

Immunomodulatory Mechanisms and Therapeutic Potential of Mesenchymal Stem Cells

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Abstract

Mesenchymal stem cells (MSCs) are regarded as highly promising cells for allogeneic cell therapy, owing to their multipotent nature and ability to display potent and varied functions in different diseases. The functions of MSCs, including native immunomodulation, high self-renewal characteristic, and secretory and trophic properties, can be employed to improve the immune-modulatory functions in diseases. MSCs impact most immune cells by directly contacting and/or secreting positive microenvironmental factors to influence them. Previous studies have reported that the immunomodulatory role of MSCs is basically dependent on their secretion ability from MSCs. This review discusses the immunomodulatory capabilities of MSCs and the promising strategies to successfully improve the potential utilization of MSCs in clinical research.

Keywords Mesenchymal stem cells · Immunomodulation · Therapeutics · Engineered · Immune cells

Introduction

Mesenchymal stem cells (MSCs) were first defined in the late 1960s, as fibroblast-like cells derived from bone marrow (BM) possessing multipotential for high differentiation and self-renewal [1–4]. Later, Caplan et al. found that

BM-derived fibroblast-like cells also possess osteogenic and chondrogenic abilities, and named these cells "MSCs" in 1991 [5, 6]. Then, MSCs have been found to differentiate into several other mesenchymal tissues, such as myocytes and adipocytes [7, 8]. Further investigations revealed that MSCs could differentiate into different lineages, including

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epithelial, neuronal, astrocytic morphology, endothelial, and smooth muscle in vitro, depending on specific conditions of cocultured cell type [9–14]. Meanwhile, Dazzi et al. and Taechangam et al. suggested that the multipotent capacity, support of hematopoiesis in the BM, and their immunoregulatory effects positively endow MSCs treatment as a promising therapeutical strategy [15, 16].

Many studies have reported that MSCs could be isolated from adipose tissue [14, 17]. It has been shown that adipose MSCs (AMSCs) with multipotentiality give rise to adipocytes, neurocytes, chondrocytes, myocytes, and osteoblasts cell lineages [18, 19]. Other reports found that exosomes derived from AMSCs can alleviate the inflammation response to promote wound healing and suppress osteoarthritic cartilage degeneration [20, 21]. MSCs restore mucosal immunity and rejuvenate mucosal immunosenescence in the elderly [22]. Moreover, reprogramming AMSCs into islet β -cells (reprogrammed AMSCs-derived islet β -cells) with low immunogenicity achieved ideal therapeutic effects in treating canine diabetes mellitus [23].

MSCs also can be isolated from neonatal tissue, including human umbilical cord blood (HUCB-MSCs), umbilical cord MSCs (HUC-MSCs), amniotic fluid (HAF-MSCs), placenta MSCs (HP-MSCs), and amniotic membrane (HAM-MSCs), which have been utilized in treating neurological deficits in animal models and patients with intracerebral hemorrhage (ICH) [24–30]. Recently, a report from a clinical trial found that it is safe and well-tolerated to infuse HUC-MSCs intravenously for moderate and severe coronavirus disease 2019 (COVID-19) patients, which may be regarded as a promising therapy in targeting the underlying aberrant immune responses [31]. The transplantation of HUC-MSCs with immunomodulatory, and anti-inflammatory properties emerge in immune and inflammatory diseases and is known as a perspective in future therapeutic utilization [32].

It has been shown that MSCs can be isolated from synovial fluid (SF-MSCs) [33], dental pulp (DMSCs) [14, 34], peripheral blood [25, 35], lungs [36–38], muscle [39, 40] and other sources. Previous studies suggested that the administration of SF-MSCs as a viable therapy is utilized in cartilage degeneration of osteoarthritis (OA) [33, 41, 42]. DMSCs improve the immunoregulatory effects on T and B lymphocyte responses based on decreasing CD4⁺ T lymphocyte proliferation, intracellular Interferon-gamma (IFN-γ), and IL-17 secretion in primary Sjögren's syndrome [43]. MSCs display different functions for different organs, differentiating into other cell lineages to improve inflammatory response and tissue damage [44–48]. Therefore, MSCs are significant mediators in sustaining tissue homeostasis and improving tissue integrity.

MSCs derived from adult tissues, such as bone marrow, peripheral blood, adipose tissue, etc.) and prenatal tissues

(particular parts of the umbilical cord and placenta) have some differences in their cell biological properties, proliferative capacities and surface marker expression. The prenatal tissues with various advantages, including the availability, avoided invasive procedures and ethical problems are widely utilized in the preclinical and clincal research. Baksh et al. found that the HUC-MSCs have a higher proliferation capacity compared with the BMSCs [49]. Moreover, more reports suggested that HUC-MSCs show a higher proliferation capacity than BMSCs [50, 51]. It has been reported that HP-MSCs exhibit a better engraftment and expansion and capacity compared with BMSCs [52]. In recent years, MSCs from other sources, such as AMSCs, HUCB-MSCs, HUC-MSCs, and HPMSCs, have received considerable attention due to better immunomodulatory characteristics and proliferation rate as compared to BMSCs [53, 54]. MSCs can be identified by distinguishing a specific panel of positive cell surface antigens (CD29, CD73, CD90, and CD105) and negative cell surface markers (CD14, CD11b, CD19, CD34, CD45, CD79α, and HLA-DR). Moreover, multiple differentiation potentials to the osteoblasts, adipocytes, and chondroblasts is a central standard for defining MSCs in vitro [55–57]. MSCs display crucial function in the repairation of damaged tissue by mechanisms of immunosuppression, anti-apoptotic, anti-inflammatory, anti-fibrotic, pro-angiogenic, antitumorigenic, neuroprotective, antibacterial, and chemo-attractive effects [58–60]. These unique characteristics endow MSCs as a promising candidate in regenerative medicine [61], and with absorbing therapeutic potential in inflammatory diseases [58, 62], and cancer [63, 64].

Initially, MSCs were primarily utilized as a promising cell therapy for tissue repair in regenerative medicine [59, 65]. It has been shown that they have been approved in clinical research for autoimmune diseases, including Crohn's disease, lupus, and graft-versus-host disease (GVHD) [58, 66]. Likewise, MSCs have been investigated for therapeutic potential against different diseases, including stroke, myocardial infarction, liver cirrhosis, multiple sclerosis, diabetes, lung injuries, and cancer, both in preclinical and human translational studies [58, 59]. Moreover, MSCs as potential treatments possess immunomodulatory functions except for being multi-potent, which have been investigated for various immune diseases. MSCs robustly exert their immunomodulation through direct cell-cell contact and their secretion ability with immune cells in the innate and adaptive immune systems [67]. Up to now, the immunomodulatory function of MSCs across tissue sources and species has been well-reproduced and clinically relevant [68]. In this review, we summarise the recent immunomodulatory properties of MSCs in different diseases and discuss the potential therapeutic targets in various diseases.



Immunomodulation of MSCs

Recent findings indicated that MSCs conduct immunomodulatory activity, associated with monocytes and regulatory T cells (Tregs) through the special cytokine-independent mechanism [58, 69-71]. AMSCs display more potential immunomodulatory effects than BMSCs, suggesting that AMSCs could serve as a better alternative for immunomodulatory therapy [54, 69]. In comparison, HUC-MSCs have shown a minimal response to the allogeneic immune risk in vivo, regarding HUC-MSCs as a suitable therapeutic candidate [72]. Some reports also found that xenogenic human MSCs have the potential to display positive ability in mouse tumor diseases, sheep tibia bone defect, porcine osteochondral reconstitution, and cartilage regeneration in canines [73-75]. Xenogenic human MSCs with lower weakly immunogenicity possess the therapeutic potential in preclinical and clincal studies. Therefore, we summarize the immunomodulatory effects of naive and modified MSCs and review the current understanding of their immunomodulatory aspects.

Immunomodulation via Intercellular Liaison

Many preclinical studies have reported that the administration of MSCs is involved in the innate and adaptive immune responses through communication with the immune cells comprised of T cells, B cells, neutrophils monocytes, macrophages, natural killer (NK) cells, and dendritic cells (DCs), exerting immunomodulatory effects (Fig. 1) through intercellular liaison and paracrine action [76–78].

Intercellular Liaison in Adaptive Immunity

As shown in in-vitro experiments, BMSCs physically hindered naive and memory-T cell reactions from associating with antigen-presenting cells (APCs) in a noncognate fashion [79]. This function was exhibited through the secretory enhancement of vascular cell adhesion molecule-1 (VCAM-1) and intercellular adhesion molecule-1 (ICAM-1), in favor of T cells activation and leukocytes recruitment to the inflammation site, which plays a critical role in immunosuppressive capacity [80, 81]. Furthermore, BMSCs cocultured with activated T cells induce Th17 lymphocytes

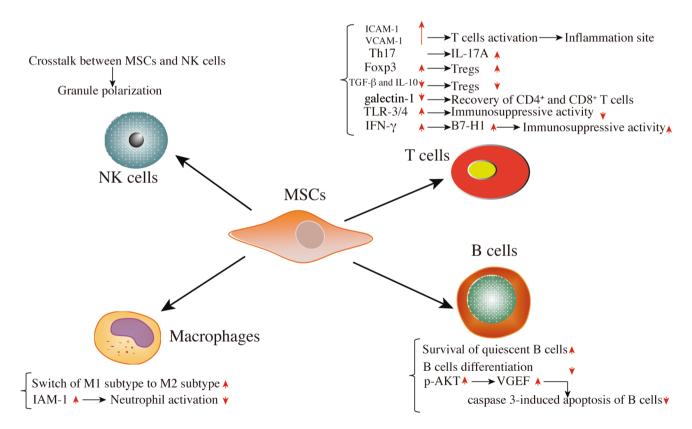


Fig. 1 Cell-to-cell contact of immunomodulation in adaptive immunity and innate immunity. MSCs display the immunomodulatory effects mainly via mutual effect with immune cells such as T cells, B cells, natural killer (NK) cells, macrophages, etc. and cell-to-cell contact is associated with the modulation of protein expression. Abbreviations: Foxp3, Forkhead box P3; ICAM-1, intercellular adhesion

molecule-1; VCAM-1, vascular cell adhesion molecule-1; TLR, Toll-like receptors; IFN, interferon; B7-H1, an inhibitory surface molecule in stem cells; VEGF, vascular endothelial growth factor; IAM-1, intercellular adhesion molecule-1; p-AKT, phosphorylated protein kinase B; TGF- β , transforming growth factor- β ; IL, interleukin; B7-H1, an inhibitory surface molecule in stem cells



expressing interleukin-17 A (IL-17 A) [82]. The activation of the Notch1/forkhead box P3 (FOXP3) pathway was observed in CD4⁺ T cells co-cultured with MSCs, mediating regulatory T cells (Tregs) induction by increasing the amount of CD4+ CD25(high) FOXP3+ cells [83, 84]. However, inhibition of TGF-β and IL-10 simultaneously suppressed Tregs induction in co-cultured cells, indicating that the two aspects display an essential effect in the immunetolerance mechanism [84]. Furthermore, galectin-1 protein is primarily expressed in MSCs, influencing cytokine secretion in GVHD and autoimmunity. The immunomodulatory properties of MSCs to the allogeneic T cells were lost after the knockdown of galectin-1, contributing to the recovery of CD4⁺ and CD8⁺ T cells [85, 86]. Also, the high Tolllike receptors (TLRs), including TLR-3 and TLR-4, are expressed on the membrane of MSCs, the function of whose ligands is responsible for the activity of nuclear factor kappaB (NF-κB) and the secretion of pro-inflammatory cytokines, including CXCL10, IL-6, and IL-8. Expression of TLR-3 and TLR-4 in MSCs effectively inhibited the immunosuppressive activity of MSCs, resulting in efficient T cells response in dangerous infections, such as RNA viruses and bacteria infections [87].

It has been shown that the function of T cells is critical to the immunomodulation of MSCs in vivo mice models. MSCs derived from compact bone (CB) have synergistic anti-tumor efficacy in combination with an immune-activating fusion protein which is associated with the activation of CD4⁺ and CD8⁺ T cells and the inhibition of Tregs in the microenvironment of a syngeneic orthotopic ovarian cancer mouse model [88, 89]. In the lipopolysaccharide (LPS) and immune response-mediated abortion models, MSCs through cell-to-cell contact with the proinflammatory macrophages boost the inhibitive adjusting and proliferation of T cells and switch macrophages to M2 phenotype, mediating abortion relief, demonstrating MSCs potential application in treating the recurrent miscarriages in clinical research [90, 91]. IFN-γ is derived from activated T cells and is regarded as a renowned proinflammatory cytokine. The IFN-γ on MSCs induces the enhancement of an inhibitory surface molecule named B7-H1 on MSCs, enhancing the MSCs' immunosuppressive properties. On the contrary, primed MSCs with activated T cells from IFN-γ ^{-/-} mice showed a remarkably weakened capacity to restrain the proliferation of T cells in MSCs' immunosuppressive function through the mechanism of intercellular liaison [92].

Besides T cells, there is an important connection between MSCs and B cells through intercellular liaison. Humanderived AMSCs are based on contact-dependent mechanisms to improve the survival of quiescent B cells and suppress B cells differentiation dependent on T cells [93, 94]. Based on improved phosphorylated protein kinase B

(p-AKT) by MSCs within the B cells, AMSCs enhance the secretion of vascular endothelial growth factor (VEGF) to inhibit the caspase 3-induced apoptosis of B cells [95]. Besides, MSCs can block the proliferation of B cells by inhibiting the cell G0/G1 phase cycle of B cells, associated with the phosphorylated extracellular response kinase (p-ERK) and p38 pathways [96].

Intercellular Liaison in Innate Immunity

Except for the adaptive immune system, MSCs also act on the innate immune system based on the mechanism of intercellular liaison. Tracking studies indicated that viable HUC-MSCs appeared in the lungs directly after intravenous injection. Most HUC-MSCs were dead and then quickly phagocytosed by monocytes after 24 h. Cocultured studies of MSCs either using a transwell or directly with two kinds of NK cell lines demonstrated that granule polarization is either induced or restrained through the special crosstalk of MSCs on the two NK cell lines [97]. Moreover, some studies found that AMSCs can switch an activated-M1 proinflammatory phenotype of macrophages to its anti-inflammatory M2 phenotype associated with the specific function of prostaglandin E2 (PGE2) [98, 99]. Furthermore, MSCs hold substantial therapeutic promise to prevent unrestrained neutrophil activation-induced tissue damage depending on the neutrophils' engulfment of intercellular adhesion molecule-1 (IAM-1) derived from MSCs [100, 101].

Various known mechanisms through which MSCs' exert immunomodulatory activity are illustrated in Fig. 1. However, it require extensive preclinical studies to be translated in clinical research.

Immunomodulation via Paracrine Activity

The Secretomes of MSCs

MSCs also exert or convey regulatory messages displaying their immunomodulatory functions by releasing secretomes [76, 78, 102]. This secretome derived from MSCs possessing immunomodulatory functions consists of a battery of cytokines, growth factors, chemokines, and extracellular vesicles (MSCs-EVs), which interact with immune cells of the innate and adaptive systems and regulate the immune and cancer-cell function [67, 103–106]. According to their size and origin, encapsulated paracrine molecules in MSCs-EVs are usually divided into exosomes (30-120 nm) from the multivesicular endosomes, microvesicles (MVs) (100–1000 nm) originating from the plasma membrane, and apoptotic bodies [67, 102, 107]. Interactions between MSC-EVs and immune cells can be regarded as an ideal therapeutic prospect for inflammatory, infectious, and autoimmune



diseases [108]. Although MSC-EVs are similar to the parent MSCs in the immunoregulatory functions [108], the paracrine function of the MSCs is based on the appropriate cultured conditions [109]. Therefore, it is critical to know the paracrine function of MSCs to improve their immunomodulatory effects (Table 1).

The Paracrine Function of MSCs in the Adaptive Immune System

MSCs exert immunomodulatory properties on the adaptive immune system depending on their paracrine secretion. MSCs restrain the differentiation of T helper 17 (Th17) cells by mediating the secretion of IL-10, PGE2, and trimethylation of histone to regulate inflammation, which was enhanced in coculturing with Th17 cells and inhibiting expression of TNF-α, IL-17, IL-22, and IFN-γ derived from Th17 cells [110, 111]. It has been shown that both BMSCs priming with cytokines and cell ratio (BMSCs/T cells), moderated their cytokine profiles on the generation of Th17 lymphocytes [82]. Nevertheless, the mechanisms of the mutual effect of MSCs with Th17 cells are not incompletely resolved. Wang et al. have found that modified MSCs with IL-25 knockdown did not mediate Th17 suppression in vitro and in vivo experiments, further verifying that the regulation of the IL-25/STAT3/PD-L1 axis can be identified as a candidate therapeutic target [112, 113]. Indoleamine 2,3-dioxygenase (IDO), secreted by MSCs, is known to suppress T cells and mediate Tregs generation, partly in charge of inducing tolerance of kidney allograft [114]. Moreover, PD-1 ligands (PD-L1 and PD-L2) derived from MSCs restrain the secretion of IL-2 and activation of CD4⁺ T cells and mediate irreversible hyporesponsiveness and cell death to exert immunosuppressive potential on induction of peripheral tolerance and T cell behavior for the treatment of immunological disorders [115].

Table 1 The secretomes of MSCs

cytokines	chemokines	Growth factors	Other factors
	-		
IFN-γ, TGF-β,	CCL1, CCL2,	BDGF,	phospholipids,
TNF-α, LIF,	CCL5, CCL8,	NGF,	Adrenomedul-
MIF, OSM,	CCL11, CCL15,	GDNF,	lin, Adiponectin,
G-CSF,	CCL16, CCL18,	PIGF,	Osteoprotegerin,
M-CSF, GM-	CCL22~24,	PDGF,	miRNAs,mRNAs,
CSF FLT3LG,	CCL26, CXCL1-	VEGF,	long non-coding
SCF, Throm-	3, CXCL5,	HGF, EGF,	RNAs, CXCR3,
bopoietin,	CXCL6, CXCL8,	IGF-1,	PGE2, PAI-1,
TSG-6, IL-1α,	CXCL11~13,	FGF-2,	MMP1, MMP3,
IL-1β, IL-2,	CX3CL1, XCL1	FGF-4,	MMP9, MMP10,
IL-3, IL-6~8,		FGF-7,	MMP13, TIMP-
IL-10-13,		FGF-9,	1~4, Leptin,
IL-16,		BMP-7	IGFBP-1~4

The Paracrine Function of MSCs in the Innate Immune System

MSCs communicate with NK cells through the acute suppression of IL-2-mediated resting NK cell proliferation and improve NK function of the innate immune system [116, 117]. Moreover, MSCs block the cytotoxic activity or cytokine production associated with the key mediators, including IDO and PGE2 [118, 119]. It has also been shown that MSCs improve the capacity of IL-12/IL-18-motivated NK cells secreting IFN-y that can potentially contribute to defending infections for tissue reconstruction at the injured spot [120]. Additionally, MSC-derived IL-6 protects neutrophils from apoptosis, signals via the activation of STAT-3 to preserve their effector functions, and prevents oxidative metabolism in the BM niche [121]. MSC-derived exosomes (MSCs-EX) mainly improve neutrophil viability, whereas MSCs-conditioned media (MSCs-CM) essentially improve the neutrophil's function, suggesting that both MSCs-EX and MSCs-CM are beneficial for increasing immunity through the improvement of neutrophil's function and survival [122]. MSCs derived from human BM and salivary gland stimulated by LPS boost the anti-microbial functions of neutrophils by releasing high levels of macrophage migration inhibitory factor (MIF) and inflammatory cytokines (IL-6 and IL-8) and eliminating the infection and inflammation [123].

MSCs show more potent immunomodulatory effects due to having higher levels of multiple cytokines secretion, including IL-6 and TGF-β1, to suppress the proliferation of stimulated PBMCs, and the differentiation of immature DCs derived from monocyte and induce the IL-10 secretion by monocytes [124, 125]. However, MSCs were empowered by MSCs-derived PGE2 to restrain the differentiation of monocytes to mature myeloid DCs, and NS-398, the PGE2 inhibitor, inhibited PGE2 production and restored DC differentiation and function [126]. It has been shown that MSCs-EVs prevent DC maturation and inhibit antigen uptake by immature DCs resulting in the downregulated expression and secretion of mature and activated DCs markers (CD38, CD80, and CD83), upregulated production of anti-inflammatory cytokine TGF-β, and pro-inflammatory cytokines (IL-6 and IL-12p70) [78]. MSCs-EVs, known as effective therapeutic agents, display a crucial function in eliciting macrophage to anti-inflammatory M2 polarization possessing effective anti-inflammatory properties by decreasing expression levels of IL-23 and IL-22 [127, 128]. Activated BMSCs with TNF-α or LPS reprogram macrophages via the higher release of PGE2, affecting the macrophages associated with the receptors of PGEP2 and PGEP4, whereas, both PGEP2 and PGEP4 receptor antagonists inhibited the increased secretion of IL-10 [129]. Human PMSCs can



transit macrophages into a typical anti-inflammatory M2 phenotype from a pro-inflammatory M1, which is associated with the secretion changes of IL-1 β , IL-10, IL-12p70, and macrophage inflammatory protein-1 alpha (MIP-1 α) and partly induced by soluble molecules, including progesterone and glucocorticoid receptors [130].

Different secretomes derived from MSCs comprised of cytokines, growth factors, and chemokines have participated in adaptive and innate immune systems to exert immunomodulatory effects (Fig. 2).

Improvement of Immunomodulatory Capabilities and Therapeutic Efficacy by Modified MSCs

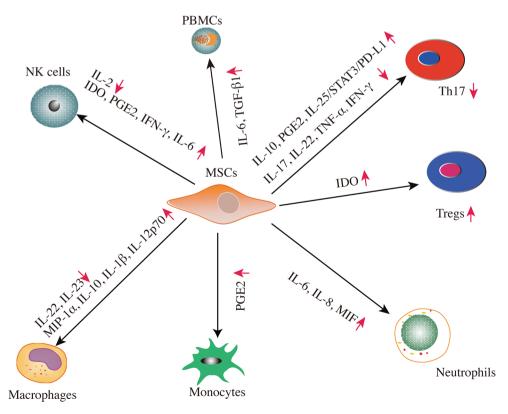
Increasing evidence has demonstrated that preconditioned MSCs can improve their paracrine potency and therapeutic abilities. It has been shown that MSCs pretreated with hypoxia, growth factors, heat shock protein (Hsp), LPS, pharmacological agents, chemical agents, serum deprivation, or inflammatory stimulation, have the ability to improve their potency and survival by increasing the expression levels of cytoprotective genes, and secretion of reparative factors [131, 132]. As such, preconditioning conditions can modulate MSCs' secretomes in vitro culture [102]. It is critical to analyze the influence of secretome on varied biological processes, including immunomodulation, angiogenesis,

Fig. 2 Paracrine activity of immunomodulation in adaptive immunity and innate immunity. MSCs exert immunomodulatory effects through the interactions with immune cells such as T cells, natural killer (NK) cells, macrophages, monocytes, PBMCs and neutrophils, through paracrine activity. MSCs' secretome secret a battery of cytokines, growth factors, and chemokines for playing their immunomodulatory function. Abbreviations: PBMCs, peripheral blood mononuclear cells; IL, interleukin; TGF-β1, transforming growth factor-β1; PGE2, prostaglandin E2; IFN, interferon; MIF, macrophage migration inhibitory factor; IDO, Indoleamine 2,3-dioxygenase; Th17, T helper 17; Tregs, regulatory T cells; MIP-1α, macrophage inflammatory protein-1 alpha

anti-fibrotic, neurogenesis, wound healing, tissue repair, and anti-tumor for tissue repairation and regeneration [107].

MSCs Preconditioning with Hypoxia

Preconditioned MSCs treatment with hypoxia has shown to have the rapeutic potential on the MSCs' immune phenotype. Specifically, MSCs preconditioned with hypoxia increase paracrine effects, especially the production of angiogenic factors and HGF observed in pulmonary fibrosis and acute kidney injury [133, 134]. Overexpressing hypoxia-inducible factor 1 α (HIF-1 α) of dental MSCs (HIF-MSCs) can inhibit DCs differentiation, resist NK-cell-induced lysis to facilitate the therapeutic ability of HIF-MSCs, and improve more monocytes migration to differentiate into suppressor macrophages [135]. Hypoxia-preconditioned CB MSCs (HP-CB-MSCs) possess effective potency to restrain T cell proliferation, including total (CD3⁺), CD4⁺, and CD8⁺ T cells. HP-CB-MSCs, accompanied by preconditioning of selective proinflammatory cytokines, such as IL-17 A TNFα, and IFN-γ, increase superior inhibition of CD8⁺ T cells compared with CD4⁺ T cells [136]. Under hypoxia conditions, MSCs improve the production of SDF-1α and VEGF, and alter the secretion of angiogenic signals (IL-8, uPA, Angiogenin, and Serpin-1) for the response of hypoxia, which contributes to more endothelial cells infiltrating into an MSCs bio-artificial scaffold for dermal regeneration [137]. Moreover, MSCs from human anticoagulated





whole BM are positively impacted by hypoxic culturing on the transcriptome and cell fitness, possibly boosting inherent properties for developing cellular therapies in treating the orthopedic diseases [138]. MSCs derived from rat BM facilitate the characteristics of stem cells accompanied by upregulation of IDO and promote the expansion of Tregs for immunosuppressive function when MSCs are cocultured with CD4⁺/allogeneic endothelial cells [139]. Additionally, primed MSCs by hypoxia and calcium ions (HC-MSCs) have demonstrated a resistance to passage-dependent senescence, which is induced through the p53/p21 cascade and monocyte chemoattractant protein-1 (MCP-1) and improved a large secretion of pro-angiogenic and immunomodulatory factors, contributing to inhibiting the proliferation of T cell. Transplantation of HC-MSCs effectively moderated allogeneic conflicts in the utilization of a humanized GVHD mice model by improving survival and weight loss and reducing histopathologic damage in target organs of GVHD [140, 141].

MSCs Preconditioned with Immunomodulatory Factors

Another positive approach for pretreatment is priming with immunomodulatory factors. Modified MSCs with TNF-α and IFN-γ can totally reverse the immunomodulatory potency associated with the MSCs' pro-inflammatory effect in palmitate in vitro [142]. TNF- α and IFN- γ , the key cytokines involved in MSC activation, could improve the expression of immunoregulatory molecules for communication through cell-cell contact and EVs [143]. Three-dimensional spheroidal MSCs primed with the heparin-microparticle transfer of IFN-γ induce continuing expression of IDO, enhancing the immunomodulatory capability, which suppresses the activation and proliferation of T cells [144]. MSCs primed with IFN-y have enhanced immunosuppressive capacity and immunosuppressive function, suppressing the proliferation of T cells through the IFN-γ/JAK/STAT1 pathway [145, 146]. Moreover, membrane particles (MPs) derived from pretreated MSCs by IFN-γ (MP-MSCs-γ) can enhance the percentage of CD90⁺ and anti-inflammatory PD-L1 monocytes, and the mRNA expression of PD-L1 in monocytes. The results demonstrated that MSCs-derived MPs possess the immunomodulatory capability and potential and can be regarded as a novel therapeutic strategy in treating immunological disorders [147]. Recently, Bolhassani et al. have demonstrated that BMSCs transfected with small Hsp 27 (sHsp27) and E7 oncoprotein DNA vaccinations (E7ODV) restrain tumor growth and increase the responses of E7-specific T cells in a tumor mouse model. This study suggested that MSCs-based vaccinations with particular modifications have the potential to treat HPV-associated cancers and can be an effective strategy for immunotherapy and protective function [148].

Engineered MSCs Improve the Immunomodulation

Many studies have reported that MSCs possessing intrinsic capacities can be used in immune-associated diseases. As for utilizing MSCs for targeting diseases, investigators have focused mainly on gene-delivery vehicles, like IFNs and ILs, and oncolytic viruses (OV) to improve the immuno-modulatory function of MSCs, empowering them to target biologics.

It has been shown that engineered MSCs with the IFN-β improve the survival of glioblastoma (GBM) mice, resistant CNS malignancies, via enhancing CD8 T cells' selective postsurgical infiltration and directly inducing tumor cells' cell-cycle arrest [149, 150]. It has demonstrated that it is essential to develop cancer immunotherapies syngeneic for tumor resection models and highlight the translational potential of local delivery of cellular immunotherapy for cancer treatment [149]. Similarly, Relation et al. also found that engineered MSCs with pro-inflammatory cytokine IFN-γ not only deliver IFN-γ directly to the tumor microenvironment but also avoid systemic toxicity, which decreases tumor growth rate and increases survival via inflammatory M1 macrophage polarization (the increased IL-17 and IL-23p19, M1 polarization markers) in vivo and in vitro [151]. Likewise, spheroidal MSCs transfected with a potent antiinflammatory cytokine IL-4 enhanced anti-inflammatory effects and chondroprotective in OA rats model and in an OA chondrocyte model [152], whereas MSCs engineered anti-tumor activity cytokine, IL-12, prolong the tumor-bearing mouse survival and decrease tumor growth after intravenous injection, including carcinoma, renal cell carcinoma (RCC), breast tumor, and melanoma [73, 153]. Systemic administration of MSCs engineered with lentivirus vector expressing murine IL-15 significantly improved the retardation of syngeneic mouse pancreatic tumor and melanoma growth and prolonged the survival in tumor-bearing mice. Additionally, such cured mice resisted the pancreatic tumor rechallenge [154, 155]. Likewise, genetically engineered MSCs with IL-21 promoted the induction of NK cells and effector T cells and potently restrained the immune suppressor cells, which prevented the tumor nodules in a mouse model of disseminated B cells lymphoma [156]. Additionally, it has been shown that in the streptozotocin-induced diabetic rats model, the injection of stromal cell-derived factor-1 (SDF-1)-modified MSCs (SDF-1-MSCs) can moderate erectile dysfunction. The beneficial effects of SDF-1-MSCs were associated with the enhancement of protein expression levels, including p-AKT, AKT, phosphorylated neuronal nitric oxide synthase (p-nNOS), nNOS, VEGF,



B-cell lymphoma-2 (Bcl-2), basic fibroblast growth factor (bFGF), and decline of protein expression levels of apoptosis factors, including the caspase-3 and Bcl-2-associated x (Bax) [157]. Besides, exosomes of AMSCs infected by miR-199a lentivirus have successfully delivered miR-199a to hepatocellular carcinoma (HCC) cells, and remarkably improved sensitivity of chemotherapeutic agents to cancer cells for the improvement of HCC chemosensitivity via the inhibition of the mammalian target of rapamycin (mTOR) signaling pathway [158].

Additionally, MSCs as a possible means can deliver oncolytic viruses (OVs), which can overcome host antiviral immunity [159]. MSCs armed by oncolytic herpes simplex virus (oHSV) successfully delivered viral progeny to target melanoma brain metastasis, ultimately inhibiting tumor growth rate and prolonging survival of brain tumor-bearing mice [159, 160]. Although many research examples of immunomodulation and delivery are successful, we still need more work. The promising strategies that are commonly being addressed are displayed in Fig. 3.

MSCs-induced Immunomodulation in Clinical Studies and Trials

39 studies have been performed to investigate the immunomodulation of MSCs in clinical trials. The diseases they focus on are infections, diabetes, knee osteoarthritis, relapsing-remitting multiple sclerosis (RRMS), Crohn's Disease and etc. which represented the most common immunes system diseases in the clinical (Table 2). A recent phase I/IIa trial (Identifier: https://clinicaltrials.gov/ct2/show/ NCT02351011) utilized a 50 million dose of BMSCs per person to 12 patients with late-stage Kellgren-Lawrence knee osteoarthritis. The results suggested that the administration of autologous BMSCs derived from BM of the posterior superior iliac spine is effective in quality of life, safe for intra-articular injection with no serious adverse events, and contribute to the improvements of synovial inflammation and pain [161]. Likewise, a conducted phase II/ III study (Clinical Trial Number: (UMIN-CTR Numberiii: UMIN000006719) indicated that BMSC transplantation enhanced the overall survival rate in twenty-five patients with acute GVHD (steroid-refractory grade III or IV) with no observed adverse events [162]. In an open-label phase IIa study (Clinical Trial Number: NCT00395200), autologous BMSCs with a dose of 1.6×10⁶ cells per kg bodyweight were safely administrated to patients with secondary progressive multiple sclerosis (SPMS) and the improvement of structure, function, and physiology demonstrated the neuroprotective effects [163]. In phase I/IIa studies for 34 adult patients with atopic dermatitis from moderate to severe (AD), subcutaneously infused high-dose HUC-MSCs $(5.0 \times 10^7 \text{cells})$ therapeutically improved the disease features, suggesting the significant reduction of severity scoring and eczema area, severity index for atopic dermatitis, pruritus score, serum IgE levels and the number of blood eosinophils [164]. A pilot study associated with serial intrafistular administration of autologous BMSCs for 12 consecutive outpatients with refractory fistulizing Crohn's disease demonstrated that local BMSCs therapy is feasible, safe, and beneficial for this disease by modulating mucosal T cell apoptotic rate and 7 patients with sustained complete closure of fistula tracks were observed in 12 patients [165]. A triumphant randomized and double-blind controlled phase III clinical trial (ClinicalTrials.gov, registered number NCT01541579) utilizing expanded allogeneic AMSCs (a single intralesional injection of 1.2×10^9 cells) to treat 212 Crohn's disease patients possessing refractory complex perianal fistulas suggested that a restorative period of at least 24 h post-thawing culture promoted MSCs' quality [166–169].

However, it is evident that the induction of some side effects may be mediated by MSCs-related treatments. For instance, recent preclinical research demonstrated that the acute hypothermia of the brain microenvironment moderated the function of intranasal administered MSCs, enhancing long-lasting motor-cognitive deficits and endothelial activation, resulting in a pro-inflammatory environment, increasing the peripheral immune cell infiltration, which exacerbated the brain injury in the hypoxic-ischemic brain [170].

Hopeful Strategies have the Potential to Improve the MSCs Efficacy of Future Trials

Preclinical reports have suggested that different administrated ways and modified factors of MSCs have the therapeutical potential for different diseases. Recent studies have demonstrated that MSCs homing to injury sites by intravenously delivering MSCs or depending on migration and mobilization of endogenous MSCs was investigated in a feeder layer-based transwell-based model. The overexpression of fibroblast growth factor 21 (FGF21) in MSCs (FGF21-MSC) exhibited the improvement of the MSCs3 homing ability to injury sites in a traumatic brain injury (TBI) mouse model [181]. Analogously, colony-stimulating factor 2 (CSF-2), a hematopoietic growth factor, improved the differentiation and migratory capacity of MSCs and promoted the therapeutic effects via PI3K/AKT and/or FAK/ ERK1/2 signal axis [182]. A proinflammation cytokine IL-1β-mediated more production of matrix metalloproteinase-1 (MMP-1) improved the migration of MSCs through



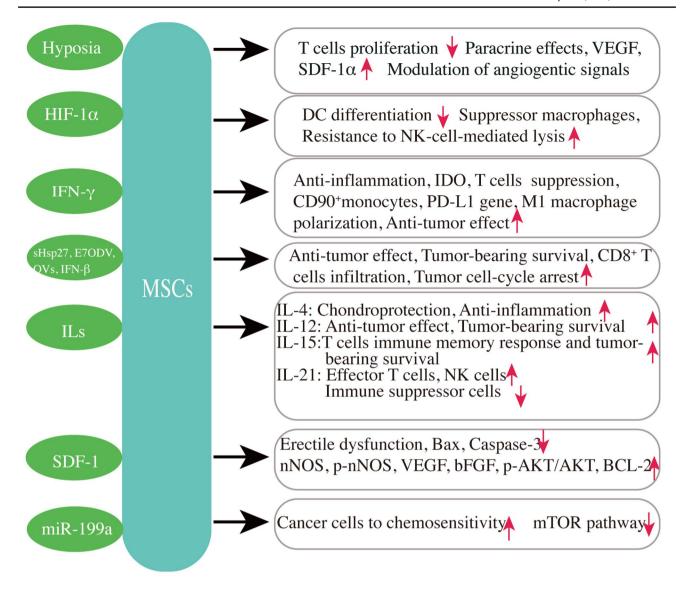


Fig. 3 Modified MSCs to increase their immunomodulatory functions and therapeutic efficacy. MSCs exert better immunomodulatory effects through modification compared with signal MSCs administration. MSCs pretreated with hypoxia. heat shock. inflammatory stimulation, and et al. have the poten-tial to increase their therapeutic potency, the expression of cytoprotective genes, and secretion of repara-tive factor. Abbreviations: IL, intericukin; IFN, interferon;

IDO, Indolcamine 23-dioxygenase; Bax, 13cl-2-associated x; nNOS, neuronal nitric oxide synthase; VEGF, vascular endothelial growth factor; bFGF, basic fibroblast growth factor; BCL-2, B-cell lymphoma-2; AKT, protein kinase B; HIF-1α, hypoxia-inducible factor 1α; sHsp27, heat shock proteins 27; E7ODV, E7 oncoprotein DNA vaccinations; OVs, oncolytic viruses; SDF-1, stromal cell-derived factor-1

the activation of the protease-activated receptor 1 (PAR1) and G-protein-coupled signaling pathways [183]. Intriguingly, intranasally administered exosomes derived from MSCs particularly accumulate as an inflammatory-driven style in injured brain sites of relevant murine models for up to 4 days post-administration, which is highly associated with the neuro-inflammatory signals in the various brain pathological regions of different diseases, such as stroke, autism, Parkinson's disease, and Alzheimer's disease [184]. Hypoxic preconditioning as a boosting factor enhanced MSCs' migration ability and survival. Preconditioned

MSCs with hypoxia evoked an increased LincRNA-p21 production, pivotal in MSCs homing through the HIF- 1α /CXCR4 and CXCR7 pathway in vitro [185].

Another strategy to improve the MSCs' effectiveness is to facilitate the ways of their cryopreservation. More specifically, freshly thawed MSCs following cryopreservation stunted functional characteristics, including the upregulation of metabolic activity and apoptosis and downregulation of cell proliferation, immunosuppressive capabilities, clonogenic capacity, and critical regenerative genes [14, 186, 187]. Notably, a 24-hour acclimation period reactivated



	Table 2	The immunomodulator	behaviour of MSCs in recent	clinical and	preclinical studi
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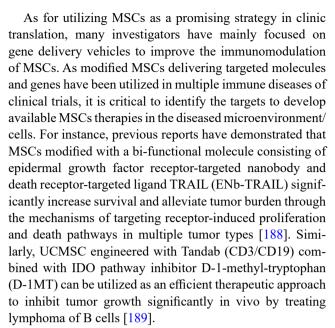
Disease	Origin of MSCs	Treatment routes	Identified Immunomodulatory Effect		References	
			Preclinical research			
Ovarian cancer mice model	Mice compact bone MSCs	Intraperitoneal injection	Activation of CD4 ⁺ and CD8 ⁺ T cells and the inhibition of Tregs		[88, 89]	
kidney allograft	Mice BMSCs	Intravenous injection	Suppression of T cells and improvement of Tregs generation		[114]	
Septic Mice	HUC-MSCs	Intravenous injection	Improving anti-inflammatory effects, increasing the Treg cells, promoting macrophage phagocytosis, enhancing the reparative macrophage, secreting more VEGF		[171]	
Asthmatic mice	Mice BMSCs	Intraperitoneal injection	Better immunomodulatory effect		[172]	
Mice With acute liver failure	Mice AMSCs	intravenous injection	Removeing debris and improving IL-6 liver regeneration		[173]	
Mice inflamma- tory bowel disease (IBD)	Intestine Peyer's patch MSCs	Intravenous injection	Improve the symptoms of mice IBD model		[174]	
GVHD mice	Human MSCs	intravenous injection	Engagement of PD-1 ligands, reduction of the clinical score and prolonging the survival of mice		[175]	
Acute respiratory distress syndrome (ARDS) mice	HUC-MSCs	Intravenous injection	Ameliorated lung injury of ARDS and inactived YAP to facilitate type II alveolar epithelial cells (AECII) differentiation.		[176]	
Mice alkali-burn injury model	Mice BMSCs	Local injection	Enhancement of their immunomodulatory and pro-angiogenic capacity, decrease of pro-inflammatory IL-1β		[177, 178]	
Host's immune system	HUC-MSCs	Intravenous injection	Elevation of neutrophil percentage and concentration of neutrophil chemoattractants Clinical Trials		[179]	
Disease	Origin of MSCs	Treatment routes	Clinical Trial Number	Phase	Identified Immuno- modulatory Effect	Refer- ences
Osteoarthritis	Autologous BMSCs	Intra-articular infusion	NCT02351011	I/II clinical trial	Decrease of synovial inflammation and pain	[161]
Steroid-refractory grade III or IV acute GVHD	Autologous BMSCs	Intravenous injection	UMIN000006719	II/III clinical trial	Enhancement of the overall survival rate	[162]
Multiple sclerosis	Autologous BMSCs	Intravenous injection	NCT00395200	IIa clinical trial	Improvement of structure, function, and physiology	[163]
Moderate to severe atopic dermatitis	HUC-MSCs	Subcutaneous injection	NCT01927705	I/II clinical trial	Reduction of severity	[164]
Crohn's disease	Autologous BMSCs	Intrafistular injections	1	/	A feasible, safe and beneficial therapy	[165]
Crohn's disease	Allogeneic AMSCs	Intrafistular injections	NCT01541579	III clinical trial	Long-term Efficacy and	[166– 169]
Gastrointestinal adenocarcinoma	Autologous BMSCs	Intravenous injection	European Union Clinical Trials Register 2012-003741-15	I/II clinical trial	Favorable safety and tolerability	[180]



the thawed cells, diminishing apoptosis and enhancing the high-mobility group box (HMOX-1) and tumor necrosis factor-stimulated gene-6 (TSG-6) genes expression with anti-inflammatory effects and VEGF gene expression with angiogenic effects [186]. Consequently, MSCs should be standardized to implement robust quality and reliability, particularly in large-scale frozen MSCs products. Recent research results concluded that administering two freezing steps accompanied by a preceding cell-culture phase of at least one passage did not impair the essential quality or parameter attributes of the MSCs' ultimate cryopreserved and thawed product [187]. A triumphant randomized and double-blind controlled phase III clinical trial (ClinicalTrials.gov, registered number NCT01541579) utilizing expanded allogeneic AMSCs (a single intralesional injection of 1.2×10^9 cells) to treat 212 Crohn's disease patients possessing refractory complex perianal fistulas suggested that a restorative period of at least 24 h post-thawing culture promoted MSCs' quality [166–169]. Hence, a 24-hour acclimation period of culture for cryopreserved cells is crucial to reactivate the thawed MSCs for restoration of their impaired MSCs' capabilities.

Conclusions and Future Perspectives

Due to MSCs' tropism with other cell types and immunomodulatory attributes, the administration of MSCs has been a novel therapeutic strategy in preclinical and clinical research. Due to the immunomodulatory capability of MSCs modified by the mutual effect and various inflammatory cytokines with other immune cells, MSCs could mediate the recovery of various inflammatory disorders. The major mechanisms associated with the immunomodulatory capability of MSCs include intercellular liaison and paracrine activity, in which process MSCs are modified by inflammatory stimuli, extracellular vesicles, cytokines, chemokines, or co-culture with other immune cells. Consequently, the transplantation of MSCs can be made an administrable, potential, and feasible method of cell-free therapy. However, it is still hard to understand the mechanisms of how the variability of MSC impacts their immunomodulatory capability. The administration of MSCs likely mediates highly intricate immunomodulatory mechanisms through their secretome. The following work should explore the effects of chronic inflammatory cytokines and other factors on MSCs-induced immunomodulatory effects. These methods can identify new preconditioned approaches in promoting the therapeutic efficacy of MSCs and weaken the variation of their paracrine capability, especially in clinical application.



A real challenge faced by MSCs research is the conflict between in vitro and in vivo results. Liu et al. reported that osteogenic cells (OCs) differentiated from MSCs possessed their immunomodulatory ability and were immuneprivileged in vitro. However, such immunomodulatory and immune-privileged abilities were lost after OCs implantation in vivo experiments following an osteogenesis model of New Zealand white rabbit [190]. A study from Isakova et al. found that the engraftment of allogeneic MSCs had weakly immunogenic after post-transplantation compared with autologous MSCs in a rhesus macaque model, which limited their durable engraftment levels [191]. These discrepancies indicate the significance of observing the MSCs' immunogenicity after transferring from in vitro to in vivo experiments and it is critical to maintaining an effective therapy in further clinical applications. Eventually, by enhancing immunomodulatory potential, further investigating homing mechanisms, and improving cryopreservation techniques accompanied by detailed preclinical and clinical studies, MSCs with promising potential can be utilized in future clinical settings.

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Code Availability Not applicable.

Declarations

Conflict of interest The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Ethics Approval Not applicable.

Consent to Participate Not applicable.

Consent for publication Not applicable.

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References

- Salem, H. K., & Thiemermann, C. (2010). Mesenchymal stromal cells: Current understanding and clinical status. *Stem Cells*, 28(3), 585–596.
- Wei, X., Yang, X., Han, Z. P., Qu, F. F., Shao, L., & Shi, Y. F. (2013). Mesenchymal stem cells: A new trend for cell therapy. Acta Pharmacologica Sinica, 34(6), 747–754.
- Friedenstein, A. J., Piatetzky, S., & II, Petrakova, K. V. (1966). Osteogenesis in transplants of bone marrow cells. *Journal Of Embryology And Experimental Morphology*, 16(3), 381–390.
- Friedenstein, A. J., Chailakhjan, R. K., & Lalykina, K. S. (1970). The development of fibroblast colonies in monolayer cultures of guinea-pig bone marrow and spleen cells. *Cell Tissue Kinet*, 3(4), 393–403.
- Caplan, A. I. (1991). Mesenchymal stem cells. *Journal Of Ortho*paedic Research, 9(5), 641–650.
- Goshima, J., Goldberg, V. M., & Caplan, A. I. (1991). The osteogenic potential of culture-expanded rat marrow mesenchymal cells assayed in vivo in calcium phosphate ceramic blocks. Clin Orthop Relat Res. (262):298–311.
- Prockop, D. J. (1997). Marrow stromal cells as stem cells for nonhematopoietic tissues. Science, 276(5309), 71–74.

- 8. Pittenger, M. F., Mackay, A. M., Beck, S. C., Jaiswal, R. K., Douglas, R., Mosca, J. D., et al. (1999). Multilineage potential of adult human mesenchymal stem cells. *Science*, 284(5411), 143–147.
- Spees, J. L., Olson, S. D., Ylostalo, J., Lynch, P. J., Smith, J., Perry, A., et al. (2003). Differentiation, cell fusion, and nuclear fusion during ex vivo repair of epithelium by human adult stem cells from bone marrow stroma. *Proc Natl Acad Sci USA*, 100(5), 2397–2402.
- Phinney, D. G., & Isakova, I. (2005). Plasticity and therapeutic potential of mesenchymal stem cells in the nervous system. *Curr Pharm Des*, 11(10), 1255–1265.
- Tropel, P., Platet, N., Platel, J. C., Noel, D., Albrieux, M., Benabid, A. L., et al. (2006). Functional neuronal differentiation of bone marrow-derived mesenchymal stem cells. *Stem Cells*, 24(12), 2868–2876.
- Sivamani, R. K., Schwartz, M. P., Anseth, K. S., & Isseroff, R. R. (2011). Keratinocyte proximity and contact can play a significant role in determining mesenchymal stem cell fate in human tissue. *The Faseb Journal*, 25(1), 122–131.
- Reyhani, S., Abbaspanah, B., & Mousavi, S. H. (2020). Umbilical cord-derived mesenchymal stem cells in neurodegenerative disorders: From literature to clinical practice. *Regenerative Medicine*, 15(4), 1561–1578.
- Yang, G., Fan, X., Mazhar, M., Yang, S., Xu, H., Dechsupa, N., et al. (2022). Mesenchymal stem cell application and its therapeutic mechanisms in Intracerebral Hemorrhage. Frontiers In Cellular Neuroscience, 16, 898497.
- Dazzi, F., Ramasamy, R., Glennie, S., Jones, S. P., & Roberts, I. (2006). The role of mesenchymal stem cells in haemopoiesis. *Blood Reviews*, 20(3), 161–171.
- Taechangam, N., Kol, A., Arzi, B., & Borjesson, D. L. (2022).
 Multipotent stromal cells and viral Interaction: Current implications for Therapy. Stem Cell Rev Rep, 18(1), 214–227.
- Namiot, E. D., Niemi, J. V. L., Chubarev, V. N., Tarasov, V. V., & Schioth, H. B. (2022). Stem cells in clinical trials on neurological Disorders: Trends in Stem cells Origins, indications, and Status of the clinical trials. *International Journal Of Molecular Sciences*, 23, 19.
- 18. Miana, V. V., & Gonzalez, E. A. P. (2018). Adipose tissue stem cells in regenerative medicine. *Ecancermedicalscience*, 12, 822.
- Tang, Q., Zhao, X. S., Guo, A., Cui, R. T., Song, H. L., Qi, Z. Y., et al. (2022). Therapeutic applications of adipose-derived stromal vascular fractions in osteoarthritis. World J Stem Cells, 14(10), 744–755
- Heo, J. S., Kim, S., Yang, C. E., Choi, Y., Song, S. Y., & Kim, H. O. (2021). Human adipose mesenchymal stem cell-derived Exosomes: A key player in Wound Healing. *Tissue Eng Regen Med*, 18(4), 537–548.
- Xu, Y., Wang, Q., Wang, X. X., Xiang, X. N., Peng, J. L., He, C. Q., et al. (2022). The Effect of different frequencies of Pulsed Electromagnetic Fields on Cartilage repair of adipose mesenchymal stem cell-derived Exosomes in Osteoarthritis. *Cartilage*, 13(4), 200–212.
- 22. Tsuruhara, A., Aso, K., Tokuhara, D., Ohori, J., Kawabata, M., Kurono, Y., et al. (2017). Rejuvenation of mucosal immunosenescence by adipose tissue-derived mesenchymal stem cells. *International Immunology*, 29(1), 5–10.
- 23. Dai, P., Qi, G., Xu, H., Zhu, M., Li, J., Chen, Y., et al. (2022). Reprogramming adipose mesenchymal stem cells into islet betacells for the treatment of canine diabetes mellitus. *Stem Cell Research & Therapy*, *13*(1), 370.
- Nan, Z., Grande, A., Sanberg, C. D., Sanberg, P. R., & Low, W. C. (2005). Infusion of human umbilical cord blood ameliorates neurologic deficits in rats with hemorrhagic brain injury. *Annals Of The New York Academy Of Sciences*, 1049, 84–96.



- Trivanovic, D., Kocic, J., Mojsilovic, S., Krstic, A., Ilic, V., Djordjevic, I. O., et al. (2013). Mesenchymal stem cells isolated from peripheral blood and umbilical cord Wharton's jelly. *Srpski Arhiv Za Celokupno Lekarstvo*, 141(3–4), 178–186.
- Hassan, G., Kasem, I., Soukkarieh, C., & Aljamali, M. (2017).
 A simple method to Isolate and Expand Human umbilical cord derived mesenchymal stem cells: Using explant method and umbilical cord blood serum. *Int J Stem Cells*, 10(2), 184–192.
- Beeravolu, N., McKee, C., Alamri, A., Mikhael, S., Brown, C., Perez-Cruet, M. Isolation and Characterization of Mesenchymal Stromal Cells from Human Umbilical Cord and Fetal Placenta.J Vis Exp. 2017(122).
- Vellasamy, S., Sandrasaigaran, P., Vidyadaran, S., George, E., & Ramasamy, R. (2012). Isolation and characterisation of mesenchymal stem cells derived from human placenta tissue. World J Stem Cells, 4(6), 53–61.
- Pelekanos, R. A., Sardesai, V. S., Futrega, K., Lott, W. B., Kuhn, M., & Doran, M. R. Isolation and Expansion of Mesenchymal Stem/Stromal Cells Derived from Human Placenta Tissue. J Vis Exp. 2016(112).
- 30. Chang, Z., Mao, G., Sun, L., Ao, Q., Gu, Y., & Liu, Y. (2016). Cell therapy for cerebral hemorrhage: Five year follow-up report. *Exp Ther Med*, 12(6), 3535–3540.
- 31. Meng, F., Xu, R., Wang, S., Xu, Z., Zhang, C., Li, Y., et al. (2020). Human umbilical cord-derived mesenchymal stem cell therapy in patients with COVID-19: A phase 1 clinical trial. *Signal Transduct Target Ther*, 5(1), 172.
- Mebarki, M., Abadie, C., Larghero, J., & Cras, A. (2021). Human umbilical cord-derived mesenchymal stem/stromal cells: A promising candidate for the development of advanced therapy medicinal products. Stem Cell Research & Therapy, 12(1), 152.
- Xu, X., Liang, Y., Li, X., Ouyang, K., Wang, M., Cao, T., et al. (2021). Exosome-mediated delivery of kartogenin for chondrogenesis of synovial fluid-derived mesenchymal stem cells and cartilage regeneration. *Biomaterials*, 269, 120539.
- Ridge, S. M., Sullivan, F. J., & Glynn, S. A. (2017). Mesenchymal stem cells: Key players in cancer progression. *Molecular Cancer*, 16(1), 31.
- Li, S., Huang, K. J., Wu, J. C., Hu, M. S., Sanyal, M., Hu, M., et al. (2015). Peripheral blood-derived mesenchymal stem cells: Candidate cells responsible for healing critical-sized calvarial bone defects. Stem Cells Transl Med, 4(4), 359–368.
- Jarvinen, L., Badri, L., Wettlaufer, S., Ohtsuka, T., Standiford, T. J., Toews, G. B., et al. (2008). Lung resident mesenchymal stem cells isolated from human lung allografts inhibit T cell proliferation via a soluble mediator. *The Journal Of Immunology*, 181(6), 4389–4396.
- Gong, X., Sun, Z., Cui, D., Xu, X., Zhu, H., Wang, L., et al. (2014). Isolation and characterization of lung resident mesenchymal stem cells capable of differentiating into alveolar epithelial type II cells. *Cell Biology International*, 38(4), 405–411.
- Walker, N. M., Badri, L. N., Wadhwa, A., Wettlaufer, S., Peters-Golden, M., & Lama, V. N. (2012). Prostaglandin E2 as an inhibitory modulator of fibrogenesis in human lung allografts. American Journal Of Respiratory And Critical Care Medicine, 185(1), 77–84.
- Lecourt, S., Marolleau, J. P., Fromigue, O., Vauchez, K., Andriamanalijaona, R., Ternaux, B., et al. (2010). Characterization of distinct mesenchymal-like cell populations from human skeletal muscle in situ and in vitro. *Experimental Cell Research*, 316(15), 2513–2526.
- Jackson, W. M., Nesti, L. J., & Tuan, R. S. (2010). Potential therapeutic applications of muscle-derived mesenchymal stem and progenitor cells. *Expert Opinion On Biological Therapy*, 10(4), 505–517.

- de Sousa, E. B., Casado, P. L., Moura Neto, V., Duarte, M. E., & Aguiar, D. P. (2014). Synovial fluid and synovial membrane mesenchymal stem cells: Latest discoveries and therapeutic perspectives. Stem Cell Research & Therapy, 5(5), 112.
- Zeng, W. N., Zhang, Y., Wang, D., Zeng, Y. P., Yang, H., Li, J., et al. (2021). Intra-articular injection of kartogenin-enhanced bone marrow-derived mesenchymal stem cells in the treatment of knee osteoarthritis in a rat model. *American Journal Of Sports Medi*cine, 49(10), 2795–2809.
- Genc, D., Gunaydin, B., Sezgin, S., Aladag, A., & Tarhan, E. F. (2022). Immunoregulatory effects of dental mesenchymal stem cells on T and B lymphocyte responses in primary Sjogren's syndrome. *Immunotherapy*, 14(4), 225–247.
- Leuning, D. G., Beijer, N. R. M., du Fosse, N. A., Vermeulen, S., Lievers, E., van Kooten, C., et al. (2018). The cytokine secretion profile of mesenchymal stromal cells is determined by surface structure of the microenvironment. *Scientific Reports*, 8(1), 7716.
- Klingemann, H., Matzilevich, D., & Marchand, J. (2008). Mesenchymal stem cells sources and clinical applications. *Transfusion Medicine And Hemotherapy: Offizielles Organ Der Deutschen Gesellschaft Fur Transfusionsmedizin Und ImmunhamAtologie*, 35(4), 272–277.
- Hwang, N. S., Zhang, C., Hwang, Y. S., & Varghese, S. (2009).
 Mesenchymal stem cell differentiation and roles in regenerative medicine. Wiley Interdisciplinary Reviews. Systems Biology And Medicine, 1(1), 97–106.
- Fitzsimmons, R. E. B., Mazurek, M. S., Soos, A., & Simmons, C. A. (2018). Mesenchymal Stromal/Stem cells in Regenerative Medicine and tissue Engineering. Stem Cells Int, 2018, 8031718.
- Dimarino, A. M., Caplan, A. I., & Bonfield, T. L. (2013). Mesenchymal stem cells in tissue repair. Frontiers In Immunology, 4, 201.
- Baksh, D., Song, L., & Tuan, R. S. (2004). Adult mesenchymal stem cells: Characterization, differentiation, and application in cell and gene therapy. *Journal Of Cellular And Molecular Medicine*, 8(3), 301–316.
- Baksh, D., Yao, R., & Tuan, R. S. (2007). Comparison of proliferative and multilineage differentiation potential of human mesenchymal stem cells derived from umbilical cord and bone marrow. *Stem Cells*, 25(6), 1384–1392.
- Lu, L. L., Liu, Y. J., Yang, S. G., Zhao, Q. J., Wang, X., Gong, W., et al. (2006). Isolation and characterization of human umbilical cord mesenchymal stem cells with hematopoiesis-supportive function and other potentials. *Haematologica*, 91(8), 1017–1026.
- Brooke, G., Tong, H., Levesque, J. P., & Atkinson, K. (2008).
 Molecular trafficking mechanisms of multipotent mesenchymal stem cells derived from human bone marrow and placenta. Stem Cells And Development, 17(5), 929–940.
- 53. Shah, K. (2016). Stem cell-based therapies for tumors in the brain: Are we there yet? *Neuro Oncol*, 18(8), 1066–1078.
- Yudintceva, N., Mikhailova, N., Fedorov, V., Samochernych, K., Vinogradova, T., Muraviov, A. (2022). Mesenchymal Stem Cells and MSCs-Derived Extracellular Vesicles in Infectious Diseases: From Basic Research to Clinical Practice. Bioengineering (Basel). ;9(11).
- Sierra-Sanchez, A., Ordonez-Luque, A., Espinosa-Ibanez, O., Ruiz-Garcia, A., & Arias-Santiago, S. (2018). Epithelial in vitro differentiation of mesenchymal stem cells. *Curr Stem Cell Res Ther*, 13(6), 409–422.
- Sarsenova, M., Kim, Y., Raziyeva, K., Kazybay, B., Ogay, V., & Saparov, A. (2022). Recent advances to enhance the immunomodulatory potential of mesenchymal stem cells. *Frontiers In Immunology*, 13, 1010399.
- Uder, C., Bruckner, S., Winkler, S., Tautenhahn, H. M., & Christ,
 B. (2018). Mammalian MSC from selected species: Features and applications. *Cytometry. Part A*, 93(1), 32–49.



- Galipeau, J., & Sensebe, L. (2018). Mesenchymal stromal cells: Clinical Challenges and Therapeutic Opportunities. *Cell Stem Cell*, 22(6), 824–833.
- Timaner, M., Tsai, K. K., & Shaked, Y. (2020). The multifaceted role of mesenchymal stem cells in cancer. *Semin Cancer Biol*, 60, 225–237.
- Zhou, T., Yuan, Z., Weng, J., Pei, D., Du, X., He, C., et al. (2021).
 Challenges and advances in clinical applications of mesenchymal stromal cells. *Journal Of Hematology & Oncology*, 14(1), 24.
- Chen, Y., Shao, J. Z., Xiang, L. X., Dong, X. J., & Zhang, G. R. (2008). Mesenchymal stem cells: A promising candidate in regenerative medicine. *International Journal Of Biochemistry & Cell Biology*, 40(5), 815–820.
- 62. Yang, G., Fan, X., Mazhar, M., Guo, W., Zou, Y., Dechsupa, N., et al. (2022). Neuroinflammation of microglia polarization in intracerebral hemorrhage and its potential targets for intervention. *Frontiers In Molecular Neuroscience*, 15, 1013706.
- Dai, L. J., Moniri, M. R., Zeng, Z. R., Zhou, J. X., Rayat, J., & Warnock, G. L. (2011). Potential implications of mesenchymal stem cells in cancer therapy. *Cancer Letters*, 305(1), 8–20.
- 64. Shah, K. (2012). Mesenchymal stem cells engineered for cancer therapy. *Advanced Drug Delivery Reviews*, 64(8), 739–748.
- Liang, W., Chen, X., Zhang, S., Fang, J., Chen, M., Xu, Y., et al. (2021). Mesenchymal stem cells as a double-edged sword in tumor growth: Focusing on MSC-derived cytokines. *Cellular & Molecular Biology Letters*, 26(1), 3.
- Krampera, M., & Le Blanc, K. (2021). Mesenchymal stromal cells: Putative microenvironmental modulators become cell therapy. *Cell Stem Cell*, 28(10), 1708–1725.
- 67. Li, N., & Hua, J. (2017). Interactions between mesenchymal stem cells and the immune system. *Cellular And Molecular Life Sciences*, 74(13), 2345–2360.
- Wang, L. T., Ting, C. H., Yen, M. L., Liu, K. J., Sytwu, H. K., Wu, K. K., et al. (2016). Human mesenchymal stem cells (MSCs) for treatment towards immune- and inflammation-mediated diseases: Review of current clinical trials. *Journal Of Biomedical Science*, 23(1), 76.
- Weiss, A. R. R., & Dahlke, M. H. (2019). Immunomodulation by Mesenchymal Stem cells (MSCs): Mechanisms of action of living, apoptotic, and dead MSCs. Frontiers In Immunology, 10, 1191
- Song, N., Scholtemeijer, M., & Shah, K. (2020). Mesenchymal stem cell immunomodulation: Mechanisms and therapeutic potential. *Trends In Pharmacological Sciences*, 41(9), 653–664.
- Weiss, A. R. R., Lee, O., Eggenhofer, E., Geissler, E., Korevaar, S. S., Soeder, Y., et al. (2020). Differential effects of heat-inactivated, secretome-deficient MSC and metabolically active MSC in sepsis and allogenic heart transplantation. *Stem Cells*, 38(6), 797–807.
- Kim, J. H., Jo, C. H., Kim, H. R., & Hwang, Y. I. (2018). Comparison of immunological characteristics of mesenchymal stem cells from the Periodontal ligament, umbilical cord, and adipose tissue. Stem Cells Int, 2018, 8429042.
- 73. Gao, P., Ding, Q., Wu, Z., Jiang, H., & Fang, Z. (2010). Therapeutic potential of human mesenchymal stem cells producing IL-12 in a mouse xenograft model of renal cell carcinoma. *Cancer Letters*, 290(2), 157–166.
- Niemeyer, P., Schonberger, T. S., Hahn, J., Kasten, P., Fellenberg, J., Suedkamp, N., et al. (2010). Xenogenic transplantation of human mesenchymal stem cells in a critical size defect of the sheep tibia for bone regeneration. *Tissue Engineering Part A*, 16(1), 33–43.
- Liu, T. P., Ha, P., Xiao, C. Y., Kim, S. Y., Jensen, A. R., Easley, J., et al. (2022). Updates on mesenchymal stem cell therapies for articular cartilage regeneration in large animal models. *Front Cell Dev Biol*, 10, 982199.

- Zhou, Y., Yamamoto, Y., Xiao, Z., & Ochiya, T. (2019). The Immunomodulatory Functions of Mesenchymal Stromal/Stem Cells Mediated via Paracrine Activity. J Clin Med.;8(7).
- Chen, J., Zheng, C. X., Jin, Y., & Hu, C. H. (2021). Mesenchymal stromal cell-mediated immune regulation: A promising remedy in the therapy of type 2 diabetes mellitus. *Stem Cells*, 39(7), 838–852.
- Reis, M., Mavin, E., Nicholson, L., Green, K., Dickinson, A. M., & Wang, X. N. (2018). Mesenchymal stromal cell-derived extracellular vesicles attenuate dendritic cell maturation and function. Frontiers In Immunology, 9, 2538.
- Krampera, M., Glennie, S., Dyson, J., Scott, D., Laylor, R., Simpson, E., et al. (2003). Bone marrow mesenchymal stem cells inhibit the response of naive and memory antigen-specific T cells to their cognate peptide. *Blood*, 101(9), 3722–3729.
- Ren, G., Zhao, X., Zhang, L., Zhang, J., L'Huillier, A., Ling, W., et al. (2010). Inflammatory cytokine-induced intercellular adhesion molecule-1 and vascular cell adhesion molecule-1 in mesenchymal stem cells are critical for immunosuppression. *The Journal Of Immunology*, 184(5), 2321–2328.
- Luz-Crawford, P., Ipseiz, N., Espinosa-Carrasco, G., Caicedo, A., Tejedor, G., Toupet, K., et al. (2016). PPARbeta/delta directs the therapeutic potential of mesenchymal stem cells in arthritis. *Annals Of The Rheumatic Diseases*, 75(12), 2166–2174.
- Najar, M., Fayyad-Kazan, H., Faour, W. H., Merimi, M., Sokal, E. M., Lombard, C. A., et al. (2019). Immunological modulation following bone marrow-derived mesenchymal stromal cells and Th17 lymphocyte co-cultures. *Inflammation Research*, 68(3), 203–213.
- 83. Del Papa, B., Sportoletti, P., Cecchini, D., Rosati, E., Balucani, C., Baldoni, S., et al. (2013). Notch1 modulates mesenchymal stem cells mediated regulatory T-cell induction. *European Journal Of Immunology*, 43(1), 182–187.
- 84. Hong, J. W., Lim, J. H., Chung, C. J., Kang, T. J., Kim, T. Y., Kim, Y. S., et al. (2017). Immune Tolerance of Human Dental Pulp-Derived mesenchymal stem cells mediated by CD4(+)CD25(+) FoxP3(+) Regulatory T-Cells and Induced by TGF-beta1 and IL-10. *Yonsei Medical Journal*, 58(5), 1031–1039.
- 85. Gieseke, F., Bohringer, J., Bussolari, R., Dominici, M., Handgretinger, R., & Muller, I. (2010). Human multipotent mesenchymal stromal cells use galectin-1 to inhibit immune effector cells. *Blood*, *116*(19), 3770–3779.
- Laranjeira, P., Pedrosa, M., Pedreiro, S., Gomes, J., Martinho, A., Antunes, B., et al. (2015). Effect of human bone marrow mesenchymal stromal cells on cytokine production by peripheral blood naive, memory, and effector T cells. Stem Cell Research & Therapy, 6(1), 3.
- 87. Liotta, F., Angeli, R., Cosmi, L., Fili, L., Manuelli, C., Frosali, F., et al. (2008). Toll-like receptors 3 and 4 are expressed by human bone marrow-derived mesenchymal stem cells and can inhibit their T-cell modulatory activity by impairing notch signaling. *Stem Cells*, 26(1), 279–289.
- 88. Zeng, Y., Li, B., Li, T., Liu, W., Ran, C., Penson, R. T., et al. (2019). CD90(low) MSCs modulate intratumoral immunity to confer antitumor activity in a mouse model of ovarian cancer. *Oncotarget*, 10(43), 4479–4491.
- Li, B., Zeng, Y., Reeves, P. M., Ran, C., Liu, Q., Qu, X., et al. (2018). AMD3100 augments the efficacy of Mesothelin-Targeted, Immune-Activating VIC-008 in Mesothelioma by modulating Intratumoral Immunosuppression. *Cancer Immunology Research*, 6(5), 539–551.
- Li, Y., Zhang, D., Xu, L., Dong, L., Zheng, J., Lin, Y., et al. (2019). Cell-cell contact with proinflammatory macrophages enhances the immunotherapeutic effect of mesenchymal stem cells in two abortion models. *Cellular & Molecular Immunology*, 16(12), 908–920.



- Li, Y. H., Zhang, D., & Du, M. R. (2021). Advances and challenges of mesenchymal stem cells for pregnancy-related diseases. Cellular & Molecular Immunology, 18(8), 2075–2077.
- 92. Sheng, H., Wang, Y., Jin, Y., Zhang, Q., Zhang, Y., Wang, L., et al. (2008). A critical role of IFNgamma in priming MSC-mediated suppression of T cell proliferation through up-regulation of B7-H1. *Cell Research*, 18(8), 846–857.
- Franquesa, M., Mensah, F. K., Huizinga, R., Strini, T., Boon, L., Lombardo, E., et al. (2015). Human adipose tissue-derived mesenchymal stem cells abrogate plasmablast formation and induce regulatory B cells independently of T helper cells. *Stem Cells*, 33(3), 880–891.
- 94. Budoni, M., Fierabracci, A., Luciano, R., Petrini, S., Di Ciommo, V., & Muraca, M. (2013). The immunosuppressive effect of mesenchymal stromal cells on B lymphocytes is mediated by membrane vesicles. *Cell Transplantation*, 22(2), 369–379.
- Healy, M. E., Bergin, R., Mahon, B. P., & English, K. (2015).
 Mesenchymal stromal cells protect against caspase 3-mediated apoptosis of CD19(+) peripheral B cells through contact-dependent upregulation of VEGF. Stem Cells And Development, 24(20), 2391–2402.
- Tabera, S., Perez-Simon, J. A., Diez-Campelo, M., Sanchez-Abarca, L. I., Blanco, B., Lopez, A., et al. (2008). The effect of mesenchymal stem cells on the viability, proliferation and differentiation of B-lymphocytes. *Haematologica*, 93(9), 1301–1309.
- Hu, C. D., Kosaka, Y., Marcus, P., Rashedi, I., & Keating, A. (2019). Differential Immunomodulatory Effects of human bone marrow-derived mesenchymal stromal cells on natural killer cells. Stem Cells And Development, 28(14), 933–943.
- Manferdini, C., Paolella, F., Gabusi, E., Gambari, L., Piacentini, A., Filardo, G., et al. (2017). Adipose stromal cells mediated switching of the pro-inflammatory profile of M1-like macrophages is facilitated by PGE2: In vitro evaluation. *Osteoarthritis Cartilage*, 25(7), 1161–1171.
- Colombini, A., Libonati, F., Cangelosi, D., Lopa, S., De Luca, P., Coviello, D. A., et al. (2022). Inflammatory priming with IL-1beta promotes the immunomodulatory behavior of adipose derived stem cells. Frontiers In Bioengineering And Biotechnology, 10, 1000879.
- 100. Jiang, D., Muschhammer, J., Qi, Y., Kugler, A., de Vries, J. C., Saffarzadeh, M., et al. (2016). Suppression of neutrophil-mediated tissue Damage-A novel skill of mesenchymal stem cells. *Stem Cells*, 34(9), 2393–2406.
- Thieblemont, N., Wright, H. L., Edwards, S. W., & Witko-Sarsat,
 V. (2016). Human neutrophils in auto-immunity. *Seminars In Immunology*, 28(2), 159–173.
- 102. Ferreira, J. R., Teixeira, G. Q., Santos, S. G., Barbosa, M. A., Almeida-Porada, G., & Goncalves, R. M. (2018). Mesenchymal stromal cell secretome: Influencing therapeutic potential by Cellular Pre-conditioning. Frontiers In Immunology, 9, 2837.
- 103. Salgado, A. J., Reis, R. L., Sousa, N. J., & Gimble, J. M. (2010). Adipose tissue derived stem cells secretome: Soluble factors and their roles in regenerative medicine. *Curr Stem Cell Res Ther*, 5(2), 103–110.
- 104. Huang, Y., Wu, Q., & Tam, P. K. H. (2022). Immunomodulatory Mechanisms of Mesenchymal Stem Cells and Their Potential Clinical Applications.Int J Mol Sci.;23(17).
- 105. Qiu, G., Zheng, G., Ge, M., Wang, J., Huang, R., Shu, Q., et al. (2019). Functional proteins of mesenchymal stem cell-derived extracellular vesicles. Stem Cell Research & Therapy, 10(1), 359.
- 106. Tsuji, K., Kitamura, S., & Wada, J. (2018). Secretomes from mesenchymal stem cells against Acute kidney Injury: Possible heterogeneity. Stem Cells Int, 2018, 8693137.
- 107. Kandoi, L. P. K., Misra, S., & Verma, R. S. V. K. R. (2019). The mesenchymal stem cell secretome: A new paradigm towards

- cell-free therapeutic mode in regenerative medicine. *Cytokine & Growth Factor Reviews*, 46, 1–9.
- 108. Mardpour, S., Hamidieh, A. A., Taleahmad, S., Sharifzad, F., Taghikhani, A., & Baharvand, H. (2019). Interaction between mesenchymal stromal cell-derived extracellular vesicles and immune cells by distinct protein content. *Journal Of Cellular Physiology*, 234(6), 8249–8258.
- 109. Madrigal, M., Rao, K. S., & Riordan, N. H. (2014). A review of therapeutic effects of mesenchymal stem cell secretions and induction of secretory modification by different culture methods. *J Transl Med*, 12, 260.
- 110. Ghannam, S., Pene, J., Moquet-Torcy, G., Jorgensen, C., & Yssel, H. (2010). Mesenchymal stem cells inhibit human Th17 cell differentiation and function and induce a T regulatory cell phenotype. *The Journal Of Immunology*, 185(1), 302–312.
- 111. Yu, W., Li, C., Zhang, D., Li, Z., Xia, P., Liu, X., et al. (2022). Advances in T cells based on inflammation in metabolic Diseases. *Cells*, 11, 22.
- 112. Wang, W. B., Yen, M. L., Liu, K. J., Hsu, P. J., Lin, M. H., Chen, P. M., et al. (2015). Interleukin-25 mediates Transcriptional Control of PD-L1 via STAT3 in Multipotent Human mesenchymal stromal cells (hMSCs) to suppress Th17 responses. *Stem Cell Reports*, 5(3), 392–404.
- 113. Luz-Crawford, P., Noel, D., Fernandez, X., Khoury, M., Figueroa, F., Carrion, F., et al. (2012). Mesenchymal stem cells repress Th17 molecular program through the PD-1 pathway. *PLoS One*, 7(9), e45272.
- 114. Ge, W., Jiang, J., Arp, J., Liu, W., Garcia, B., & Wang, H. (2010). Regulatory T-cell generation and kidney allograft tolerance induced by mesenchymal stem cells associated with indoleamine 2,3-dioxygenase expression. *Transplantation*, 90(12), 1312–1320.
- 115. Davies, L. C., Heldring, N., Kadri, N., & Le Blanc, K. (2017). Mesenchymal stromal cell secretion of programmed Death-1 Ligands regulates T cell mediated Immunosuppression. *Stem Cells*, 35(3), 766–776.
- 116. Spaggiari, G. M., Capobianco, A., Becchetti, S., Mingari, M. C., & Moretta, L. (2006). Mesenchymal stem cell-natural killer cell interactions: Evidence that activated NK cells are capable of killing MSCs, whereas MSCs can inhibit IL-2-induced NK-cell proliferation. *Blood*, 107(4), 1484–1490.
- 117. Cui, R., Rekasi, H., Hepner-Schefczyk, M., Fessmann, K., Petri, R. M., Bruderek, K., et al. (2016). Human mesenchymal stromal/stem cells acquire immunostimulatory capacity upon cross-talk with natural killer cells and might improve the NK cell function of immunocompromised patients. Stem Cell Research & Therapy, 7(1), 88.
- 118. Spaggiari, G. M., Capobianco, A., Abdelrazik, H., Becchetti, F., Mingari, M. C., & Moretta, L. (2008). Mesenchymal stem cells inhibit natural killer-cell proliferation, cytotoxicity, and cytokine production: Role of indoleamine 2,3-dioxygenase and prostaglandin E2. *Blood*, 111(3), 1327–1333.
- 119. Pietra, G., Manzini, C., Rivara, S., Vitale, M., Cantoni, C., Petretto, A., et al. (2012). Melanoma cells inhibit natural killer cell function by modulating the expression of activating receptors and cytolytic activity. *Cancer Research*, 72(6), 1407–1415.
- 120. Thomas, H., Jager, M., Mauel, K., Brandau, S., Lask, S., & Flohe, S. B. (2014). Interaction with mesenchymal stem cells provokes natural killer cells for enhanced IL-12/IL-18-induced interferongamma secretion. *Mediators Inflamm*, 2014, 143463.
- 121. Raffaghello, L., Bianchi, G., Bertolotto, M., Montecucco, F., Busca, A., Dallegri, F., et al. (2008). Human mesenchymal stem cells inhibit neutrophil apoptosis: A model for neutrophil preservation in the bone marrow niche. *Stem Cells*, 26(1), 151–162.
- 122. Mahmoudi, M., Taghavi-Farahabadi, M., Rezaei, N., & Hashemi, S. M. (2019). Comparison of the effects of adipose tissue



- mesenchymal stromal cell-derived exosomes with conditioned media on neutrophil function and apoptosis. *International Immunopharmacology*, 74, 105689.
- 123. Brandau, S., Jakob, M., Bruderek, K., Bootz, F., Giebel, B., Radtke, S., et al. (2014). Mesenchymal stem cells augment the anti-bacterial activity of neutrophil granulocytes. *PLoS One*, 9(9), e106903.
- 124. Melief, S. M., Zwaginga, J. J., Fibbe, W. E., & Roelofs, H. (2013). Adipose tissue-derived multipotent stromal cells have a higher immunomodulatory capacity than their bone marrow-derived counterparts. Stem Cells Transl Med, 2(6), 455–463.
- 125. Garcia-Bernal, D., Blanquer, M., Martinez, C. M., Garcia-Guillen, A. I., Garcia-Hernandez, A. M., Carmen Alguero, M., et al. (2022). Enforced mesenchymal stem cell tissue colonization counteracts immunopathology. NPJ Regen Med, 7(1), 61.
- 126. Spaggiari, G. M., Abdelrazik, H., Becchetti, F., & Moretta, L. (2009). MSCs inhibit monocyte-derived DC maturation and function by selectively interfering with the generation of immature DCs: Central role of MSC-derived prostaglandin E2. *Blood*, 113(26), 6576–6583.
- 127. Hyvarinen, K., Holopainen, M., Skirdenko, V., Ruhanen, H., Lehenkari, P., Korhonen, M., et al. (2018). Mesenchymal stromal cells and their extracellular vesicles enhance the anti-inflammatory phenotype of Regulatory Macrophages by downregulating the production of Interleukin (IL)-23 and IL-22. Frontiers In Immunology, 9, 771.
- 128. Lo Sicco, C., Reverberi, D., Balbi, C., Ulivi, V., Principi, E., Pascucci, L., et al. (2017). Mesenchymal stem cell-derived extracellular vesicles as mediators of anti-inflammatory Effects: Endorsement of macrophage polarization. *Stem Cells Transl Med*, 6(3), 1018–1028.
- 129. Nemeth, K., Leelahavanichkul, A., Yuen, P. S., Mayer, B., Parmelee, A., Doi, K., et al. (2009). Bone marrow stromal cells attenuate sepsis via prostaglandin E(2)-dependent reprogramming of host macrophages to increase their interleukin-10 production. *Nature Medicine*, 15(1), 42–49.
- 130. Abumaree, M. H., Al Jumah, M. A., Kalionis, B., Jawdat, D., Al Khaldi, A., Abomaray, F. M., et al. (2013). Human placental mesenchymal stem cells (pMSCs) play a role as immune suppressive cells by shifting macrophage differentiation from inflammatory M1 to anti-inflammatory M2 macrophages. Stem Cell Rev Rep, 9(5), 620–641.
- 131. Silva, L. H. A., Antunes, M. A., Dos Santos, C. C., Weiss, D. J., Cruz, F. F., & Rocco, P. R. M. (2018). Strategies to improve the therapeutic effects of mesenchymal stromal cells in respiratory diseases. Stem Cell Research & Therapy, 9(1), 45.
- 132. Jung, S. Y., Kim, Y. E., Park, W. S., Ahn, S. Y., Sung, D. K., Sung, S. I. (2022). Thrombin Preconditioning Improves the Therapeutic Efficacy of Mesenchymal Stem Cells in Severe Intraventricular Hemorrhage Induced Neonatal Rats.Int J Mol Sci.;23(8).
- 133. Zhang, W., Liu, L., Huo, Y., Yang, Y., & Wang, Y. (2014). Hypoxia-pretreated human MSCs attenuate acute kidney injury through enhanced angiogenic and antioxidative capacities. *Biomed Research International*, 2014, 462472.
- 134. Lan, Y. W., Choo, K. B., Chen, C. M., Hung, T. H., Chen, Y. B., Hsieh, C. H., et al. (2015). Hypoxia-preconditioned mesenchymal stem cells attenuate bleomycin-induced pulmonary fibrosis. *Stem Cell Research & Therapy*, 6, 97.
- 135. Martinez, V. G., Ontoria-Oviedo, I., Ricardo, C. P., Harding, S. E., Sacedon, R., Varas, A., et al. (2017). Overexpression of hypoxiainducible factor 1 alpha improves immunomodulation by dental mesenchymal stem cells. Stem Cell Research & Therapy, 8(1), 208.
- 136. Sivanathan, K. N., Gronthos, S., Grey, S. T., Rojas-Canales, D., & Coates, P. T. (2017). Immunodepletion and Hypoxia Preconditioning of Mouse Compact Bone cells as a Novel Protocol to

- isolate highly immunosuppressive mesenchymal stem cells. *Stem Cells And Development*, 26(7), 512–527.
- 137. Fierro, F. A., O'Neal, A. J., Beegle, J. R., Chavez, M. N., Peavy, T. R., Isseroff, R. R., et al. (2015). Hypoxic pre-conditioning increases the infiltration of endothelial cells into scaffolds for dermal regeneration pre-seeded with mesenchymal stem cells. Front Cell Dev Biol, 3, 68.
- 138. Elabd, C., Ichim, T. E., Miller, K., Anneling, A., Grinstein, V., Vargas, V., et al. (2018). Comparing atmospheric and hypoxic cultured mesenchymal stem cell transcriptome: Implication for stem cell therapies targeting intervertebral discs. *J Transl Med*, 16(1), 222.
- 139. Kadle, R. L., Abdou, S. A., Villarreal-Ponce, A. P., Soares, M. A., Sultan, D. L., David, J. A., et al. (2018). Microenvironmental cues enhance mesenchymal stem cell-mediated immunomodulation and regulatory T-cell expansion. *PLoS One*, 13(3), e0193178.
- 140. Kim, Y., Jin, H. J., Heo, J., Ju, H., Lee, H. Y., Kim, S., et al. (2018). Small hypoxia-primed mesenchymal stem cells attenuate graft-versus-host disease. *Leukemia*, 32(12), 2672–2684.
- 141. Introna, M., & Golay, J. (2020). Tolerance to bone marrow transplantation: Do mesenchymal stromal cells still have a future for Acute or Chronic GvHD? Frontiers In Immunology, 11, 609063.
- 142. Boland, L., Burand, A. J., Brown, A. J., Boyt, D., Lira, V. A., & Ankrum, J. A. (2018). IFN-gamma and TNF-alpha pre-licensing protects mesenchymal stromal cells from the pro-inflammatory Effects of palmitate. *Molecular Therapy*, 26(3), 860–873.
- 143. Lopez-Garcia, L., & Castro-Manrreza, M. E. (2021). TNF-alpha and IFN-gamma Participate in Improving the Immunoregulatory Capacity of Mesenchymal Stem/Stromal Cells: Importance of Cell-Cell Contact and Extracellular Vesicles.Int J Mol Sci.;22(17).
- 144. Zimmermann, J. A., Hettiaratchi, M. H., & McDevitt, T. C. (2017). Enhanced immunosuppression of T cells by sustained presentation of Bioactive Interferon-gamma within three-dimensional mesenchymal stem cell constructs. *Stem Cells Transl Med*, 6(1), 223–237.
- 145. Klinker, M. W., Marklein, R. A., Lo Surdo, J. L., Wei, C. H., & Bauer, S. R. (2017). Morphological features of IFN-gammastimulated mesenchymal stromal cells predict overall immunosuppressive capacity. *Proc Natl Acad Sci U S A*, 114(13), E2598–E607.
- 146. Kim, D. S., Jang, I. K., Lee, M. W., Ko, Y. J., Lee, D. H., Lee, J. W., et al. (2018). Enhanced Immunosuppressive Properties of Human mesenchymal stem cells primed by Interferon-gamma. *EBioMedicine*, 28, 261–273.
- 147. Goncalves, F. D. C., Luk, F., Korevaar, S. S., Bouzid, R., Paz, A. H., Lopez-Iglesias, C., et al. (2017). Membrane particles generated from mesenchymal stromal cells modulate immune responses by selective targeting of pro-inflammatory monocytes. *Scientific Reports*, 7(1), 12100.
- 148. Bolhassani, A., Shahbazi, S., Agi, E., Haghighipour, N., Hadi, A., & Asgari, F. (2019). Modified DCs and MSCs with HPV E7 antigen and small hsps: Which one is the most potent strategy for eradication of tumors? *Molecular Immunology*, 108, 102–110.
- 149. Choi, S. H., Stuckey, D. W., Pignatta, S., Reinshagen, C., Khalsa, J. K., Roozendaal, N., et al. (2017). Tumor resection recruits effector T cells and boosts therapeutic efficacy of encapsulated stem cells expressing IFNbeta in Glioblastomas. *Clinical Cancer Research*, 23(22), 7047–7058.
- 150. van Solinge, T. S., Nieland, L., Chiocca, E. A., & Broekman, M. L. D. (2022). Advances in local therapy for glioblastoma taking the fight to the tumour. *Nat Rev Neurol*, 18(4), 221–236.
- 151. Relation, T., Yi, T., Guess, A. J., La Perle, K., Otsuru, S., Hasgur, S., et al. (2018). Intratumoral Delivery of Interferongamma-Secreting mesenchymal stromal cells repolarizes



- Tumor-Associated Macrophages and suppresses neuroblastoma proliferation in vivo. *Stem Cells*, *36*(6), 915–924.
- 152. Song, S. Y., Hong, J., Go, S., Lim, S., Sohn, H. S., Kang, M., et al. (2020). Interleukin-4 gene transfection and spheroid formation Potentiate Therapeutic Efficacy of Mesenchymal Stem cells for Osteoarthritis. Adv Healthc Mater, 9(5), e1901612.
- 153. Chen, X., Lin, X., Zhao, J., Shi, W., Zhang, H., Wang, Y., et al. (2008). A tumor-selective biotherapy with prolonged impact on established metastases based on cytokine gene-engineered MSCs. *Molecular Therapy*, 16(4), 749–756.
- 154. Jing, W., Chen, Y., Lu, L., Hu, X., Shao, C., Zhang, Y., et al. (2014). Human umbilical cord blood-derived mesenchymal stem cells producing IL15 eradicate established pancreatic tumor in syngeneic mice. *Molecular Cancer Therapeutics*, 13(8), 2127–2137.
- 155. Elzaouk, L., Moelling, K., & Pavlovic, J. (2006). Anti-tumor activity of mesenchymal stem cells producing IL-12 in a mouse melanoma model. *Experimental Dermatology*, 15(11), 865–874.
- 156. Kim, N., Nam, Y. S., Im, K. I., Lim, J. Y., Lee, E. S., Jeon, Y. W., et al. (2015). IL-21-Expressing mesenchymal stem cells prevent Lethal B-Cell Lymphoma through efficient delivery of IL-21, which redirects the Immune System to Target the Tumor. Stem Cells And Development, 24(23), 2808–2821.
- 157. Jeon, S. H., Zhu, G. Q., Bae, W. J., Choi, S. W., Jeong, H. C., Cho, H. J. (2018). Engineered Mesenchymal Stem Cells Expressing Stromal Cell-derived Factor-1 Improve Erectile Dysfunction in Streptozotocin-Induced Diabetic Rats.Int J Mol Sci.;19(12).
- 158. Lou, G., Chen, L., Xia, C., Wang, W., Qi, J., Li, A., et al. (2020). MiR-199a-modified exosomes from adipose tissue-derived mesenchymal stem cells improve hepatocellular carcinoma chemosensitivity through mTOR pathway. *Journal Of Experimental & Clinical Cancer Research : Cr*, 39(1), 4.
- Kavari, S. L., & Shah, K. (2020). Engineered stem cells targeting multiple cell surface receptors in tumors. *Stem Cells*, 38(1), 34–44.
- 160. Du, W., Seah, I., Bougazzoul, O., Choi, G., Meeth, K., Bosenberg, M. W., et al. (2017). Stem cell-released oncolytic herpes simplex virus has therapeutic efficacy in brain metastatic melanomas. *Proc Natl Acad Sci U S A*, 114(30), E6157–E65.
- 161. Chahal, J., Gomez-Aristizabal, A., Shestopaloff, K., Bhatt, S., Chaboureau, A., Fazio, A., et al. (2019). Bone marrow mesenchymal stromal cell treatment in patients with Osteoarthritis results in overall improvement in Pain and symptoms and reduces synovial inflammation. Stem Cells Transl Med, 8(8), 746–757.
- 162. Muroi, K., Miyamura, K., Okada, M., Yamashita, T., Murata, M., Ishikawa, T., et al. (2016). Bone marrow-derived mesenchymal stem cells (JR-031) for steroid-refractory grade III or IV acute graft-versus-host disease: A phase II/III study. *International Journal Of Hematology*, 103(2), 243–250.
- 163. Connick, P., Kolappan, M., Crawley, C., Webber, D. J., Patani, R., Michell, A. W., et al. (2012). Autologous mesenchymal stem cells for the treatment of secondary progressive multiple sclerosis: An open-label phase 2a proof-of-concept study. *Lancet Neurology*, 11(2), 150–156.
- 164. Kim, H. S., Lee, J. H., Roh, K. H., Jun, H. J., Kang, K. S., & Kim, T. Y. (2017). Clinical trial of human umbilical cord blood-derived stem cells for the treatment of moderate-to-severe atopic dermatitis: Phase I/IIa studies. Stem Cells, 35(1), 248–255.
- 165. Ciccocioppo, R., Bernardo, M. E., Sgarella, A., Maccario, R., Avanzini, M. A., Ubezio, C., et al. (2011). Autologous bone marrow-derived mesenchymal stromal cells in the treatment of fistulising Crohn's disease. *Gut*, 60(6), 788–798.
- 166. Panes, J., Garcia-Olmo, D., Van Assche, G., Colombel, J. F., Reinisch, W., Baumgart, D. C., et al. (2016). Expanded allogeneic adipose-derived mesenchymal stem cells (Cx601) for complex

- perianal fistulas in Crohn's disease: A phase 3 randomised, double-blind controlled trial. *Lancet*, 388(10051), 1281–1290.
- 167. Ciccocioppo, R., & Corazza, G. R. (2016). Mesenchymal stem cells for fistulising Crohn's disease. *Lancet*, 388(10051), 1251–1252.
- Thomas, H., & Therapy (2016). MSCs promote fistula closure in Crohn's disease. *Nature Reviews. Gastroenterology & Hepatology*, 13(10), 560.
- 169. Panes, J., Garcia-Olmo, D., Van Assche, G., Colombel, J. F., Reinisch, W., Baumgart, D. C., et al. (2018). Long-term efficacy and safety of Stem Cell Therapy (Cx601) for Complex Perianal Fistulas in patients with Crohn's Disease. *Gastroenterology*, 154(5), 1334–1342. e4.
- 170. Herz, J., Koster, C., Reinboth, B. S., Dzietko, M., Hansen, W., Sabir, H., et al. (2018). Interaction between hypothermia and delayed mesenchymal stem cell therapy in neonatal hypoxic-ischemic brain injury. *Brain, Behavior, And Immunity*, 70, 118–130.
- 171. Zhang, L., Zhang, X., Liu, Y., Zhang, W., Wu, C. T., & Wang, L. (2023). CD146 + umbilical cord mesenchymal stem cells exhibit high immunomodulatory activity and therapeutic efficacy in septic mice. *J Inflamm Res*, 16, 579–594.
- 172. Bao, X. H., Gao, F., Athari, S. S., & Wang, H. (2023). Immuno-modulatory effect of IL-35 gene-transfected mesenchymal stem cells on allergic asthma. *Fundamental & Clinical Pharmacology*, 37(1), 116–124.
- 173. Wang, C. H., Chen, C. Y., Wang, K. H., Kao, A. P., Chen, Y. J., Lin, P. H., et al. (2023). Comparing the therapeutic mechanism and Immune Response of Human and Mouse Mesenchymal Stem cells in Immunocompetent mice with Acute Liver failure. Stem Cells Transl Med. 12(1), 39–53.
- 174. Chen, J., Huang, J., Shi, J., Li, M., Zhao, E., Li, G., et al. (2023). Nestin + Peyer's patch resident MSCs enhance healing of inflammatory bowel disease through IL-22-mediated intestinal epithelial repair. *Cell Proliferation*, *56*(2), e13363.
- 175. Hackel, A., Vollmer, S., Bruderek, K., Lang, S., & Brandau, S. (2023). Immunological priming of mesenchymal stromal/stem cells and their extracellular vesicles augments their therapeutic benefits in experimental graft-versus-host disease via engagement of PD-1 ligands. Frontiers In Immunology, 14, 1078551.
- 176. Chen, X. Y., Chen, K. Y., Feng, P. H., Lee, K. Y., Fang, Y. T., Chen, Y. Y., et al. (2023). YAP-regulated type II alveolar epithelial cell differentiation mediated by human umbilical cord-derived mesenchymal stem cells in acute respiratory distress syndrome. *Biomedicine & Pharmacotherapy*, 159, 114302.
- 177. Volkova, M. V., Shen, N., Polyanskaya, A., Qi, X., Boyarintsev, V. V., Kovaleva, E. V. (2023). Tissue-Oxygen-Adaptation of Bone Marrow-Derived Mesenchymal Stromal Cells Enhances Their Immunomodulatory and Pro-Angiogenic Capacity, Resulting in Accelerated Healing of Chemical Burns. Int J Mol Sci.;24(4).
- 178. Volkova, M. V., Boyarintsev, V. V., Trofimenko, A. V., Kovaleva, E. V., Othman, A. A., Melerzanov, A. V., et al. (2023). Local injection of bone-marrow derived mesenchymal stromal cells alters a molecular expression profile of a contact frostbite injury wound and improves healing in a rat model. *Burns: Journal Of The International Society For Burn Injuries*, 49(2), 432–443.
- 179. Amadeo, F., Hanson, V., Liptrott, N. J., Wilm, B., Murray, P., & Taylor, A. (2023). Fate of intravenously administered umbilical cord mesenchymal stromal cells and interactions with the host's immune system. *Biomedicine & Pharmacotherapy*, 159, 114191.
- 180. Niess, H., von Einem, J. C., Thomas, M. N., Michl, M., Angele, M. K., Huss, R., et al. (2015). Treatment of advanced gastrointestinal tumors with genetically modified autologous mesenchymal stromal cells (TREAT-ME1): Study protocol of a phase I/II clinical trial. *Bmc Cancer*, 15, 237.
- 181. Shahror, R. A., Ali, A. A. A., Wu, C. C., Chiang, Y. H., & Chen, K. Y. (2019). Enhanced Homing of Mesenchymal Stem Cells



- Overexpressing Fibroblast Growth Factor 21 to Injury Site in a Mouse Model of Traumatic Brain Injury.Int J Mol Sci.;20(11).
- 182. Park, S. R., Cho, A., Kim, J. W., Lee, H. Y., & Hong, I. S. (2019). A novel endogenous damage Signal, CSF-2, activates multiple beneficial functions of adipose tissue-derived mesenchymal stem cells. *Molecular Therapy*, 27(6), 1087–1100.
- 183. Chen, M. S., Lin, C. Y., Chiu, Y. H., Chen, C. P., Tsai, P. J., & Wang, H. S. (2018). IL-1beta-Induced Matrix Metalloprotease-1 promotes mesenchymal stem cell Migration via PAR1 and G-Protein-coupled signaling pathway. Stem Cells Int, 2018, 3524759.
- 184. Perets, N., Betzer, O., Shapira, R., Brenstein, S., Angel, A., Sadan, T., et al. (2019). Golden Exosomes selectively target brain pathologies in neurodegenerative and neurodevelopmental Disorders. *Nano Letters*, 19(6), 3422–3431.
- 185. Meng, S. S., Xu, X. P., Chang, W., Lu, Z. H., Huang, L. L., Xu, J. Y., et al. (2018). LincRNA-p21 promotes mesenchymal stem cell migration capacity and survival through hypoxic preconditioning. Stem Cell Research & Therapy, 9(1), 280.
- 186. Antebi, B., Asher, A. M., Rodriguez, L. A. 2nd, Moore, R. K., Mohammadipoor, A., & Cancio, L. C. (2019). Cryopreserved mesenchymal stem cells regain functional potency following a 24-h acclimation period. *J Transl Med*, 17(1), 297.
- 187. Oja, S., Kaartinen, T., Ahti, M., Korhonen, M., Laitinen, A., & Nystedt, J. (2019). The utilization of freezing Steps in Mesenchymal Stromal Cell (MSC) Manufacturing: Potential impact on quality and cell functionality attributes. Frontiers In Immunology, 10, 1627.
- 188. Zhu, Y., Bassoff, N., Reinshagen, C., Bhere, D., Nowicki, M. O., Lawler, S. E., et al. (2017). Bi-specific molecule against EGFR

- and death receptors simultaneously targets proliferation and death pathways in tumors. *Scientific Reports*, 7(1), 2602.
- 189. Zhang, X., Yang, Y., Zhang, L., Lu, Y., Zhang, Q., Fan, D., et al. (2017). Mesenchymal stromal cells as vehicles of tetravalent bispecific Tandab (CD3/CD19) for the treatment of B cell lymphoma combined with IDO pathway inhibitor D-1-methyl-tryptophan. *Journal Of Hematology & Oncology*, 10(1), 56.
- 190. Liu, H., Kemeny, D. M., Heng, B. C., Ouyang, H. W., Melendez, A. J., & Cao, T. (2006). The immunogenicity and immunomodulatory function of osteogenic cells differentiated from mesenchymal stem cells. *The Journal Of Immunology*, 176(5), 2864–2871.
- 191. Isakova, I. A., Lanclos, C., Bruhn, J., Kuroda, M. J., Baker, K. C., Krishnappa, V., et al. (2014). Allo-reactivity of mesenchymal stem cells in rhesus macaques is dose and haplotype dependent and limits durable cell engraftment in vivo. *PLoS One*, 9(1), e87238.

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