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OPEN Opposite correlation of 25-hydroxy-vitamin Dand 1,25-dihydroxy-vitamin **D**-metabolites with gestational age, bone- and lipid-biomarkers in pregnant women

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25-Hydroxyvitamin D (25OHD) and 1,25-dihydroxyvitamin D (1,25(OH)₂D) need to be bound to carrier proteins to be transported to their target cells. The majority of either 25OHD or 1,25(OH)₂D is bound to vitamin D-binding protein (DBP), a smaller fraction is bound to albumin and only very small amounts of 25OHD or 1,25(OH)₂D are free. Albumin-bound 25OHD or 1,25(OH)₂D is relatively easily available after dissociation from albumin. Thus, the sum of free and albumin-bound forms is called bioavailable 250HD and bioavailable 1,25(OH)₂D. Total 250HD and 1,25(OH)₂D are defined as the sum of free, albumin-bound and DBP-bound 25OHD and 1,25(OH)₂D, respectively. This crosssectional study in 427 pregnant women compared the correlation of the six vitamin D compounds with biomarkers of bone health, lipid metabolism, kidney function, endocrine parameters, and group B water-soluble vitamins. Among the 250HD metabolites analysed, total 1,25(OH)₂D showed clearly the best correlation with calcium, bone-specific alkaline phosphatase, adiponectin, LDL, HDL, urea, thyroxine, and group B water-soluble vitamins. When comparing the three 25OHD metabolites, both free 25OHD and bioavailable 25OHD showed overall good correlations with calcium, bone-specific alkaline phosphatase, adiponectin, LDL, HDL, urea, thyroxine, triiodothyronine, and group B watersoluble vitamins, The correlations of 1,25(OH)₂D and 25OHD metabolites went always in opposite directions. Only PTH correlates always inversely with all six vitamin D compounds. In conclusion, free 25(OH)D and bioavailable 25(OH)D are more precise determinants of the vitamin D status than total 25(OH)D in normal pregnancy, whereas total 1,25(OH)₂D is superior to free and bioavailable 1,25(OH),D. Except for PTH, correlations of 25(OH)D and 1,25(OH),D metabolites with typical clinical chemistry readouts go in opposite directions.

Abbreviations

125(OH) ₂ D	1,25-Dihydroxyvitamin D
25OHD	25-Hydroxyvitamin D
ANOVA	Analysis of variance
B25OHD	Bioavailable 25-hydroxyvitamin D
B125(OH) ₂ D	Bioavailable 1,25-dihydroxyvitamin D
BSAP	Bone-specific alkaline phosphatase

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CE	Conformité Européene
DBP	Vitamin D-binding protein
ELISA	Enzyme-linked immunosorbent assay
F25OHD	Free 25-hydroxyvitamin D
F125(OH) ₂ D	Free 1,25-dihydroxyvitamin D
fT3	Free triiodothyronine
fT4	Free thyroxine
HDL	High-density lipoprotein cholesterol
IVD	In-Vitro-Diagnostic Medical Device
LC-MS/MS	Liquid chromatography-tandem mass spectrometry
LDL	Low-density lipoprotein cholesterol
MCH	Mean corpuscular hemoglobin
MCHC	Mean corpuscular hemoglobin concentration
MCV	Mean corpuscular volume of red blood cells
PTH	Parathyroid hormone
RBC	Red blood cell
RDW	Red blood cell distribution width
SD	Standard deviation
T25OHD	Total 25-hydroxyvitamin D
T125(OH) ₂ D	Total 1,25-dihydroxyvitamin D
VDR	Vitamin D receptor
WBC	White blood cell

Both 25OHD and $1,25(OH)_2D$, being lipophilic molecules, have a special carrying protein, enabling their transport in the circulation, protection from hepatic degradation and accumulation of circulating reserves, called vitamin D-binding protein (DBP). In the serum, 85–90% of the whole 25OHD/1,25(OH)₂D pool is present in the DBP-bound condition. The remaining 10–15% are loosely bound to serum albumin and lipoproteins (10–15%), and just a tiny fraction of 25OHD/1,25(OH)₂D (less than 0.1% of 25OHD and about 0.4% of 1,25(OH)₂D^{1,2}) is available in the free, unbound condition. Thus, from this point of view, the terminology "total 25OHD" "total 1,25(OH)₂D", which will be used throughout this manuscript, refers to a sum of three 25OHD/1,25(OH)₂D fractions: DBP-bound, albumin-bound (the latter fraction is also called bioavailable, since this binding is very weak and spontaneous dissociation of vitamin D metabolites from albumin is very easy to occur), and free compounds. Since serum levels of liver-produced DBP is subject to substantiate changes under numerous physiologic and pathologic conditions (genetic background, endocrine status in particular with regards to estrogens, liver diseases and kidney diseases), the values of total 25OHD—the current gold standard vitamin D status marker—might not precisely clarify the status of vitamin D, particularly in such conditions as normal pregnancy, estrogen-containing oral contraceptive intake, as well as liver and renal pathologies.

Since the mid-1980s it is known that free $25OHD/1,25(OH)_2D$ determination is possible based on the laws of protein–ligand binding kinetics using a formula employing serum concentrations of total 25OHD, DBP, albumin, as well as $25OHD/1,25(OH)_2D$ -DBP and $25OHD/1,25(OH)_2D$ -albumin binding affinity constants, which was originally suggested by Bikle et al.³.

The present study was designed to perform for the first time a systematic head-to-head comparison of three ways of measuring 25-hydroxyvitamin D (free, bioactive, ant total 25OHD) with three ways to measure 1,25-dihydroxyvitamin D (free, bioactive, ant total 1,25(OH)₂D) using pregnant women as a test system, because vitamin D binding protein is incresed in this population.

Materials and methods

Study population. The study was a part of the Berlin Birth Cohort (BBC) study^{4,5}. This study was approved by the local ethical committee of the Charité—Universitätsmedizin Berlin, Campus Mitte, Berlin, Germany and written informed consent was obtained from all participants. The study cohort consisted of 427 Caucasian pregnant healthy women aged > 18 years. All methods were carried out in accordance with relevant German guide-lines and regulations.

The study participants were categorized into three trimester groups according to their gestational age: the 1st trimester (≤ 12 weeks+6 days), the 2nd trimester (13 weeks-27 weeks+6 days), and the 3rd trimester (≥ 28 weeks until delivery). The mean gestational age of the entire study cohort was 18.49 weeks (n=427). The group sizes were 211, 78, and 135 participants in 1st, 2nd, and 3rd pregnancy trimesters, respectively.

The study was carried out at clinical laboratory medical institution in Berlin, Germany. Venous blood samples were collected in the period from May till June 2016. The study was approved by the local ethics committee.

Measurement of 25OHD and 1,25(OH)₂**D metabolites, DBP, and albumin.** All analytes were measured in serum samples. Total 25OHD (hereafter "25OHD" is referred to as a sum of 25OHD₂ and 25OHD₃ metabolites) was measured by Abbott Architect i2000 (Abbott Laboratories, Wiesbaden, Germany) using Abbott Architect 25OHD automated chemiluminescent microparticle immunoassay (Abbott Laboratories, Wiesbaden, Germany). As declared by the assay manufacturer, the kit has the following cross-reactivities with vitamin D metabolites: 98.6–101.1% with 25OHD₃, 54.0% with 25OHD₂, 101.9–189.2% with 24,25OH₂D₃, 71.4–114.2% with 24,25OH₂D₂, and <1% with other vitamin D forms (vitamin D₃, vitamin D₂, 1,25(OH)₂D₃, and 1,25(OH)₂D₂). Total 1,25(OH)₂D (hereafter "1,25(OH)₂D" is referred to as a sum of 1,25(OH)₂D₂ and 1,25(OH)₂D₃ metabolites) was determined using IDS-iSYS 1,25 VitD^{XP} chemiluminescence-immunoassay on

the IDS-iSYS Multi-Discipline Automated system (both by IDS Immunodiagnostic Systems GmbH, Frankfurt am Main, Germany). According to the manufacturer's data sheet, this assay has the following cross-reactivities with $1,25(OH)_2D$ metabolites: 97% with $1,25(OH)_2D_3$, 72% with $1,25(OH)_2D_2$, 106.7% with $1,24,25OH_3D_3$, and <1% with other vitamin D metabolites. Serum DBP was determined by a polyclonal antibody immunoassay (Immundiagnostik AG, Bensheim, Germany). Serum albumin was assessed using an automated analyzer (Beckman Coulter Inc., Krefeld, Germany). Regarding the quality control in total 25OHD, total $1,25(OH)_2D$ and albumin measurement, our lab constantly fulfill the requirements of the round robin tests organized by the Reference Institute for Bioanalytics (RfB)—one of the two German proficiency testing organizations which have been officially charged by the German Medical Association, in which our lab participates regularly. Regarding DBP, this analyte does not belong to the list of routine clinical biomarkers and was measured exclusively for the present study. However, this kit is IVD- ("In-Vitro-Diagnostic Medical Device") and CE- ("*Conformité Européene*") certified, which means that it can be used in-vitro for the examination of specimens, including bloodand tissue donations, derived from the human body within the European Economic Area.

Free 25OHD was measured based on the laws of protein–ligand binding kinetics by calculation method using a formula employing serum concentrations of total 25OHD, DBP, albumin, as well as 25OHD-DBP and 25OHD-albumin binding affinity constants, which was originally suggested by Bikle et al.⁶:

Free 25(OH)D =
$$\frac{Total 25(OH)D}{1 + K_{alb} \times albumin + K_{DBP} \times DBP}$$

where Free 25OHD = concentration of free 25OHD in mol/L; K_{alb} = affinity constant between 25OHD and albumin equal to 6×10^5 M⁻¹; K_{DBP} = affinity constant between 25OHD and DBP, equal to 7×10^8 M^{-16,7}; albumin = concentration of total serum albumin in mol/L; DBP = concentration of total vitamin D-binding protein in mol/L; Total 25OHD = concentration of total 25OHD in mol/L.

Utilizing exactly the same principle, but using $1,25(OH)_2D$ -specific binding affinity constants— $K_{DBP} = 4.0 \times 10^7 M^{-1}$ and $K_{alb} = 5.4 \times 10^4 M^{-1}$ —we calculated free $1,25(OH)_2D^{6.7}$:

Free1,
$$25(OH)_2D = \frac{Total \ 1, 25(OH)_2D}{1 + K_{alb} \times albumin + K_{DBP} \times DBP}$$

Bioavailable 25OHD and bioavailable $1,25(OH)_2D$ were calculated using the following formula based on precalculated free 25OHD or free $1,25(OH)_2D$ levels, respectively^{8,9}:

Bioavailable $D = Free D + albumin - bound D = Free D + K_{alb} \times albumin \times Free D$

where D = concentration of vitamin D metabolite (25OHD or $1,25(OH)_2D$) in mol/L; K_{alb} = affinity constant between a respective vitamin D metabolite ($6 \times 10^5 \text{ M}^{-1}$ for 25OHD and $5.4 \times 10^4 \text{ M}^{-1}$ for $1,25(OH)_2D$) and albumin in mol⁻¹; albumin = concentration of serum albumin in mol/L.

Measurement of other serum analytes. The detailed data on diagnostic assays and instrumental devices used for measurement of other serum parameters (calcium, PTH, bone-specific alkaline phosphatase, LDL, HDL, hemoglobin, etc.) are provided in Supplementary Table 1.

Statistical analyses. Statistical analyses were performed using SPSS 20.0 (IBM, New York, NY, USA) and GraphPad Prism version 5 software (GraphPad Software, San Diego, CA, USA) as recently described^{10,11}. All relevant parameters were tested for normality using a D'Agostino and Pearson omnibus normality test and were found to be not normally distributed (p < 0.05) in the majority of the cases. For this reason, we used Spearman's rank correlation analysis to assess bivariate associations between vitamin D metabolites with each other and with other analytes. The correlation strength (measured as Spearman's rho values) was classified as follows: ≤ 0.1 no correlation; 0.1-0.3 weak to modest; 0.3-0.49 moderate; > 0.5 strong correlation. To assess statistical differences among three trimester groups we used either one-way ANOVA followed by Tukey post hoc test (in cases when all three trimester groups were normally distributed, as determined by D'Agostino and Pearson omnibus normality text), or Kruskal–Wallis analysis with Dunn's multiple comparison test (in cases when at least one group out of three analyzed groups was found to be not normally distributed). In both correlation and group comparison analyses, correlations/group differences were considered significant if *p* values were < 0.05.

Results

Study participant characteristics. The mean values of participant's age and gestational age, as well as selected serum biomarkers for the pregnancy overall and for early, mid-, and late pregnancy are presented in Table 1.

Changes in serum parameters throughout pregnancy. The data on changes of biochemical and biochemical parameters throughout pregnancy (split by three trimesters) are presented in Table 1. Importantly, a progressive increase in DBP during pregnancy was observed (43% increase in 3rd trimester compared to the 1st trimester, p < 0.001, Table 1).

Regarding 25(OH)D metabolites, we found no significant changes in total 25OHD throughout pregnancy. In contrast, free 25OHD and bioavailable 25OHD were both significantly decreased with gestational age (Table 1). The dynamics in changes of $1,25(OH)_2D$ isoform levels was totally different compared to the behavior of 25OHD compounds. Indeed, total $1,25(OH)_2D$ showed a progressive increase throughout pregnancy. The same trend

Variable, units	1st trimester of pregnancy, mean±SD	2nd trimester of pregnancy, mean±SD	3rd trimester of pregnancy, mean ± SD	Changes throughout pregnancy	
T25OHD, ng/mL	18.80 ± 9.96	20.26±11.09	20.55±11.56	\leftrightarrow	
F25OHD, pg/mL	2.84 ± 1.54	2.41±1.41	2.31±1.48***	Ļ	
B25OHD, ng/mL	1.05 ± 0.57	0.76±0.47***	0.69±0.44***	Ļ	
T1,25(OH) ₂ D, pg/mL	105.80±42.59	150.95±47.28***	163.28±43.68***	1	
F1,25(OH) ₂ D, pg/mL	0.27±0.10	0.30±0.10**	0.31±0.10***	1	
B1,25(OH) ₂ D, pg/mL	9.05±3.47	8.84±2.93	8.58±2.68	\leftrightarrow	
Age, years	30.63±6.36	29.54±5.28	29.87±5.86	\leftrightarrow	
Gestational age, weeks	7.98 ± 1.84	21.29±4.06***	33.25±2.19 ^{\$\$\$} , ***	1	
PTH, pg/mL	15.37±10.76	14.60±13.48	17.93±13.98	\leftrightarrow	
BSAP, μg/mL	12.93±5.12	13.97±5.86	18.01±8.21 ^{\$§§,***}	1	
Calcium, mmol/L	2.38±0.12	2.25±0.13***	2.24±0.10***	Ļ	
Phosphate, U/L	4.39±1.65	4.07 ± 1.24	4.33±2.23	\leftrightarrow	
DBP, mg/L	532.56±138.73	690.00±171.15***	762.70±254.08***	1	
LDL, mg/dL	118.04±34.00	158.52±39.12***	184.16±40.60 ^{\$\$,***}	1	
HDL, mg/dL	59.36±13.25	65.10±12.40**	67.48±12.64***	1	
LDL/HDL ratio	2.05 ± 0.61	2.49±0.63***	2.83±0.82***	1	
Adiponectin, µg/mL	9.28±4.15	8.06±3.73*	8.41±3.97	Ļ	
Free T3, pg/mL	3.10±0.38	3.15±0.38	3.03±0.38 [§]	Ļ	
Free T4, ng/dL	1.02 ± 0.14	0.87±0.12***	0.85±0.11***	Ļ	
TSH, uU/mL	1.48 ± 0.97	1.54 ± 0.79	1.45 ± 0.66	\leftrightarrow	
Hemoglobin, g/dL	12.55 ± 0.94	11.53±0.83***	11.66±0.93***	Ļ	
RBC count, 106/µL	4.34±0.39	3.90±0.35***	4.00±0.34***	\downarrow	
WBC count, 103/µL	8.54±2.14	9.55±2.83*	9.75±2.33***	1	
Platelet count, 103/µL	232.55±50.51	214.12±42.94	220.68±54.10	\leftrightarrow	
MCV, fL	89.13±5.42	89.77±5.67	89.34±5.39	\leftrightarrow	
MCH, pg	29.18±1.96	29.73±2.12	29.32±1.95	\leftrightarrow	
MCHC, g/dL	32.75±0.97	33.15±0.96*	32.93±1.11	\leftrightarrow	
RDW, %	13.97±1.23	13.99±1.11	13.87±1.16	\leftrightarrow	
Hematocrit, %	38.57±2.85	34.86±2.39***	35.51±2.96***	Ļ	
Urea, mg/dL	23.07±5.85	17.97±4.89***	17.93±4.58***	Ļ	
Albumin, g/dL	4.09 ± 0.38	3.45±0.30***	3.30±0.24***	\downarrow	
Vitamin B6, ng/mL	14.43 ± 18.70	7.34±7.78***	5.55±13.11 ^{\$,***}	\downarrow	
Vitamin B12, pg/mL	368.63±157.81	279.23±123.68***	275.75±121.83***	Ļ	
Sodium, mmol/L	137.80 ± 4.46	138.03±2.80	137.77±3.43	\leftrightarrow	
Zinc, µmol/L	12.50±2.83	10.13±2.21**	9.86±2.13***	Ļ	

Table 1. Biochemical characteristics of the study population. Mean values ± standard deviation of age and gestational age, vitamin D metabolites and biochemical, hematological, and endocrinological serum parameters in 1st, 2nd, and 3rd pregnancy trimesters. B25OHD, bioavailable 25OHD; B1,25(OH)₂D, bioavailable 1,25(OH)₂D; BSAP, bone-specific alkaline phosphatase; F25OHD, free 25OHD; F1,25(OH)₂D, free 1,25(OH)₂D; fT3, free triiodothyronine; fT4, free thyroxine; HDL, high-density lipoprotein cholesterol; LDL, low-density lipoprotein cholesterol; MCH, mean corpuscular hemoglobin; MCHC, mean corpuscular hemoglobin concentration; MCV, mean corpuscular volume of red blood cells; PTH, parathyroid hormone; RBC, red blood cell; RDW, red blood cell distribution width; T25OHD, total 25OHD; T1,25(OH)₂D, total 1,25(OH)₂D; WBC, white blood cell. **p* < 0.05 versus 1st trimester; ***p* < 0.01 versus 1st trimester; ***p* < 0.001 versus 2nd trimester.

revealed free $1,25(OH)_2D$ with however weaker numerical rise. In contrast, bioavailable $1,25(OH)_2D$ showed no significant differences between early and late gestation groups (Table 1).

Associations between 25OHD and 1,25(OH)₂**D metabolites.** In general, absolutely all analyzed compounds of 25OHD and 1,25(OH)₂D showed a significant positive correlation with each other, however, the degree of such association (characterized by Spearman's rho) varied from 0.15 until 0.98 (p < 0.001 for all pairs of comparison, except of the pair total 1,25(OH)₂D versus bioavailable 25OHD with p = 0.002, Table 2). Regarding 25OHD isoforms, total 25OHD, free 25OHD, and bioavailable 25OHD revealed very strong correlation between each other (0.813 < rho < 0.976, Table 2). Regarding interrelations between 1,25(OH)₂D compounds, the correlation was substantially weaker (0.588 < rho < 0.923, p < 0.001 for all pairs of comparison) compared to the ones between 25OHD isoforms and more variable depending on certain pair of comparison. In addition,

	T25OHD, ng/mL	F25OHD, pg/mL	B 25OHD, ng/mL	T1,25(OH) ₂ D, pg/mL	F1,25(OH) ₂ D, pg/mL	B1,25(OH) ₂ D, pg/mL	Spearman's rho color gradient
T25OHD, ng/mL	-	rho = 0.886, p < 0.001	rho = 0.813, p < 0.001	rho = 0.572, p < 0.001	rho = 0.560, p < 0.001	rho = 0.543, p < 0.001	0.1 < rho < 0.2 0.2 < rho < 0.3
F25OHD, pg/mL		-	rho = 0.976, p < 0.001	rho = 0.290, p < 0.001	rho = 0.551, p < 0.001	rho = 0.628, p < 0.001	0.3 < rho < 0.4 0.4 < rho < 0.5
B25OHD, ng/mL			-	rho = 0.152, p = 0.002	rho = 0.455, p < 0.001	rho = 0.603, p < 0.001	0.5 < rho < 0.6 0.6 < rho < 0.7
T1,25(OH) ₂ D, pg/mL				-	rho = 0.775, p < 0.001	rho = 0.588, p < 0.001	0.7 < rho < 0.8 0.8 < rho < 0.9
F1,25(OH) ₂ D, pg/mL					-	rho = 0.923, p < 0.001	0.9 < rho < 1.0
B1,25(OH) ₂ D, pg/mL						-	

Table 2. Interrelations between 25OHD and $1,25(OH)_2D$ isoforms. The correlation strength is defined by Spearman's rank correlation coefficient rho. Abbreviations: B25OHD, bioavailable 25OHD; B1,25(OH)_2D, bioavailable 1,25(OH)_2D; F25OHD, free 25OHD; F1,25(OH)_2D, free 1,25(OH)_2D; T25OHD, total 25OHD; T1,25(OH)_2D, total 1,25(OH)_2D.

total 25OHD showed a medium correlation with all three $1,25(OH)_2D$ compounds of nearly equal strength (rho around 0.55). In contrast, free 25OHD and bioavailable 25OHD showed much weaked correlation with total $1,25(OH)_2D$ than with free $1,25(OH)_2D$ and bioavailable $1,25(OH)_2D$ (rho=0.29, 0.55, and 0.63, respectively, see Table 3). The relationship between bioavailable 25OHD and $1,25(OH)_2D$ compounds was very similar to interrelations between free 25OHD and $1,25(OH)_2D$.

Correlations of free and total 25(OH)D with serum biochemical and hematological parame-ters. Overall, free 25OHD and, particularly bioavailable 25OHD, showed stronger associations with distinct parameters than total 25OHD (Table 3).

Total 25OHD showed no correlation with serum calcium—a key parameter of bone health (rho=0.080, p=0.098), whereas free 25OHD revealed weak but significant positive correlation with calcium (rho=0.234, p<0.001, Table 3). Bioavailable 25OHD showed even stronger association with calcium (rho=0.369, p<0.001, Table 3).

Total 25(OH)D showed no correlation with serum albumin, free thyroxin, urea, adiponectin, LDL, LDL/HDL ratio, vitamin B12, hemoglobin, white blood cell count, and red blood cell distribution width, in contrast to free 25(OH)D, which did show weak but significant correlations with these analytes (Table 3). In addition, despite the presence of the correlation between total 25(OH)D and bone-specific alkaline phosphatase and vitamin B6, it was borderline significant, in contrast to free 25(OH)D, which was correlated with these parameters with p < 0.001. The associations of total and free 25(OH)D with other biochemical and hematological parameters are presented in Table 3.

Discussion

Summary of the key findings. To the best of our knowledge, the present cross-sectional study is the first report comparing the behavior of the six vitamin D compounds—free, bioavailable, and total 25-hydroxyvitamin D as well as free, bioavailable, and total 1,25-dihydroxyvitamin D—in a relatively large population of healthy pregnant women (n=427). Among the 1,25-dihydroxyvitamin D metabolites analyzed, total 1,25(OH)₂D showed clearly the best correlation with components of the endocrine vitamin D system and/or potential cellular targets of activation of the vitamin D receptor (see Table 3). When comparing the three 25OHD metabolites, both free 25OHD and bioavailable 25OHD showed overall good correlations with the analyzed clinical laboratory parameters, bioavailable 25OHD might perform even better then free 25OHD. The only exception from the above described overall findings is PTH. It correlates well with total 25OHD most likely because the parathyroid cells have a highly specific uptake mechanism for bound 25OHD—megalin/cubilin-uptake system, see below.

[Total 25(OH)D,	Free 25(OH)D,	Bioavailable 25(OH)D,	Total 1,25(OH) ₂ D,	Free 1,25(OH) ₂ D,	Bioavailable 1,25(OH) ₂ D,	Spearman's rho color
	ng/mL	pg/mL	ng/mL	pg/mL	pg/mL	pg/mL	gradient
Age and gestational age							-0.5 > rho > -0.6
Age, years	0.218, p < 0.001	0.235, p < 0.001	0.227, p < 0.001	0.027, p = 0.572	0.078, p = 0.109	0.091, p = 0.062	-0.4 > rho > -0.5
Gestational age, days	0.095, p = 0.081	-0.198, p < 0.001	-0.350, p < 0.001	0.597, p < 0.001	0.244, p < 0.001	-0.045, p = 0.413	-0.3 > rho > -0.4
Bone turnover biomarkers							-0.2 > rho > -0.3
PTH, pg/mL	-0.292, p < 0.001	-0.266, p < 0.001	-0.256, p < 0.001	-0.163, p = 0.001	-0.184, p < 0.00	–0.210, p < 0.001	-0.1 > rho > -0.2
BSAP, μg/mL	-0.155, p = 0.001	–0.253, p < 0.001	-0.287, p < 0.001	0.145, p = 0.003	0.021, p = 0.673	-0.067, p = 0.172	0.0 > rho > -0.1
Calcium, mmol/L	0.080, p = 0.098	0.234, p < 0.001	0.369, p < 0.001	–0.278, p < 0.001	-0.074, p = 0.131	0.182, p < 0.001	0.0 < rho < 0.1
Phosphate, U/L	0.097, p = 0.046	0.041, p = 0.403	0.050, p = 0.305	0.003, p = 0.946	-0.096, p = 0.050	-0.090, p = 0.066	0.1 < rho < 0.2
Lipid metabolism biomarkers							0.2 < rho < 0.3
LDL, mg/dL	0.049, p = 0.315	–0.174, p < 0.001	-0.300, p < 0.001	0.464, p < 0.001	0.193, p < 0.001	-0.004, p = 0.420	0.3 < rho < 0.4
HDL, mg/dL	0.228, p < 0.001	0.094, p = 0.057	0.031, p = 0.527	0.333, p < 0.001	0.178, p < 0.001	0.093, p = 0.059	0.4 < rho < 0.5
LDL/HDL ratio	-0.099, p = 0.044	–0.228, p < 0.001	-0.308, p < 0.001	0.232, p < 0.001	0.071, p = 0.153	-0.104, p = 0.036	0.5 < rho < 0.6
Adiponectin, μg/mL	0.063, p = 0.202	0.140, p = 0.005	0.168, p = 0.001	-0.123, p = 0.012	-0.045, p = 0.362	0.018, p = 0.718	
Thyroid gland-associated biomarkers							
Free T3, pg/mL	-0.114, p = 0.018	-0.144, p = 0.003	-0.120, p = 0.013	-0.040, p = 0.415	-0.088, p = 0.073	-0.070, p = 0.153	
Free T4, ng/dL	-0.095, p = 0.049	0.087, p = 0.074	0.216, p < 0.001	–0.413, p < 0.001	-0.203, p < 0.001	0.028, p = 0.571	
TSH, uU/mL	0.021, p = 0.670	0.000, p = 0.996	-0.005, p = 0.916	-0.020, p = 0.674	-0.087, p = 0.075	-0.095, p = 0.052	
Hematological biomarkers							
Hemoglobin, g/dL	0.063, p = 0.252	0.203, p < 0.001	0.298, p < 0.001	-0.226, p < 0.001	-0.008, p = 0.881	0.177, p = 0.001	
RBC count, 10 ⁶ /μL	0.016, p = 0.784	0.119, p = 0.042	0.208, p < 0.001	-0.215, p < 0.001	-0.056, p = 0.338	0.104, p = 0.074	
WBC count, 10 ³ /µL	0.051, p = 0.379	-0.062, p = 0.287	-0.098, p = 0.092	0.231, p < 0.001	0.088, p = 0.130	0.031, p = 0.600	
Platelet count, 10³/µL	-0.084, p = 0.148	-0.062, p = 0.295	-0.026, p = 0.654	-0.109, p = 0.060	–0.117, p = 0.045	-0.060, p = 0.310	
MCV, fL	0.116, p = 0.046	0.145, p = 0.013	0.146, p = 0.013	0.031, p = 0.598	0.084, p = 0.153	0.110, p = 0.060	
MCH, pg	0.101, p = 0.083	0.123, p = 0.035	0.110, p = 0.061	0.065, p = 0.263	0.122, p = 0.037	0.135, p = 0.021	
MCHC, g/dL	0.004, p = 0.940	0.004, p = 0.945	-0.027, p = 0.649	0.091, p = 0.118	0.134, p = 0.022	0.115, p = 0.049	
RDW, %	-0.095, p = 0.101	-0.102, p = 0.081	-0.094, p = 0.110	-0.073, p = 0.207	-0.109, p = 0.062	-0.129, p = 0.028	
Hematocrit, %	0.090, p = 0.120	0.228, p < 0.001	0.333, p < 0.001	–0.239, p < 0.001	-0.034, p = 0.562	0.158, p = 0.007	
Other biomarkers							
Urea, mg/dL	0.026, p = 0.597	0.161, p = 0.001	0.240, p < 0.001	-0.360, p < 0.001	–0.224, p < 0.001	-0.082, p = 0.098	
Albumin, g/dL	-0.056, p = 0.247	0.185, p < 0.001	0.377, p < 0.001	-0.533, p < 0.001	-0.272, p < 0.001	0.075, p = 0.126	
Vitamin B6, ng/mL	0.178, p < 0.001	0.367, p < 0.001	0.458, p < 0.001	–0.277, p < 0.001	-0.025, p = 0.623	0.164, p = 0.001	
Vitamin B12, pg/mL	0.130, p = 0.008	0.280, p < 0.001	0.363, p < 0.001	–0.246, p < 0.001	-0.069, p = 0.164	0.102, p = 0.038	//
Sodium, mmol/L	0.086, p = 0.081	0.081, p = 0.104	0.082, p = 0.098	0.043, p = 0.380	0.024, p = 0 629	0.032, p = 0.526	
Zinc, μmol/L	-0.020, p = 0.867	0.163, p = 0.171	0.275, p = 0.020	-0.396, p = 0.001	-0.297, p = 0.011	-0.001, p = 0.992	

Table 3. Interrelations between 25OHD and 1,25(OH) and $1,25(OH)_2D$ isoforms with age, gestational age, biochemical, hematological, and endocrinological serum parameters. The correlation strength is defined by Spearman's rank correlation coefficient rho (appears first in each cell). Abbreviations: B25OHD, bioavailable 25OHD; B1,25(OH)₂D, bioavaialble $1,25(OH)_2D$; BSAP, bone-specific alkaline phosphatase; F25OHD, free 25OHD; F1,25(OH)₂D, free $1,25(OH)_2D$; fT3, free triiodothyronine; fT4, free thyroxine; HDL, high-density lipoprotein cholesterol; LDL, low-density lipoprotein cholesterol; MCH, mean corpuscular hemoglobin; MCHC, mean corpuscular hemoglobin concentration; MCV, mean corpuscular volume of red blood cells; PTH, parathyroid hormone; RBC, red blood cell; RDW, red blood cell distribution width; T25OHD, total 25OHD; T1,25(OH)₂D; WBC, white blood cell.

With the exception of PTH, associations of 25(OH)D metabolites and $1,25(OH)_2D$ metabolites with typical clinical chemistry readouts goes in opposite directions.

Moreover, our study confirmed the finding of the study by Bouillon et al.¹² that serum total 25OHD doesn't markedly change during pregnancy, whereas serum total $1,25(OH)_2D$ increased significantly along with serum DBP, which in turn resulted in the increase in free vitamin D metabolites: free 25OHD was declined and free $1,25(OH)_2D$ was slightly elevated as total $1,25(OH)_2D$ was increased slightly more than DBP.

Free 250HD and bioavailable 250HD were found to be superior to traditional total 250HD. Our data suggest that in normal human pregnancy both free 250HD and bioavailable 250HD (the latter is tendentially slightly superior to free 250HD) appear to be more precise indicators of vitamin D status during pregnancy than routine total 250HD in terms of overall better correlations with gestational age, calcium, bone-specific alkaline phosphatase, adiponectin, hemoglobin and red blood cell count, vitamins B6 and B12 and other analytes (Table 3). In other words, our data are in favor of clinical relevance of free hormone hypothesis for 250HD.

To date, the data on changes in free/bioavailable 25OHD with total 25OHD throughout pregnancy were analyzed in nine studies^{9-11,13-18}, including our own^{10,11}. Overall, the data regarding the comparison of the diagnostic potential of free/bioavailable 25OHD versus total 25OHD, reported by these authors, are conflicting due to low sample sizes (varying between 10 and 60 participants), with the exception of two studies by Powe et al.¹⁸ and Gustafsson et al.¹⁷, which was based on 170 and 855 pregnant women, respectively. This discrepancy was discussed in details in our recent publications^{11,19}. This study by Powe et al.¹⁸ differs from ours: just three-fourth of the participating women were healthy and only around 60% were of White ethnicity. Although F25OHD was assessed by calculation method using vitamin D-binding protein (DBP) levels like in our study, DBP in this study was measured using a monoclonal antibody-based ELISA. This DBP measurement technique is considered unreliable, especially when having populations with different ethnic backgrounds due to DBP polymorphism²⁰⁻²³. Our study is larger, conducted in an ethnically homogenous cohort and by using the state-of-the-art way to measure DBP. We thus believe that our data are more relevant. Moreover, these results are in good agreement with the findings of our previous study¹¹, where we compared total 25OHD with directly measured free 25OHD.

Regarding the large longitudinal study by Gustafsson et al.¹⁷, since the authors did not present the data of bivariate correlation analysis (free 25OHD/total 25OHD versus PTH), a one-to-one comparison of our results with their results is impossible.

Neither free $1,25(OH)_2D$ nor bioavailable $1,25(OH)_2D$ showed superiority to total $1,25(OH)_2D$. In contrast to 25OHD—vitamin D's precursor and reservoir—our data do not support the free hormone hypothesis for $1,25(OH)_2D$ —vitamin D's biologically active metabolite. Indeed, with the exception of PTH, total $1,25(OH)_2D$ showed constantly stronger correlations with all analyzed serum biomarkers than free $1,25(OH)_2D$ and bioavailable $1,25(OH)_2D$ (Table 3).

According to the data of numerous cross-sectional and longitudinal studies, total $1,25(OH)_2D$ is known to rise up to threefold versus non-pregnant state²⁴. This is the result of the increased production of $1,25(OH)_2D$ by 25-hydroxyvitamin D-1 α -hydroxylase (CYP27B1) by not only maternal kidneys, but also, although to a lesser extent, by the fetoplacental unit (maternal decidual and fetal trophoblast tissues of the placenta and fetal kidney)^{24–26}. We likewise detected total $1,25(OH)_2D$ increase throughout gestation. Notably, free $1,25(OH)_2D$ but not bioavailable $1,25(OH)_2D$ was found to rise throughout pregnancy course. Three other studies^{3,17,27} likewise reported an increase in free $1,25(OH)_2D$ with pregnancy course.

Correlation with PTH stays apart: all six vitamin D isoforms showed nearly equal associations with PTH. PTH—a major secretory product of parathyroid gland and a second, along with $1,25(OH)_2D$, hormonal regulator of calcium homeostasis—was found to be the only biomarker which showed the same direction of correlation (negative correlation) and strength (weak but still significant negative correlation) with all six analysed vitamin D metabolites. The most likely explanation of this finding could be that parathyroid gland, along with kidney and placenta, are exceptional organs, which have megalin/cubilin-mediated system of $25OHD/1,25(OH)_2D$ uptake from 25OHD-DBP and $1,25(OH)_2D-DBP$ complexes^{28,29} and thus the cells of parathyroid gland are influenced by all circulating isoforms of $25OHD/1,25(OH)_2D$ at nearly the equal extent. This mechanism is absent in most of the other cell types and organs, leaving for those cells only the option of the freely circulating $25OHD/1,25(OH)_2D$ uptake via passive diffusion through plasma membranes.

25OHD and 1,25(OH)₂D correlate in opposite directions with biochemical and endocrinological biomarkers. The direction of correlation (i.e., positive vs negative) between 25OHD metabolites on one hand, and 1,25(OH)₂D metabolites on the other hand, with other serum biomarkers was found to be opposite. The most prominent examples are the differences in the correlation directions with such biomarkers as LDL, free thyroxin, urea, vitamins B6, B12, and zinc (Table 3). This emphasizes the different biological roles of total 1,25(OH)₂D-vitamin D's fully active isoform, which (total 1,25(OH)₂D)-is present in the circulation in a picomolar range and have a half-life of 5-80 h³⁰ and total 25OHD—vitamin D's precursor, which is present in the circulation in nearly thousand-fold higher concentration range and have a half-live of 10-40 days³⁰ and which is well known to serve as a reservoir/storage fraction of vitamin D, responsible for balancing/minimization of short-time (intraday) vitamin D isoform fluctuations. The 10-100-fold lower affinity between DBP and free 1,25(OH)₂D compared to the affinity between DBP and free 25OHD (to the lesser extent this is valid likewise for albumin) as well as much higher (up to 100-fold) affinity of nuclear VDR for free 1,25(OH)₂D than for free 25OHD are factors influencing concentrations of the vitamin D metabolites at the nuclear receptor. Whether or not the strength of the binding of either free $1,25(OH)_2D$ or free 25OHD to the nuclear receptor is affecting gene expression in an opposite manner is unknown so far. It is conceivable that hormone receptor ligands with low binding affinity for the receptor may act as partial receptor antagonists, whereas ligands with high affinity fully stimulate the receptor. However, this needs to be proven in adequately designed basic science experiments. In any case, this is the first study reporting differences in correlation strength between $250HD/1,25(OH)_2D$ isoforms versus key clinical parameters. This for sure needs to be repeated in independent cohorts and the underlying molecular mechanisms (see our hypothesis above) need to be explored in future studies.

Study limitations. The data on free and bioavailable 25OHD and $1,25(OH)_2D$ were based on calculations using the affinity constants between 25OHD/1,25(OH)_2D and DBP and 25OHD/1,25(OH)_2D and albumin. These affinity constants were measured experimentally in serum samples of non-pregnant individuals and has not been verified in pregnant women so far. However, the possible inaccuracies (a systematic shift in calculated free/bioavailable 25OHD/1,25(OH)_2D absolute levels *versus* their real values) are not influencing the key findings of our study, because our conclusions depend on the relative changes of free, bioavailable and total 25OHD and 1,25(OH)_2D, respectively rather than absolute changes of these parameters.

It is very well known that during pregnancy, especially in the 3rd trimester, the blood volume of the pregnant women increases and thus the blood is diluted to a certain extent³¹. However, the influence of hemodilution as a confounding factor in bivariate associations presented in this study will not change the correlation direction. Hemodilution might have just flatten the correlation strength (as presented by Spearman's rho values), but it will not reverse the direction of correlation.

The accuracy of total 25OHD levels produced by fully-automated commercial immunoassays, especially in the conditions of pregnancy-induced increased DBP levels, is a well-known issue³². However, we used the total 25OHD kit with the best diagnostic precision characteristics in pregnancy among the other tested commercial kits when comparing to total 25OHD concentrations measured by mass-spectrometry—a technique considered the gold standard for the measurement of 25OHD³². Undoubtedly, the usage of LC–MS/MS might have further improved the data quality.



Figure 1. Key findings of our study are: (i) Both free 25(OH)D and bioavailable 25(OH)D showed better correlations with gestational age, bone and lipid metabolism biomarkers than total 25(OH)D. (ii) Total $1,25(OH)_2D$ is superior to free $1,25(OH)_2D$ and bioavailable $1,25(OH)_2D$. (iii) 25(OH)D metabolites and $1,25(OH)_2D$ metabolites correlated with gestational age, bone and lipid biomarkers in opposite directions.

DBP, being a highly polymorphic protein, was reported to have genotype differences (up to threefold) in affinity constants with 25OHD³³. Although these data were not confirmed by three other studies³⁴⁻³⁶, we are on the safe side with our key conclusions, because we used quite a monogenic (Caucasian) cohort and we measured DBP with a polyclonal-antibody-based ELISA kit. Finally, we conducted a single-centered study and thus single-centered effects cannot be excluded.

Conclusions. The present study compared the performance of the six vitamin D compounds—free, bioavailable, and total 25-hydroxyvitamin D (free 25OHD, bioavailable 25OHD, total 25OHD) as well as free, bioavailable, and total 1,25-dihydroxyvitamin D (free 1,25(OH)₂D, bioavailable 1,25(OH)₂D, total 1,25(OH)₂D). Among the 1,25-dihydroxyvitamin D metabolites, total 1,25(OH)₂D clearly showed the best correlation with components of the endocrine vitamin D system and/or potential cellular targets of activation of the vitamin D receptor. When comparing the three 25OHD metabolites, both free 25OHD and bioavailable 25OHD showed an overall good correlation with the analyzed clinical laboratory parameters, bioavailable 25OHD might perform even better then free 25OHD. The only exception is PTH. It correlates well with total 25OHD most likely because the parathyroid cells have a highly specific uptake mechanism for total 25OHD—the megalin/cubilin-uptake system. This specific system acts much faster as compared to the uptake via the cell membrane. With the exception of PTH, correlations of 25(OH)D and 1,25(OH)₂D metabolites with typical clinical chemistry readouts go in opposite directions suggesting potentially opposite effects of either 25(OH)D or 1,25(OH)₂D on the nuclear vitamin D receptor in some target cells of vitamin D (see also Fig. 1 summarizing the key findings of our study). This hypothesis based on clinical data needs to be confirmed in adequately designed basic science studies.

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Author contributions

All the authors have accepted responsibility for the entire content of this submitted manuscript and approved submission. BH designed the study. O.T., B.K.K. and B.H. wrote the manuscript. B.H. and O.T. did statistics, O.T., C.C., C.B. and S.E. collected the data.

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Additional information

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