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Original article

Prevalence and impact factors of recurrent positive SARS-CoV-2 detection in 599 hospitalized COVID-19 patients

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ABSTRACT

Objectives: Repeat-positive tests for severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) in individuals with coronavirus disease 2019 (COVID-19) were common. We aimed to investigate the rate and risk factors of recurrent positive detection of SARS-CoV-2 in hospitalized individuals with COVID-19. *Methods*: Oropharyngeal and nasopharyngeal swabs (n = 3513) were collected to detect SARS-CoV-2 during the hospitalization. We analysed the recurrent positive rate after consecutive negative results and its relationship to demographic characteristics.

Results: Among 599 enrolled individuals with COVID-19, the median time for viral RNA shedding was 24 days (interquartile range 19–33 days). The positive rates of RT-PCR were 35.9% (215/599), 17.0% (65/383) and 12.4% (23/185) after one, two and three consecutive negative RT-PCR test results, respectively. Medians of Ct values of initial positive test, rebound positive test after two consecutive negative results, and rebound positive after three consecutive negative results were 28.8, 32.8 and 36.1, respectively. Compared with male patients, females had a significantly higher rate of recurrent positive RT-PCR after three consecutive negative results (18.2%, 18/99, versus 5.8%, 5/86; p 0.013). Older individuals (\geq 55 years) had a significantly higher rate of recurrent positive rate than oropharyngeal swab tests (37.3%, 152/408, versus 35.8%, 1111/3105).

Conclusion: Our study revealed the prevalence and dynamic characteristics of recurrent positive RT-PCR to SARS-CoV-2. We showed that around 17.0% (65/383) of patients tested positive for SARS-CoV-2 after two consecutive negative results. Patients with a rebound positive RT-PCR test had a low viral load. Older age and being female were risk factors for recurrent positive results. **Chun Gao, Clin Microbiol Infect 2021;27:785.e1–785.e7**

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Introduction

To date, coronavirus disease 2019 (COVID-19) associated with severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) has caused a global pandemic [1,2]. At the time of drafting the manuscript (Jan 5th, 2021), over 85 500 000 cases were confirmed worldwide with a case fatality rate of approximately 2.2% [3].

The Chinese National Health Committee (seventh version) updated their guidelines regarding discharge and discontinuing isolation to include resolution of fever, improvement in respiratory symptoms and radiography findings, and with two consecutive negative real-time RT-PCR test results for SARS-COV-2 RNA (from two consecutive respiratory specimens collected \geq 24 hours apart) [4]. The US CDC updated its criteria for discontinuation of transmission-based precautions for individuals with confirmed COVID-19; a negative RT-PCR test is no longer recommended [5]. Nonetheless, increasing reports on recurrent positive RT-PCR tests for SARS-COV-2 have aroused wide concern [6–8]. Prolonged viral RNA shedding in certain infected individuals and relatively high false negatives for the viral test by RT-PCR may be responsible for the recurrence. A false-negative RT-PCR result is defined as the negative test result followed by a recurrent positive test. False-negative results were mainly caused by errors in sampling and detection methods.

The prevalence of recurrent positive results of RT-PCR tests to SARS-CoV-2 were reported in several studies. Here, we conducted a

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retrospective study to investigate the dynamic viral RNA shedding and impact factors for recurrent positive test results of RT-PCR to SARS-CoV-2.

Materials and methods

Study design and participants

We performed a retrospective study with 599 hospitalized individuals with COVID-19 in Tongji Hospital of Huazhong University of Science and Technology in Wuhan, China. All the patients enrolled were confirmed as being diagnosed with COVID-19 according to the diagnosis and treatment guideline for SARS-CoV-2 from the Chinese National Health Committee (seventh version) [4]. The inclusion criteria of this study were as follows: (a) laboratory-confirmed diagnosis of COVID-19; (b) moderate or severe illness; and (c) receive at least one RT-PCR test after two consecutive negative RT-PCR test results during hospitalization. Patients included for analysis in our previous publication were excluded to avoid duplication of data [9] (see Supplementary material, Fig. S1). This study was approved by the ethics committee of Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology. Written informed consent was waived by the ethics commission of the designated hospital for emerging infectious disease. All data were collected up to the final follow-up date (21 April 2020).

Data collection and definitions

We collected demographic and clinical data of patients from the electronic medical record system. We included patients with moderate or severe illness according to the guidelines-Moderate: individuals who have evidence of lower respiratory disease by clinical assessment or imaging and a saturation of oxygen $(Spo_2) \ge 94\%$ on room air at sea level and Severe: individuals who have respiratory frequency >30 breaths per minute, Spo₂ <94% on room air at sea level (or, for patients with chronic hypoxemia, a decrease from baseline of >3%), ratio of arterial partial pressure of oxygen to fraction of inspired oxygen (Pao₂/ Fio₂) <300 mmHg, or lung infiltrates >50%) [4,5]. Oropharyngeal and nasopharyngeal swabs were collected both upon admission and during hospitalization by qualified medical professionals through standard procedures under level 3 biosafety protection. Real-time RT-PCR assay were performed on swabs to detect SARS-CoV-2 using COVID-19 test kits (Shanghai Huirui Biotechnology Co., Ltd, Shanghai, China). Once two consecutive negative results were collected, the period between symptom onset and the date of first negative RT-PCR test result was considered as the length of viral RNA shedding.

RT-PCR assay for SARS-CoV-2

Oropharyngeal swab samples or nasopharyngeal swab samples were collected to extract RNA to confirm the diagnosis of COVID-19. Total RNA was extracted using magnetic beads (Tianlong, Xi'an, China). Two target genes, including open reading frame 1ab (ORF1ab) and nucleocapsid protein (N), were simultaneously amplified and tested during the real-time RT-PCR assay. The real-time RT-PCR assay was performed using a COVID-19 nucleic acid detection kit according to the manufacturer's protocol (Shanghai Huirui Biotechnology Co., Ltd). The results of the RT-PCR assay were expressed as the cycle threshold (Ct) value. As recommend by the instruction, a Ct value < 37 was defined as a positive test result, and a Ct value \geq 39.2 was considered as a negative test. Ct values

between 37 and 39.2 suggested that a confirmatory RT-PCR should be obtained using the same sample.

Ethical approval

This study was approved by the ethics committee of Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology. All procedures followed in this study were in accordance with the 1964 Helsinki Declaration and later versions. Written informed consent was waived by the ethics commission of the designated hospital for emerging infectious disease.

Statistical analysis

We performed statistical analyses using SPSS version 24.0 (IBM, Armonk, NY, USA). Continuous variables were present as mean \pm standard error of the mean or medians (interquartile range, IQR) and were analysed with Mann–Whitney *U* test. We reported categorical variables as whole numbers and percentages. The cutoff value for age was determined from the receiver operating characteristic curve according to prolonged viral shedding (>24 days). A p value < 0.05 was considered statistically significant.

Results

Demographic and clinical characteristics of patients with recurrent positive RT-PCR results

We included a total of 599 individuals with a confirmed diagnosis of COVID-19 in our study (Table 1). In detail, the median age of the included patients was 61 years (IQR 49–68 years) with 291 men (48.6%, 291/599) and 308 women (51.4%, 308/599). A total of 569 (95.0%, 569/599) were classified as having moderate disease. No

Table 1

The demographic and clinical characteristics of patients with COVID-19

Variables	All patients ($n = 599$)
Age (years), median (IQR)	61 (49-68)
Gender, male, n (%)	291 (48.6%)
Smoking, Yes, n (%)	24 (4.0%)
Co-morbidities, n (%)	
Hypertension	122 (20.3%)
Diabetes	68 (11.3%)
Pulmonary diseases	13 (2.2%)
Malignancy	5 (0.8%)
Renal failure	6 (1.0%)
Cerebrovascular diseases	9 (1.5%)
Severity on admission, n (%)	
Moderate	569 (95.0%)
Severe	30 (5.0%)
RT-PCR test, median (IQR)	
Onset of symptom to first RT-PCR test (days)	7 (4–11)
Length of viral RNA shedding (days)	24 (19-33)
No. of RT-PCR tests of each patients	5 (4-8)
Clinical characteristics, median (IQR)	
Onset of symptom to admission (days)	10 (7-14)
Length of hospital stay (days)	30 (23-40)
Positive rate of SARS-CoV-2 RT-PCR (weeks after onset)	
Week 2	90.2%
Week 3	59.6%
Week 4	35.4%
Week 5	20.7%
Week 6	11.5%
Week 7	5.3%
Week 8	1.0%
Week 9	0.5%
\geq Week 10	0%

Abbreviations: COVID-19, coronavirus disease 2019; IQR, interquartile range; SARS-CoV-2, severe acute respiratory syndrome coronavirus 2.

patient was transferred to an intensive care unit. The median time from onset of symptoms to admission was 10 days (IQR 7–14 days) and the median length of hospital stay was 30 days (23–40 days). The median numbers of RT-PCR tests per patient was 5 (IQR 4–8). The median length for viral shedding was 24 days (IQR 19–33 days). The percentages of patients who shed virus for less than 3 weeks, between 3 and 6 weeks and more than 6 weeks were 40.4%, 48.1% and 11.5%, respectively (see Supplementary material, Fig. S2).

Positive RT-PCR rates were 90.2% (540/599), 59.6% (357/599), 35.4% (212/599), 20.7% (124/599), 11.5% (68/599), 5.3% (32/599), 1.0% (6/599), 0.5% (3/599) and 0% (0/599) on weeks 2, 3, 4, 5, 6, 7, 8, 9 and 10 or later after symptoms onset, respectively (Fig. 1a).

Table 2 shows the demographic characteristics of individuals with recurrent positive RT-PCR results; 35.9% (215/599) of patients had positive RT-PCR results after one negative result. Among 383 individuals with two consecutive negative RT-PCR results, 17.0% (65/383) had recurrent positive test results. Among 185 individuals with three consecutive negative RT-PCR results, 12.4% (23/185) had a recurrent positive test result (Fig. 1b). As demonstrated in Fig. 1c, individuals older than 55 years had a significantly higher recurrence rate after one negative result (42.3%, 165/390, versus 23.9%, 50/209; p < 0.001); however, no significant difference in recurrent positive rate was found after two or three consecutive negative RT-PCR results regardless of age. Women had a significantly higher rate of recurrent positive RT-PCR after three consecutive negative results than men (18.2%, 18/99, versus 5.8%, 5/86; p 0.013) (Fig. 1d). The rate of recurrent positive tests between genders showed no significant difference after one or two consecutive negative RT-PCR results. Fig. 2 shows the dynamic of RT-PCR Ct values. Medians of Ct values of initial positive test, rebound positive test after two consecutive negative results and rebound positive test after three consecutive negative results were 28.8 (95% CI 28.0-29.7), 32.8 (95% CI 32.0-34.4) and 36.1 (95% CI 33.6-37.1), respectively.

Comparison of viral RNA shedding time in individuals with COVID-19 by age and gender

We analysed the viral RNA shedding time of 599 individuals grouped by age and gender (Table 3). As demonstrated in Fig. 3a, individuals <55 years had a shorter period of viral RNA shedding than those aged \geq 55 years, with a significantly higher percentage of end of viral shedding on week 2 (13.9%, 29/209, versus 7.7%, 30/390; p 0.02) and week 3 (33.5%, 70/209, versus 29.0%, 113/390; p < 0.01) after symptom onset. Individuals aged \geq 55 years had a significantly higher percentage for length of viral shedding on week 7 after symptom onset (9.2%, 36/390, versus 0.5%, 1/209; p 0.03). Men had a shorter period of viral shedding than women, with a higher percentage of viral shedding during the early stages (from week 2 to 5 after symptom onset) (Fig. 3b).

Comparison of positive rate for SARS-CoV-2 detection by oropharyngeal and nasopharyngeal swabs

From week 1 to week 6 after symptom onset, nasopharyngeal swabs produced a higher positive rate for SARS-CoV-2 than oropharyngeal swabs (see Supplementary material, Fig. S3a). In weeks 3, 4 and 5 after symptom onset, positive rates from nasopharyngeal swabs were significantly higher than those from oropharyngeal swabs (65.0%, 13/20, versus 30.1%, 195/647, 44.6%, 37/83, versus 22.9%, 155/677, and 36.3%, 33/91, versus 17.9%, 85/474; p < 0.05). Medians of the Ct values of initial positive test, rebound positive test after two consecutive negative results and rebound positive test after three consecutive negative results from nasopharyngeal swabs were higher than those from oropharyngeal swabs (see Supplementary material, Fig, S3b). From week 6 to week 10 and beyond after onset of symptoms, positive rates for viral detection using nasopharyngeal and oropharyngeal tests were not significantly different (see Supplementary material, Table S1).



Fig. 1. Positive rate of RT-PCR test and recurrent positive result. (a) Positive rate of RT-PCR to SARS-CoV-2 from symptom onset. (b) Recurrent positive rate of RT-PCR after consecutive negative results. (c) Impact of age on recurrent positive rate of RT-PCR. (d) Impact of gender on recurrent positive rate of RT-PCR.

Variables	RT-PCR after one negative result ($n = 599$)		p value	RT-PCR after two consecutive negative results ($n = 383$)		p value	RT-PCR after 3 consecutive negative results ($n = 185$)		p value
	Positive	Negative		Positive	Negative		Positive	Negative	
Age (years)			< 0.001			0.553			0.484
<55	50/209 (23.9%)	159/209 (76.1%)		17/115 (14.8%)	98/115 (85.2%)		6/64 (9.4%)	58/64 (90.6%)	
≥55	165/390 (42.3%)	225/390 (57.7%)		48/268 (17.9%)	220/268 (82.1%)		17/121 (14.0%)	104/121 (86.0%)	
Gender			0.088			0.341			0.013
Male	94/291 (32.3%)	197/291 (67.7%)		28/188 (14.9%)	160/188 (85.1%)		5/86 (5.8%)	81/86 (94.2%)	
Female	121/308 (39.3%)	187/308 (60.7%)		37/195 (19.0%)	158/195 (81.0%)		18/99 (18.2%)	81/99 (81.8%)	
Total	215/599 (35.9%)	384/599 (64.1%)		65/383 (17.0%)	318/383 (83.0%)		23/185 (12.4%)	162/185 (87.6%)	

Table 2
The demographic characteristics of patients with recurrent positive RT-PCR result for SARS-CoV-2

Abbreviations: SARS-CoV-2, severe acute respiratory syndrome coronavirus 2.

Discussion

With more than 85 500 000 infections, the world is now facing a pandemic of COVID-19. In this study, we collected data from 599 individuals with COVID-19 and summarized the duration of viral RNA shedding. Meanwhile, we analysed the rate of recurrent positive RT-PCR results and their impact factors.

According to various reports, the median duration of viral RNA shedding ranged from 12 to 31 days from illness onset [10-12]. These reports had a small sample size or a relatively short observation period. In our study, we demonstrated the dynamics of viral

RNA shedding after symptom onset in 599 individuals with COVID-19. We found that the median time from symptom onset to the end of viral RNA shedding was 24 days (IQR 19–33 days); 40.4% (242/ 599) of patients had viral shedding within 3 weeks after symptom onset, and 11.5% (69/599) of patients for over 6 weeks (see Supplementary material, Fig. S2).

Prolonged viral RNA shedding of COVID-19 was reported in various studies [7,13,14]. Although the potential mechanism for prolonged viral shedding was not yet elucidated, studies showed that impaired immune function and a high level of proinflammatory cytokines in the serum of individuals with COVID-



Fig. 2. Dynamic characteristics of Ct value by RT-PCR (a), and initial and rebound Ct value (medians, interquartile range) after negative RT-PCR tests (b).

Table 3
Viral RNA shedding time of patients with COVID-19

Viral RNA shedding time	Age		p value	Gender	p value	
	<55 years	\geq 55 years		Male	Female	
Week 2	29/209 (13.9%)	30/390 (7.7%)	0.020	32/291 (11.0%)	27/308 (8.8%)	0.36
Week 3	70/209 (33.5%)	113/390 (29.0%)	< 0.001	94/291 (32.3%)	89/308 (28.9%)	0.37
Week 4	57/209 (27.3%)	88/390 (22.6%)	0.200	72/291 (24.7%)	73/308 (23.7%)	0.77
Week 5	29/209 (13.9%)	59/390 (15.1%)	0.680	46/291 (15.8%)	42/308 (13.6%)	0.45
Week 6	14/209 (6.7%)	41/390 (10.5%)	0.130	23/291 (7.9%)	32/308 (10.4%)	0.29
Week 7	1/209 (0.5%)	36/390 (9.2%)	0.003	13/291 (4.5%)	24/308 (7.8%)	0.1
Week 8	7/209 (3.3%)	19/390 (4.9%)	0.390	9/291 (3.1%)	17/308 (5.5%)	0.15
Week 9	1/209 (0.5%)	2/390 (0.5%)	0.950	2/291 (0.7%)	1/308 (0.3%)	0.54
\geq Week 10	1/209 (0.5%)	2/390 (0.5%)	0.950	0/291 (0.0%)	3/308 (1.0%)	0.21

Abbreviations: COVID-19, coronavirus disease 2019.

19 were risk factors for prolonged shedding [13,14]. Our previous study also revealed distinct characteristics between patients with or without prolonged viral RNA shedding. We found that hypertension, older age, lymphopenia and high levels of interleukin-2 receptor were independent risk factors for prolonged viral RNA shedding [15]. Patients with prolonged RNA shedding were generally asymptomatic. However, a recent study reported a case of recurrent symptomatic pneumonia with rebound positive RT-PCR test after discharge from hospital. The authors speculate that low titres of anti-SARS-CoV-2 antibodies may be the reason for COVID-19 relapse [16].

Recurrent detection of SARS-CoV-2 viral RNA by RT-PCR was reported in recent studies [6–8,17,18]. The large-scale study including 257 individuals with COVID-19 by Zou et al. revealed that 53 patients (20.6%) suffered with recurrence of positive SARS-CoV-2 RNA detection after two consecutive negative results [6]. Zou et al. also demonstrated that the positive rate of RT-PCR test for SARS-CoV-2 was 5.4% after three negative detections. Another study, by Hao et al. [14] including 104 individuals with COVID-19 found that the positive rates of RT-PCR viral detection were 30.5%, 16.4% and 4.8% after one, two and three consecutive negative RT-PCR test results, respectively. In our study we found that the positive rates of RT-PCR tests were 35.9%, 17.0% and 12.4% after one, two and three consecutive negative rate (17.0%) after two negative tests was close to those of previous reports.

The causes of recurrent positive tests for SARS-CoV-2 were unclear. Currently, false-negative RT-PCR tests were considered as the main reason for recurrent positivity, and they could result from errors in sampling or non-infectious viral RNA remnants. However, the false-negative results did not include the negative results achieved when the viral load was very low or reached the limit of the detection assay. Recurrent positives of COVID-19 have raised increasing public concern regarding the infectivity of individuals with recurrent positive tests. In theory, viral transmission is determined by viral replication. Recurrent-positive patients may be a potential source of transmission. However, accumulating evidence has suggested that patients with COVID-19 who have retested positive are barely infectious. In late-phase, non-infectious remnant viral RNA may still be detectable by RT-PCR. According to different reports, the Ct values retrieved from re-positive patients were generally higher than those from initial positive tests. A viral dynamics study by He et al. revealed that the highest viral load was detected at the time of symptom onset. The authors inferred that transmissibility of COVID-19 peaked on or even before symptom onset [19]. The viral load in nasopharyngeal swabs decreased to extremely low limits on day 21 after infection [19]. A recent report by Bullard et al. indicated no viral growth in samples with a Ct

value > 24 or symptom onset to test of >8 days [20]. Another study also proved that samples retrieved from positive RT-PCR tests following negative results with Ct values > 29.5 were not associated with virus culture in vitro [21]. A study from England showed that the probability of culturing virus declined to 8% in samples with Ct value > 35 and to 6% 10 days after symptom onset [22]. In a large-scale multicentre investigation, researchers found that none of the 209 close contacts of 69 re-positive patients developed COVID-19 [23]. Ct values are inversely related to viral RNA copy number. According to the literature, Ct values of 35 may correspond to a viral load between 1.0×10^2 and 1.0×10^3 copies/mL [24]. In our study, we also noted a trend of increased Ct values in individuals with re-positive tests after two (32.8) and three (36.1) negative tests. At this point, without standardization of different RT-PCR assays, we do not have a precise threshold value of infectivity. Nonetheless, according to the above reports, we can deduce that patients with Ct values > 35 (viral load $< 10^3$ copies/ml) were hardly infectious. In our study, the median Ct values of recurrent positive (32.8 and 36.1) suggested that the transmission risk of these patients was low, even if viral shedding could still be detected by RT-PCR in late phase. Until now, no evidence indicating aggressive treatment or public isolation were benefit for these patients. Our results suggested that patients with Ct value > 36 may be considered for discharge.

Furthermore, we investigate the impact of age and gender on false-negative rates of RT-PCR to SARS-CoV-2. We found that older individuals and women tended to have a higher false-negative rate of RT-PCR tests. Notably, female patients had a significantly higher rate of recurrent positive RT-PCR after three consecutive negative results than male patients (18.2%, 18/99, versus 5.8%, 5/86; p 0.013). In clinical practice, the above findings may provide information for repeat viral testing in selected patients. Studies also showed that older age, co-morbidity, low level of antibody response and host immunity status may be potential risk factors for recurrent positivity of RT-PCR tests for SARS-CoV-2, while comprehensive intervention may be a protective factor [25,26].

Various studies investigate the dynamics of viral shedding in individuals with COVID-19. Patients with severe disease had a longer duration of viral shedding [9,10]. Our study showed that 36.7% of patients aged \geq 55 years, compared with 47.4% of younger patients (age <55 years), had a median viral shedding time of <3 weeks. Male patients had a shorter viral RNA shedding period than females (43.3%, 126/291 versus 37.7%, 116/308 in 3 weeks) (Table 3). In our study, samples from nasopharyngeal swabs produced a higher positive rate than samples from oropharyngeal swabs. The mean RT-PCR Ct value was higher from oropharyngeal specimens than from nasopharyngeal specimens (see



Fig. 3. Dynamic characteristics of viral RNA shedding by RT-PCR grouped by (a) age and (b) gender.

Supplementary material, Fig. S3). Hence, a false-negative test result of RT-PCR may be related to the sampling site. We suggest that nasopharyngeal swabs should be the standard procedure to obtain respiratory specimens for viral tests.

This large-scale study revealed the false-negative rate and dynamic profile of RT-PCR tests for SARS-CoV-2. We also investigated the different viral shedding dynamics of gender and age. The present study has several limitations. First, some data, such as laboratory characteristics, serological data and treatments, were incomplete and not included for analysis. Second, we only included symptomatic hospitalized patients with moderate to severe illness. The results may not be applicable for asymptomatic infected individuals or critical cases. Third, patients included in our study had not received identical treatments, which might have an impact on the duration of viral shedding.

Conclusions

In summary, in this study we for the first time performed a large-scale investigation of the dynamic characteristics of viral RNA shedding in 599 hospitalized individuals with COVID-19. We showed that the median length of viral RNA shedding was 24 days from symptom onset. The positive rate of RT-PCR tests to SARS-

CoV-2 was 17% after two consecutive negative results. Recurrent positive patients with a relatively high Ct value were unlikely to be infectious.

Transparency declaration

All authors declare that there are no conflicts of interest.

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We thank Cheng Chen for correcting the English in this manuscript.

Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.cmi.2021.01.028.

Author contributions

All authors participated in the study design. CG and SZ contributed to conceptualization and writing the original draft. LZ and ATX contributed to formal analysis and project administration.

CCJ contributed to data curation, methodology and software. YXT contributed to methodology and software. All authors have agreed on the final version and meet the major criteria recommended by the ICMJE (http://www.icmje.org/).

Availability of data and materials

The database used and/or analysed during the current study is not publicly available (to maintain privacy) but can be available from the corresponding author on reasonable request.

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