- 1 **Title:** Photoreceptor outer segment reflectivity with ultrahigh resolution visible light
- 2 optical coherence tomography in systemic hydroxychloroquine use
- 3

4 **Authors:** Anupam K. Garg, M.D., Ph.D.^{1,*}; Jingyu Wang, Ph.D.^{1,2,*}, Bailee Alonzo,

- 5 B.S.¹, Ji Yi, Ph.D.^{1,2}, Amir H. Kashani, M.D., Ph.D.^{1,3}
- 6

7 Institutions:

- ⁸ ¹Wilmer Eye Institute, Johns Hopkins University, Baltimore, MD, USA
- ⁹ ²Department of Biomedical Engineering, Johns Hopkins University, Baltimore, MD, USA
- ³Department of Biomedical Engineering, Johns Hopkins Hospital, Baltimore, MD, USA
- 11 *These authors contributed equally.
- 12

13 Corresponding Author:

- 14 Amir H. Kashani, M.D., Ph.D.
- 15 Boone Pickens Professor of Ophthalmology and Biomedical Engineering
- 16 Wilmer Eye Institute
- 17 600 N Wolfe Street
- 18 <u>akashan1@jhmi.edu</u>
- 19
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- 28
- 29 Abbreviations:
- 30 **OCT**, optical coherence tomography; **VIS**, visible light; **SS**, swept source; **SD**, spectral
- 31 domain; HCQ, hydroxychloroquine; ELM, external limiting membrane; EZ, ellipsoid
- 32 zone; **COST**, cone outer segment tips; **ROST**, rod outer segment tips; **RPE**, retinal
- 33 pigment epithelium; **BM**, Bruch's membrane
- 34

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35 Abstract

36 <u>Purpose</u>: To evaluate outer retinal organization in normal subjects and those using

37 hydroxychloroquine (HCQ) with ultrahigh resolution visible light optical coherence

tomography (VIS-OCT).

Methods: Forty eyes of 22 adult subjects were recruited from a tertiary care retina
practice including controls (20 eyes, 12 subjects, mean age 40±22yrs, mean logMAR
BCVA 0.19, 90% female) and subjects with a history of HCQ use (20 eyes, 10 subjects,
mean age 62±17yrs, mean logMAR BCVA 0.03, 67% female). Each subject was
imaged using a custom-built VIS-OCT device (axial resolution 1.3µm) and FDAapproved OCT devices.

45 Results: Using VIS-OCT, control subjects demonstrate 5 and 6 hyperreflective bands in 46 the foveal and parafoveal regions, respectively, between the outer nuclear layer and 47 Bruch's membrane. These bands demonstrate intensity profiles complementary to the known histopathologic distribution of rods and cones. In comparison to controls, 48 49 subjects taking HCQ demonstrate blunting of all bands, particularly bands 2-4. In all 50 cases of suspected or known toxicity, VIS-OCT demonstrated attenuation of band 3i and in no cases was there attenuation of other bands that was more severe than band 51 3i, suggesting that changes in the reflectivity of Band 3i may be the earliest identifiable 52 53 sign of HCQ toxicity.

54 <u>Conclusions</u>: VIS-OCT of the outer retina demonstrates a unique outer retinal banding 55 pattern corresponding to photoreceptor density profiles. There is a notable attenuation 56 of the photoreceptor outer segment reflectivity profile associated with early HCQ

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- 57 toxicity. This finding may be an early, and possibly reversible, sign of HCQ toxicity,
- 58 primarily impacting the cones.

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59 Introduction

Hydroxychloroquine (HCQ) is a well-described anti-inflammatory and anti-malarial 60 61 medication with relatively rare but devastating toxic effects on the retina¹. Although a 62 benign drug in general, the potential for this serious side-effect causes significant 63 problems in the management of patients who are on the medication. The exact 64 pathophysiology of the side-effect is not well understood however several lines of 65 evidence suggest that toxicity at the level of the retinal pigment epithelium and, 66 secondarily the photoreceptor, is likely. In vitro studies have demonstrated that HCQ 67 inhibits protein synthesis² and lysosomal function³ in RPE. Toxicity manifests in 68 subjects with prolonged HCQ exposure as thinning of the outer retina and loss of the retinal pigment epithelium in a bulls-eye pattern⁴. However, even subjects without any 69 70 clinical findings or subjective symptoms can demonstrate subclinical signs of toxicity such as modest retinal thinning on OCT and decreased signal amplitude on mfERG^{5,6}. 71 72 Careful analysis of OCT images has suggested these changes are caused by loss of 73 the ellipsoid zone⁷ and/or changes in the photoreceptor outer segments⁸. While severe changes may be seen with retinal thickness maps⁹, the resolution of commercially 74 75 available OCT devices is not sufficient to easily and reliably assess subclinical changes in the outer segment banding patterns on single OCT volumes¹⁰. To overcome this 76 77 limitation, serial OCT imaging has been used to detect very subtle paracentral thinning 78 patterns that seem strongly correlated with toxicity⁶. This finding suggests that higher 79 resolution imaging of the photoreceptor/RPE complex may reveal even earlier signs of 80 toxicity.

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82 While most commercial devices utilize near-IR wavelength light, ultrahigh resolution visible light OCT (VIS-OCT) utilizes shorter wavelengths of light to enable higher axial 83 resolution (~1 µm) compared to conventional OCT devices¹¹. Recent technological 84 advances have enabled the development of VIS-OCT devices for human retinal 85 imaging, enabling characterization of both inner and outer retinal layers in greater detail 86 than previously possible in human subjects^{12–15} and animal models^{16,17}. Prior work has 87 88 demonstrated that VIS-OCT reveals outer retinal banding morphology that was not readily visible with conventional devices in control subjects without a known history of 89 90 retinal pathology, including Bruch's membrane and sub-bands in photoreceptor outer 91 segments¹⁸. Similar banding morphology has also been recently demonstrated using ultrahigh resolution SD-OCT prototype devices^{19,20}. *In vivo* characterization of these fine 92 93 details and their changes in retinal disease may provide valuable insights into disease pathophysiology and management. Given that the outer retina is primarily affected in 94 95 early HCQ toxicity, VIS-OCT is a promising imaging modality to enable early detection 96 of HCQ toxicity. In this study we used a custom-built VIS-OCT device to identify subtle outer retinal changes in HCQ toxicity not visible with conventional OCT devices. 97

98

99 Methods

100 *Recruitment of Subjects*

Both eyes of adult subjects with a history of hydroxychloroquine use were prospectively recruited from a tertiary care retina practice for retinal imaging with a custom-built dualchannel VIS-OCT device with an axial resolution of 1.3µm (see Wang et al., 2022 for technical details^{18,21}) (**Figure 1**). Each subject was also imaged using a commercial

| 105 | swept-source OCT imaging device. The most recent hepatic and renal function testing |
|-----|--|
| 106 | results of each subject were also obtained. Control subjects were recruited from the |
| 107 | retina clinics if they had no vision threatening retinal or ocular disease in at least one |
| 108 | eye. Information regarding subjects is summarized in Table 1 . All subjects provided |
| 109 | informed consent according to a human subject protocol approved by the Johns |
| 110 | Hopkins Medicine Institutional Review Boards (IRB) and in accordance with the |
| 111 | principles of the Declaration of Helsinki. Subjects with significant media opacity, poor |
| 112 | signal quality, or inability to fixate sufficiently to obtain at least one high quality foveal |
| 113 | line scan were excluded. |
| 114 | |
| 115 | Imaging Device |
| 116 | The dual-channel VIS-OCT features a visible-light bandwidth ranging from 500 to 650 |
| 117 | nm and a near-infrared light bandwidth spanning from 750 nm to 900 nm, with power |

118 levels of 0.25 mW and 0.8 mW on the cornea, respectively. Subjects were instructed to

119 fixate using one of two LED displays that served as an external fixation target. A tunable

120 lens was utilized to correct spherical errors. For each eye, the image was initially

121 aligned and optimized under the NIR channel and then promptly imaged with the visible122 light channel.

123

124 Imaging Protocol

All subjects had standard SD-OCT imaging performed as part of their standard of care
 assessment. In addition, subjects were scanned using two research OCT devices
 including a custom-built ultrahigh resolution visible light OCT (VIS-OCT) system and a

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| 128 | commercially available SS-OCT system (PlexElite, Carl Zeiss Meditec, Dublin, CA. |
|-----|---|
| 129 | Imaging on the SS-OCT was performed using a standard 6mm x 6mm raster scan |
| 130 | pattern centered on the fovea. The VIS-OCT imaging was performed using a high- |
| 131 | definition 4-line radial scanning pattern (32x2048 A-lines x 4 B-scans) as well as |
| 132 | multiple high-definition single line scans (64x1024 A-lines x 1 B-scan) in the region of |
| 133 | the raster scan pattern from the SS- and SD-OCT devices. For all VIS-OCT scans, 32 |
| 134 | or 64 modulated A-lines over ~0.1 mm orthogonal to the B-scan direction were |
| 135 | averaged to produce high-quality images with reduced speckle noise ^{21,22} . The line rate |
| 136 | was 100kHz with a total acquisition time of 2.62 seconds per scan pattern. Images |
| 137 | were flattened using a custom algorithm. VIS-OCT and SS-OCT foveal scans were |
| 138 | manually registered by using the depth of the foveal center. |

139

140 Image Processing for VIS-OCT

141 We employed a per-A-line noise cancellation algorithm to eliminate baseline light source 142 spectrum, reduce noise, and enhance the signal-to-noise ratio. Dispersion 143 compensation and fast Fourier transform (FFT) were then applied to generate B-scans. 144 To establish the display range, we defined the intensity of background after FFT as the 145 lower boundary and selected the intensity value at 0.05% from the sorted values (from high to low) as the upper boundary. Utilizing these upper and lower boundaries, we 146 147 applied a logarithmic scale and normalization for image display. Given the substantial 148 dynamic range (exceeding 60 dB for control subjects) between the inner and outer 149 retina, achieving a balanced visual effect proved challenging. Therefore, we increased 150 the image brightness by 40% to enhance the visibility of the inner retina, although this

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resulted in saturation of the outer retina. Additionally, we manually flattened the B-scans based on band 5 (Bruch's membrane) **(Supplemental Figure 1**). The intensity of each outer retinal layer was quantified by manually segmenting the boundaries of each layer and calculating the cumulative intensity between the boundaries. Manual segmentation was performed independently by two separate graders and differences were adjudicated by a third grader.

157

158 **Results**

159 As described in Table 1, a total of 20 eyes from 12 control subjects (without known 160 retinal pathology) were recruited into this study. VIS-OCT imaging of these subjects 161 demonstrates outer retinal anatomy in finer detail than previously possible with 162 conventional OCT technology. Figure 1 and Supplemental Figure 2 demonstrate representative VIS-OCT images from control subjects. Within the foveola, five hyper-163 164 reflective outer retinal bands are visualized, labeled as bands 1-5 (**Figure 1C**). These 165 bands are putatively identified as the external limiting membrane (ELM, band 1), 166 ellipsoid zone (EZ, band 2), cone photoreceptor outer segment tips (COST, band 3), 167 retinal pigment epithelium (RPE, band 4) and Bruch's Membrane (BM, band 5). This banding pattern was observed in all control subjects irrespective of patient age. Henle's 168 fiber layer was visible as a slightly darker region above the outer nuclear layer (Figure 169 170 **1B**, **1E**). Magnified views of the inner plexiform layer demonstrate sublayers in the 171 perifoveal region (Figure 1B and 1F), consistent with previous findings²³.

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173 In the parafoveal region, outer retinal band 3 consistently divides into two distinct 174 hyperreflective bands (labelled bands 3 inner (3i) and 3 outer (3o)) for a total of six 175 hyperreflective bands (Figure 1A and 1D). In accordance with prior anatomical studies 176 and previously published VIS-OCT imaging data^{13,21}, these bands are putatively 177 identified as the cone and rod photoreceptor outer segment tips (COST/ROST), 178 respectively. To better characterize these bands, each one was manually segmented, 179 and the mean intensity of each band averaged across three control subjects was plotted 180 as a function of the distance from the fovea (**Figure 2**). Notably, the intensity of band 3i 181 peaks in the foveal region and sharply declines with increasing retinal eccentricity. On 182 the other hand, band 3o is not visible in the foveal region and gradually appears with 183 increasing retinal eccentricity. This pattern mirrors previously published histological studies of cone and rod density^{24,25}, supporting the hypothesis that bands 3i and 3o 184 represent cone and rod photoreceptor outer segments tips, respectively. This inverse 185 186 relationship between bands 3i and 3o was consistent between two graders who 187 independently segmented the layers (Supplemental Figure 3). 188 189 A total of 20 eyes from 10 subjects with a history of hydroxychloroquine use were 190 recruited (**Table 1**). These subjects demonstrated a range in severity of 191 hydroxychloroquine toxicity findings (including subjects with suspected toxicity or no 192 evidence of toxicity on prior testing) and were imaged using VIS-OCT. These subjects 193 range in age (33 to 84 years) and length of hydroxychloroguine use (1 to 35 years). The 194 approximate cumulative lifetime hydroxychloroguine dose was calculated, and each

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patient's most recent renal and hepatic function testing was recorded if available fromtheir medical records.

197

198 **Figure 3** illustrates data from a 61-65 age range female subject with an 11-year history 199 of hydroxychloroguine use and gualitatively unremarkable SD-OCT and SS-OCT scans 200 (Figure 3B). The patient had excellent visual acuity (20/20 in both eyes) and denied any subjective vision changes at the time of examination. Hydroxychloroguine toxicity was 201 suspected based on serial SD-OCT mean central subfield thickness measurements 202 203 showing decreasing thickness in the nasal and temporal ETDRS subfield sectors as 204 described in Melles et al., 2022 (Figure 3B, 3F). VIS-OCT images (Figure 3A, 3C-D) were compared to a control subject (Figure 3E) and demonstrated blunting of most 205 206 band boundaries as well as essentially complete loss of Band 3i in the parafoveal region. Figure 3G illustrates the average intensity profile of the outer retina in the nasal 207 208 parafovea from 3mm to 6mm eccentricity, corresponding to the outer parafoveal ETDRS 209 subfield (indicated with vertical white dashed lines in **Figure 3A**). There is a marked 210 decrease in the pixel intensity of regions corresponding to band 3i and 3o (red line) 211 relative to the average of three control subjects (black line) as well as in comparison to 212 the intensity of bands 4 and 5, suggestive of damage to the photoreceptor bands. This decrease in band reflectivity (~5x decrease) is much larger proportionately than the 213 214 gualitative decrease in the thickness from ELM to Bruch's membrane illustrated in 215 Figure 3G. Also, the mean retinal thickness from the same 3mm to 6mm nasal 216 parafoveal ETDRS subfield is within the normal range relative to a normative database 217 (279 µm in **Figure 3F**).

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218

219 Figure 4 illustrates data from a 71-75 age range female subject with a 5-year history of 220 hydroxychloroguine use and excellent best corrected visual acuity of 20/25 in the right 221 eye and 20/20 in the left eye. She denied any subjective vision changes at the time of 222 the examination. SD and SS-OCT images suggested a very mild parafoveal attenuation 223 of the external limiting membrane and/or ellipsoid zone. Serial SD-OCT central subfield 224 thickness assessments suggested thinning in the nasal and temporal parafoveal 225 ETDRS subfields as described in Melles et al., 2022 (Figure 4F). These findings were 226 concerning for early hydroxychloroguine toxicity and the patient was closely monitored 227 while encouraging minimizing the dose of hydroxychloroguine. VIS-OCT imaging 228 demonstrates diffuse attenuation of band 3i and patchy parafoveal attenuation of band 229 30, which were not clearly apparent on individual b-scans scans in commercially available devices (Figure 4A versus 4B) nor in control subjects (Figure 4C,D 230 231 compared to Figure 4E). Similar to the subject demonstrated in **Figure 3**, there was a 232 marked decrease in the intensity of the photoreceptor bands (particularly bands 3i and 233 30) relative to the intensity of bands 4 and 5 (Figure 4G). This decrease in band 234 reflectivity was present despite of any qualitative decrease in the thickness from ELM to 235 Bruch's membrane, as illustrated in Figure 4G. The subject ultimately stopped HCQ due 236 to the significant concern for early toxicity.

237

238 VIS-OCT imaging of subjects with symptomatic and severe HCQ retinal toxicity

demonstrated marked parafoveal attenuation of bands 2, 3i, 3o, and 4 (Figure 5A).

240 Given the severity of damage, this attenuation was also evident with SS-OCT (Figure

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241 **5B**) and with a marked decrease in SD-OCT central subfield retinal thickness as well 242 (Figure 5F). At the time of imaging, the patient's best-corrected visual acuity was 20/63 243 in the right eye and 20/80 in the left eye and the patient had been previously diagnosed 244 with severe HCQ toxicity. The medication had been discontinued prior to our 245 examination. Notably, even in this subject, there was more severe attenuation of band 246 3i compared to any other band with increasing eccentricity beyond the parafovea where 247 clear atrophy was not evident. While all bands were less distinct than normal, bands 1, 2 and 30 were particularly diffuse when compared with a control subject (Figure 5C, D 248 249 compared for E). In no case was the attenuation of any band more severe than the 250 attenuation of Band 3i (Figure 5G). As in the previous case, this decrease in band 251 reflectivity was present despite of any qualitative decrease in the thickness from ELM to 252 Bruch's membrane, as illustrated in Figure 5G.

253

254 Discussion

255 Using ultrahigh resolution visible light OCT, we demonstrate outer retinal banding 256 patterns that are not clearly or reliably visible with commercially available SD or SS-257 OCT. Most importantly, these changes in banding pattern reflectivity are much larger in magnitude than changes in retinal thickness measured qualitatively or quantitatively on 258 259 SD-OCT or SS-OCT. We also demonstrate that the outer retinal band intensity profiles 260 on VIS-OCT in healthy controls are similar to the known density profiles of rods and 261 cones from histologic studies. In control subjects, we consistently identify five outer retinal bands in the foveola (Bands 1, 2, 3, 4, and 5) and six bands in the parafovea 262 263 (Bands 1, 2, 3i, 3o, 4, and 5). These bands putatively represent (1) the external limiting

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264 membrane, (2) the ellipsoid zone, (3i) the cone outer segment tips, (3o) the rod outer 265 segment tips, (4) the retinal pigment epithelium and (5) Bruch's membrane. These data 266 strongly suggest that VIS-OCT imaging can distinguish rod- and cone-specific retinal 267 anatomy non-invasively.

268

269 We demonstrate the utility of ultrahigh resolution VIS-OCT to detect sub-clinical 270 changes in the outer retinal band reflectivity corresponding to photoreceptor outer 271 segments in asymptomatic subjects at high risk of hydroxychloroquine toxicity. 272 Specifically, we observe that Band 3i (corresponding to the putative cone outer segment 273 tips) is consistently and most severely attenuated in subjects at high risk of toxicity and 274 in whom serial SD-OCT measurements from commercial devices demonstrate retinal 275 thinning. Notably, this latter finding of serial thinning has been implicated in HCQ 276 toxicity⁶. We hypothesize that attenuation of Band 3i is the earliest sign of HCQ toxicity 277 and may be readily detectable on a single visit VIS-OCT while serial SD-OCT 278 measurements over months or years are needed to detect decreasing thickness trends. 279 In our individual VIS-OCT scans, the attenuation of VIS-OCT banding reflectivity is 280 present despite normal retinal thickness as measured with central subfield thickness 281 using SD-OCT when compared with age-matched control subjects. We hypothesize that 282 this outer retinal attenuation of band 3i is the earliest known marker of 283 hydroxychloroquine toxicity. Our analyses assessing the cumulative intensity of bands 3i and 3o at increasing retinal eccentricities in **Figure 2** closely align with previously 284 published anatomical studies quantifying cone and rod densities²⁴, supporting the 285

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hypothesis that these bands represent the cone and rod photoreceptor outer segments,respectively.

288

289 The mechanism of hydroxychloroguine retinopathy is not clearly understood, though 290 prior studies have suggested multiple potential mechanisms²⁶. A study by Xu et al. 291 demonstrates that both chloroquine and hydroxychloroquine inhibit organic anion transporting peptide 1A2 (OATP1A2), which mediates uptake of all-trans-retinoic acid in 292 the retinal pigment epithelium in the visual cycle.²⁷ This may lead to toxicity of both 293 294 photoreceptors as well as the retinal pigment epithelium. Animal studies have revealed 295 the binding of chloroquine to pigmented retinal structures, including the retinal pigment epithelium²⁸. It is unclear why band 3i appears to be attenuated first in subjects on HCQ 296 297 followed by band 3o, though this finding suggests a tendency for cone photoreceptors to 298 be preferentially over rods. In more severe stages, as shown in Figure 5, there appears to be attenuation of the remaining outer retinal bands including the retinal pigment 299 300 epithelium, favoring a mechanism of toxicity affecting both the RPE as well as the 301 photoreceptors.

302

Recent work has measured ellipsoid zone attenuation with SD-OCT to detect and quantify HCQ toxicity^{29,30}. Our results demonstrate the ability of VIS-OCT imaging to detect early HCQ toxicity with attenuation of the photoreceptor bands, which are not easily visible with SD-OCT or SS-OCT, prior to EZ attenuation. We demonstrate that early HCQ toxicity is characterized mainly by attenuation of bands 3i and 3o, with more severe toxicity affecting the EZ and the retinal pigment epithelium. This corroborates

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earlier studies that suggest damage occurs to the photoreceptors^{4,28}. As demonstrated
in Figures 3 and 4, early toxicity is often not apparent on individual line scans with SSOCT (or SD-OCT), requiring averaging of retinal thickness across a large region to
observe retinal thinning on serial thickness maps. Using VIS-OCT, these changes are
apparent on foveal line scans, and may assist with earlier diagnosis of HCQ toxicity.
It is important to note that there are limitations to VIS-OCT imaging compared to SS-

OCT and conventional FDA approved devices. First, the brightness of the visible light used to acquire VIS-OCT images can be distracting to patients, particularly when utilizing lengthy imaging protocols or in patients who are light sensitive. Additionally, while its increased resolution allows for visualization of Bruch's membrane, visible light is limited in its ability to penetrate beyond Bruch's membrane and visualize structures within the choroid, which may limit its utility in the diagnosis of choroidal pathologies.

323 Due to our small sample size and our study being limited to subjects with HCQ use and 324 toxicity, it remains unclear whether the pattern of changes described can also be seen 325 in other retinal diseases or whether they are specific to HCQ toxicity. Our findings 326 support previous work suggesting that anatomically detectable damage to photoreceptors precedes similar damage to the RPE^{1,4}. Prospective studies and 327 328 analyses with larger sample sizes will be necessary to further characterize these 329 changes. Additionally, as previously described, the pattern of retinal toxicity and vision 330 loss associated with HCQ use varies in patients of Asian descent, and additional work is 331 necessary to determine whether VIS-OCT can demonstrate this difference or provide

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- further insight into its pathogenesis. Prior work has also demonstrated that in a minority
- 333 of patients, early HCQ toxicity is detected on visual field testing prior to clear SD-OCT
- 334 changes although this is likely due to the limited resolution of single SD-OCT scans to
- reliably demonstrate changes in the outer retinal layers as we have shown above.³¹
- 336 Further work with a larger sample size is necessary to determine whether changes on
- 337 VIS-OCT can reliably be detected prior to visual field changes.

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434 Tables

435 <u>Table 1</u>: Demographic information of study subjects

| Subject | Diagnosis | Age range | Race | Sex | Years HCQ Use | Known Toxicity? | Cumulative HCQ Dose, Renal/Hepatic Function | Visual Acuity |
|---------|------------------|--------------|------|-----|---------------------|--------------------|--|------------------|
| 271 | SLE | 66-70 | WNH | F | 22 | suspected | 3212 gm | 20/20 |
| | | | | | | | Creatine: 0.9 mg/dl | 00 |
| | | | | | | | AST/ALT: 25/19 U/L | |
| 308 | SLE | 66-70 | BNH | F | 27 | no | 4088 gm | 20/20 |
| | | | | | | | Creatine: 0.89 mg/dl | 00 |
| | | | | | | | AST/ALT: 25/17 U/L | |
| 178 | Sjogren's | 51-55 | WNH | F | 4 | yes | 584 gm | 20/50 |
| | Syndrome and SLE | | | | | | Creatine: 0.56 mg/dl | 0D 20/80 OS |
| | | | | | | | AST/ALT: 29/26 u/L | |
| 347 | RA | 76-80 | WNH | F | 31 | suspected | 4526 gm | 20/50 |
| | | | | | | | Creatine: 1.5 mg/dL | 20/40 OS |
| | | | | | | | AST/ALT: 26/19 U/L | |
| 370 | Undifferentiated | 31-35 | WNH | F | 1 | no | 146 gm | 20/20 |
| | disease | | | | | | Creatine: 0.7 mg/dL | 00 |
| | | | | | | | AST/ALT: 17/26 U/L | |
| 374 | SLE | 31-35 | BNH | F | 2 | no | 292 gm | 20/20 |
| | | | | | | | Creatine: 0.7 mg/dL | 00 |
| | | | | | | | AST/ALT: 22/20 U/L | |
| 417 | RA | 76-80 | WNH | F | 35 | yes | unknown cumulative dose | 20/50 OD |
| | | | | | | | Creatine: 0.9 mg/dL AST/ALT: 40/25 U/L | 20/250 OS |
| 439 | SLE | 61-65 | WNH | F | 11 | no | 1204.5 gm | 20/20 OU |
| | | | | | | | Creatine: 1.5 mg/dL | |

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| | | | | | | | AST/ALT: 21/10 U/L | |
|--|--|---------------------------|-----------------------|--------------------|--------------------------|------------------------------------|---|-----------------|
| 115 | PA and | 81.85 | W/NH | M | 5 | no | 730 gm | 20/40 |
| 440 | Sarcoidosis | 01-05 | VVINI I | IVI | 5 | ΠU | | 0D |
| | | | | | | | Creatine: 0.99 | 20/25 OS |
| | | | | | | | AST/ALT: 18/7 U/L | |
| 378 | SLE | 71-75 | BNH | F | 5 | suspected | 730 gm | 20/25 |
| | | | | | | | Creatine: 0.9 mg/dL | 00 |
| | | | | | | | AST/ALT: 19/15 | |
| | | | | | | | U/L | |
| Subject | Diagnosis | Age | Race | Sex | Eye(s) Imaged | | Diagnosis | |
| 451 | Control | 26-30 | BNH | М | | OU | N/A | - |
| 456 | Control | 21-25 | WNH | F | | OU | N/A | 20/20 |
| | | | | | | | | 00 |
| 446 | Control | 21-25 | WNH | F | | OU | N/A | 20/20 OU |
| 464 | Control | 21.25 | | | | 011 | NI/A | |
| 404 | Control | 21-25 | VVINH | Г | 00 | | N/A | - |
| 465 | Control | 21-25 | BNH | F | | OU | N/A | 20/20 OU |
| 402 | Control | 66-70 | WNH | M | | 00 | N/A | 20/20 |
| | | | | | | | | OU |
| 399 | Control | 26-30 | ANH | F | | OD | Optic disc pit OS | 20/20 |
| | | | | | | | | OD |
| 395 | Control | 76-80 | WNH | F | | OU | PVD OU | 20/32 |
| | | | | | | | | 20/25 OS |
| 385 | Control | 26-30 | WH | M | | OU | N/A | - |
| 303 | Control | 61-65 | WNH | F | | | Choroidal nevus | 20/32 |
| | | | | | | 00 | OS | OD |
| 409 | Control | 31-35 | WNH | F | <u> </u> | OU | CRVO OS | 20/32 |
| | | | | | | | | OD |
| | | | | | | | | 0S |
| 333 | Control | 56-60 | BNH | M | | OU | PVD OD | 20/20 |
| | | | | | | | | OU |
| The final of | column ("Notes") inclu | ides the cu | umulative | e HCQ | dose and | renal/hepatic f | unction testing results f | or subjects |
| with a history of hydroxychloroquine use. For control subjects, the primary diagnosis resulting in examination in the retina clinic is listed ("N/A" indicates subjects who were recruited as a control subject without any ocular | | | | | | | | |
| complaint |). AST: aspartate trar | isaminase | ; ALT: al | anine ti | ransamina | ase; BNH: blac | k/non-Hispanic; CRVO: | central |
| retinal vei vitreous d | n occlusion; HCQ: hy letachment: RA: rheu | droxychlor matoid arth | oquine; hritis: SL | OD: rig E: svst | ht eye; OS emic lupus | 5: left eye; OU: s ervthematosા | : both eyes; PVD: poste us: WNH: white/non-His | erior spanic |

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437 Figures



| 439 | Figure 1: (a) Contrast-adjusted VIS-OCT image of a 36-40 age range Caucasian male |
|-----|--|
| 440 | subject with no known ocular history or retinal pathology highlighting outer retinal |
| 441 | features. (b) Contrast-adjusted image highlighting inner retinal features. (c) Magnified |
| 442 | view of foveal outer retinal features seen in panel (a) demonstrating outer retinal |
| 443 | banding pattern with outer retinal bands labeled. (d) Magnified view of parafoveal outer |
| 444 | retina. (e) Magnified view of foveal inner retinal features seen in panel (c) (see |
| 445 | abbreviations below). (f) Magnified view of parafoveal inner retinal features. |

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- 446 RNFL: retinal nerve fiber layer, GCL: ganglion cell layer, IPL: inner plexiform layer, INL:
- 447 inner nuclear layer, OPL: outer plexiform layer, ONL: outer nuclear layer, HFL: Henle's
- 448 fiber layer



- 450 Figure 2: (a) VIS-OCT image of control subject demonstrating manual segmentation of
- 451 outer retinal layers (red). (b) Mean intensity of each outer retinal band (summation of
- 452 intensity between manually segmented outer retinal bands) averaged across three
- 453 control subjects shown relative to distance from the foveal pit.

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454

Figure 3: (a) Contrast-adjusted VIS-OCT image of a 61-65 age range subject with a 455 456 history of systemic lupus erythematosus and 11 years of hydroxychloroquine use. (b) Swept-source OCT image of same subject in (a). (c-d) Magnified view of parafoveal 457 458 outer retinal features (dashed lines in (a)) demonstrating broadening of band 2 and loss 459 of layer 3i. (e) Magnified view of outer retinal features from control subject at similar 460 distance to fove as c-d, scaled to align with panels c-d. (f) Central subfield thickness 461 map of total retinal thickness obtained from swept-source OCT, measurements listed in 462 micrometers. Coloring indicates normal distribution percentile per device manufacturer. 463 (g) Averaged A-line signal over nasal 3mm-6mm eccentricity from Panel A between two

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- 464 white lines. The A-line trace signal is extracted from the same region in normal eyes in
- Figure 1. Both A-lines have zero means and were normalized to RPE band.



<u>Figure 4</u>: (a) Contrast-adjusted VIS-OCT image of 71-75 age range patient with a 5-year
history of HCQ use and suspected HCQ retinal toxicity. (c-d) Magnified view of
parafoveal outer retinal features (dashed lines in (a)) demonstrating broadening of band
2 and loss of band 3i and patchy attenuation of band 3o. (e) Magnified view of outer
retinal features from control subject at similar distance to fovea as c-d, scaled to align
with panels c-d. (f) Central subfield thickness map of total retinal thickness obtained
from swept-source OCT, measurements listed in micrometers. (g) Averaged A-line

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- 474 signal over nasal 3mm-6mm eccentricity from Panel A between two white lines. The A-
- 475 line trace signal is extracted from the same region in normal eyes in Figure 1. Both A-
- 476 lines have zero means and were normalized to RPE band.



Figure 5: (a) Contrast-adjusted VIS-OCT image of 51-55 age range patient with a 4-year
history of HCQ use with severe HCQ retinal toxicity and no other known retinal
pathology. (c-d) Magnified view of parafoveal outer retinal features (dashed lines in (a))
demonstrating diffuse loss of bands 2, 3i, 3o, and 4. (e) Magnified view of outer retinal
features from control subject at similar distance to fovea as c-d, scaled to align with
panels c-d. (f) Central subfield thickness map of total retinal thickness obtained from

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- 484 swept-source OCT, measurements listed in micrometers. Normative data not available.
- (g) Averaged A-line signal over nasal 3mm-6mm eccentricity from Panel A between two
- 486 white lines. The A-line trace signal is extracted from the same region in normal eyes in
- 487 Figure 1. Both A-lines have zero means and were normalized to RPE band.





- 489 <u>Supplemental Figure 1</u>: (a) Sample raw output of foveal line scan from VIS-OCT device
- 490 prior to image processing. (b) Foveal line scan following image flattening and contrast
- 491 adjustment as described in Methods.

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| 493 | Supplemental Figure 2: (a) Contrast-adjusted VIS-OCT image of a 26-30 age range |
|-----|---|
| 494 | Caucasian male subject with no known ocular history or retinal pathology highlighting |
| 495 | outer retinal features. (b) Contrast- adjusted image highlighting inner retinal features. (c) |
| 496 | Magnified view of foveal outer retinal features seen in panel (a) demonstrating outer |
| 497 | retinal banding pattern with outer retinal bands labeled. (d) Magnified view of parafoveal |
| 498 | outer retina. (e) Magnified view of foveal inner retinal features seen in panel (c) (see |
| 499 | abbreviations below). (f) Magnified view of parafoveal inner retinal features. |

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500

501 <u>Supplemental Figure 3</u>: Mean intensity of each outer retinal band (summation of 502 intensity between manually segmented outer retinal bands) averaged across three 503 control subjects shown relative to distance from the foveal pit for VIS-OCT scan 504 displayed in Figure 2A. Manual segmentation of outer retinal bands performed 505 independently by second grader.