#### PROKARYOTES



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# Complete Genome Sequence of Dehalobacterium formicoaceticum Strain DMC, a Strictly Anaerobic Dichloromethane-Degrading Bacterium

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**ABSTRACT** Dehalobacterium formicoaceticum utilizes dichloromethane as the sole energy source in defined anoxic bicarbonate-buffered mineral salt medium. The products are formate, acetate, inorganic chloride, and biomass. The bacterium's genome was sequenced using PacBio, assembled, and annotated. The complete genome consists of one 3.77-Mb circular chromosome harboring 3,935 predicted protein-encoding genes.

Dichloromethane (DCM) is both naturally occurring (1) and synthesized by industry. Whereas aerobic DCM degradation has been studied in detail (2–4), DCM degradation under anoxic conditions is unclear (5–9). *Dehalobacterium formicoaceticum* is the only published isolate utilizing DCM as the sole energy source under anoxic conditions (7). *D. formicoaceticum* is a strictly anaerobic Gram-positive rod-shaped spore-forming bacterium affiliated with the *Peptococcaceae* family (7, 10). Previous physiological and biochemical studies suggested that DCM metabolism involves initial dechlorination and the formation of methylene tetrahydrofolate, which is funneled into the Wood-Ljungdahl pathway (11). The draft genome of *"Candidatus* Dichloromethanomonas elyunquensis," identified as the DCM degrader in an anaerobic consortium, has been published (12), and here we report genomic information for axenic *D. formicoaceticum*.

D. formicoaceticum was obtained from the American Type Culture Collection (ATCC 700118) and cultivated in defined anoxic bicarbonate-buffered mineral salt medium (13, 14) containing DCM as the sole energy source. Genomic DNA was isolated using the cetyltrimethylammonium bromide method (15). The long-insert library for sequencing on the RS II platform (Pacific Biosciences, Menlo Park, CA, USA) was prepared by shearing DNA with a g-TUBE (Covaris, Woburn, MA, USA), targeting an average fragment size of 20 kb. The SMRTbell template preparation kit (Pacific Biosciences) was used to ligate hair-pin adapters to the fragmented DNA. The final library was size selected (BluePippin, Sage Science, Beverly, MA, USA) and sequenced on a single SMRT cell using PacBio P6-C4 chemistry with one 240-min movie. PacBio raw data were error corrected and assembled using the HGAP (SMRT Analysis version 2.3.0), Canu version 1.2 (16), and Celera version 8.2 assemblers with default parameters for bacterial genome assembly. The resulting assemblies were assessed for inconsistencies and misassembly using NUCmer version 3.0 whole-genome alignments (17) and Circleator plots (GC-skew) (18). Canu version 1.2 (16) generated a single contig representing the chromosome, which was polished using Quiver (SMRT Analysis version 2.3.0) to generate the final consensus genome sequence. The IGS prokaryotic annotation pipeline

Received 20 July 2017 Accepted 26 July 2017 Published 14 September 2017

**Citation** Chen G, Murdoch RW, Mack EE, Seger ES, Löffler FE. 2017. Complete genome sequence of *Dehalobacterium formicoaceticum* strain DMC, a strictly anaerobic dichloromethane-degrading bacterium. Genome Announc 5:e00897-17. https://doi .org/10.1128/genomeA.00897-17.

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was used for coding gene prediction and functional annotation (19). The output annotations were loaded into a MySQL Chado relational database and accessed through the visualization tool Manatee (http://manatee.sourceforge.net). Protein-coding genes were predicted using Glimmer version 3 (20), and noncoding RNA genes were predicted using tRNAscan-SE (21) and RNAmmer version 1.2 (22).

The complete genome of *D. formicoaceticum* comprises one circular chromosome (3,766,545 bp) with an overall G+C content of 43.17%. A total of 3,935 predicted protein-encoding genes were identified in the genome. In total, 55 tRNAs and 17 rRNAs were identified, including six 5S rRNAs, five 16S rRNAs, and six 23S rRNAs. The genome harbors genes encoding all enzymes involved in the Wood-Ljungdahl pathway with a featured core acetyl coenzyme A synthase (*acs*) gene cluster (23). Genes encoding *c*-type cytochromes (7 genes), a complete NADH:ubiquinone oxidoreductase (Nuo) complex (11 genes), and one  $F_1F_0$ -ATPase (11 genes) were identified, suggesting chemiosmotic energy conservation. A complete set of sporulation genes is consistent with microscopic observations of spores (7). Genes encoding reductive dehalogenases were not found.

**Accession number(s).** The complete genome sequence of *D. formicoaceticum* has been deposited in GenBank under accession no. CP022121.

# **ACKNOWLEDGMENTS**

Sequencing, genome assembly, and functional annotation were performed by the Institute for Genome Sciences, University of Maryland School of Medicine. This work was supported by the Chemours Company.

#### REFERENCES

- 1. Gribble GW. 2010. Naturally occurring organohalogen compounds—a comprehensive update, p 12–13. *In* Progress in the chemistry of organic natural products, vol 91. Springer, Vienna, Austria.
- Brunner W, Staub D, Leisinger T. 1980. Bacterial degradation of dichloromethane. Appl Environ Microbiol 40:950–958.
- Kohler-Staub D, Leisinger T. 1985. Dichloromethane dehalogenase of Hyphomicrobium sp. strain DM2. J Bacteriol 162:676–681.
- Leisinger T, Braus-Stromeyer SA. 1995. Bacterial growth with chlorinated methanes. Environ Health Perspect 103(suppl 5):33–36.
- Freedman DL, Gossett JM. 1991. Biodegradation of dichloromethane and its utilization as a growth substrate under methanogenic conditions. Appl Environ Microbiol 57:2847–2857.
- Stromeyer SA, Winkelbauer W, Kohler H, Cook AM, Leisinger T. 1991. Dichloromethane utilized by an anaerobic mixed culture: acetogenesis and methanogenesis. Biodegradation 2:129–137. https://doi.org/10 .1007/BF00114603.
- Mägli A, Wendt M, Leisinger T. 1996. Isolation and characterization of Dehalobacterium formicoaceticum gen. nov. sp. nov., a strictly anaerobic bacterium utilizing dichloromethane as source of carbon and energy. Arch Microbiol 166:101–108. https://doi.org/10.1007/s002030050362.
- Justicia-Leon SD, Ritalahti KM, Mack EE, Löffler FE. 2012. Dichloromethane fermentation by a *Dehalobacter* sp. in an enrichment culture derived from pristine river sediment. Appl Environ Microbiol 78:1288–1291. https://doi.org/10.1128/AEM.07325-11.
- Kleindienst S, Higgins SA, Tsementzi D, Chen G, Konstantinidis KT, Mack EE, Löffler FE. 2017. 'Candidatus Dichloromethanomonas elyunquensis' gen. nov., sp. nov., a dichloromethane-degrading anaerobe of the *Pep-tococcaceae* family. Syst Appl Microbiol 40:150–159. https://doi.org/10 .1016/j.syapm.2016.12.001.
- Mägli A, Rainey FA, Leisinger T. 1995. Acetogenesis from dichloromethane by a two-component mixed culture comprising a novel bacterium. Appl Environ Microbiol 61:2943–2949.
- 11. Mägli A, Messmer M, Leisinger T. 1998. Metabolism of dichloromethane by the strict anaerobe *Dehalobacterium formicoaceticum*. Appl Environ Microbiol 64:646–650.
- Kleindienst S, Higgins SA, Tsementzi D, Konstantinidis KT, Mack EE, Löffler FE. 2016. Draft genome sequence of a strictly anaerobic dichloromethanedegrading bacterium. Genome Announc 4(2):e00037-16. https://doi.org/10 .1128/genomeA.00037-16.
- 13. Löffler FE, Sanford RA, Tiedje JM. 1996. Initial characterization of a

reductive dehalogenase from *Desulfitobacterium chlororespirans* Co23. Appl Environ Microbiol 62:3809–3813.

- Löffler FE, Sanford RA, Ritalahti KM. 2005. Enrichment, cultivation, and detection of reductively dechlorinating bacteria, p 77–111. *In* Methods in enzymology, vol 397. Academic Press, New York, NY. https://doi.org/ 10.1016/S0076-6879(05)97005-5.
- Joint Genome Institute. 2012. Bacterial genomic DNA isolation using CTAB. http://jgi.doe.gov/wp-content/uploads/2014/02/JGI-Bacterial-DNA-isolation -CTAB-Protocol-2012.pdf.
- Koren S, Walenz BP, Berlin K, Miller JR, Bergman NH, Phillippy AM. 2017. Canu: scalable and accurate long-read assembly via adaptive k-mer weighting and repeat separation. Genome Res 27:722–736. https://doi .org/10.1101/gr.215087.116.
- Kurtz S, Phillippy A, Delcher AL, Smoot M, Shumway M, Antonescu C, Salzberg SL. 2004. Versatile and open software for comparing large genomes. Genome Biol 5:R12. https://doi.org/10.1186/gb-2004-5-2-r12.
- Crabtree J, Agrawal S, Mahurkar A, Myers GS, Rasko DA, White O. 2014. Circleator: flexible circular visualization of genome-associated data with BioPerl and SVG. Bioinformatics 30:3125–3127. https://doi.org/10.1093/ bioinformatics/btu505.
- Galens K, Orvis J, Daugherty S, Creasy HH, Angiuoli S, White O, Wortman J, Mahurkar A, Giglio MG. 2011. The IGS standard operating procedure for automated prokaryotic annotation. Stand Genomic Sci 4:244–251. https://doi.org/10.4056/sigs.1223234.
- Delcher AL, Bratke KA, Powers EC, Salzberg SL. 2007. Identifying bacterial genes and endosymbiont DNA with Glimmer. Bioinformatics 23: 673–679. https://doi.org/10.1093/bioinformatics/btm009.
- Lowe TM, Eddy SR. 1997. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. Nucleic Acids Res 25:955–964.
- Lagesen K, Hallin P, Rødland EA, Staerfeldt HH, Rognes T, Ussery DW. 2007. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. Nucleic Acids Res 35:3100–3108. https://doi.org/10.1093/nar/ gkm160.
- Pierce E, Xie G, Barabote RD, Saunders E, Han CS, Detter JC, Richardson P, Brettin TS, Das A, Ljungdahl LG, Ragsdale SW. 2008. The complete genome sequence of *Moorella thermoacetica* (f. *Clostridium thermoaceticum*). Environ Microbiol 10:2550–2573. https://doi.org/10.1111/j.1462 -2920.2008.01679.x.