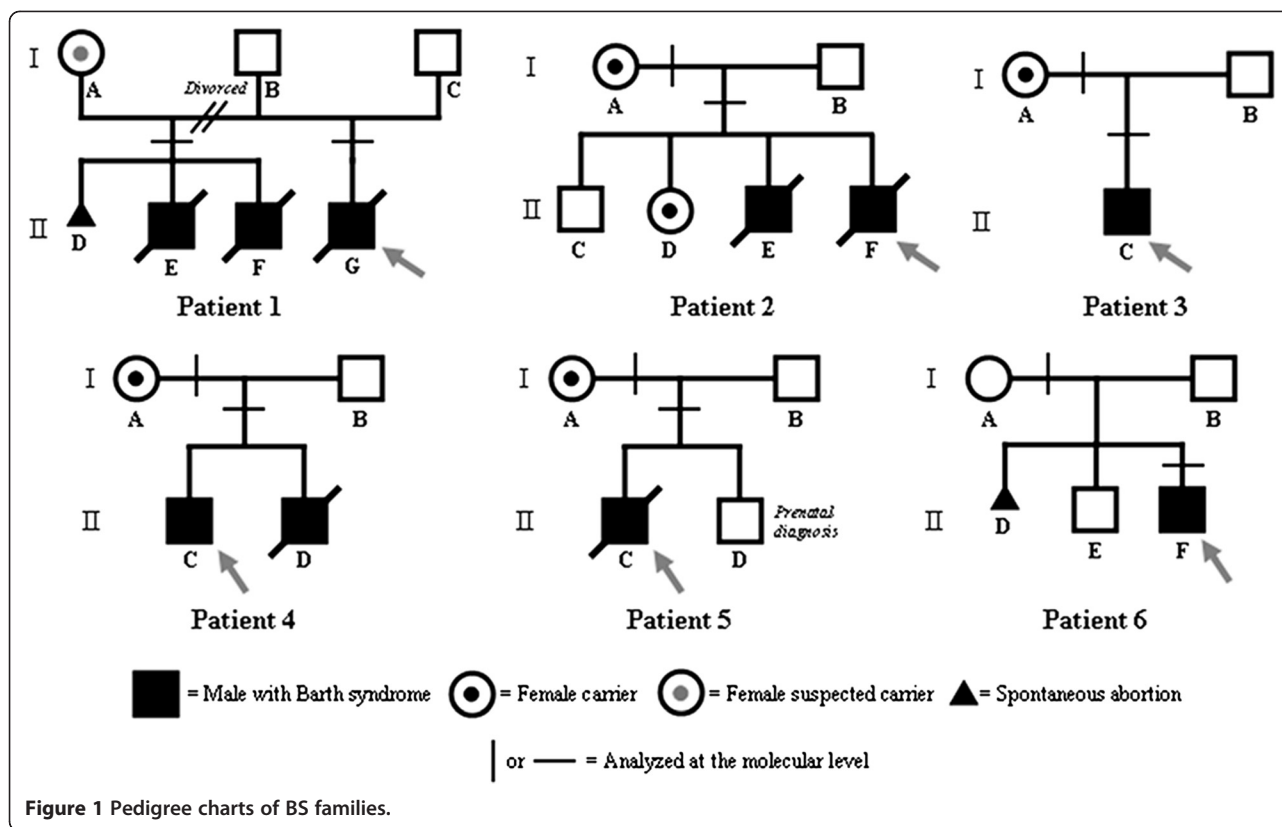
### Patients and controls

Whole blood DNA samples from six BS patients were examined. The heterozygous carrier status of the patients' mothers was evaluated after informed consent was obtained for all patients, in accordance with local ethical committee recommendations. The clinical histories of the patients analyzed are as follows and the pedigree charts are shown in Figure 1:

**Patient 1 (Pt1)** was the first child born to non-consanguineous parents after 36 weeks of gestation. Weight at birth was 2.25 kg with length 47 cm. On his first day of life, Pt1 manifested respiratory distress, lactic acidosis and relative neutropenia (neutrophil count, 900/mm<sup>3</sup>). Two days later, a severe dilated cardiomyopathy was diagnosed by heart ultrasound. Since severe hypotonia has also been presented, muscle biopsy was performed and revealed clearly reduced staining for cytochrome C oxidase, which was confirmed by a reduction in cytochrome c oxidase activity measured spectrophotometrically in muscle homogenate (44% reduction in activity by reference to the lowest range). This child died 10 days after birth due to heart failure. His mother was on her second marriage. During her first marriage, she had experienced a spontaneous abortion during her first pregnancy whilst two sons had subsequently died from heart failure at the ages of 2½ months and 40 days. Suspicion of BS, and the subsequent confirmation of the diagnosis by cardiolipin/monolysocardiolipin analysis [16] (MLCL/CL ratio 253 on patient's fibroblasts; normal 0–0.3), arose only after the birth of Pt1.

**Patient 2 (Pt2)** was born in the 39th week of gestation, with a spontaneous delivery after an uneventful pregnancy. APGAR was 1': 8; 5': 9; weight was 2.7 kg and length 47 cm. After 12 h of life, the newborn exhibited mild cyanosis with O<sub>2</sub> Sat of 88-89%, which was moderately responsive to O<sub>2</sub> administration (O<sub>2</sub> Sat 92-94 %), lactic acidosis, neutropenia and dilated cardiomyopathy with left ventricular non-compaction. Biochemical analysis of urinary organic acids revealed 3-methylglutaconic aciduria. This patient died at the age of one year due to heart failure. He was the youngest of four siblings, one healthy male, one sister and a brother who had died aged 40 days due to staphylococcal pneumonia. Autopsy of this patient's brother also revealed dilated cardiomyopathy. The clinical features of Pt2 and his family history led to suspicion of BS.

**Patient 3 (Pt3)** was the first child born with spontaneous delivery to non-consanguineous parents in the 39th week of gestation. Dilated cardiomyopathy with increased



thickness of the left ventricle had been revealed during pregnancy as a result of a routine check-up. At birth, APGAR was 1': 8; 5': 9. Further medical investigations confirmed dilated cardiomyopathy with left ventricular non-compaction, but the patient exhibited good cardiovascular compensation. However, lactic acidosis was present and urinary organic acids analysis revealed 3-methylglutaconic aciduria, with abnormally high levels of pyruvic acid, 3-hydroxybutyric acid, methylglutaric acid, 4-hydroxyphenyl lactic acid and p-hydroxyphenyl pyruvic acid. Neutropenia was noticed, according to age appropriate ranges, on several full blood counts over a period of eight years. Diagnosis of BS was eventually confirmed by molecular and biochemical analysis (MLCL/CL ratio 70 on patient's fibroblasts; normal 0–0.3). Pt3 exhibits intermittent neutropenia and dilated cardiomyopathy, but stable cardiovascular compensation. Occasionally, infections have occurred with fever but have been successfully treated with specific antibiotic therapies. To date, the treatment with G-CSF has not been carried out because the patient has never presented severe infections resistant to antibiotic therapy. He is currently eight years old, 20 kg (3<sup>rd</sup> percentile) in weight and 115.5 cm in height (under the 2<sup>nd</sup> percentile).

**Patient 4 (Pt4)** was the first child of non-consanguineous parents. Weight at birth was 2.8 kg. At the age of five months, he had a first episode of hypotonia and heart

failure with gallop rhythm and his weight was under the 3<sup>rd</sup> percentile. Dilated cardiomyopathy was diagnosed. Early therapeutic treatment allowed stabilization of clinical conditions. Severe neutropenia and urinary 3-methylglutaconic aciduria were also detected and confirmed during subsequent evaluations. During his life, the child has been in a good physical condition, but neurological milestones were delayed. He is currently eleven years old, 34.2 kg in weight (25<sup>th</sup> percentile), 142 cm in height (25<sup>th</sup> percentile) and exhibits dilated cardiomyopathy. BS was suspected and then confirmed by molecular analysis during his first physical examination, when he was 10 years old. A second son was born when Pt4 was 5 years old. The younger brother weighed 3.76 kg at birth and presented with severe sepsis at 3 days. Clinical and biochemical investigations revealed dilated cardiomyopathy, hypotonia, growth retardation, severe neutropenia and 3-methylglutaconic aciduria. At 25 months of age, he again, suffered a severe episode of sepsis due to pyelonephritis. The child died at the age of two years - ten months due to heart failure.

**Patient 5 (Pt5)** was the first child of non-consanguineous parents. He was born at 41 weeks of gestation, weighing 3.3 kg. APGAR was 1': 9; 5': 10 after a pregnancy complicated by gestational diabetes treated with diet therapy. Five hours after birth, he exhibited severe

oxygen desaturation with cyanosis and bradycardia, requiring intubation. Hemodynamic instability was persistent with a systolic murmur of 2/6. Mild hepatomegaly was present. Echocardiography showed non-compaction of the left ventricular myocardium with reduced pump function. Blood gas analysis revealed lactic acidosis. Inotropic and diuretic therapy helped obtaining clinical and hemodynamic stabilization, followed by improvement of cardiac function. On the 8<sup>th</sup> day of life, he had a first episode of cardiac arrest from which he was rescued by long resuscitation but deep coma ensued. Over the following days, clinical and hemodynamic conditions remained critical with persistence of anuria. The child died 12 days after birth. *TAZ* gene mutational analysis confirmed the diagnosis of BS in proband and the heterozygous status of the mother. A prenatal diagnosis was performed nine months later and revealed that the foetus in question did not have BS.

**Patient 6 (Pt6)** was the second child born to healthy non-consanguineous parents. Family history includes one healthy male first born and one natural abortion. APGAR was 1': 9; 5': 10, 3.0 kg weight, 48 cm in length. Pt6 displayed frequent regurgitation and vomiting during the first month of life with growth delay (0.1 kg per month). An episode of gastroenteritis with dehydration occurred during his seventh month of life. Further clinical investigations revealed dilated cardiomyopathy with left ventricular non-compaction, alterations in ventricular repolarization, hypokinesia, gallop rhythm, mitral and tricuspid regurgitation, and diastolic murmur 2-3/6 in the mesocardial area. Urinary 3-methylglutaconic aciduria, neutropenia and lactic acidosis were present. Acquired microcephaly with mild cerebral ventricular asymmetry and hypotonia were also evident. Cardioplin/monolysocardioplin analysis was performed on a dried blood spot [16] and led to the diagnosis of BS, MLCL/CL ratio was 7 (normal 0–0.3). As a consequence of a good response to therapy, at the fifteenth month of life, his physical condition was good, albeit with mild hypotrophy of the legs. He was clinically stable until two years of age at which time he presented with an episode of bronchitis and otitis with fever, which was resolved with antibiotic therapy. As in the case of Pt 3, the treatment with G-CSF has not been carried out to date, because the patient has never presented severe infections resistant to antibiotic therapy. At last follow-up he was two years old, 10.43 kg in weight (between 3<sup>rd</sup>-5<sup>th</sup> percentile) and 86.2 cm in height (under the 3<sup>rd</sup> percentile). His clinical condition is generally good with cardiovascular compensation and systolic function at the lower end of the normal range.

#### Analysis of genomic DNA

Genomic DNA was isolated using the EZ1 DNA Blood 350 µl Kit (QIAGEN, Hilden, Germany). The entire coding

region and intron-exon boundaries of the *TAZ* genes of the six enrolled patients were amplified and sequenced by PCR using the oligonucleotides listed in Table 1. PCR conditions are available on request. Sequencing PCR reactions were performed using the BigDye Terminator v1.1 Cycle Sequencing Kit (Applied Biosystems; Carlsbad, CA, USA) and purification was carried out using Sephadex G-50 Fine (GE Healthcare; Little Chalfont, UK). Capillary electrophoresis was performed using ABI PRISM 3130 Genetic Analyser (Applied Biosystems) as recommended by the manufacturer.

All nucleotide positions given in this work are based upon the human *TAZ* gene nucleotide sequence [GenBank: NG\_009634.1].

#### Multiplex PCR analysis

Multiplex PCR analysis was performed using the primer pairs listed in Table 1. Different combinations of PCR fragments were tested in order to confirm multi-exon deletions in the *TAZ* genes of patients Pt1 and Pt2. PCR fragment combinations Taz5 + Taz8-9, Taz5 + Taz6-7 + Taz10-11, Taz5 + Taz10-11 and Taz1-2 + Taz10-11 were successfully assayed. Multiplexed PCR products were checked by agarose gel electrophoresis using 2% Agarose Low Melting (Eurobio, France) + 1% Standard Agarose (AB Analytica, Italy).

#### Deletion breakpoint determination

In patients Pt1 and Pt2, who harbored large gene deletions verified by multiplex PCR, we performed long-range PCR (LR-PCR) using TaKaRa La Taq DNA polymerase (TaKaRa Bio Inc, Japan) to determine the precise locations of breakpoint boundaries. We also performed LR-PCRs using primer TAZ 5fw (Table 1) as the primer binding upstream of the deleted region, coupled with a primer binding to exon 1 of the *ATP6API* gene located about 7000 bp downstream of the *TAZ* gene. LR-PCR products were purified from a 0.8% agarose gel and sequenced by primer-walking [34]. The Clustal W program [35] was used to align sequences located at the breakpoints.

#### Screening for the novel missense mutation p.His214Arg

The *TAZ* genes of 120 male controls were analysed by sequencing fragment Taz8-9 containing the region encoding codon 214.

#### Bioinformatic analysis of interspersed repeated sequences

The *TAZ* gene region was screened for interspersed repeated sequences (SINEs and LINEs) and low complexity DNA sequences using RepeatMasker [36].

**Table 1 Oligonucleotides used for TAZ gene mutation analysis**

Fragment name	Primer	Sequence 5'-3'	Included exons	Amplified gene region*	Length (bp)
Taz 1-2	Taz 1-2fw	agtcaggggccagtgctc	1 and 2	g.5224_5736	552
	Taz 1-2rv	gaaggggtttgttctgacga			
Taz 3-4	Taz 3-4fw	ctgggatatgggaagtgg	3 and 4	g.6642_7095	494
	Taz 3-4rv	ccataggtccctccaaaaca			
Taz 5	Taz 5fw	aaggtcatgggtaggaggt	5	g.7519_7714	236
	Taz 5rv	aaactcctggcttgagtga			
Taz 6-7	Taz 6-7fw	ccccgagaatggttactgat	6 and 7	g.12983_13340	398
	Taz 6-7rv	aggcctagtctcagcacctg			
Taz 8-9	Taz 8-9fw	gggagctgaattgaactgga	8 and 9	g.13404_13753	390
	Taz 8-9rv	agacagcagacaggcagaca			
Taz 10-11	Taz 10-11fw	ggcactcctactgctcctca	10 and 11	g.14097_14513	457
	Taz 10-11rv	agcatcagtcacatccctcag			

\*based upon the reference nucleotide sequence NG\_009634.1. Positions do not include the primer sequences.

### Bioinformatic analysis of the missense mutation p.His214Arg

Tafazzin protein sequences from 13 different species (*Macaca mulatta* GenBank:NP\_001028086.1, *Mus musculus* GenBank:NP\_852657.1, *Rattus norvegicus* GenBank:NP\_001020919.1, *Bos taurus* GenBank:DAA13200.1, *Oryctolagus cuniculus* GenBank:NP\_001164847.1, *Rhinolophus ferrumequinum* GenBank:ACC68906.1, *Salmo salar* GenBank:ACN11123.1, *Danio rerio* GenBank:NP\_0010-01814.1, *Ictalurus punctatus* GenBank:NP\_001187914.1, *Aedes aegypti* GenBank:XP\_001653587.1, *Drosophila melanogaster* GenBank:NP\_477432.3, *Ascaris suum* GenBank:ADY45663.1, *Schistosoma japonicum* GenBank:CAX73490.1) were aligned with the human TAZ sequence (GenBank:NP\_000107.1) using the Clustal W program [35]; multialignments were checked to determine the degree of evolutionary conservation of the amino acid residue (His214) involved. Further, the His214Arg amino acid substitution was also analysed by PolyPhen [37], which makes a prediction as to whether an amino acid change is likely to be deleterious to protein function [38]. Profile scores of >2.0 indicate that the polymorphism is probably damaging to protein function whereas scores of 1.5–2.0 are possibly damaging, and scores of <1.5 are indicative of a benign variant. To confirm results from PolyPhen, we used (i) the MutPred Server [39], which generates a probability for a given mutation to be deleterious to protein function (scores that range from 0 to 1), and the likelihood of it being pathogenic (scores over 0.5) [40] and (ii) SIFT [41], which yields a score of ≤0.05 if the amino acid substitution is predicted to be damaging and >0.05 if it is predicted to be tolerated.

### Results

#### Identification of the TAZ mutations in six new unrelated BS patients

Molecular analysis of the TAZ gene in the six patients confirmed the clinical diagnosis by revealing that each one of them carried a mutated TAZ allele. Six different TAZ mutations were identified, including the previously reported c.367C>T (p.Arg123Term) nonsense mutation [42] and five new mutations: a c.641A>G (p.His214Arg) missense variant, the c.284dupG (p.Thr96Aspfs\*37) microinsertion, the c.678\_691del14 (p.Tyr227Trpfs\*79) microdeletion and the two g.8009\_16445del8437 and g.[9777\_9814del38; 9911-?\_14402del] large rearrangements. The TAZ gene variants identified in the BS patients and their related clinical features are summarized in Table 2.

#### Confirmation of the likely pathogenicity of p.His214Arg

The p.His214Arg amino acid substitution was not found in 120 TAZ alleles from healthy male controls.

ClustalW analysis revealed a very high degree of evolutionary conservation of histidine at position 214 (His214) in the tafazzin protein (Figure 2). Not only do all vertebrate tafazzins contain histidine at the analogous location but also do the tafazzins of two insects (*Aedes aegypti* and *Drosophila melanogaster*) and a flatworm (*Schistosoma japonicum*). This implies that this histidine residue must have been conserved for the last ~700 Ma, since prior to the divergence of deuterostomes from protostomes. Such a degree of evolutionary conservation implies a key function for this residue.

PolyPhen predicted that this mutation is probably damaging with a score of 2.862 for TAZ-FL (GenBank:NP\_000107.1) and 3.123 for the TAZ-Δ5 isoform (GenBank:NP\_851828.1, [25]). This prediction was

**Table 2 Genotypic and phenotypic data for the BS patients under study**

Patient	TAZ gene mutation	Reference	Age of onset	Manifestations at onset	Exitus
Pt1	g.8009_16445del8437	this work	first day of life	respiratory distress, lactic acidosis, neutropenia	ten days, heart failure
Pt2	g.[9777_9814del38; 9911-?_14402del]	this work	twelve hours of life	mild cyanosis, neutropenia, lactic acidosis, dilated cardiomyopathy	one year, heart failure
Pt3	c.641A>G (p.His214Arg)	this work	<i>in utero</i>	dilated cardiomyopathy	living, 8 years old
Pt4	c.367C>T (p.Arg123Term)	[42]	five months	hypotonia and heart failure	living, 11 years old
Pt5	c.678_691del14 (p.Tyr227Trpfs*79)	this work	five hours	oxygen desaturation with cyanosis and bradycardia, lactic acidosis	twelve days, heart failure
Pt6	c.284dupG (p.Thr96Aspfs*37)	this work	first month	frequent regurgitation and vomiting, growth delay, lactic acidosis	living, 3 years old

confirmed by MutPred (TAZ-FL, score  $g = 0.732$ ; TAZ- $\Delta 5$ , score  $g = 0.677$ ) and SIFT (score of 0.01 for TAZ-FL; 0.02 for TAZ- $\Delta 5$ ).

**Interspersed repeated sequences in the TAZ gene**

Results obtained from the analysis of the human TAZ gene sequence (GenBank:NG\_009634.1) with RepeatMasker are summarized in Table 3.

**Characterization of the large deletion breakpoints**

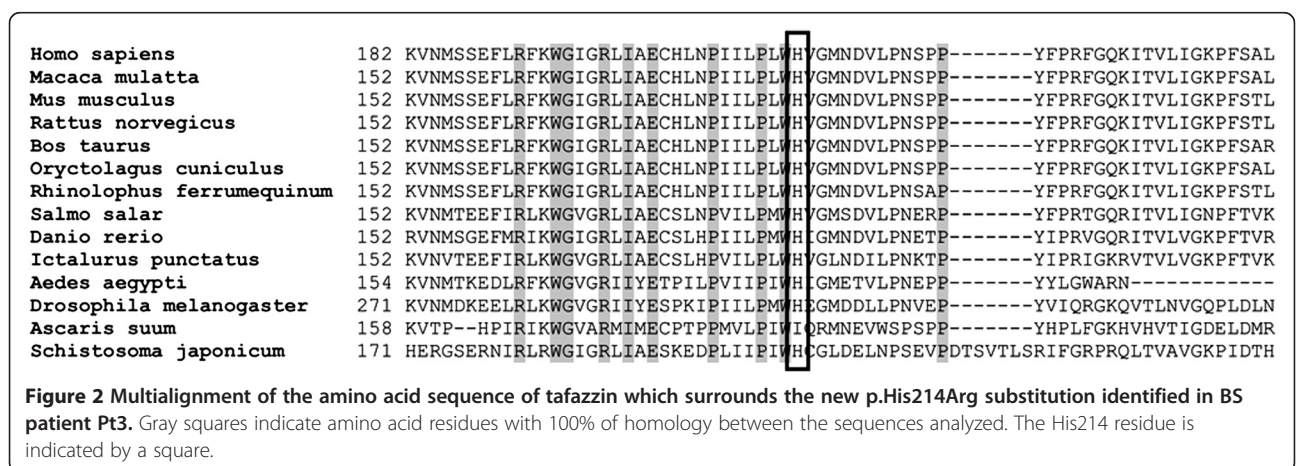
In patients 1 and 2, PCR analysis of the TAZ gene exonic sequence could not amplify exons 6 to 11. Multiplex PCR analysis was then performed, which resulted in the amplification of exons 1 to 5 but not exons 6 to 11, indicating that gross deletions spanned a large region of the TAZ gene in both patients. LR-PCR, performed to determine deletion breakpoints, yielded amplicons that were shorter in length than the PCR products obtained from healthy controls. Characterization of the breakpoint junctions by sequencing analysis in Pt1 revealed the deletion of an 8437 bp genomic fragment (g.8009\_16445del8437) spanning exons 6 to 11 of the TAZ gene inclusive. The breakpoint junctions mapped to within two *AluY* repeats which were in the same orientation and located respectively 0.357 kb downstream of the exon 5 donor splice site, and

1.979 kb downstream of the exon 11 donor splice site (Figure 3A). The g.8009\_16445del8437 deletion gave rise to a new hybrid *AluY* element containing the breakpoint (Figure 3A).

The analysis of the breakpoint junctions in Pt2 revealed the g.[9777\_9814del38; 9911-?\_14402del] complex allele. A deletion of 38 bp between nucleotides 9777 and 9814 was accompanied by a larger deletion of more than 4 kb (Figure 3B). The upstream breakpoint junction of the larger deletion is located around 2246–2260 bp downstream of the exon 5 donor splice site (nucleotides 9898\_9911 in GenBank:NG\_009634.1), within an A-rich sequence (14 A in the Ref Seq). One characteristic of our sequencing method was that, after a repeat structure such as a polyA stretch of >9-10 nucleotides, the sequencing signal became unreadable making it impossible to determine the number of As in the repeated sequence. Hence we could not determine precisely at which nucleotide, between 9898\_9911 in NG\_009634.1, the upstream breakpoint of Pt2 occurred. However, the downstream junction was located within exon 11 (nucleotide c.814).

**Determination of heterozygous carrier status**

Since the TAZ allele of Pt2 harbors a portion of exon 11, we were able to amplify across his deletion junctions



**Table 3 Interspersed repeat sequences within the TAZ gene (output data by Repeatmasker)**

	Number of elements	Total length (bp)	Percentage of sequence* (%)
Total interspersed repeats	33	7673	44.64
- SINEs:	27	6487	37.74
ALUs	25	6143	35.74
MIRs	2	344	2.00
- LINEs:	6	1197	6.96

\* TAZ gene reference sequence NG\_009634.1.

with the primer pair TAZ 5fw and TAZ 10-11rv, obtaining a fragment of almost 2.6 kb. The same PCR reaction yielded a fragment of around 7 kb from the wild-type TAZ allele. Such LR-PCR amplification was extended to female relatives of Pt2, which allowed confirmation of the heterozygous carrier status of both his mother and sister (Figure 4).

The mother of Pt1 declined further investigation.

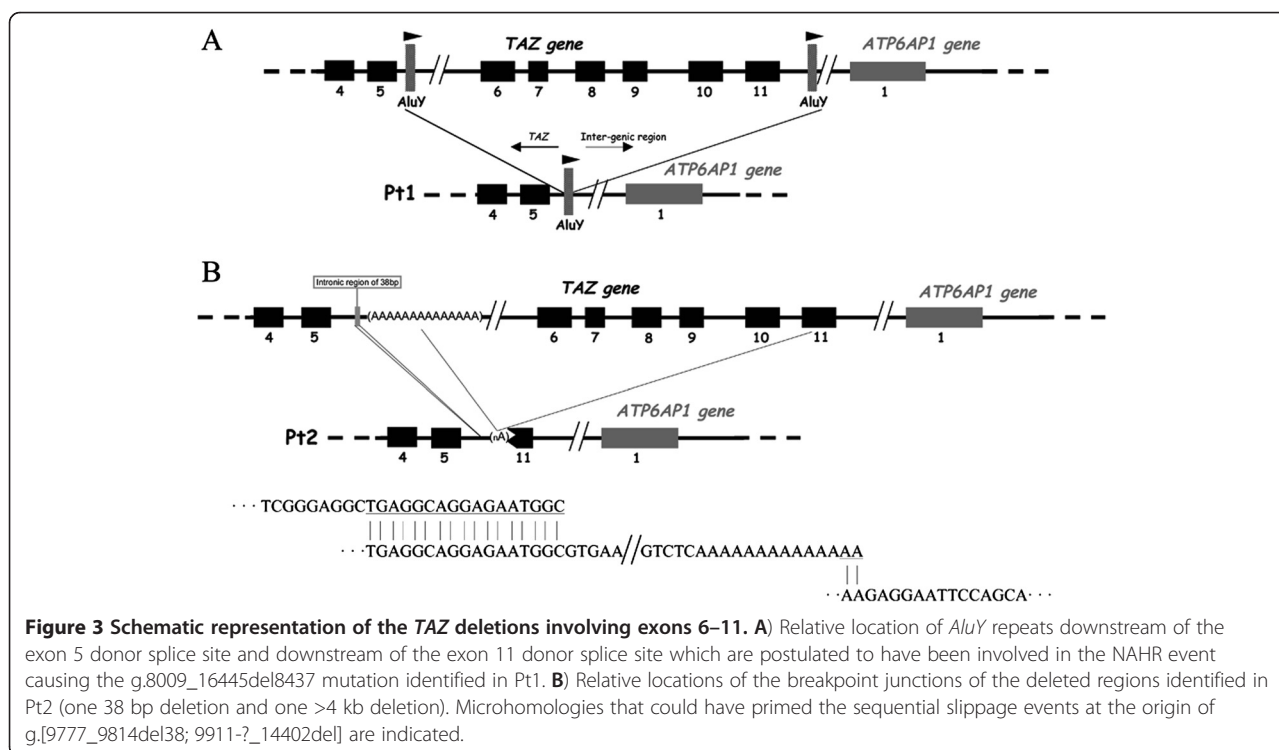
### Discussion

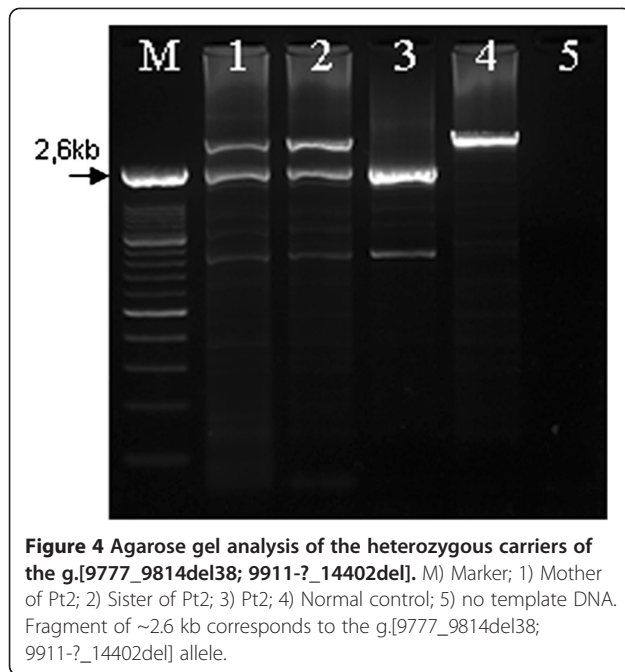
Mutations underlying human inherited disease vary greatly in size, from gross rearrangements to single base-pair substitutions. They can also originate via a number of different molecular mechanisms. Irrespective of the type of lesion, the nature, location and frequency of human gene mutations are shaped to a considerable extent, and

often in remarkably predictable ways, by the local DNA sequence environment (for a review, see [43]).

Almost 45% of the genomic sequence of the human TAZ gene is represented by interspersed repeated sequences (SINES and LINES) and 76% of these (35% of the TAZ gene sequence) are *Alu* sequences. The proportion of the whole human genome that is *Alu* sequence is estimated to be around 10.6% [44]. A high proportion of *Alu* repeats within a gene sequence can promote gross gene rearrangements [45], as reported for example for the *PAFAH1B1* (LIS1) gene [46]. We report here an intragenic deletion in the TAZ gene, which appears to have been mediated by recombination between two *AluY* repeats. The deletion junctions in Pt1 were located within the two highly homologous and similarly oriented *AluY* repeats (Figure 3A). Non-allelic homologous recombination (NAHR) between *Alu* elements is a likely mechanism of mutation in this patient [45]. An *Alu*-mediated TAZ gene deletion that ablates the entire gene has also been reported [47]. These data suggest that the TAZ gene is located within a genomic environment that is particularly rich in repeats that can promote genetic rearrangements.

HGMD Professional v.2012.2 ([30] update 29th June 2012) reports a total of 104 TAZ gene mutations of which 35.6% are missense, 12.5% are nonsense, 18.3% affect splicing, 19.2% are microdeletions, 7.7% are microinsertions and 6.7% are large gene rearrangements. Of the eight gross gene rearrangements reported, three were published [31,47,48] whereas the remaining five can be retrieved





from the “Human Tafazzin Gene Mutation and Variation Database” [33] to which they were submitted directly, without supporting details. One mutation in the latter group is reported to involve the deletion of exons 6–11. In our patient sample, we identified deletions of exons 6–11 in two patients (Pt1 and Pt2) who harbored two different genetic rearrangements. Defining the exact breakpoints of such rearrangements is a prerequisite to identify heterozygous carriers of *TAZ* gene rearrangements, and for pre-natal diagnosis.

Despite the high density of *Alu* repeats in the *TAZ* gene, the deletion observed in Pt2 was not mediated by NAHR between *Alu* elements. In this patient, the genetic rearrangement was found to be more complex than in Pt1 (one 38 bp deletion and one >4 kb deletion) and might have been prompted by and Serial Replication Slippage (SRS) [49]. Consistently with this assertion, the sequences flanking the breakpoints exhibit microhomologies as depicted in Figure 3B. Sequential slippage events at the 3' end of the nascent strand during DNA replication could have been primed by these microhomologies [49]. We postulate that the SRS model, instead of the more recently proposed FoSTeS model [50], is the more likely mechanism of the rearrangement in Pt2 since the SRS model assumes that the replication slippage occurs at closely adjacent sites and gives rise to relatively small rearrangements (up to 10 kb), whereas the FoSTeS model is better suited to explain template switching that occurs over long distances, generating much larger DNA rearrangements (120 to 550 kb) [49]. Whatever the underlying mechanism, characterization of gross rearrangements in the *TAZ* gene is essential for

identifying heterozygous carriers among female relatives of the probands and, as it is the case for Pt2, to confirm the carrier status of close female relatives.

In addition to these gross gene rearrangements, we identified two frameshift variants in the *TAZ* genes of our patients: a small 14 bp deletion (Pt5) and a small duplication (Pt6). Ball et al. [51] suggested that a combination of slipped mispairing mediated by direct repeats and secondary structure formation promoted by symmetric elements could account for most microdeletions and microinsertions. We analyzed the sequence environment as suggested [51], considering the 10 bp of DNA sequence flanking the c.678\_691del14 deletion detected in Pt5 (ACAGTCCGCCctactccccgctTTGGACAGGT). This region is G/C rich and the deletion is flanked by CCGC repeats, consistently with a slipped mispairing model. At the protein level, the c.678\_691del14 deletion is predicted to lead to a p.Tyr227Trpfs\*79 frameshift.

The c.284dupG duplication in Pt6 (GTTGATGC GTTGgGTGAGGAGGA) might be explained by the modified slipped mispairing model [52], whereby misalignment occurs between a specific contiguous sequence on one strand and a second partially homologous, yet interrupted, sequence on the other strand. One repeat copy only becomes capable of base-pairing once an additional base has been inserted or deleted. In the case of Pt6, the first repeat is GTTGA whilst the second repeat is GTTGGA; slipped mispairing would therefore account for the duplication of a G in this first repeat. At the protein level, the c.284dupG mutation is predicted to lead to a frameshift p.Thr96Aspfs\*37 with premature termination of translation.

We also detected two single nucleotide substitutions in the *TAZ* genes: c.641A>G (Pt3) and c.367C>T (Pt4). c.641A>G leads to the substitution p.His214Arg, which, although not predicted to change the polarity of the involved residue, occurs in a residue which is evolutionarily conserved, suggesting an essential role in protein function. We are unable to infer about the possible impact of the missense change directly on the three-dimensional structure of human tafazzin, as the crystal structure is not available, however molecular characterization of mutant *TAZ* alleles has been performed in a *Saccharomyces cerevisiae* model of Barth syndrome [53]. While the corresponding residue in the *S. cerevisiae* Taz1p ortholog is not conserved (it is an Alanine), the p.His214Arg mutation occurs in a putative unusual membrane anchor region in yeast Taz1p [53]. Interestingly, mutations in this membrane anchor result in two distinct biochemical fates: low fidelity sorting with increased rates of aggregation and aberrant macromolecular assembly [53].

Pt4 carried the p.Arg123Term nonsense mutation previously reported by van Werkhoven et al. [42]. According to the Human Tafazzin Gene Mutation and Variation



Database [33], multiple examples of the p.Arg123Term mutation have been reported, suggesting recurrent mutation at this CpG dinucleotide mutation hotspot.

We found lactic acidosis to be present in 5/6 patients at the time they were brought to clinical attention, suggesting that, in addition to 3-methylglutaconic aciduria and neutropenia, lactic acidosis is an important biochemical marker for suspecting BS. In addition, a deep metabolic decompensation during the first hours of life, including respiratory distress and cyanosis with oxygen desaturation, was associated with the most severe outcome (Pt1, P2 and Pt5). Deep metabolic decompensation associated to these other signs may thus correlate with the severity of the disease more than it does the time of onset of cardiomyopathy that, for instance, was already detected *in utero* in Pt3 who was still alive at last follow up [54,55]. The alteration of cardiolipin metabolism in BS leads to an abnormal lipid structure of the inner mitochondrial membrane. This may result in mitochondrial leakage of metabolites, such as the lactate as we observed in our BS patients. Thus, we could hypothesize that an acute metabolic decompensation during the first hours of life, associated with respiratory distress and oxygen desaturation, or with other signs such as mild hyperammonaemia, hypoglycaemia and elevated transaminases [54] could be correlated to a deeper impairment of the mitochondrial function at the onset of BS and thus to an earlier and deeper cardioskeletal tissue damage. However, it cannot be excluded that the severity of the metabolic decompensation can be related also to other still undefined factors, as it has been reported that the pathophysiology of Barth syndrome can unlikely be explained solely on the basis of cardiolipin abnormalities and additional factors could play a role in modifying the disease expression [56]. Anyhow, these assertions remains to be proven.

Several cases of clinical manifestations of BS being detected *in utero*, as in the case of Pt3, have been reported [57,58]. Therefore, cardiolipin analysis or mutational analysis of the *TAZ* gene should be considered in cases of heart failure or sepsis during the neonatal period or of *in utero* cardiomyopathy. Such analysis should also be considered when investigating the causes of natural abortions (e.g. Pt1 and Pt6) as also previously reported [58].

Early diagnosis of BS is a prerequisite for early and effective therapy and follow-up, and for preventing crisis due to sepsis, as suggested by the clinical histories of Pt3 and Pt6. Moreover, prompt molecular characterization of BS patients is vital for prenatal diagnosis and for preventing the recurrence of BS in the same family (e.g. Pt5). Late diagnosis of BS can lead to familial recurrence of the disease as observed in the families of Pt1, Pt2 and Pt4.

## Conclusions

The human *TAZ* gene sequence is rich in interspersed repeats that may promote the occurrence of gross gene rearrangements albeit with different mutational mechanisms. Therefore, gross *TAZ* rearrangements, which may appear to be similar in different patients on the basis of ablated exons, may actually represent private mutations.

Lactic acidosis is an early biochemical marker of BS, together with 3-methylglutaconic aciduria, such as are metabolic decompensation, cardiomyopathy, heart failure and hypotonia. The severity of the metabolic decompensation at onset of BS correlates with the severity of the clinical course of the disease, and is therefore important for prognostic assessment. Cardiomyopathy may be detectable *in utero*, but its time of onset does not seem to be necessarily correlated with the severity of the prognosis.

Mutational analysis of the *TAZ* gene is necessary to confirm the clinical and biochemical diagnosis in probands, to identify heterozygous carriers and for prenatal diagnosis, but should also be considered in the presence of cardiomyopathy *in utero* and of natural abortion together with cardiolipin ratio analysis.

## Competing interests

The authors declare no competing interests.

## Authors' contributions

LF performed the molecular genetic studies. AM coordinated the research. SM and SC contributed to the *TAZ* gene sequencing analysis of BS patients. SF performed urinary organic acid analysis of BS patients. FMV performed cardiolipin/monolysocardiolipin analysis of BS patients. LF and AM wrote the manuscript. MAD contributed to the discussion of clinical findings. DNC contributed to the discussion of genetic findings and to critical revision of the manuscript. MAD, LL, LR, EB, who are referent physicians for the patients, contributed to description of clinical cases. RG contributed to critical revision of the manuscript. All authors read and approved the final manuscript.

## Acknowledgments

The Fondazione Meyer ONLUS, Firenze, Italia and the Regione Toscana, Italia are gratefully acknowledged for their financial support (grant "POR CRO FSE 2007-2013, MEYER AON-CHAP"). This work was also partially supported by AMMeC (Associazione Malattie Metaboliche Congenite, Italia). A grant Ricerca Corrente from the Italian Ministry of Health to EB is gratefully acknowledged.

## Author details

<sup>1</sup>Department of Neurosciences, Psychology, Pharmacology and Child Health, University of Florence and Paediatric Neurology Unit and Laboratories, Meyer Children's Hospital, Viale Pieraccini n. 24, Florence 50139, Italy. <sup>2</sup>Metabolic and Muscular Unit, Neuroscience Department, Meyer Children's Hospital, Florence, Italy. <sup>3</sup>Paediatric Neurology Unit and Laboratories, Neuroscience Department, Meyer Children's Hospital, Florence, Italy. <sup>4</sup>Neonatology Unit, Department of Mother & Child, University of Modena, Modena, Italy. <sup>5</sup>University of Bologna and Azienda Ospedaliera S. Orsola Malpighi, Bologna, Italy. <sup>6</sup>Department of Neurosciences, Laboratory of Molecular Medicine, Bambino Gesù Children's Research Hospital, Rome, Italy. <sup>7</sup>Department of Clinical Chemistry and Paediatrics, Amsterdam, Netherlands. <sup>8</sup>Institute of Medical Genetics, School of Medicine, Cardiff University, Cardiff, UK. <sup>9</sup>IRCCS, Stella Maris, Pisa, Italy.

Received: 19 October 2012 Accepted: 30 January 2013

Published: 14 February 2013

## References

- Barth PG, Scholte HR, Berden JA, Van der Klei-Van Moorsel JM, Luyt-Houwen IE, Van 't Veer-Korthof ET, Van der Harten JJ, Sobotka-Plojhar MA: **An X-linked mitochondrial disease affecting cardiac muscle, skeletal muscle and neutrophil leucocytes.** *J Neurol Sci* 1983, **62**:327–355.
- Spencer CT, Bryant RM, Day J, Gonzalez IL, Colan SD, Thompson WR, Berthry J, Redfearn SP, Byrne BJ: **Cardiac and clinical phenotype in Barth syndrome.** *Pediatrics* 2006, **118**:e337–346.
- Kelley RI, Cheatham JP, Clark BJ, Nigro MA, Powell BR, Sherwood GW, Sladky JT, Swisher WP: **X-linked dilated cardiomyopathy with neutropenia, growth retardation, and 3-methylglutaconic aciduria.** *J Pediatr* 1991, **119**:738–747.
- Finsterer J, Stollberger C, Fazio G: **Neuromuscular disorders in left ventricular hypertrabeculation/noncompaction.** *Curr Pharm Des* 2010, **16**:2895–2904.
- Takeda A, Sudo A, Yamada M, Yamazawa H, Izumi G, Nishino I, Ariga T: **Eponym: Barth syndrome.** *Eur J Pediatr* 2011, **170**:1365–1367.
- Makaryan V, Kulik W, Vaz FM, Allen C, Dror Y, Dale DC, Aprikyan AA: **The cellular and molecular mechanisms for neutropenia in Barth syndrome.** *Eur J Haematol* 2012, **88**:195–209.
- Barth PG, Valianpour F, Bowen VM, Lam J, Duran M, Vaz FM, Wanders RJ: **X-linked cardioskeletal myopathy and neutropenia (Barth syndrome): an update.** *Am J Med Genet A* 2004, **126A**:349–354.
- Houtkooper RH, Vaz FM: **Cardiolipin, the heart of mitochondrial metabolism.** *Cell Mol Life Sci* 2008, **65**:2493–2506.
- Vreken P, Valianpour F, Nijtmans LG, Grivell LA, Plecko B, Wanders RJ, Barth PG: **Defective remodeling of cardiolipin and phosphatidylglycerol in Barth syndrome.** *Biochem Biophys Res Commun* 2000, **279**:378–382.
- Acehan D, Malhotra A, Xu Y, Ren M, Stokes DL, Schlame M: **Cardiolipin affects the supramolecular organization of ATP synthase in mitochondria.** *Biophys J* 2011, **100**:2184–2192.
- Claypool SM, Boontheung P, McCaffery JM, Loo JA, Koehler CM: **The cardiolipin transacylase, tafazzin, associates with two distinct respiratory components providing insight into Barth syndrome.** *Mol Biol Cell* 2008, **19**:5143–5155.
- Claypool SM, Oktay Y, Boontheung P, Loo JA, Koehler CM: **Cardiolipin defines the interactome of the major ADP/ATP carrier protein of the mitochondrial inner membrane.** *J Cell Biol* 2008, **182**:937–950.
- Koshkin V, Greenberg ML: **Oxidative phosphorylation in cardiolipin-lacking yeast mitochondria.** *Biochem J* 2000, **347**(Pt 3):687–691.
- Schlame M, Towbin JA, Heerdt PM, Jehle R, DiMauro S, Blanck TJ: **Deficiency of tetralinoleoyl-cardiolipin in Barth syndrome.** *Ann Neurol* 2002, **51**:634–637.
- Valianpour F, Mitsakos V, Schlemmer D, Towbin JA, Taylor JM, Ekert PG, Thorburn DR, Munnich A, Wanders RJ, Barth PG, Vaz FM: **Monolysocardiolipins accumulate in Barth syndrome but do not lead to enhanced apoptosis.** *J Lipid Res* 2005, **46**:1182–1195.
- Houtkooper RH, Turkenburg M, Poll-The BT, Karall D, Perez-Cerda C, Morrone A, Malvagia S, Wanders RJ, Kulik W, Vaz FM: **The enigmatic role of tafazzin in cardiolipin metabolism.** *Biochim Biophys Acta* 2009, **1788**:2003–2014.
- Malhotra A, Xu Y, Ren M, Schlame M: **Formation of molecular species of mitochondrial cardiolipin. 1. A novel transacylation mechanism to shuttle fatty acids between sn-1 and sn-2 positions of multiple phospholipid species.** *Biochim Biophys Acta* 2009, **1791**:314–320.
- Acehan D, Khuchua Z, Houtkooper RH, Malhotra A, Kaufman J, Vaz FM, Ren M, Rockman HA, Stokes DL, Schlame M: **Distinct effects of tafazzin deletion in differentiated and undifferentiated mitochondria.** *Mitochondrion* 2009, **9**:86–95.
- Schlame M, Ren M: **Barth syndrome, a human disorder of cardiolipin metabolism.** *FEBS Lett* 2006, **580**:5450–5455.
- Khuchua Z, Yue Z, Batts L, Strauss AW: **A zebrafish model of human Barth syndrome reveals the essential role of tafazzin in cardiac development and function.** *Circ Res* 2006, **99**:201–208.
- He Q: **Tafazzin knockdown causes hypertrophy of neonatal ventricular myocytes.** *Am J Physiol Heart Circ Physiol* 2010, **299**:H210–216.
- Acehan D, Vaz F, Houtkooper RH, James J, Moore V, Tokunaga C, Kulik W, Wansapura J, Toth MJ, Strauss A, Khuchua Z: **Cardiac and skeletal muscle defects in a mouse model of human Barth syndrome.** *J Biol Chem* 2011, **286**:899–908.
- Soustek MS, Falk DJ, Mah CS, Toth MJ, Schlame M, Lewin AS, Byrne BJ: **Characterization of a transgenic short hairpin RNA-induced murine model of Tafazzin deficiency.** *Hum Gene Ther* 2011, **22**:865–871.
- Bione S, D'Adamo P, Maestrini E, Gedeon AK, Bolhuis PA, Toniolo D: **A novel X-linked gene, G4.5, is responsible for Barth syndrome.** *Nat Genet* 1996, **12**:385–389.
- Vaz FM, Houtkooper RH, Valianpour F, Barth PG, Wanders RJ: **Only one splice variant of the human TAZ gene encodes a functional protein with a role in cardiolipin metabolism.** *J Biol Chem* 2003, **278**:43089–43094.
- Gonzalez IL: **Barth syndrome: TAZ gene mutations, mRNAs, and evolution.** *Am J Med Genet A* 2005, **134**:409–414.
- Lu B, Kelher MR, Lee DP, Lewin TM, Coleman RA, Choy PC, Hatch GM: **Complex expression pattern of the Barth syndrome gene product tafazzin in human cell lines and murine tissues.** *Biochem Cell Biol* 2004, **82**:569–576.
- Xu Y, Zhang S, Malhotra A, Edelman-Novemsky I, Ma J, Kruppa A, Cernicica C, Blais S, Neubert TA, Ren M, Schlame M: **Characterization of tafazzin splice variants from humans and fruit flies.** *J Biol Chem* 2009, **284**:29230–29239.
- D'Adamo P, Fassone L, Gedeon A, Janssen EA, Bione S, Bolhuis PA, Barth PG, Wilson M, Haan E, Orstavik KH, et al: **The X-linked gene G4.5 is responsible for different infantile dilated cardiomyopathies.** *Am J Hum Genet* 1997, **61**:862–867.
- HGMD Professional v.2012.2*. 2012. <https://portal.biobase-international.com>.
- Aradhya S, Lewis R, Bonaga T, Nwokekeh N, Stafford A, Boggs B, Hruska K, Smaoui N, Compton JG, Richard G, Suchy S: **Exon-level array CGH in a large clinical cohort demonstrates increased sensitivity of diagnostic testing for Mendelian disorders.** *Genet Med* 2012, **14**:594–603.
- Moric-Janiszewska E, Markiewicz-Loskot G: **Genetic heterogeneity of left-ventricular noncompaction cardiomyopathy.** *Clin Cardiol* 2008, **31**:201–204.
- Barth Syndrome Foundation*. <http://www.barthsyndrome.org/>.
- Strauss EC, Kobori JA, Siu G, Hood LE: **Specific-primer-directed DNA sequencing.** *Anal Biochem* 1986, **154**:353–360.
- Thompson JD, Higgins DG, Gibson TJ: **CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice.** *Nucleic Acids Res* 1994, **22**:4673–4680.
- RepeatMasker program: *Institute for System Biology*. Seattle, WA: [www.repeatmasker.org](http://www.repeatmasker.org).
- PolyPhen: *Prediction of functional effect of human nsSNPs*. <http://genetics.bwh.harvard.edu/pph/>.
- Sunyaev S, Ramensky V, Koch I, Lathe W III, Kondrashov AS, Bork P: **Prediction of deleterious human alleles.** *Hum Mol Genet* 2001, **10**:591–597.
- MutPred application tool: *Mooney laboratory at the Buck Institute for Age Research*. <http://mutpred.mutdb.org/>.
- Li B, Krishnan VG, Mort ME, Xin F, Kamati KK, Cooper DN, Mooney SD, Radivojac P: **Automated inference of molecular mechanisms of disease from amino acid substitutions.** *Bioinformatics* 2009, **25**:2744–2750.
- SIFT application (Sorting Intolerant from Tolerant)*. <http://sift.bii.a-star.edu.sg/>.
- van Werkhoven MA, Thorburn DR, Gedeon AK, Pitt JJ: **Monolysocardiolipin in cultured fibroblasts is a sensitive and specific marker for Barth Syndrome.** *J Lipid Res* 2006, **47**:2346–2351.
- Cooper DN, Bacolla A, Ferec C, Vasquez KM, Kehrer-Sawatzki H, Chen JM: **On the sequence-directed nature of human gene mutation: the role of genomic architecture and the local DNA sequence environment in mediating gene mutations underlying human inherited disease.** *Hum Mutat* 2011, **32**:1075–1099.
- Lander ES, Linton LM, Birren B, Nusbaum C, Zody MC, Baldwin J, Devon K, Dewar K, Doyle M, FitzHugh W, et al: **Initial sequencing and analysis of the human genome.** *Nature* 2001, **409**:860–921.
- Batzler MA, Deininger PL: **Alu repeats and human genomic diversity.** *Nat Rev Genet* 2002, **3**:370–379.
- Mei D, Lewis R, Parrini E, Lazarou LP, Marini C, Pilz DT, Guerrini R: **High frequency of genomic deletions—and a duplication—in the LIS1 gene in lissencephaly: implications for molecular diagnosis.** *J Med Genet* 2008, **45**:355–361.
- Singh HR, Yang Z, Siddiqui S, Pena LS, Westerfield BH, Fan Y, Towbin JA, Vatta M: **A novel Alu-mediated Xq28 microdeletion ablates TAZ and**

- partially deletes DNL1L in a patient with Barth syndrome. *Am J Med Genet A* 2009, **149A**:1082–1085.
48. Cosson L, Toutain A, Simard G, Kulik W, Matyas G, Guichet A, Blasco H, Maakaroun-Vermesse Z, Vaillant MC, Le Caignec C, *et al*: **Barth syndrome in a female patient.** *Mol Genet Metab* 2012, **106**:115–120.
  49. Gu W, Zhang F, Lupski JR: **Mechanisms for human genomic rearrangements.** *Pathogenetics* 2008, **1**:4.
  50. Lee JA, Carvalho CM, Lupski JR: **A DNA replication mechanism for generating nonrecurrent rearrangements associated with genomic disorders.** *Cell* 2007, **131**:1235–1247.
  51. Ball EV, Stenson PD, Abeyasinghe SS, Krawczak M, Cooper DN, Chuzhanova NA: **Microdeletions and microinsertions causing human genetic disease: common mechanisms of mutagenesis and the role of local DNA sequence complexity.** *Hum Mutat* 2005, **26**:205–213.
  52. Cooper DN, Krawczak M: **Mechanisms of insertional mutagenesis in human genes causing genetic disease.** *Hum Genet* 1991, **87**:409–415.
  53. Claypool SM, McCaffery JM, Koehler CM: **Mitochondrial mislocalization and altered assembly of a cluster of Barth syndrome mutant tafazzins.** *J Cell Biol* 2006, **174**:379–390.
  54. Donati MA, Malvagia S, Pasquini E, Morrone A, La Marca G, Garavaglia B, Toniolo D, Zammarchi E: **Barth syndrome presenting with acute metabolic decompensation in the neonatal period.** *J Inherit Metab Dis* 2006, **29**:684.
  55. Yen TY, Hwu WL, Chien YH, Wu MH, Lin MT, Tsao LY, Hsieh WS, Lee NC: **Acute metabolic decompensation and sudden death in Barth syndrome: report of a family and a literature review.** *Eur J Pediatr* 2008, **167**:941–944.
  56. Schlame M, Kelley RI, Feigenbaum A, Towbin JA, Heerdt PM, Schieble T, Wanders RJ, DiMauro S, Blanck TJ: **Phospholipid abnormalities in children with Barth syndrome.** *J Am Coll Cardiol* 2003, **42**:1994–1999.
  57. Brady AN, Shehata BM, Fernhoff PM: **X-linked fetal cardiomyopathy caused by a novel mutation in the TAZ gene.** *Prenat Diagn* 2006, **26**:462–465.
  58. Steward CG, Newbury-Ecob RA, Hastings R, Smithson SF, Tsai-Goodman B, Quarrell OW, Kulik W, Wanders R, Pennock M, Williams M, *et al*: **Barth syndrome: an X-linked cause of fetal cardiomyopathy and stillbirth.** *Prenat Diagn* 2010, **30**:970–976.

doi:10.1186/1750-1172-8-27

**Cite this article as:** Ferri *et al.*: New clinical and molecular insights on Barth syndrome. *Orphanet Journal of Rare Diseases* 2013 **8**:27.

**Submit your next manuscript to BioMed Central and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at  
[www.biomedcentral.com/submit](http://www.biomedcentral.com/submit)

