



# Sixty Years (1957–2017) of Research on Toxoplasmosis in China—An Overview

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Toxoplasma gondii is a ubiquitous zoonotic pathogen belonging to apicomplexan parasites. Infection in humans and animals may cause abortion and other severe symptoms under certain circumstances, leading to great economical losses and public health problems. T. gondii was first discovered in China in 1955 and the corresponding work was published in 1957. Since then, a lot of work has been done on this parasite and the diseases it causes. This review summarizes the major progress made by Chinese scientists over the last 60 years, and gives our perspectives on what should be done in the near future. A wide variety of diagnostic approaches were designed, including the ones to detect T. gondii specific antibodies in host sera, and T. gondii specific antigens or DNA in tissue and environmental samples. Further work will be needed to translate some of the laboratory assays into reliable products for clinic uses. Epidemiological studies were extensively done in China and the sero-prevalence in humans increased over the years, but is still below the world average, likely due to the unique eating and cooking habits. Infection rates were shown to be fairly high in meat producing animals such as, pigs, sheep, and chickens, as well as in the definitive host cats. Numerous subunit vaccines in the form of recombinant proteins or DNA vaccines were developed, but none of them is satisfactory in the current form. Live attenuated parasites using genetically modified strains may be a better option for vaccine design. The strains isolated from China are dominated by the ToxoDB #9 genotype, but it likely contains multiple subtypes since different ToxoDB #9 strains exhibited phenotypic differences. Further studies are needed to understand the general biology, as well as the unique features of strains prevalent in China.

Keywords: Toxoplasma gondii, epidemiology, China, vaccine, genotype

# **INTRODUCTION**

*Toxoplasma gondii* is an obligate intracellular parasitic protozoan widely distributed around the world. One-third of the world's population was estimated to be infected by this parasite (Dubey, 2010). In addition to humans, it can also infect more than 200 species of animals and causes toxoplasmosis in them (Gao, 2014). As an opportunistic pathogen, *T. gondii* infection rarely causes severe symptoms in healthy humans and most other hosts. The main risks are congenital infections during pregnancy, which may cause abortion, stillbirth, developmental defects, and other serious

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#### Specialty section:

This article was submitted to Infectious Diseases, a section of the journal Frontiers in Microbiology

Received: 23 June 2017 Accepted: 06 September 2017 Published: 25 September 2017

#### Citation:

Pan M, Lyu C, Zhao J and Shen B (2017) Sixty Years (1957–2017) of Research on Toxoplasmosis in China—An Overview. Front. Microbiol. 8:1825. doi: 10.3389/fmicb.2017.01825

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diseases to the fetus. Congenital infections and subsequent complications are common in humans and farm animals such as, pigs and sheep, leading to great economical losses and social problems (Dubey, 2009).

One important reason for the wide spread of *T. gondii* is its complex life cycle, which consists a sexual phase and an asexual phase. Sexual reproduction occurs in definitive hosts, the felids. After infection, they shed oocysts in feces to contaminate water and environment and pass the infection to other hosts if oocysts are ingested. In intermediate hosts, the parasites propagate asexually and they can be transmitted between intermediate hosts through predation. A large portion of human toxoplasmosis cases were thought to be derived from consumption of undercooked meat that was infected (Montoya and Liesenfeld, 2004).

China has a long history of T. gondii research, which dates back to 1955 when Yu et al first isolated T. gondii from cats and rabbits in Fujian Province. This work was published in the Chinese journal Acta Microbiologica Sinica in 1957 (Yu et al., 1957). After this discovery, more and more work has been done to understand the epidemiology and biology of this parasite in China. When the key words "Toxoplasma" (or "toxoplasmosis") and "China" were used to search the PubMed database (English articles) and the China National Knowledge Infrastructure (CNKI) database (Chinese articles), we estimated the number of publications on Toxoplasma or toxoplasmosis in each year from the Chinese research community and the results are summarized in Figure 1. There were only sporadic reports published before 1978, but since then the research activities became more active (Figure 1). There are at least two reasons for that: first, during late 1970s, many pigs in Shanghai and the nearby area were reported to have a fatal syndrome called "high fever with unknown reason," and later it was shown to be caused by T. gondii infection (Ren and Bao, 1982), which promoted studies on this parasite; second, the implementation of the Economic Reform and Open up policies in 1978 in general boosted scientific researches. In this review, we collected all the studies published in English and Chinese journals and summarized the work that have been done on *Toxoplasma* and toxoplasmosis in China over the last 60 years. A perspective was also outlined for areas of research in the future.

# DEVELOPMENT OF DIAGNOSTIC APPROACHES

Because of the lack of unique symptoms or traits, diagnosis of toxoplasmosis is challenging. This is particularly true for human infections during pregnancy, since the risks of which depend on when exactly the infection is acquired. Most European and North American countries have reference laboratories for the diagnosis and risk assessments. This concept has been introduced into China, but it never really works the way as it should, since there is no bona fide reference labs operating at the national level. Nonetheless, over the last 40 years, Chinese scientists have done a lot of work to develop diagnostic strategies to detect *T. gondii* infections. They can be mainly grouped into three types: serological tests detecting *T. gondii* specific antibodies

post infection, immunological methods detecting *T. gondii* specific antigen, and nucleic acids based pathogen detection. Representative methods for each type were listed in **Table 1**.

A large collection of serological tests have been designed, using a wide variety of antigens. Most often, these tests were developed to detect T. gondii specific IgM or IgG antibodies in the sera of humans and animals. Although different formats of serological tests were designed, enzyme-linked immunosorbent assays (ELISA) and agglutination tests were the most common. In general, ELISA tests are more sensitive than agglutination tests, but they often require a species specific secondary antibody that recognize the IgM or IgG antibody of the animals to be tested. Agglutination tests are superior to ELISA in that they do not need any secondary antibodies, therefore are suitable for on-site diagnosis. Among the different ELISA formats, indirect ELISAs with native or recombinant T. gondii antigens to catch specific antibodies from infected hosts are the most common (Wu et al., 2009; Dai et al., 2013; Wang Z. et al., 2014). Many different antigens have been tried for this purpose, which included native antigens such as, the whole tachyzoite soluble extract and excretory-secretory antigens, as well as recombinant antigens such as, surface proteins (SAG1, SAG2, etc.), secretory proteins from different organelles (MIC3, GRA1, GRA7, etc.), and others (listed in Table 1 with corresponding references). In addition to single antigens, chimeric antigens containing epitopes from multiple proteins have also been tried and thought to be superior to single antigens (Dai et al., 2013; Wang Z. et al., 2014; Feng et al., 2016). To bypass the requirement of species specific secondary antibodies in indirect ELISAs, protein A or protein G from Staphylococcal aureus were used to functionally replace those secondary antibodies, due to their high binding affinities to immunoglobulins from a wide range of animals. Such ELISAs were called AG-ELISAs (Zhang et al., 2010). Besides indirect ELISAs, other formats of ELISAs, such as, double-sandwich ELISA were also developed (Lu et al., 2006). Although there was no solid and direct comparison between these different types of ELISA tests, from literature it seems like SAG and GRA7 based indirect ELISAs are the most reliable. The sensitivity and specificity were claimed to be nearly 100% and above 96%, respectively (Lu et al., 2006; Sun et al., 2015).

Like ELISAs, agglutination tests were also designed in a number of different ways (Table 1). Traditional agglutination tests (called modified agglutination tests) use fixed tachyzoites as agglutogen, which are widely used but difficult to standardize, due to the variations during tachyzoites preparation (Al-Adhami et al., 2016; Feng et al., 2016). To improve, different recombinant antigens derived from T. gondii were used to coat and sensitize red blood cells or latex particles, which were then used as agglutogen. Such testes are called indirect hemagglutination assays (IHA) and latex agglutination tests (LAT), respectively (Feng et al., 2016). The antigens used to sensitize agglutination particles were more or less the same as the ones used in indirect ELISAs (Chen et al., 1980; Yu, 2001; Jiang et al., 2008). In addition to agglutination tests and ELISAs, there are other serological methods reported, such as, indirect fluorescent assays (IFA) (Wang Y. B. et al., 2012). More recently, rapid diagnostic

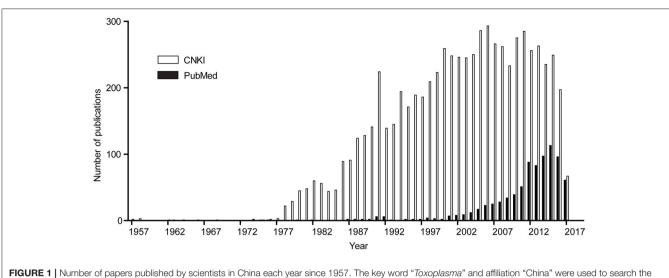


FIGURE 1 Number of papers published by scientists in China each year since 1957. The key word "*Toxoplasma*" and affiliation "China" were used to search the PubMed database (https://www.ncbi.nlm.nih.gov/pubmed/) for papers published by scientists in China in international journals. The keywords "*Toxoplasma*" or "toxoplasmosis" or the Chinese equivalent of the two were used to search the Chinese library CNKI (http://www.cnki.net/) to estimate the papers in Chinese journals. Subsequently the numbers of papers from the two searches were reported by year.

#### TABLE 1 | Diagnostic methods developed by scientists in China.

Detecting	Methods	Sensitivity and specificity (host)	References
<i>T. gondii</i> specific Antibodies	Indirect-ELISA, using: Truncated recombinant SAG1	93.9% (31/33) positive concordance compared with Western blot (100%) (human)	Wu et al., 2009
	Multi-epitope peptide derived from SAG1, SAG2 and SAG3	The total concordance was 93.2 and 95.7% for the detection of IgG and IgM antibodies, respectively, compared with commercial ELISA tests (human)	Dai et al., 2013
	GRA1, GRA7 MIC3	Highly sensitive (93.2%) and specific (94.0%) (dog) Higher positive detections than MAT in pigs and goats	Wang Z. et al., 2014 Zhang et al., 2010
	Dynamic flow immunochromatographic Test (DFICT) using SAG1 and SAG2	The lowest detectable limit was 1:320 dilution of positive serum (dog and cat)	Jiang W. et al., 2015
	Latex agglutination tests (LAT) using MIC3	After 8 days infection, <i>T. gondii</i> specific antibodies could be detected (pig)	Jiang et al., 2008
	Indirect hemagglutination assays (IHA) using soluble parasite antigen sensitized sheep red blood cells	<i>T. gondii</i> specific antibodies could be detected on the 7th days after initial infection (rabbit)	Chen et al., 1980
	Piezoelectric immunoagglutination assay using total antigen of the RH strain	The detectable limit was 1:5,500 dilution of positive serum (rabbit)	Wang et al., 2004
	Fast dipstick dye immunoassay (DDIA) using total soluble antigen of the RH strain	Sensitivity and specificity of IgG were 100 and 96%, respectively, while sensitivity and specificity of IgM were 100 and 94%, respectively (human)	Jin et al., 2005
<i>T. gondii</i> Antige <b>n</b> s	Modified rapid sandwich ELISA using rabbit polyclonal antibodies against soluble <i>T. gondii</i> antigens	Detected circulating antigens at the concentration of 31.2 ng/mL, and no cross-reaction with other protozoa (human and rabbit)	Chen R. et al., 2008
	Immunochromatographic strip using sheep antisera against excretory/secretory antigens of <i>T. gondii</i> circulating antigens	Circulating antigens could be detected from day 2 to day 14 (or even longer) after parasite infection (pig, goat, and sheep)	Wang et al., 2011
T. gondii DNA	<b>LAMP</b> targeting 529 bp repeat sequence, SAG1 gene or B1 gene	Superior sensitivity than the conventional PCR, can detected <i>T. gondii</i> at the concentration of 2–3 parasites/mL, highly specific (pig, sheep, mouse, rabbit)	Yang et al., 2008; Lin et al., 2012; Lai et al., 2014
	RT-LAMP targeting 18s RNA gene	Showed higher sensitivity than RT-PCR with the detection limit of 1 tachyzoite in 1 g pork (pig)	Qu et al., 2013b
	<b>RT-PCR</b> targeting 529 bp repeat sequence	The sensitivity was as higher (1 fg DNA), compared to conventional PCR (100 fg) (pig, sheep, cattle, and dog)	Lin et al., 2011

methods using immune colloidal gold technique were developed (Feng et al., 2016).

Although numerous T. gondii antibody detecting methods were designed in the lab, the majority of them were not subject to extensive evaluation to determine their value of clinic use. This is particularly the case for the assays to detect animal toxoplasmosis. Because of the high demand for TORCH (Toxoplasma, Rubella virus, Cytomegalovirus, and Herpes simplex virus) screens, China Food and Drug Administration (CFDA) issued certificates to a number of manufactures to make Toxoplasma specific IgM and IgG detecting kits for the diagnosis of human toxoplasmosis. However, published work assessing their sensitivity, specificity and Youden index indicated that, except for a few, the majority of these domestic kits were not of high enough quality to be useful in clinic (He et al., 2008). Especially for IgM detecting kits, the Youden index for many of them were close to zero (Jiang et al., 2003), therefore were of no value for clinic use. For the diagnosis of animal toxoplasmosis, so far there is no method certified by the China Institute of Veterinary Drug Control. Therefore, efforts are still needed to translate some of the laboratory assays to reliable and commercially available products for the diagnosis of toxoplasmosis in China. For the same reason, the sero-prevalence results discussed below only represent rough estimates of T. gondii infections in China.

In addition to the antibody detecting methods, there were also reports using immunological methods to detect *T. gondii* antigens in sera or other tissue extracts. For instance, a rapid modified ELISA was developed to test *T. gondii* antigens in humans, which included 5,428 sera, 548 cerebrospinal fluid and two breast milk samples. The lowest concentration of antigen detected was as low as 31.2 ng/mL without cross-reaction with antigens from other protozoa, trematodes, or nematodes (Chen R. et al., 2008). It is worth noting that detecting *T. gondii* antigens by immunological methods has limited success, due to the low levels of antigens exist and limited sensitivity of the tests.

To increase the sensitivity of pathogen detection, molecular biology methods based on nucleic acids detection were developed in recent years. Most of them were based on specific amplification of *T. gondii* DNA sequences, which include: conventional PCR, loop-mediated isothermal amplification (LAMP), nested-PCR, and real-time PCR (RT-PCR) etc. (Lin et al., 2012; Qu et al., 2013b). The target sequences for amplification often include the 529 bp repeat fragment, ITS-1, B1 gene, etc. Among these tests, LAMP is the most sensitive and does not require expensive equipments (Yang et al., 2008; Lin et al., 2012; Lai et al., 2014), therefore is suitable for on-site uses. However, it can be contaminated easily and give high rates of false positive reports.

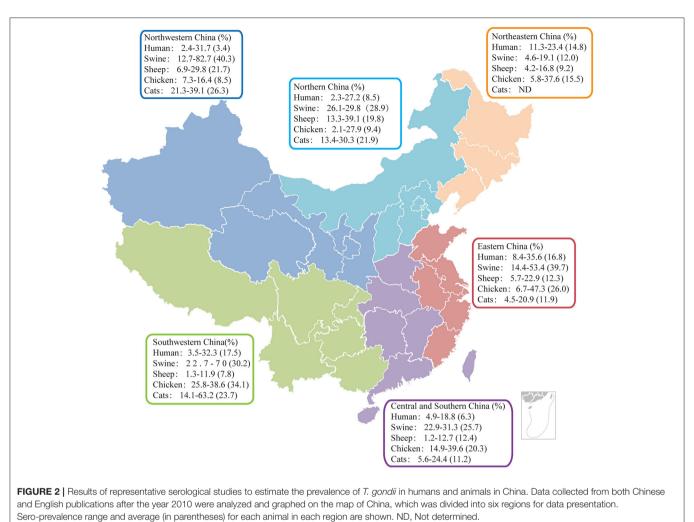
### SERO-PREVALENCE STUDIES IN HUMANS

As an opportunistic pathogen, *T. gondii* posts health threats to the public, particularly to individuals with reduced immune functions. China has a large susceptible population: the number of AIDS patients increased from 650,000 in 2005 to 780,000 in 2011 (Wu, 2011; Ding et al., 2017), the number of women

at childbearing age was estimated to be around 375.8 million in 2013 (Ding et al., 2017). Over the last a few decades, epidemiological surveys have been frequently done to monitor the prevalence of *T. gondii* in China. The average infection rate in humans is close to 10% but it varies significantly between different geographic regions and different populations (Xiao et al., 2010; Li et al., 2011; Sun et al., 2013; Cong et al., 2015a; Li H. L. et al., 2015; Wang L. et al., 2015; Wang S. et al., 2015), etc. Some of the representative results are shown in **Figure 2**. It is worth noting that the sero-prevalence of human toxoplasmosis in China is lower than the world average (10 vs. 25–30%), which is likely due to the unique eating habits (boil before eating/drinking) of Chinese people (Howe and Sibley, 1995; Yu, 2000).

Years of sero-prevalence studies seem to suggest that T. gondii infection rates in humans increased over the last 30 years in China. For example, the overall infection rate was 5.2% in the first national survey implemented between 1988 and 1992 (Yu et al., 1994), then rose to about 8% between 2001 and 2004 (Zhou et al., 2008). Possible reasons for such change include increased meat consumption and pet cat numbers as a consequence of economy development. At present, it is estimated that 10% of humans in China are infected, but the distribution is not even. For example, due to the unique culture and eating habits, people from certain ethnic groups have significant higher infection rates than others. Thirty percent of the Bai people in Yunnan province was seropositive, compared to 10% in Han people in the same region, which is likely due to the raw pork eating culture of the Bai people (Li H. L. et al., 2015). For the same reason, the prevalences in Miao (25.4%), Buyi (25.3%), and Mongol (17.1%) ethnic groups are also higher than other groups (Xu et al., 2005). In addition, people with certain careers are also at higher risk, herdsmen, veterinary, meat processing workers, and slaughterhouse workers tend to be more frequently infected than general public (Chen et al., 2005; Huo et al., 2007; Yu et al., 2007).

Serological tests targeting special groups of people were also frequently done, the best known of which is the TORCH examination in pregnant women. TORCH test is highly recommended by many hospitals, and is mandatory in some cities in China. It should be pointed out that the Toxoplasma examination part of TORCH is over simplified, only testing the presence of T. gondii IgM and IgG antibodies in a single test to inform the infection status. Since IgM may last more than a year, IgM positive women do not necessarily acquire the infection during pregnancy (Feng et al., 2016). As such, medical decisions made from a single IgM/IgG test are not always appropriate. Nevertheless, routine TORCH screens found that on average 0.3% of pregnant women were IgM positive, and 5.3% were IgG positive for T. gondii (Ding et al., 2009, 2015; Yang et al., 2014; Liu T. et al., 2017). A certain portion (although the exact number is not clear) of the IgM positive women were predicted to be infected after conception, which is high likely to lead to poor obstetric outcomes, or increase the risk congenital toxoplasmosis to newborns (Zhou et al., 2011a; Peng et al., 2015; Guan, 2017). Surveys also suggested that seroprevalence of T. gondii in infertile couples is higher than that in healthy couples (Zhou et al., 2002), although the relationship



between *T. gondii* infection and infertility is currently unknown. **Figure 2**, which provide a rough estimate for the current status

Sero-prevalence studies in hospitalized patients suggested that infection rates in AIDS patients, cancer patients, and patients in intensive care units were almost two-fold of that of health people (Xiao et al., 2010; Jiang C. et al., 2015; Zhang et al., 2015). The exact reasons for the higher sero-prevalence in such patients are currently unknown, but these results imply that patients with compromised immune functions may pick up infections and sero-convert more frequently. A recent meta-analysis on AIDS patients worldwide also showed the same trend (Liu L. et al., 2017; Liu T. et al., 2017).

# ANIMAL TOXOPLASMOSIS

China has one of the highest biodiversities in the world, with a wide variety of landscapes and animal populations. Using mostly indirect ELISAs and agglutination tests, sero-prevalence of *T. gondii* in farm and wild animals was estimated over the years. In general, the prevalence in animals was much higher than that in humans. Representative results from epidemiological studies performed after the year 2010 were summarized in of animal toxoplasmosis in China. Among the major animals monitored, pigs were the most frequently infected by T. gondii. Roughly 30-50% of pigs raised in China are sero-positive for T. gondii (Yu et al., 2011; Wu et al., 2012; Xu Y. et al., 2014), and this number can reach 70% in some areas and farms (Li Y. N. et al., 2015). Due to the facts that China is the biggest pig-raising country (more than one billion pigs were raised and butchered each year) and porkconsuming country in the world, high T. gondii prevalence in pigs lead to serious economic lossess and public health concerns. First of all, As mentioned above, pigs are susceptible to T. gondii infection. In late 1970s and early 1980s, the "high fever with unknown reason" syndrome caused by T. gondii killed a large number of pigs in Yangtze River delta (Shanghai, Jiangsu, and Zhejiang) (Ren and Bao, 1982). Even nowadays, abortions caused by congenital swine toxoplasmosis are still very common, leading to huge economic losses (Fan, 2017; Wu, 2017). In addition, high T. gondii prevalence in pigs also increases the risks of human infection through meat consumption (Wang H. et al., 2012), since pork is the main meat Chinese people consume.

Among other meat producing animals, sheep and chickens are at the top of the list in terms of T. gondii infection rates. Northern and northwestern parts of China are the main regions for sheep production. Sero-prevalence in sheep is also the highest in these regions, reaching 20% (Figure 2), likely as a consequence of the widespread of T. gondii oocysts in the environment (Yang et al., 2017). In other regions, infection rates in sheep are about 10% on average (Wu et al., 2011a; Xu P. et al., 2014; Zou et al., 2015; Li et al., 2016; Zhang N. et al., 2016). Given the fact that among all farm animals, sheep are probably the most susceptible to T. gondii infection, economic losses caused by abortions and other diseases from sheep toxoplasmosis are predicted to be huge, but there is no reliable estimate so far. In addition, lamb meat is the major meat used in barbecue in China. Because of the unique processing style, barbecue often does not kill all pathogens in the meat. As such, high prevalence of T. gondii in sheep may make sheep an important source for human infections. Interestingly, T. gondii is also fairly prevalent in chickens, particularly in the south of China, over 20% of chickens are sero-positive (Zhao et al., 2012; Liu R. Z. et al., 2013). Beef is another major meat people consumed in China, but the sero-prevalence of T. gondii in cattle (below 10% on average) is significantly lower than that in pigs or chickens (Yu et al., 2006; Zhu et al., 2017). In addition, cattle are generally fairly resistant to *T. gondii* infection. Therefore, beef is probably not a significant source of human infections.

Cats, as definitive hosts for T. gondii, play a critical role in its transmission. Pet cats in particular, are important sources for human infections due to their intimate association with humans. Sero-prevalence studies, using mostly ELISAs and agglutination tests, suggested that cats were also infected with T. gondii with relatively high frequencies. The infection rates vary significantly, depending on the regions and cat types. In general, stray cats were much more frequently infected than pet cats (Ding et al., 2017). In one study, 43 stray cats in Shanghai examined by IHA were found to be infected at 100% (Chen, 2010). Judging from reported results, the average sero-positivity of housed cats were somewhere between 15 and 25% (Zhang et al., 2009; Wu et al., 2011c; Ding et al., 2017). But it should be pointed out that, unlike animals (pigs, chickens, etc.) that are concentrated in farms, cats are more randomly distributed and sero-prevalence studies in cats from selected regions may not reflect the overall picture. In our opinion, the sero-positivity was likely underestimated, due to the fact that most of the studies did not include samples from rural areas, where the prevalence is predicted to be higher than urban regions. There were also sporadic reports on the sero-prevalence of T. gondii in pet dogs. Generally speaking, infection in pet dogs is common but is not as frequent as in pet cats, and is around 10% on average (Wu et al., 2011b; Yang et al., 2013; Gao Y. M. et al., 2016).

There are also studies reporting sero-prevalence of *T. gondii* in wild animals such as, rodents, bats, and gulls etc. (Yuan et al., 2013a; Zhang Y. et al., 2013; Miao et al., 2014). However, these studies are often regional and static. Nonetheless, these sporadic studies suggested that wild animals were frequently infected. For example, sero-prevalence in rodents like *Microtus* 

*fortis* was as high as 50% in Jilin province (Zhang Y. et al., 2013), and is 29% in Hunan province (Zhang et al., 2004). Similarly, infection rates in bats and black-headed gulls are around 20% (Yuan et al., 2013a; Miao et al., 2014). These results, along with the epidemiological studies in farm animals and pets, indicated that *T. gondii* is present in the environments and food chain in China, at similar frequencies as in other parts of the world (Dos Santos et al., 2010; De Berardinis et al., 2017). However, human infection rates in China are lower than the world average, which likely ascribe to the unique eating habits of the Chinese people.

# T. GONDII IN ENVIRONMENTS

High prevalence of T. gondii in animals is correlated with the wide spreading of parasites in environments, most likely in the form of oocysts. Indeed, when nucleic acids based detection methods were used to assess the presence of T. gondii DNA, environmental samples were examined as positive at fairly high frequencies. In 2012, two studies looked at the presence of T. gondii DNA in soil samples collected from pig farms and public parks in central China by PCR and LAMP, and found T. gondii contamination in 16-23% of the samples from parks, and 21-38% of the samples from pig farms, depending the methods used (Du et al., 2012a,b). Similarly, more than 9,000 soil samples collected from northeastern China were analyzed by real-time quantitative PCR and found over 30% of them to be positive (Gao X. et al., 2016). Presumably the detected T. gondii specific DNA was derived from oocysts, however it remains unknown how often viable oocysts can be isolated from such environmental samples. Given the environmental resistance of oocysts, a significant portion of the DNA positive samples were likely to contain viable oocysts, which may explain the high infection rates in animals. On the other hand, the PCR based methods to detect oocysts in environments are very sensitive to contaminations, extra caution needs to be taken in interpreting such data. Alternative methods such as, oocyst enrichment and isolation are necessary to estimate the spread of oocysts in environmental samples.

# GENOTYPES OF *T. GONDII* STRAINS CIRCULATING IN CHINA

Numerous efforts were taken to understand the genotypes and population structures of the *T. gondii* strains prevalent in China. Generally, two methods were used for this purpose. Either isolated live strains and genotyped them, or extracted DNA from infected tissues and then used the DNA samples for genotyping, with the latter being much more common. Either way, the isolates were often genotyped by PCR-RFLP based on 11 genetic markers (SAG1, 5'-SAG2 and 3'-SAG2, alternative SAG2, SAG3, BTUB, GRA6, c22-8, c29-2, L358, PK1, and an apicoplast locus Apico; Su et al., 2006). Consistent with the large territory and great biodiversities of hosts in China, many different genotypes were discovered, which were summarized in **Table 2**. It is obvious that the dominating genotype in China is ToxoDB #9 (also

#### **TABLE 2** | Genotypes of *T. gondii* parasites prevalent in China.

Host	Tissue origin	Type (Number)	Frequency	References
Cancer patients	Diseased tissues	ToxoDB #9 (8) ToxoDB #10 (9)	ToxoDB #9: 65%	Cong et al., 2015b
	Serum samples	ToxoDB #9 (9)	ToxoDB #10: 35%	Wang L. et al., 2015
Pigs	Hilar lymph nodes	ToxoDB #10 (5 ToxoDB #3 (1) ToxoDB #9 (21)	ToxoDB #9: 83%	Zhou et al., 2010; Zhou et al., 2011b; Jiang H. H. et al., 2013 Jiang W. et al., 2013; Jiang et al., 2016
	Retail meat	ToxoDB #9 (1) ToxoDB #213 (1)	ToxoDB #10: 11% ToxoDB #3: 4% Others: 2%	Wang H. et al., 2012
	Blood, heart and brain	ToxoDB #9 (3) ToxoDB #3 (1) ToxoDB #9 (13)		Li Y. N. et al., 2015; Wang D. et al., 2016
Cats	Brain, tongue, Heart, liver, Blood, feces	ToxoDB #9 (29) ToxoDB #225 (2) ToxoDB #1 (4) ToxoDB #2 (1) ToxoDB #3 (1) ToxoDB #20 (1) ToxoDB #17 (1)	ToxoDB #9: 74% ToxoDB #1: 10% ToxoDB #225: 5% Others: 11%	Qian et al., 2012; Liu R. Z. et al., 2013; Liu T. et al., 2013; Wang et al., 2013; Li et al., 2014; Tian et al., 2014; Li Y. N. et al., 2015; Yang et al., 2015; Cong et al., 2016a
RODENTS				
Microtus fortis	Lung	ToxoDB #10 (2) ToxoDB #9 (2)	ToxoDB #9: 53%	Zhang et al., 2014
Qinghai vole Plateau pika Tibetan ground-tit	Brain	ToxoDB #10 (4) New genotype (2)	ToxoDB #10: 35% Others: 12%	Zhang X. X. et al., 2013
Rats and mice	Brain	ToxoDB#9 (7)		Yan et al., 2014
OTHERS				
Wild birds	Breast, muscle	ToxoDB #10 (1) ToxoDB #1 (2)		Huang et al., 2012
Pet birds	Brain	ToxoDB #3 (1)		Cong et al., 2014
	heart	ToxoDB#9 (1) ToxoDB#2 (1) ToxoDB#10 (1)		Chen et al., 2015
House sparrows	Heart, brain, lung	ToxoDB #3 (1) New genotype (1)	ToxoDB #9: 44%	Cong et al., 2013
Domestic rabbit	Brain, spleen, liver	ToxoDB #2 (1)	ToxoDB #10: 37%	Zhou et al., 2013
Bats	Lung, heart, liver, spleen, stomach, intestine or kidney	ToxoDB #10 (4) ToxoDB #9 (1)	ToxoDB #3: 7% ToxoDB #2: 5% Others: 7%	Jiang et al., 2014
	liver	ToxoDB#10 (3) ToxoDB#9 (5)		Qin et al., 2014
Black goats	Liver, lung, lymph nodes	ToxoDB#10 (7) ToxoDB#9 (1)		Miao et al., 2015
Sika deer	Liver, lung, muscle	ToxoDB#9 (6)		Cong et al., 2016b
Farmed minks	Brain	ToxoDB#9 (5) ToxoDB#3 (1)		Zheng et al., 2016

called Chinese I or China I), moren than half of the isolates belong to this genotype (Li Y. N. et al., 2015). ToxoDB #9 is the dominating strain type in almost all examined hosts (**Table 2**). The next common one is ToxoDB #10 (Type I), which is the classic type 1 genotype (Zhou et al., 2009; Wang et al., 2013). However, it should be pointed out that most studies only reported the detection of DNA corresponding to ToxoDB #10 strains, only a few live type I parasites (TgHuZS2 and TgCtxz1) were isolated and preserved (Wang et al., 2013). Therefore, the actual frequency of such strains in China remain to be determined. ToxoDB #9 and #10 strains probably account for 90% of the strains examined in China. This situation is similar to North

America and Europe in that a few genotypes dominated the population (Minot et al., 2012). However, unlike the clonal nature of the type I, II, and III strains in north America and Europe, there are probably multiple subtypes in the ToxoDB #9 group, since different ToxoDB #9 isolates have quite different properties (such as, acute virulence in mice). Using multilocus sequence typing, a most recent study suggested that ToxoDB #9 isolates could be divided into at least two (potentially more) groups: Chinese I and Chinese III, with Chinese III being more closely related to classic type 1 strains (Gao et al., 2017). In addition, the dominating ToxoDB #9 strains seem to be distinct from the strains isolated from other parts of the world. Using three independent sets of polymorphic DNA markers to estimate the phylogenetic distances among 950 isolates collected from all over the world, the TgCtPRC04 strain representing ToxoDB #9 isolated from China was separated from other strains and formed a unique haplogroup (Su et al., 2012). Whole genome sequencing was done on another ToxoDB #9 strain TgCtPRC2 (which was isolated from a cat and also belonged to the same haplogroup as TgCtPRC04) and the data of which was deposited in ToxoDB, which facilitated the biological studies of ToxoDB #9 strains.

# **VACCINE DEVELOPMENTS**

Starting in 1990s, with the rapid progress of molecular biology techniques, a significant amount of T. gondii research work was on vaccine developments, particularly subunit vaccines. Some of the representative studies were summarized in Table 3, which shows that there are mainly three types of vaccines tried: recombinant protein vaccines, DNA vaccines, and live vaccines. The core antigens used for recombinant protein vaccines and DNA vaccines are very similar, mostly surface proteins (SAGs) and proteins from secretory organelles such as, dense granules (GRAs), rhoptries (ROPs), and micronemes (MICs) (see Table 3 for examples and references). These two types of vaccines only differ in the way the core antigens being delivered: for recombinant protein vaccines, the core antigens were administrated in the form of purified proteins, often along with chemical adjuvant to boost the immune response; DNA vaccines were administrated in the form of DNA (plasmids or viral vectors) that expressed the core antigens in hosts, sometimes using cytokine (IL-12, IL-15, etc.) expressing constructs as adjuvants (Zhang et al., 2007; Cui et al., 2008; Chen et al., 2014a). Most of these vaccines elicited Th1 immune responses and had some degree of protections against parasite infections, increasing survival rates, and/or time or reducing cyst burdens (Yuan et al., 2011b; Zheng et al., 2017). Nonetheless, none of these subunit vaccines in the current form offered enough protection to be used in clinic.

Live vaccines were thought to be the most effective against *T. gondii*, with the good example of "Toxovax," although this vaccine is also not perfect (Buxton and Innes, 1995; Burrells et al., 2015). Design of *T. gondii* vaccines using live parasites were also tried in China, but only with limited success. For example, A temperature-sensitive mutant of RH (ts-4) was tested as a live vaccine for ocular toxoplasmosis in a mouse

model. After intracameral inoculation, this strain did provide protective immunity against wild type parasite infection (Lu et al., 2009). However, the strain by itself also caused ocular pathology, preventing its use as a vaccine. Live vaccines were also designed using other parasite species as antigen delivery vectors. For example, transgenic *Eimeria tenella* expressing TgSAG1 was tested as vaccine to protect *T. gondii* infection in chickens and mice. Although it did elicit a Th1-dominant immune response that restricted *T. gondii* proliferation, the protective immunity it offered was limited (Tang et al., 2016). More recently, with the wide use of CRISPR/CAS9 based genome editing system, generating gene deletion mutants to be used as live vaccines became popular, which is also a feasible approach given the success of uracil auxotrophs (Fox and Bzik, 2002).

### **MOLECULAR BASIS FOR PATHOGENESIS**

Before the year 2000, the majority of T. gondii research in China is about epidemiology and vaccine design, very little was done toward the understanding of parasite biology or mechanisms of pathogenesis. After entering the new century, basic research became more active. By looking at the papers Chinese scientists published during the last 15 years, it is obvious that the vast majority of the limited work was about hostparasite interactions, or what influence parasites had on host and host cells. Quite a few transcriptomic and proteomic studies examined the gene expression and protein abundance changes of the hosts upon parasite infection. For examples, RNA-Seq was used to look at the gene expression changes in cat liver, mouse liver and pig peripheral blood mononuclear cells (PBMCs) after infection (He et al., 2016a; Zhou et al., 2016a,b). Similarly, iTRAQ-based proteomic analysis on mice liver suggested that the relative abundances of hundreds of proteins were changed upon parasite infection, many of which were involved in key metabolic pathways (He et al., 2016b). These omics studies did generate a large amount of data that were informative, but followup studies are required to figure out the biological significances of these data. More recently, with the development of genome editing techniques, increasing amount of work started to address the biological properties of the parasite. Using the CRISPR/Cas9 technology, leucine aminopeptidase, calcium-dependent protein kinases, and rhoptry proteins were inactivated to check the functions of these genes during parasite growth and development (Zheng et al., 2015; Wang J. L. et al., 2016, 2017). Today, studies in China cover most fields of T. gondii research. In this review, we only focused on the studies of ToxoDB #9 strains, which are almost specific to China.

As mentioned above, strains isolated from China are dominated by the ToxoDB #9 genotype. However, strains belonging to this genotype may display different properties. TgCtwh3 and TgCtwh6 are probably the best characterized ToxoDB #9 strains, and they show different virulence in mice, with TgCtwh3 being significantly more virulent than TgCtwh6 (Wang et al., 2013). In addition, TgCtwh3 was shown to induce alternative activation of mouse macrophages with STAT6 phosphorylation, whereas the TgCtwh6 elicited classical

### TABLE 3 | T. gondii vaccine candidates designed by scientists in China.

	Antigens/parasite strains	Effect	References
DNA and vector vaccines	SAG1	Increased survival time to 20.38 $\pm$ 3.38 days vs. control (13.25 $\pm$ 1.16 days)	Chen et al., 1999
	SAG1, SAG2 linked to A2/B subunits of cholera toxin	40% survival rate in immunized mice	Cong et al., 2005
	SAG1, ROP2 with IL-12 as adjuvant	Increased survival time to 22 days vs. control (4-8 days)	Zhang et al., 2007
	SAG1-ROP2-SAG2 co-deliveried with IL-12	Increased survival rate	Cui et al., 2008
	SAG1, GRA1, GRA2, GRA4 antigen segments	The survival rate of BALB/c and C57BL/6 mice were 100 and 40%, respectively	Liu et al., 2009
	SAG2C, SAG2D, SAG2X	Reduction of cyst burden (77%) in the brain	Zhang M. et al., 2013
	MIC3 (suicidal vector pSCA1)	Increased the survival time to 15 days vs. control mice (5 days)	Fang et al., 2009
	MIC3 (recombinant pseudorabies virus rPRV)	The survival rates of BALB/c mice injected with rPRV-MIC3 alone was 50%	Nie et al., 2011
	MIC3, SAG1 (baculovirus vaccine BV-MIC3+BV-SAG1)	50% of the mice survived	Fang et al., 2012
	MIC6	Increased survival time to 13.3 $\pm$ 1.2 days vs. control mice (7 days)	Peng et al., 2009
	MIC8	Increased survival time to 10.3 $\pm$ 0.9 days vs. control mice (5 days)	Liu M. M. et al., 2010; Liu Y. et al., 2010
	MIC11	Increased the survival time to 15 days vs. control mice (8–10 days)	Tao et al., 2013
	MIC13	Increased survival time to $21.3 \pm 11.3$ days vs. control mice (5–10 days) and reduced number of cysts in brain of mice (57.14%)	Yuan et al., 2013b
	ROP9	Increased survival time to 12.9 $\pm$ 2.9 days vs. control mice (6 days)	Chen et al., 2014b
	ROP16	Increased survival time to 21.6 $\pm$ 9.9 days vs. control mice (7 days)	Yuan et al., 2011a
	ROP18	Increased survival time to 27.9 $\pm$ 15.1 days vs. control mice (7 days)	Yuan et al., 2011b
	ROM1	Increased survival time to 12.5 $\pm$ 0.7 days vs. control mice (5 days)	Li et al., 2012
	GRA6 with levamisole as adjuvant	53.3% survival	Sun et al., 2011
	elF4A	Increased survival time to 23.0 $\pm$ 5.5 days compared to control mice (7 days)	Chen et al., 2013
	MIC3, GRA1	Increased survival time to 12–19 days (15.7 $\pm$ 1.88) vs. control group survived for 3–5 days (4.5 $\pm$ 0.38)	Gong et al., 2016
	MIC3, ROP18	Increased survival time to 14–19 days vs. control mice (7 days)	Qu et al., 2013a
	Aspartic protease 1	Increased the survival time to 16 days vs. control (7 days)	Zhao et al., 2013
	CDPK1 with a plasmid encoding IL-15 and IL-21	Increased survival time to 19.2 $\pm$ 5.1 days vs. control (6 days) and reduced the number of brain cysts (72.7%)	Chen et al., 2014a
	NTPase-II (pDREP)	71.4% reduction in brain cysts	Zheng et al., 2017
Recombinant protein vaccines	ROP5	Increased survival time to 12.1 $\pm$ 3.4 days vs. control (6 days)	Zheng et al., 2013
	ROP17	Lower liver and brain parasite burdens (59.17 and 49.08%, respectively) and increased survival time by 50%	Wang H. L. et al., 2014
	ROP18, ROP38 encapsulated in poly (lactide-co-glycolide)	81.3% reduction of tissue cysts	Xu et al., 2015
Live vaccines	T. gondii temperature-sensitive mutant	Induced protective immunity in an ocular toxoplasmosis model	Lu et al., 2009
	Eimeria tenalla sporozoites expressing TgSAG1	Induced Th1 immune response and prolonged survival of mice	Tang et al., 2016

activation of mouse macrophages with nuclear translocation of NF-κB (Zhang A. M. et al., 2013). In order to understand why the two strains with the same genotype had these differences, both strains were subject to whole genome sequencing and compared to that of type 1 strain GT1. Although single nucleotide polymorphisms (SNPs) and insertions/deletions (indels) were found in hundreds of genes in TgCtwh3 and TgCtwh6 strains, these two strains were genetically more similar to each other than to GT1. GRA3 and RON3 were shown to have drastic expression differences between the two strains, but whether or not they contribute to the phenotypic difference is still unknown (Cheng et al., 2015). A recent study using multilocus sequence typing suggested that ToxoDB #9 strains could be divided into multiple sub-groups. However, using this typing method, TgCtwh3 and TgCtwh6 were still grouped together, although they display different phenotypes. As the authors of this work implied, perhaps genotyping using neutral genetic markers is not very useful in predicting pathogenic phenotypes (Gao et al., 2017). Therefore, further studies are required to dissect the underlying molecular mechanisms for such phenotypic differences, as a way to better understand the biology of ToxoDB #9 strains.

### CHINESE HERBAL MEDICINES FOR T. GONDII INTERVENTION

For decades, inhibitors of folic acid metabolism such as, pyrimethamine and sulfonamide were the primary drugs to treat acute toxoplasmosis. But due to strong side effects and other limitations, these drugs are far from ideal. Developing new anti-Toxoplasma interventions is a long lasting task in the field and scientists in China contributed to this effort. Noticeable achievements include the studies of Chinese herbal medicine against T. gondii (Table 4). As early as 1990, the effect of artemisinin and its derivatives on T. gondii growth was tested *in vitro*, and found that at 0.4  $\mu$ g/ml artemisinin blocked plaque formation on host cell monolayers (Ke et al., 1990). Follow-up work was done to test the effect of artemisinin in controlling toxoplasmosis in vivo in animal models (Shen et al., 2003; Yang and Wan, 2006). The results were not exactly the same from different studies but all seemed to show that artemisinin was not as effective as expected, and some derivatives were more potent than the others (Dunay et al., 2009).

In addition to artemisinin, which was isolated from Artemisia annua, extracts from many other herbs such as, Glycyrrhiza, Scutellaria, and Ginkgo biloba etc, were also reported to have anti-Toxoplasma effects (Chen S. X. et al., 2008; Gong et al., 2011). However, in most cases, the active compounds in these extracts were not known and deserve further research. There are a few occasions where the active compounds were reported. For example, oxymatrine (OM) and matrine (ME), the main alkaloids in the Sophora leguminous plants, were shown to restrict T. gondii growth both in vivo and in vitro: both OM and ME showed high anti-T. gondii activity and low toxicity to Hela cells than spiramycin (SPY); treating infected mice with oxymatrine or matrine increased the survival rates, although not to 100% (Zhang X. et al., 2016). This shows the potential of using natural products to treat toxoplasmosis, but obviously we need to dig further to find out and optimize the active components.

# **CONCLUSIONS AND PERSPECTIVE**

Since the first isolation and discovery of T. gondii in China in 1955, Chinese scientists have been working on this parasite for over 60 years. Tremendous amount of work has been done on the development of diagnostic strategies, subunit vaccines, and epidemiology surveys. It is clear that T. gondii is widespread in China. In particular, pigs and chickens, the main meat source for Chinese citizens, as well as cats, a popular pet people keep, are among the most frequently infected animals, which greatly increase the chance of human infections. But thanks to the unique eating/cooking habits, human infection rates in China are below the world average. A large number of vaccine candidates in different forms were designed, but frankly speaking, none of them is good enough to be used as a vaccine in the current form. Years of research in the vaccine design field seems to suggest that live attenuated parasites may be the best way to achieve good T. gondii vaccines. With the improvements of genetic modification techniques in T. gondii, work on designing live attenuated vaccines becomes more common, significant progress in this direction is expected.

In terms of diagnostics, although lots of efforts have been taken and a variety of assays were developed, very few of them were translated into standardized commercial products or technical standards. As a consequence, many veterinary clinics use home-made protocols for *T. gondii* diagnosis. For the

Active ingredients	Dosage	Effect	References
Artemisinin	0.4 μg/mL (5 days) 1.3 μg/mL (14 days)	Inhibit plaque formation <i>in vitro</i> Eliminate all parasites <i>in vitro</i>	Ke et al., 1990
Ginkgolic acids	167.1 μg/ml	No visible parasites in HFF cells after 48 h of exposure to ginkgolic acids (isolated from the <i>Ginkgo biloba</i> sarcotesta)	Chen R. et al., 2008
nontus obliquus oolysaccharide	3 mg/10 g/d	Decreased testicular spermatogenic cell pathology damage caused by <i>Toxoplasma</i> infection	Liu Y. et al., 2010
Radix glycyrrhizae	5 g/Kg	While combined with sulfachloropyrazine-sodium (SPZ), the survival rate of mice was up to $50\%$	Jiang W. et al., 201
Oxymatrine, matrine	100 mg/Kg	Decreased the number of tachyzoite (45.2 and 53.8%) and increased survival rate of mice to 67%	Zhang X. et al., 201

TABLE 4 | Effects of herbal medicines or their active components on Toxoplasma gondi

toxoplasmosis examination in humans during pregnancy, wellestablished reference laboratories are still lacking in China. Most hospitals use a single IgM-IgG test to judge the risk, which is not reliable and may lead to false positive results and unnecessary termination of pregnancy. In addition, diagnostic products, especially IgM detecting kits, from domestic manufacturers are not yet ideal for clinic uses. Therefore, efforts are needed to translate the laboratory assays to standardized products for accurate and consistent diagnosis. Reference laboratories should be established to help local clinics and hospitals to get reliable test results for human samples. In addition to these, new diagnostic strategies, which can not only tell the infection status but also inform the source of infection (oocysts vs. tachyzoites/bradyzoites; Santana et al., 2015), may be needed for better risk factor association analysis.

On the basic research side, although so far the contribution from Chinese scientists to the understanding of biology, as well as mechanisms of pathogenesis of *T. gondii* is limited, it is obvious that the research activities are really booming after the year 2000. Currently, there are groups working on the host mechanisms that restrict parasite infections (Qin et al., 2017), parasite mechanisms of immune evasion (Xue et al., 2017), molecular basis for *T. gondii* invasion (Wang Y. et al., 2014; Wang M. et al., 2017), growth and differentiation (Shen et al., 2016), etc. Besides these common research interests shared by the whole community, studies addressing the population structure

### REFERENCES

- Al-Adhami, B. H., Simard, M., Hernández-Ortiz, A., Boireau, C., and Gajadhar, A. A. (2016). Development and evaluation of a modified agglutination test for diagnosis of *Toxoplasma* infection using tachyzoites cultivated in cell culture. *Food Waterborne Parasitol.* 2, 15–21. doi: 10.1016/j.fawpar.2015. 12.001
- Burrells, A., Benavides, J., Canton, G., Garcia, J. L., Bartley, P. M., Nath, M., et al. (2015). Vaccination of pigs with the S48 strain of *Toxoplasma gondii*–safer meat for human consumption. *Vet. Res.* 46:47. doi: 10.1186/s13567-015-0177-0
- Buxton, D., and Innes, E. A. (1995). A commercial vaccine for ovine toxoplasmosis. *Parasitology* 110(Suppl.), S11–S16. doi: 10.1017/S003118200000144X
- Chen, J. (2010). Epidemiological Study of Toxoplasmosis of Dogs and Cats in Shanghai. Master, Shanghai Jiaotong University. (Translated from Chinese).
- Chen, J., Huang, S. Y., Li, Z. Y., Yuan, Z. G., Zhou, D. H., Petersen, E., et al. (2013). Protective immunity induced by a DNA vaccine expressing eIF4A of *Toxoplasma gondii* against acute toxoplasmosis in mice. *Vaccine* 31, 1734–1739. doi: 10.1016/j.vaccine.2013.01.027
- Chen, J., Li, Z. Y., Huang, S. Y., Petersen, E., Song, H. Q., Zhou, D. H., et al. (2014a). Protective efficacy of *Toxoplasma gondii* calcium-dependent protein kinase 1 (TgCDPK1) adjuvated with recombinant IL-15 and IL-21 against experimental toxoplasmosis in mice. *BMC Infect. Dis.* 14:487. doi: 10.1186/1471-2334-14-487
- Chen, J., Zhou, D. H., Li, Z. Y., Petersen, E., Huang, S. Y., Song, H. Q., et al. (2014b). *Toxoplasma gondii*: protective immunity induced by rhoptry protein 9 (TgROP9) against acute toxoplasmosis. *Exp. Parasitol.* 139, 42–48. doi: 10.1016/j.exppara.2014.02.016
- Chen, R., Lin, X., Hu, L., Chen, X., Tang, Y., Zhang, J., et al. (2015). Genetic characterization of *Toxoplasma gondii* from zoo wildlife and pet birds in Fujian, China. *Iran. J. Parasitol.* 10, 663–668.
- Chen, R., Lu, S., Lou, D., Lin, A., Zeng, X., Ding, Z., et al. (2008). Evaluation of a rapid ELISA technique for detection of circulating antigens of *Toxoplasma gondii*. *Microbiol. Immunol.* 52, 180–187. doi: 10.1111/j.1348-0421.2008.00020.x

of strains prevalent in China, the unique biological properties of ToxoDB #9 strains, and interplays between local prevalent strains and animal hosts are also required. It is expected that, after some years, breakthroughs in related fields will be made by our Chinese colleagues.

# AUTHOR CONTRIBUTIONS

MP, CL, JZ, and BS reviewed the literature and wrote the paper.

# **FUNDING**

This work was supported the National Key Research and Development Program of China (project 2017YFD0501304), National Natural Science Foundation of China (project No. 31572508), and the Fundamental Research Funds for the Central Universities in China (Project 2662015PY104).

# ACKNOWLEDGMENTS

We thank Drs. Min Hu and Yanqin Zhou from the College of Veterinary Medicine, Huazhong Agricultural University for their critical reading and comments of the manuscript. We also thank all the scientists whose work was included in the paper but was not cited due to limited space.

- Chen, S. X., Wu, L., Jiang, X. G., Feng, Y. Y., and Cao, J. P. (2008). Anti-Toxoplasma gondii activity of GAS in vitro. J. Ethnopharmacol. 118, 503–507. doi: 10.1016/j.jep.2008.05.023
- Chen, X. G., Fung, M. C., Ma, X., Peng, H. J., Shen, S. M., and Liu, G. Z. (1999). Baculovirus expression of the major surface antigen of *Toxoplasma gondii* and the immune response of mice injected with the recombinant P30. *Southeast Asian J. Trop. Med. Public Health* 30, 42–46.
- Chen, Y. M., Li, G. D., and Li, B. C. (1980). Indirect heamagglutination assay of experimental toxoplasmosis. *J. Vet. Technol.* 4, 10–16. (Translated from Chinese).
- Chen, Z. Y., Li, A. M., Lin, G. C., and Wang, X. Z. (2005). Serological investigation of Toxoplasma infection among different populations of people in Guizhou Province. Acta Academiae Med Zunyi 28, 382–383. (Translated from Chinese).
- Cheng, W., Liu, F., Li, M., Hu, X., Chen, H., Pappoe, F., et al. (2015). Variation detection based on next-generation sequencing of type Chinese 1 strains of *Toxoplasma gondii* with different virulence from China. *BMC Genomics* 16:888. doi: 10.1186/s12864-015-2106-z
- Cong, H., Gu, Q. M., Jiang, Y., He, S. Y., Zhou, H. Y., Yang, T. T., et al. (2005). Oral immunization with a live recombinant attenuated Salmonella typhimurium protects mice against Toxoplasma gondii. Parasite Immunol. 27, 29–35. doi: 10.1111/j.1365-3024.2005.00738.x
- Cong, W., Dong, W., Bai, L., Wang, X. Y., Ni, X. T., Qian, A. D., et al. (2015a). Seroprevalence and associated risk factors of *Toxoplasma gondii* infection in psychiatric patients: a case-control study in eastern China. *Epidemiol. Infect.* 143, 3103–3109. doi: 10.1017/S0950268814003835
- Cong, W., Huang, S. Y., Zhou, D. H., Zhang, X. X., Zhang, N. Z., Zhao, Q., et al. (2013). Prevalence and genetic characterization of *Toxoplasma gondii* in house sparrows (*Passer domesticus*) in Lanzhou, China. *Korean J. Parasitol.* 51, 363–367. doi: 10.3347/kjp.2013.51.3.363
- Cong, W., Liu, G. H., Meng, Q. F., Dong, W., Qin, S. Y., Zhang, F. K., et al. (2015b). *Toxoplasma gondii* infection in cancer patients: prevalence, risk factors, genotypes and association with clinical diagnosis. *Cancer Lett.* 359, 307–313. doi: 10.1016/j.canlet.2015.01.036

- Cong, W., Meng, Q. F., Blaga, R., Villena, I., Zhu, X. Q., and Qian, A. D. (2016a). *Toxoplasma gondii*, *Dirofilaria immitis*, feline immunodeficiency virus (FIV), and feline leukemia virus (FeLV) infections in stray and pet cats (*Felis catus*) in northwest China: co-infections and risk factors. *Parasitol. Res.* 115, 217–223. doi: 10.1007/s00436-015-4738-y
- Cong, W., Meng, Q. F., Song, H. Q., Zhou, D. H., Huang, S. Y., Qian, A. D., et al. (2014). Seroprevalence and genetic characterization of *Toxoplasma gondii* in three species of pet birds in China. *Parasit. Vectors* 7:152. doi: 10.1186/1756-3305-7-152
- Cong, W., Qin, S. Y., Meng, Q. F., Zou, F. C., Qian, A. D., and Zhu, X. Q. (2016b). Molecular detection and genetic characterization of *Toxoplasma gondii* infection in sika deer (Cervus nippon) in China. *Infect. Genet. Evol.* 39, 9–11. doi: 10.1016/j.meegid.2016.01.005
- Cui, Y. L., He, S. Y., Xue, M. F., Zhang, J., Wang, H. X., and Yao, Y. (2008). Protective effect of a multiantigenic DNA vaccine against *Toxoplasma* gondii with co-delivery of IL-12 in mice. *Parasite Immunol.* 30, 309–313. doi: 10.1111/j.1365-3024.2008.01025.x
- Dai, J. F., Jiang, M., Qu, L. L., Sun, L., Wang, Y. Y., Gong, L. L., et al. (2013). *Toxoplasma gondii*: enzyme-linked immunosorbent assay based on a recombinant multi-epitope peptide for distinguishing recent from past infection in human sera. *Exp. Parasitol.* 133, 95–100. doi: 10.1016/j.exppara.2012.10.016
- De Berardinis, A., Paludi, D., Pennisi, L., and Vergara, A. (2017). *Toxoplasma gondii*, a foodborne pathogen in the swine production Chain from a European perspective. *Foodborne Pathog. Dis.* doi: 10.1089/fpd.2017.2305. [Epub ahead of print].
- Ding, H., Gao, Y. M., Deng, Y., Lamberton, P. H., and Lu, D. B. (2017). A systematic review and meta-analysis of the seroprevalence of *Toxoplasma gondii* in cats in mainland China. *Parasit. Vectors* 10:27. doi: 10.1186/s13071-017-1970-6
- Ding, H., Liu, H. L., Han, J. H., Chen, W., Xu, R. N., and Li, X. (2009). Serum TORCH test results of 7650 pregnant women. *Pract. Prev. Med.* 16, 160–162. doi: 10.3969/j.issn.1007-4287.2009.10.057. (Translated from Chinese).
- Ding, R., Zhang, Z. X., Zeng, H., Chen, M., and Wang, L. S. (2015). Analysis of TORCH test results of 6027 cases of pregnant women and newborns. *Int. J. Lab. Med.* 36, 485–486. doi: 10.3969/j.issn.1673-4130.2015.04.022. (Translated from Chinese).
- Dos Santos, T. R., Nunes, C. M., Luvizotto, M. C., De Moura, A. B., Lopes, W. D., Da Costa, A. J., et al. (2010). Detection of *Toxoplasma gondii* oocysts in environmental samples from public schools. *Vet. Parasitol.* 171, 53–57. doi: 10.1016/j.vetpar.2010.02.045
- Du, F., Feng, H. L., Nie, H., Tu, P., Zhang, Q. L., Hu, M., et al. (2012a). Survey on the contamination of *Toxoplasma gondii* oocysts in the soil of public parks of Wuhan, China. *Vet. Parasitol.* 184, 141–146. doi: 10.1016/j.vetpar.2011. 08.025
- Du, F., Zhang, Q., Yu, Q., Hu, M., Zhou, Y., and Zhao, J. (2012b). Soil contamination of *Toxoplasma gondii* oocysts in pig farms in central China. *Vet. Parasitol.* 187, 53–56. doi: 10.1016/j.vetpar.2011.12.036
- Dubey, J. P. (2009). Toxoplasmosis in pigs-the last 20 years. Vet. Parasitol. 164, 89-103. doi: 10.1016/j.vetpar.2009.05.018
- Dubey, J. P. (2010). *Toxoplasmosis of Animals and Humans*. Boca Raton, FL: CRC press.
- Dunay, I. R., Chan, W. C., Haynes, R. K., and Sibley, L. D. (2009). Artemisone and artemiside control acute and reactivated toxoplasmosis in a murine model. *Antimicrob. Agents Chemother*, 53, 4450–4456. doi: 10.1128/AAC.00502-09
- Fan, Y. H. (2017). A case of abortion caused by porcine toxoplasmosis. *Heilongjiang J. Anim. Reprod.* 25. doi: 10.3969/j.issn.1005-2739.2017.01.025. (Translated from Chinese).
- Fang, R., Feng, H., Hu, M., Khan, M. K., Wang, L., Zhou, Y., et al. (2012). Evaluation of immune responses induced by SAG1 and MIC3 vaccine cocktails against *Toxoplasma gondii*. *Vet. Parasitol.* 187, 140–146. doi: 10.1016/j.vetpar.2011.12.007
- Fang, R., Nie, H., Wang, Z., Tu, P., Zhou, D., Wang, L., et al. (2009). Protective immune response in BALB/c mice induced by a suicidal DNA vaccine of the MIC3 gene of *Toxoplasma gondii*. Vet. Parasitol. 164, 134–140. doi: 10.1016/j.vetpar.2009.06.012
- Feng, J. X., Zhao, Y. K., Meng, F. P., Wu, Z. M., Liu, Z., Li, N., et al. (2016). Research progress in diagnosis of toxoplasmosis. *Infect. Dis. Inform.*

29, 139-143. doi: 10.3969/j.issn.1007-8134.2016.03.004. (Translated from Chinese).

- Fox, B. A., and Bzik, D. J. (2002). De novo pyrimidine biosynthesis is required for virulence of *Toxoplasma gondii*. Nature 415, 926–929. doi: 10.1038/415926a
- Gao, G. (2014). Recent advances in clinical characteristicks and diagnosis and treatment of toxoplasmosis. *Chin. J. Pathog. Biol.* 9, 848–851. doi: 10.13350/j.cjpb.14092. (Translated from Chinese).
- Gao, J. M., Xie, Y. T., Xu, Z. S., Chen, H., Hide, G., Yang, T. B., et al. (2017). Genetic analyses of Chinese isolates of *Toxoplasma gondii* reveal a new genotype with high virulence to murine hosts. *Vet. Parasitol.* 241, 52–60. doi: 10.1016/j.vetpar.2017.05.007
- Gao, X., Wang, H., Wang, H., Qin, H., and Xiao, J. (2016). Land use and soil contamination with *Toxoplasma gondii* oocysts in urban areas. *Sci. Total Environ.* 568, 1086–1091. doi: 10.1016/j.scitotenv.2016.06.165
- Gao, Y. M., Ding, H., Lamberton, P. H., and Lu, D. B. (2016). Prevalence of *Toxoplasma gondii* in pet dogs in mainland China: a meta-analysis. *Vet. Parasitol.* 229, 126–130. doi: 10.1016/j.vetpar.2016.10.009
- Gong, G. H., Hou, J. F., Jiang, W., Liu, Y. C., Chen, Y. J., Jing, Z. Y., et al. (2011). Analysis of anti-*Toxoplasma gondii* drugs of seven kinds of drugs. *Anim. Husbandy Vet. Med.* 43, 9–10. (Translated from Chinese).
- Gong, P., Cao, L., Guo, Y., Dong, H., Yuan, S., Yao, X., et al. (2016). *Toxoplasma gondii*: protective immunity induced by a DNA vaccine expressing GRA1 and MIC3 against toxoplasmosis in BALB/c mice. *Exp. Parasitol.* 166, 131–136. doi: 10.1016/j.exppara.2016.04.003
- Guan, L. B. (2017). Analysis of TORCH test results in spontaneous abortion patients. Syst. Med. 2, 7–9. doi: 10.19368/j.cnki.2096-1782.2017.05.007
- He, J. J., Ma, J., Elsheikha, H. M., Song, H. Q., Huang, S. Y., and Zhu, X. Q. (2016a). Transcriptomic analysis of mouse liver reveals a potential hepatoenteric pathogenic mechanism in acute *Toxoplasma gondii* infection. *Parasit. Vectors* 9:427. doi: 10.1186/s13071-016-1716-x
- He, J. J., Ma, J., Elsheikha, H. M., Song, H. Q., Zhou, D. H., and Zhu, X. Q. (2016b). Proteomic profiling of mouse liver following acute *Toxoplasma gondii* infection. *PLoS ONE* 11:e0152022. doi: 10.1371/journal.pone.0152022
- He, Y. Y., Jiang, S. F., Ma, X. B., and Qiu, Q. W. (2008). Evaluation of sixteen kinds of kits inner and abroad available for detecting antibodies of *Toxoplasma* gondii. Sci. Travel Med. 14, 43–45. doi: 10.3969/j.issn.1006-7159.2008.03.022. (Translated from Chinese).
- Howe, D. K., and Sibley, L. D. (1995). *Toxoplasma gondii* comprises three clonal lineages: correlation of parasite genotype with human disease. J. Infect. Dis. 172, 1561–1566. doi: 10.1093/infdis/172.6.1561
- Huang, S. Y., Cong, W., Zhou, P., Zhou, D. H., Wu, S. M., Xu, M. J., et al. (2012). First report of genotyping of *Toxoplasma gondii* isolates from wild birds in China. J. Parasitol. 98, 681–682. doi: 10.1645/GE-3038.1
- Huo, S. L., Song, Z. Z., Zhang, B., Han, S., Bai, L., and Wu, C. P. (2007). A Serological Investigation of Toxoplasmosis in inner Mongolia. J. Med. Pest Control 23, 894–896. doi: 10.3969/j.issn.1003-6245.2007.12.006. (Translated from Chinese).
- Jiang, C., Li, Z., Chen, P., and Chen, L. (2015). The seroprevalence of *Toxoplasma* gondii in Chinese population with cancer: a systematic review and metaanalysis. *Medicine* 94:e2274. doi: 10.1097/MD.00000000002274
- Jiang, H. H., Huang, S. Y., Zhou, D. H., Zhang, X. X., Su, C., Deng, S. Z., et al. (2013). Genetic characterization of *Toxoplasma gondii* from pigs from different localities in China by PCR-RFLP. *Parasit. Vectors* 6:227. doi: 10.1186/1756-3305-6-227
- Jiang, H. H., Qin, S. Y., Wang, W., He, B., Hu, T. S., Wu, J. M., et al. (2014). Prevalence and genetic characterization of *Toxoplasma* gondii infection in bats in southern China. *Vet. Parasitol.* 203, 318–321. doi: 10.1016/j.vetpar.2014.04.016
- Jiang, H. H., Wang, S. C., Huang, S. Y., Zhao, L., Wang, Z. D., Zhu, X. Q., et al. (2016). Genetic characterization of *Toxoplasma gondii* isolates from pigs in Jilin Province, Northeastern China. *Foodborne Pathog. Dis.* 13, 88–92. doi: 10.1089/fpd.2015.2043
- Jiang, S. F., Zhang, S. Y., Pan, C. E., He, Y. Y., and Wei, M. X. (2003). Evaluation of five commercial available kits for detecting antibodies of *Toxoplasma gondii*. *Chin. J. Zoonoses* 19, 97–99. doi: 10.3969/j.issn.1002-2694.2003.01.030. (Translated from Chinese).
- Jiang, T., Gong, D., Ma, L. A., Nie, H., Zhou, Y., Yao, B., et al. (2008). Evaluation of a recombinant MIC3 based latex agglutination test for the rapid

serodiagnosis of *Toxoplasma gondii* infection in swines. *Vet. Parasitol.* 158, 51–56. doi: 10.1016/j.vetpar.2008.07.035

- Jiang, W., Chen, Y. J., Zhang, H. J., Liu, Y. C., Jing, Z. Y., Xue, J. X., et al. (2013). Treatment effects of Sulfachloropyrazine-sodium with Scutellaria Baicalensis and Glycyrrhiza against acute murine toxoplasomsis. *Chin. J. Anim. Vet. Sci.* 40, 2022–2028. doi: 10.11843/j.issn.0366-6964.2013.12.024. (Translated from Chinese).
- Jiang, W., Liu, Y., Chen, Y., Yang, Q., Chun, P., Yao, K., et al. (2015). A novel dynamic flow immunochromatographic test (DFICT) using gold nanoparticles for the serological detection of *Toxoplasma gondii* infection in dogs and cats. *Biosens. Bioelectron.* 72, 133–139. doi: 10.1016/j.bios.2015.04.035
- Jin, S., Chang, Z. Y., Ming, X., Min, C. L., Wei, H., Sheng, L. Y., et al. (2005). Fast dipstick dye immunoassay for detection of immunoglobulin G (IgG) and IgM antibodies of human toxoplasmosis. *Clin. Diagn. Lab. Immunol.* 12, 198–201. doi: 10.1128/CDLI.12.1.198-201.2005
- Ke, O. Y., Krug, E. C., Marr, J. J., and Berens, R. L. (1990). Inhibition of growth of *Toxoplasma gondii* by qinghaosu and derivatives. *Antimicrob. Agents Chemother*. 34, 1961–1965. doi: 10.1128/AAC.34.10.1961
- Lai, Z. F., Huang, H. P., Gu, J. B., Situ, C. M., Chen, R. L., Zhong, J. M., et al. (2014). The establishment of the toxoplasma detection method: LAMP. *J. Trop. Med.* 14, 1035–1037. (Translated from Chinese).
- Li, F., Wang, S. P., Wang, C. J., He, S. C., Wu, X., and Liu, G. H. (2016). Seroprevalence of *Toxoplasma gondii* in goats in Hunan province, China. *Parasite* 23:44. doi: 10.1051/parasite/2016053
- Li, H. L., Dong, L., Li, Q., Zhang, L., Chen, J., Zou, F. C., et al. (2015). Seroepidemiology of *Toxoplasma gondii* infection in Bai and Han ethnic groups in southwestern China. *Epidemiol. Infect.* 143, 881–886. doi: 10.1017/S0950268814001551
- Li, J., Han, Q., Gong, P., Yang, T., Ren, B., Li, S., et al. (2012). *Toxoplasma gondii* rhomboid protein 1 (TgROM1) is a potential vaccine candidate against toxoplasmosis. *Vet. Parasitol.* 184, 154–160. doi: 10.1016/j.vetpar.2011.08.014
- Li, M., Mo, X. W., Wang, L., Chen, H., Luo, Q. L., Wen, H. Q., et al. (2014). Phylogeny and virulence divergency analyses of *Toxoplasma gondii* isolates from China. *Parasit. Vectors* 7:133. doi: 10.1186/1756-3305-7-133
- Li, S., Cui, L., Zhao, J., Dai, P., Zong, S., Zuo, W., et al. (2011). Seroprevalence of *Toxoplasma gondii* infection in female sterility patients in China. J. Parasitol. 97, 529–530. doi: 10.1645/GE-2680.1
- Li, Y. N., Nie, X., Peng, Q. Y., Mu, X. Q., Zhang, M., Tian, M. Y., et al. (2015). Seroprevalence and genotype of *Toxoplasma gondii* in pigs, dogs and cats from Guizhou province, Southwest China. *Parasit. Vectors* 8:214. doi: 10.1186/s13071-015-0809-2
- Lin, Z. B., Zhang, H. S., Cao, J., Zhou, Y. Z., Zhang, Y. L., Li, G. Q., et al. (2011). The establishment and preliminary application of the toxoplasma detection method: real-time PCR. *Anim. Husbandry Vet. Med.* 43, 20–23. (Translated from Chinese).
- Lin, Z., Zhang, Y., Zhang, H., Zhou, Y., Cao, J., and Zhou, J. (2012). Comparison of loop-mediated isothermal amplification (LAMP) and real-time PCR method targeting a 529-bp repeat element for diagnosis of toxoplasmosis. *Vet. Parasitol.* 185, 296–300. doi: 10.1016/j.vetpar.2011.10.016
- Liu, L., Liu, L. N., Wang, P., Lv, T. T., Fan, Y. G., and Pan, H. F. (2017). Elevated seroprevalence of *toxoplasma gondii* in AIDS/HIV patients: a meta-analysis. *Acta Trop.* 176, 162–167. doi: 10.1016/j.actatropica.2017. 08.001
- Liu, M. M., Yuan, Z. G., Peng, G. H., Zhou, D. H., He, X. H., Yan, C., et al. (2010). *Toxoplasma gondii* microneme protein 8 (MIC8) is a potential vaccine candidate against toxoplasmosis. *Parasitol. Res.* 106, 1079–1084. doi: 10.1007/s00436-010-1742-0
- Liu, R. Z., Chen, Z. Q., Zhang, C. F., Zhong, W. C., and Chen, M. X. (2013). Serological investigation of *Toxoplasma gondii* infection in parts of Guangdong province. *Poultry Poultry Dis. Control* 7–9. (Translated from Chinese).
- Liu, S., Shi, L., Cheng, Y. B., Fan, G. X., Ren, H. X., and Yuan, Y. K. (2009). Evaluation of protective effect of multi-epitope DNA vaccine encoding six antigen segments of *Toxoplasma gondii* in mice. *Parasitol. Res.* 105, 267–274. doi: 10.1007/s00436-009-1393-1
- Liu, T., Meng, X. F., and Hou, X. X. (2017). Analysis of TORCH test results of 7122 women of childbearing age in Luoyang, Henan province. J. Henan Univ. Sci. Technol. 35, 63–65. doi: 10.15926/j.cnki.issn1672-688x.2017.01.021. (Translated from Chinese).

- Liu, T., Zhang, Q., Liu, L., Xu, X., Chen, H., Wang, H., et al. (2013). Trophoblast apoptosis through polarization of macrophages induced by Chinese *Toxoplasma gondii* isolates with different virulence in pregnant mice. *Parasitol. Res.* 112, 3019–3027. doi: 10.1007/s00436-01 3-3475-3
- Liu, Y., Ju, Y. L., Liu, L. B., Li, J. J., and Wang, Y. C. (2010). Effects of Inontus obliquus plosaccharide against acute *Toxoplasma gondii* infection on apoptosis of testicular gem cell in male mice. *J. Anhui Agric. Sci.* 38, 10474–10475. doi: 10.3969/j.issn.0517-6611.2010.19.196. (Translated from Chinese).
- Lu, B., Wu, S., Shi, Y., Zhang, R., Zou, L., Gao, S., et al. (2006). *Toxoplasma gondii*: expression pattern and detection of infection using full-length recombinant P35 antigen. *Exp. Parasitol.* 113, 83–90. doi: 10.1016/j.exppara.2005.12.014
- Lu, F., Huang, S., and Kasper, L. H. (2009). The temperature-sensitive mutants of *Toxoplasma gondii* and ocular toxoplasmosis. *Vaccine* 27, 573–580. doi: 10.1016/j.vaccine.2008.10.090
- Miao, Q., Han, J. Q., Xiang, X., Yuan, F. Z., Liu, Y. Z., Duan, G., et al. (2014). Prevalence of antibody to *Toxoplasma gondii* in black-headed gulls (*Chroicocephalus ridibundus*), Dianchi Lake, China. J. Wildl. Dis. 50, 717–719. doi: 10.7589/2014-01-016
- Miao, Q., Huang, S. Y., Qin, S. Y., Yu, X., Yang, Y., Yang, J. F., et al. (2015). Genetic characterization of *Toxoplasma gondii* in Yunnan black goats (*Capra hircus*) in southwest China by PCR-RFLP. *Parasit. Vectors* 8:57. doi: 10.1186/s13071-015-0673-0
- Minot, S., Melo, M. B., Li, F., Lu, D., Niedelman, W., Levine, S. S., et al. (2012). Admixture and recombination among *Toxoplasma gondii* lineages explain global genome diversity. *Proc. Natl. Acad. Sci. U.S.A.* 109, 13458–13463. doi: 10.1073/pnas.1117047109
- Montoya, J. G., and Liesenfeld, O. (2004). Toxoplasmosis. Lancet 363, 1965–1976. doi: 10.1016/S0140-6736(04)16412-X
- Nie, H., Fang, R., Xiong, B. Q., Wang, L. X., Hu, M., Zhou, Y. Q., et al. (2011). Immunogenicity and protective efficacy of two recombinant pseudorabies viruses expressing *Toxoplasma gondii* SAG1 and MIC3 proteins. *Vet. Parasitol.* 181, 215–221. doi: 10.1016/j.vetpar.2011.04.039
- Peng, G. H., Yuan, Z. G., Zhou, D. H., He, X. H., Liu, M. M., Yan, C., et al. (2009). *Toxoplasma gondii* microneme protein 6 (MIC6) is a potential vaccine candidate against toxoplasmosis in mice. *Vaccine* 27, 6570–6574. doi: 10.1016/j.vaccine.2009.08.043
- Peng, Y. Y., Zhu, G. M., Fan, H. N., Lin, Y. L., and Li, Y. (2015). Analysis of TORCH test in spontaneous abortion patients. *Adv. Modern Biomed.* 15, 3707–3710. doi: 10.13241/j.cnki.pmb.2015.19.028. (Translated from Chinese).
- Qian, W., Wang, H., Su, C., Shan, D., Cui, X., Yang, N., et al. (2012). Isolation and characterization of *Toxoplasma gondii* strains from stray cats revealed a single genotype in Beijing, China. *Vet. Parasitol.* 187, 408–413. doi: 10.1016/j.vetpar.2012.01.026
- Qin, A., Lai, D. H., Liu, Q., Huang, W., Wu, Y. P., Chen, X., et al. (2017). Guanylatebinding protein 1 (GBP1) contributes to the immunity of human mesenchymal stromal cells against *Toxoplasma gondii*. Proc. Natl. Acad. Sci. U.S.A. 114, 1365–1370. doi: 10.1073/pnas.1619665114
- Qin, S. Y., Cong, W., Liu, Y., Li, N., Wang, Z. D., Zhang, F. K., et al. (2014). Molecular detection and genotypic characterization of *Toxoplasma* gondii infection in bats in four provinces of China. *Parasit. Vectors* 7:558. doi: 10.1186/s13071-014-0558-7
- Qu, D., Han, J., and Du, A. (2013a). Evaluation of protective effect of multiantigenic DNA vaccine encoding MIC3 and ROP18 antigen segments of *Toxoplasma gondii* in mice. *Parasitol. Res.* 112, 2593–2599. doi: 10.1007/s00436-013-3425-0
- Qu, D., Zhou, H., Han, J., Tao, S., Zheng, B., Chi, N., et al. (2013b). Development of reverse transcription loop-mediated isothermal amplification (RT-LAMP) as a diagnostic tool of *Toxoplasma gondii* in pork. *Vet. Parasitol.* 192, 98–103. doi: 10.1016/j.vetpar.2012.10.010
- Ren, X. P., and Bao, J. Z. (1982). The outbreak of toxoplasmosis cases in pig in China. *Anim. Husbandry Vet. Med.* (Translated from Chinese).
- Santana, S. S., Gebrim, L. C., Carvalho, F. R., Barros, H. S., Barros, P. C., Pajuaba, A. C., et al. (2015). CCp5A protein from *Toxoplasma gondii* as a serological marker of oocyst-driven infections in humans and domestic animals. *Front. Microbiol.* 6:1305. doi: 10.3389/fmicb.2015.01305
- Shen, B., Yuan, Y., Cheng, J., Pan, M., Xia, N., Zhang, W., et al. (2016). Activation of chronic toxoplasmosis by transportation stress in a

mouse model. Oncotarget 7, 87351-87360. doi: 10.18632/oncotarget. 13568

- Shen, C. J., Zhan, X. M., Yang, S. J., He, A., Zheng, H. X., Zhang, R. L., et al. (2003). Efficacy of artesunate against toxoplasmosis. *Chin. J. Zoonoses* 19, 128–129. doi: 10.3969/j.issn.1002-2694.2003.05.041. (Translated from Chinese).
- Su, C., Khan, A., Zhou, P., Majumdar, D., Ajzenberg, D., Darde, M. L., et al. (2012). Globally diverse *Toxoplasma gondii* isolates comprise six major clades originating from a small number of distinct ancestral lineages. *Proc. Natl. Acad. Sci. U.S.A.* 109, 5844–5849. doi: 10.1073/pnas.1203190109
- Su, C., Zhang, X., and Dubey, J. P. (2006). Genotyping of *Toxoplasma gondii* by multilocus PCR-RFLP markers: a high resolution and simple method for identification of parasites. *Int. J. Parasitol.* 36, 841–848. doi: 10.1016/j.ijpara.2006.03.003
- Sun, X. M., Zou, J., Elashram Saeed, A. A., Yan, W. C., Liu, X. Y., Suo, X., et al. (2011). DNA vaccination with a gene encoding *Toxoplasma gondii* GRA6 induces partial protection against toxoplasmosis in BALB/c mice. *Parasit. Vectors* 4:213. doi: 10.1186/1756-3305-4-213
- Sun, X., Lu, H., Jia, B., Chang, Z., Peng, S., Yin, J., et al. (2013). A comparative study of *Toxoplasma gondii* seroprevalence in three healthy Chinese populations detected using native and recombinant antigens. *Parasit. Vectors* 6:241. doi: 10.1186/1756-3305-6-241
- Sun, X., Wang, Z., Li, J., Wei, F., and Liu, Q. (2015). Evaluation of an indirect ELISA using recombinant granule antigen GRA1, GRA7 and soluble antigens for serodiagnosis of *Toxoplasma gondii* infection in chickens. *Res. Vet. Sci.* 100, 161–164. doi: 10.1016/j.rvsc.2015.04.011
- Tang, X., Yin, G., Qin, M., Tao, G., Suo, J., Liu, X., et al. (2016). Transgenic Eimeria tenella as a vaccine vehicle: expressing TgSAG1 elicits protective immunity against *Toxoplasma gondii* infections in chickens and mice. *Sci. Rep.* 6:29379. doi: 10.1038/srep29379
- Tao, Q., Fang, R., Zhang, W., Wang, Y., Cheng, J., Li, Y., et al. (2013). Protective immunity induced by a DNA vaccine-encoding *Toxoplasma gondii* microneme protein 11 against acute toxoplasmosis in BALB/c mice. *Parasitol. Res.* 112, 2871–2877. doi: 10.1007/s00436-013-3458-4
- Tian, Y. M., Huang, S. Y., Miao, Q., Jiang, H. H., Yang, J. F., Su, C., et al. (2014). Genetic characterization of *Toxoplasma gondii* from cats in Yunnan Province, Southwestern China. *Parasit. Vectors* 7:178. doi: 10.1186/1756-3305-7-178
- Wang, D., Liu, Y., Jiang, T., Zhang, G., Yuan, G., He, J., et al. (2016). Seroprevalence and genotypes of *Toxoplasma gondii* isolated from pigs intended for human consumption in Liaoning province, northeastern China. *Parasit. Vectors* 9:248. doi: 10.1186/s13071-016-1525-2
- Wang, H. L., Zhang, T. E., Yin, L. T., Pang, M., Guan, L., Liu, H. L., et al. (2014). Partial protective effect of intranasal immunization with recombinant *Toxoplasma gondii* rhoptry protein 17 against toxoplasmosis in mice. *PLoS ONE* 9:e108377. doi: 10.1371/journal.pone.0108377
- Wang, H., Lei, C., Li, J., Wu, Z., Shen, G., and Yu, R. (2004). A piezoelectric immunoagglutination assay for *Toxoplasma gondii* antibodies using gold nanoparticles. *Biosens. Bioelectron.* 19, 701–709. doi: 10.1016/S0956-5663(03)00265-3
- Wang, H., Wang, T., Luo, Q., Huo, X., Wang, L., Liu, T., et al. (2012). Prevalence and genotypes of *Toxoplasma gondii* in pork from retail meat stores in Eastern China. *Int. J. Food Microbiol.* 157, 393–397. doi:10.1016/j.ijfoodmicro.2012.06.011
- Wang, J. L., Huang, S. Y., Li, T. T., Chen, K., Ning, H. R., and Zhu, X. Q. (2016). Evaluation of the basic functions of six calcium-dependent protein kinases in *Toxoplasma gondii* using CRISPR-Cas9 system. *Parasitol. Res.* 115, 697–702. doi: 10.1007/s00436-015-4791-6
- Wang, J. L., Li, T. T., Elsheikha, H. M., Chen, K., Zhu, W. N., Yue, D. M., et al. (2017). Functional characterization of rhoptry kinome in the virulent *Toxoplasma gondii* RH strain. *Front. Microbiol.* 8:84. doi: 10.3389/fmicb.2017.00084
- Wang, L., Chen, H., Liu, D., Huo, X., Gao, J., Song, X., et al. (2013). Genotypes and mouse virulence of *Toxoplasma gondii* isolates from animals and humans in China. *PLoS ONE* 8:e53483. doi: 10.1371/journal.pone.0053483
- Wang, L., He, L. Y., Meng, D. D., Chen, Z. W., Wen, H., Fang, G. S., et al. (2015). Seroprevalence and genetic characterization of *Toxoplasma gondii* in cancer patients in Anhui Province, Eastern China. *Parasit. Vectors* 8:162. doi: 10.1186/s13071-015-0778-5

- Wang, M., Cao, S., Du, N., Fu, J., Li, Z., Jia, H., et al. (2017). The moving junction protein RON4, although not critical, facilitates host cell invasion and stabilizes MJ members. *Parasitology* 144, 1490–1497. doi: 10.1017/S0031182017000968
- Wang, S., Lan, C., Zhang, L., Zhang, H., Yao, Z., Wang, D., et al. (2015). Seroprevalence of *Toxoplasma gondii* infection among patients with hand, foot and mouth disease in Henan, China: a hospital-based study. *Infect. Dis. Poverty* 4:53. doi: 10.1186/s40249-015-0088-3
- Wang, Y. B., Yang, X. D., and Yang, G. R. (2012). A survey of toxoplasmosis. *Chin. J. Trop. Med.* 12, 497–500. doi: 10.13604/j.cnki.46-1064/r.2012.04.026. (Translated from Chinese).
- Wang, Y. H., Li, X. R., Wang, G. X., Yin, H., Cai, X. P., Fu, B. Q., et al. (2011). Development of an immunochromatographic strip for the rapid detection of *Toxoplasma gondii* circulating antigens. *Parasitol. Int.* 60, 105–107. doi: 10.1016/j.parint.2010.11.002
- Wang, Y., Fang, R., Yuan, Y., Hu, M., Zhou, Y., and Zhao, J. (2014). Identification of host proteins interacting with the integrin-like A domain of *Toxoplasma* gondii micronemal protein MIC2 by yeast-two-hybrid screening. Parasit. Vectors 7:543. doi: 10.1186/s13071-014-0543-1
- Wang, Z., Ge, W., Huang, S. Y., Li, J., Zhu, X. Q., and Liu, Q. (2014). Evaluation of recombinant granule antigens GRA1 and GRA7 for serodiagnosis of *Toxoplasma gondii* infection in dogs. *BMC Vet. Res.* 10:158. doi: 10.1186/1746-6148-10-158
- Wu, D., Lv, R., Sun, X., Shu, F., Zhou, Z., Nie, K., et al. (2012). Seroprevalence of *Toxoplasma gondii* antibodies from slaughter pigs in Chongqing, China. *Trop. Anim. Health Prod.* 44, 685–687. doi: 10.1007/s11250-011-9965-3
- Wu, K., Chen, X. G., Li, H., Yan, H., Yang, P. L., Lun, Z. R., et al. (2009). Diagnosis of human toxoplasmosis by using the recombinant truncated surface antigen 1 of *Toxoplasma gondii*. *Diagn. Microbiol. Infect. Dis.* 64, 261–266. doi: 10.1016/j.diagmicrobio.2009.02.009
- Wu, S. M., Danba, C., Huang, S. Y., Zhang, D. L., Chen, J., Gong, G., et al. (2011a). Seroprevalence of *Toxoplasma gondii* infection in Tibetan sheep in Tibet, China. J. Parasitol. 97, 1188–1189. doi: 10.1645/GE-2912.1
- Wu, S. M., Huang, S. Y., Fu, B. Q., Liu, G. Y., Chen, J. X., Chen, M. X., et al. (2011b). Seroprevalence of *Toxoplasma gondii* infection in pet dogs in Lanzhou, Northwest China. *Parasit. Vectors* 4:64. doi: 10.1186/1756-3305-4-64
- Wu, S. M., Zhu, X. Q., Zhou, D. H., Fu, B. Q., Chen, J., Yang, J. F., et al. (2011c). Seroprevalence of *Toxoplasma gondii* infection in household and stray cats in Lanzhou, northwest China. *Parasit. Vectors* 4:214. doi:10.1186/1756-3305-4-214
- Wu, T. H. (2017). A case of the diagnosis, treatment and experience in porcine toxoplasmosis. *Livestock Poult. Industry* 28, 65–68. doi: 10.19567/j.cnki.1008-0414.2017.04.039. (Translated from Chinese).
- Wu, Z. Y. (2011). The new situation and challenges of prevention and control of AIDS in China. *Chin. J. Public Health* 27, 1505–1507. (Translated from Chinese).
- Xiao, Y., Yin, J., Jiang, N., Xiang, M., Hao, L., Lu, H., et al. (2010). Seroepidemiology of human *Toxoplasma gondii* infection in China. *BMC Infect. Dis.* 10:4. doi: 10.1186/1471-2334-10-4
- Xu, L. Q., Chen, Y. D., Sun, F. H., Cai, L., Fang, Y. Y., Wang, L. P., et al. (2005). A national survey on current status of the important parasitic diseases and control strategies. *Chin. J. Parasitol. Parasite Dis.* 23, 332–340. (Translated from Chinese).
- Xu, P., Li, X., Guo, L., Li, B., Wang, J., Yu, D., et al. (2014). Seroprevalence of *Toxoplasma gondii* infection in Liaoning cashmere goat from northeastern China. *Parasite* 21:22. doi: 10.1051/parasite/2014023
- Xu, Y., Li, R. C., Liu, G. H., Cong, W., Zhang, X. X., Yu, X. L., et al. (2014). Seroprevalence of *Toxoplasma gondii* infection in sows in Hunan province, China. Sci. World J. 2014:347908. doi: 10.1155/2014/347908.
- Xu, Y., Zhang, N. Z., Wang, M., Dong, H., Feng, S. Y., Guo, H. C., et al. (2015). A long-lasting protective immunity against chronic toxoplasmosis in mice induced by recombinant rhoptry proteins encapsulated in poly (lactide-co-glycolide) microparticles. *Parasitol. Res.* 114, 4195–4203. doi: 10.1007/s00436-015-4652-3
- Xue, J., Jiang, W., Chen, Y., Gong, F., Wang, M., Zeng, P., et al. (2017). Thioredoxin reductase from *Toxoplasma gondii*: an essential virulence effector with antioxidant function. *FASEB J.* doi: 10.1096/fj.201700008R. [Epub ahead of print].

- Yan, C., Liang, L. J., Zhang, B. B., Lou, Z. L., Zhang, H. F., Shen, X., et al. (2014). Prevalence and genotyping of *Toxoplasma gondii* in naturally-infected synanthropic rats (*Rattus norvegicus*) and mice (*Mus musculus*) in eastern China. *Parasit. Vectors* 7:591. doi: 10.1186/s13071-014-0591-6
- Yang, C. P., and Wan, H. J. (2006). Research progress of artemisinin and its derivatives against *Toxoplasma gondii*. Int. J. Med. Parasit. Dis. 33, 160–162. doi: 10.3760/cma.j.issn.1673-4122.2006.03.012. (Translated from Chinese).
- Yang, N., Mu, M., Li, H., Hu, J., Gao, W., Yang, S., et al. (2013). Seroprevalence of *Toxoplasma gondii* infection in pet dogs in Shenyang, northeastern China. J. Parasitol. 99, 176–177. doi: 10.1645/GE-3211.1
- Yang, Q. L., Zhang, R. S., Wu, H. P., Zhang, Y. K., and Wang, K. G. (2008). The detection of *Toxoplasma gondii* with loop-mediated isothermal amplification technology. *Chin. J. Parasitol. Parasit. Dis.* 26, 304–306. doi: 10.3969/j.issn.1000-7423.2008.04.014. (Translated from Chinese).
- Yang, Y. B., Li, S. Q., Du, T., Liang, H. S., and Chen, Q. L. (2014). Retrospective analysis of TORCH of 2642 infertile women in Guangzhou area. J. Trop. Med. 14, 1453–1455. (Translated from Chinese).
- Yang, Y., Feng, Y., Yao, Q., Wang, Y., Lu, Y., Liang, H., et al. (2017). Seroprevalence, Isolation, Genotyping, and Pathogenicity of *Toxoplasma gondii* Strains from Sheep in China. *Front. Microbiol.* 8:136. doi: 10.3389/fmicb.2017.00136
- Yang, Y., Ying, Y., Verma, S. K., Cassinelli, A. B., Kwok, O. C., Liang, H., et al. (2015). Isolation and genetic characterization of viable *Toxoplasma gondii* from tissues and feces of cats from the central region of China. *Vet. Parasitol.* 211, 283–288. doi: 10.1016/j.vetpar.2015.05.006
- Yu, E. S. (2000). *China Toxoplasmosis*. Hong Kong: Asian Medicine Press. 1–11. (Translated from Chinese).
- Yu, E. S. (2001). Evaluation and suggestion on domestic commercial available kits for the diagnosis of toxoplasmosis. *Chin. J. Zoonoses* 17, 5–6. doi: 10.3969/j.issn.1002-2694.2001.03.001. (Translated from Chinese).
- Yu, E. S., Chen, T. S., and Lin, S. C. (1957). The occurrence of Toxoplasma in cats and rabbits in Fukien, China. *Acta Microbiol. Sin.* 5, 101–110. (Translated from Chinese).
- Yu, H. J., Zhang, Z., Liu, Z., Qu, D. F., Zhang, D. F., Zhang, H. L., et al. (2011). Seroprevalence of *Toxoplasma gondii* infection in pigs, in Zhejiang Province, China. J. Parasitol. 97, 748–749. doi: 10.1645/GE-2713.1
- Yu, J. H., Liu, Q., and Xia, Z. F. (2006). Epidemiological investigations of infections with *Neospora caninum* and *Toxoplasma gondii* in cattle and water buffaloes. *Vet. Sci. China* 36, 247–251. doi: 10.3969/j.issn.1673-4696.2006. 03.017. (Translated from Chinese).
- Yu, P. L., Chen, J. S., Zhang, H. X., Zhang, C., Wang, C. X., and Xu, M. X. (2007). The serological survy of Toxoplasma infection in Wuhan city. *Chin. J. Zoonoses* 23, 393–394. doi: 10.3969/j.issn.1002-2694.2007.04.023. (Translated from Chinese).
- Yu, S. H., Xu, L. Q., Jiang, Z. X., Xu, S. H., Han, J. J., Zhu, Y. G., et al. (1994). Report on the first national wide survey of the distribution of human parasites in China I: regional distribution of parasite species. *Chin. J. Parasitol. Parasite Dis.* 12, 241–247. (Translated from Chinese).
- Yuan, Z. G., Luo, S. J., Dubey, J. P., Zhou, D. H., Zhu, Y. P., He, Y., et al. (2013a). Serological evidence of *Toxoplasma gondii* infection in five species of bats in China. *Vector Borne Zoonotic Dis.* 13, 422–424. doi: 10.1089/vbz.2012.1091
- Yuan, Z. G., Ren, D., Zhou, D. H., Zhang, X. X., Petersen, E., Li, X. Z., et al. (2013b). Evaluation of protective effect of pVAX-TgMIC13 plasmid against acute and chronic *Toxoplasma gondii* infection in a murine model. *Vaccine* 31, 3135–3139. doi: 10.1016/j.vaccine.2013.05.040
- Yuan, Z. G., Zhang, X. X., He, X. H., Petersen, E., Zhou, D. H., He, Y., et al. (2011a). Protective immunity induced by *Toxoplasma gondii* rhoptry protein 16 against toxoplasmosis in mice. *Clin. Vaccine Immunol.* 18, 119–124. doi: 10.1128/CVI.00312-10
- Yuan, Z. G., Zhang, X. X., Lin, R. Q., Petersen, E., He, S., Yu, M., et al. (2011b). Protective effect against toxoplasmosis in mice induced by DNA immunization with gene encoding *Toxoplasma gondii* ROP18. *Vaccine* 29, 6614–6619. doi: 10.1016/j.vaccine.2011.06.110
- Zhang, A. M., Shen, Q., Li, M., Xu, X. C., Chen, H., Cai, Y. H., et al. (2013). Comparative studies of macrophage-biased responses in mice to infection with *Toxoplasma gondii* ToxoDB #9 strains of different virulence isolated from China. *Parasit. Vectors* 6:308. doi: 10.1186/1756-3305-6-308
- Zhang, D., Wang, Z., Fang, R., Nie, H., Feng, H., Zhou, Y., et al. (2010). Use of protein AG in an enzyme-linked immunosorbent assay for serodiagnosis of

*Toxoplasma gondii* infection in four species of animals. *Clin. Vaccine Immunol.* 17, 485–486. doi: 10.1128/CVI.00479-09

- Zhang, H., Zhou, D. H., Zhou, P., Lun, Z. R., Chen, X. G., Lin, R. Q., et al. (2009). Seroprevalence of *Toxoplasma gondii* infection in stray and household cats in Guangzhou, China. *Zoonoses Public Health* 56, 502–505. doi: 10.1111/j.1863-2378.2008.01209.x
- Zhang, J., He, S., Jiang, H., Yang, T., Cong, H., Zhou, H., et al. (2007). Evaluation of the immune response induced by multiantigenic DNA vaccine encoding SAG1 and ROP2 of *Toxoplasma gondii* and the adjuvant properties of murine interleukin-12 plasmid in BALB/c mice. *Parasitol. Res.* 101, 331–338. doi: 10.1007/s00436-007-0465-3
- Zhang, M., Zhao, L., Song, J., Li, Y., Zhao, Q., He, S., et al. (2013). DNA vaccine encoding the *Toxoplasma gondii* bradyzoite-specific surface antigens SAG2CDX protect BALB/c mice against type II parasite infection. *Vaccine* 31, 4536–4540. doi: 10.1016/j.vaccine.2013.07.065
- Zhang, N., Wang, S., Wang, D., Li, C., Zhang, Z., Yao, Z., et al. (2016). Seroprevalence of *Toxoplasma gondii* infection and risk factors in domestic sheep in Henan province, central China. *Parasite* 23:53. doi: 10.1051/parasite/2016064
- Zhang, S. Y., Jiang, S. F., He, Y. Y., Pan, C. E., Zhu, M., and Wei, M. X. (2004). Serologic prevalence of *Toxoplasma gondii* in field mice, Microtus fortis, from Yuanjiang, Hunan Province, People's Republic of China. *J. Parasitol.* 90, 437–438. doi: 10.1645/GE-168R
- Zhang, X. X., Huang, S. Y., Zhang, Y. G., Zhang, Y., Zhu, X. Q., and Liu, Q. (2014). First report of genotyping of *Toxoplasma gondii* in free-living Microtus fortis in northeastern China. J. Parasitol. 100, 692–694. doi: 10.1645/13-381.1
- Zhang, X. X., Lou, Z. Z., Huang, S. Y., Zhou, D. H., Jia, W. Z., Su, C., et al. (2013). Genetic characterization of *Toxoplasma gondii* from Qinghai vole, Plateau pika and Tibetan ground-tit on the Qinghai-Tibet Plateau, China. *Parasit. Vectors* 6:291. doi: 10.1186/1756-3305-6-291
- Zhang, X., Jin, L., Cui, Z., Zhang, C., Wu, X., Park, H., et al. (2016). Antiparasitic effects of oxymatrine and matrine against *Toxoplasma gondii in vitro* and *in vivo*. *Exp. Parasitol*. 165, 95–102. doi: 10.1016/j.exppara.2016. 03.020
- Zhang, Y. B., Cong, W., Li, Z. T., Bi, X. G., Xian, Y., Wang, Y. H., et al. (2015). Seroprevalence of *Toxoplasma gondii* infection in patients of intensive care unit in China: a hospital based study. *Biomed Res. Int.* 2015:908217. doi: 10.1155/2015/908217
- Zhang, Y., Xu, D., Cao, L., Gao, Y., Xia, X., Zhang, Z., et al. (2013). High prevalence of *Toxoplasma gondii* infection in *Microtus fortis* by seminested PCR from Jilin Province, Northeastern China. *J. Parasitol.* 99, 580–582. doi: 10.1645/GE-3195.1
- Zhao, G., Shen, B., Xie, Q., Xu, L. X., Yan, R. F., Song, X. K., et al. (2012). Detection of *Toxoplasma gondii* in free-range chickens in China based on circulating antigens and antibodies. *Vet. Parasitol.* 185, 72–77. doi: 10.1016/j.vetpar.2011.10.031
- Zhao, G., Zhou, A., Lu, G., Meng, M., Sun, M., Bai, Y., et al. (2013). Identification and characterization of *Toxoplasma gondii* aspartic protease 1 as a novel vaccine candidate against toxoplasmosis. *Parasit. Vectors* 6:175. doi: 10.1186/1756-3305-6-175
- Zheng, B., Lu, S., Tong, Q., Kong, Q., and Lou, D. (2013). The virulencerelated rhoptry protein 5 (ROP5) of *Toxoplasma gondii* is a novel vaccine candidate against toxoplasmosis in mice. *Vaccine* 31, 4578–4584. doi: 10.1016/j.vaccine.2013.07.058
- Zheng, J., Jia, H., and Zheng, Y. (2015). Knockout of leucine aminopeptidase in *Toxoplasma gondii* using CRISPR/Cas9. Int. J. Parasitol. 45, 141–148. doi: 10.1016/j.ijpara.2014.09.003
- Zheng, L., Hu, Y., Hua, Q., Luo, F., Xie, G., Li, X., et al. (2017). Protective immune response in mice induced by a suicidal DNA vaccine encoding NTPase-II gene of *Toxoplasma gondii*. Acta Trop. 166, 336–342. doi: 10.1016/j.actatropica.2016.12.004
- Zheng, W. B., Zhang, X. X., Ma, J. G., Li, F. C., Zhao, Q., Huang, S. Y., et al. (2016). Molecular Detection and Genetic Characterization of *Toxoplasma gondii* in Farmed Minks (Neovison vison) in Northern China by PCR-RFLP. *PLoS ONE* 11:e0165308. doi: 10.1371/journal.pone.0165308
- Zhou, C. X., Zhou, D. H., Liu, G. X., Suo, X., and Zhu, X. Q. (2016a). Transcriptomic analysis of porcine PBMCs infected with *Toxoplasma gondii* RH strain. *Acta Trop.* 154, 82–88. doi: 10.1016/j.actatropica.2015.11.009

- Zhou, C. X., Zhu, X. Q., Elsheikha, H. M., He, S., Li, Q., Zhou, D. H., et al. (2016b). Global iTRAQ-based proteomic profiling of *Toxoplasma gondii* oocysts during sporulation. *J. Proteomics* 148, 12–19. doi: 10.1016/j.jprot.2016. 07.010
- Zhou, P., Chen, N., Zhang, R. L., Lin, R. Q., and Zhu, X. Q. (2008). Foodborne parasitic zoonoses in China: perspective for control. *Trends Parasitol.* 24, 190–196. doi: 10.1016/j.pt.2008.01.001
- Zhou, P., Chen, Z., Li, H. L., Zheng, H., He, S., Lin, R. Q., et al. (2011a). *Toxoplasma gondii* infection in humans in China. *Parasit. Vectors* 4:165. doi: 10.1186/1756-3305-4-165
- Zhou, P., Nie, H., Zhang, L. X., Wang, H. Y., Yin, C. C., Su, C., et al. (2010). Genetic characterization of *Toxoplasma gondii* isolates from pigs in China. J. Parasitol. 96, 1027–1029. doi: 10.1645/GE-2465.1
- Zhou, P., Sun, X. T., Yin, C. C., Yang, J. F., Yuan, Z. G., Yan, H. K., et al. (2011b). Genetic characterization of *Toxoplasma gondii* isolates from pigs in southwestern China. J. Parasitol. 97, 1193–1195. doi: 10.1645/GE-2851.1
- Zhou, P., Zhang, H., Lin, R. Q., Zhang, D. L., Song, H. Q., Su, C., et al. (2009). Genetic characterization of *Toxoplasma gondii* isolates from China. *Parasitol. Int.* 58, 193–195. doi: 10.1016/j.parint.2009.01.006
- Zhou, Y. H., Lu, Y. J., Wang, R. B., Song, L. M., Shi, F., Gao, Q. F., et al. (2002). Survey on infection of *Toxoplasma gondii* in infertile couples in Suzhou countryside. *National J. Androl.* 8, 350–352. doi: 10.3969/j.issn.1009-3591.2002. 05.012. (Translated from Chinese).

- Zhou, Y., Zhang, H., Cao, J., Gong, H., and Zhou, J. (2013). Isolation and genotyping of *Toxoplasma gondii* from domestic rabbits in China to reveal the prevalence of type III strains. *Vet. Parasitol.* 193, 270–276. doi: 10.1016/j.vetpar.2012.11.031
- Zhu, Z., Sun, Y. Y., Li, K., Zhou, Z. Y., and Wang, Z. Y. (2017). Research progress on the infection of *Toxoplasma gondii* and its influencing factors in cattle and sheep in China. *Prog. Vet. Med.* 38, 107–110. (Translated from Chinese).
- Zou, F., Yu, X., Yang, Y., Hu, S., Chang, H., Yang, J., et al. (2015). Seroprevalence and risk factors of *Toxoplasma gondii* infection in buffaloes, sheep and goats in yunnan province, Southwestern China. *Iran. J. Parasitol.* 10, 648–651.

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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