



The Variable Nature of Vitamin C—Does It Help When Dealing with Coronavirus?

Katarzyna Grudlewska-Buda¹, Natalia Wiktorczyk-Kapischke¹, Anna Budzyńska¹, Joanna Kwiecińska-Piróg¹, Jana Przekwas¹, Agnieszka Kijewska², Dominika Sabiniarz³, Eugenia Gospodarek-Komkowska¹ and Krzysztof Skowron^{1,*}

- ¹ Department of Microbiology, Ludwik Rydygier Collegium Medicum, Nicolaus Copernicus University in Toruń, 85-094 Bydgoszcz, Poland; katinkag@gazeta.pl (K.G.-B.); natalia12127@gmail.com (N.W.-K.); an.budzynska@wp.pl (A.B.); j.kwiecinska-pirog@cm.umk.pl (J.K.-P.); jana.przekwas@gmail.com (J.P.); gospodareke@cm.um.pl (E.G.-K.)
- ² Department of Immunobiology and Environmental Biology, Institute of Maritime and Tropical Medicine, Medical University of Gdansk, 80-211 Gdansk, Poland; agnieszka.p.kijewska@gumed.edu.pl
- ³ Antoni Jurasz University Hospital No. 1, 85-094 Bydgoszcz, Poland; dominika.sabiniarz@gmail.com
- * Correspondence: skowron238@wp.pl; Tel.: +48-(52)-585-38-38

Abstract: Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) is still spreading worldwide. For this reason, new treatment methods are constantly being researched. Consequently, new and already-known preparations are being investigated to potentially reduce the severe course of coronavirus disease 2019 (COVID-19). SARS-CoV-2 infection induces the production of pro-inflammatory cytokines and acute serum biomarkers in the host organism. In addition to antiviral drugs, there are other substances being used in the treatment of COVID-19, e.g., those with antioxidant properties, such as vitamin C (VC). Exciting aspects of the use of VC in antiviral therapy are its antioxidant and pro-oxidative abilities. In this review, we summarized both the positive effects of using VC in treating infections caused by SARS-CoV-2 in the light of the available research. We have tried to answer the question as to whether the use of high doses of VC brings the expected benefits in the treatment of COVID-19 and whether such treatment is the correct therapeutic choice. Each case requires individual assessment to determine whether the positives outweigh the negatives, especially in the light of populational studies concerning the genetic differentiation of genes encoding the solute carriers responsible for VC adsorption. Few data are available on the influence of VC on the course of SARS-CoV-2 infection. Deducing from already-published data, high-dose intravenous vitamin C (HDIVC) does not significantly lower the mortality or length of hospitalization. However, some data prove, among other things, its impact on the serum levels of inflammatory markers. Finally, the non-positive effect of VC administration is mainly neutral, but the negative effect is that it can result in urinary stones or nephropathies.

Keywords: SARS-CoV-2; COVID-19; ascorbic acid; natural antioxidants; inflammatory process; respiratory distress syndrome

1. Introduction

The struggle with the coronavirus disease 2019 (COVID-19) pandemic caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) has been ongoing since December 2019 [1], and the emergence of new SARS-CoV-2 variants is not conducive to ending the pandemic state. The number of COVID-19 patients continues to increase, and mortality, despite vaccination, remains high [2]. The new variants of SARS-CoV-2 represent a significant challenge in fighting the disease and reducing the spread of infection, despite current COVID-19 vaccinations. The latest variants of SARS-CoV-2 represent a significant challenge in fighting the disease and reducing the spread of infection, despite current challenge in fighting the disease and reducing the spread of infection, despite current challenge in fighting the disease and reducing the spread of infection, despite current challenge in fighting the disease and reducing the spread of infection, despite current challenge in fighting the disease and reducing the spread of infection, despite current challenge in fighting the disease and reducing the spread of infection, despite current challenge in fighting the disease and reducing the spread of infection, despite current challenge in fighting the disease and reducing the spread of infection, despite current challenge in fighting the disease and reducing the spread of infection.



Citation: Grudlewska-Buda, K.; Wiktorczyk-Kapischke, N.; Budzyńska, A.; Kwiecińska-Piróg, J.; Przekwas, J.; Kijewska, A.; Sabiniarz, D.; Gospodarek-Komkowska, E.; Skowron, K. The Variable Nature of Vitamin C—Does It Help When Dealing with Coronavirus? *Antioxidants* 2022, *11*, 1247. https://doi.org/10.3390/ antiox11071247

Academic Editors: Giovanna Mobbili, Roberta Galeazzi and Dario Rusciano

Received: 30 May 2022 Accepted: 22 June 2022 Published: 24 June 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). COVID-19 vaccinations. According to the CDC, as of 12–22 May 2022, the variants of concern are delta (B.1.617.2) and omicron (BA.1, BA.2, BA.4, and BA.5) [3].

The aggravation of disease symptoms in the form of severe pneumonia contributes to acute respiratory distress syndrome (ARDS), cytokine release syndrome, and lymphopenia [4,5]. ARDS is associated with the oxidative stress generated during infection, which results from the production of cytokines and free radicals. The uncontrolled and unregulated secretion of inflammatory and pro-inflammatory cytokines is positively associated with the severity of viral infection and mortality [6]. The secretion of various pro-inflammatory cytokines such as TNF- α , IL-1 β , and IL-6 leads to a hyperinflammatory response by recruiting macrophages, as well as T and B lymphocytes, in alveolar cells [7-10]. This process results in multi-organ failure, which can lead to death. People with reduced immunity are the most vulnerable to severe infection. These patients have low levels of inflammatory cytokines or chemokines such as IL-6 and TNF [10]. Currently, substances are being sought that would limit the effects of infection by modulating the immune system to reduce the inflammatory response or activate the antiviral response. Vitamin C (VC) is considered to be an antioxidant and is believed to strengthen immune function. It is a safe and affordable essential nutrient. This review aims to summarize the currently available scientific data on the role of VC in supporting the treatment of COVID-19 patients and the possibility of alleviating symptoms of the disease.

2. The Course of the Inflammatory Process during SARS-CoV-2 Infection

Angiotensin converting enzyme 2 (ACE2) has been identified as the primary cell receptor for SARS-CoV-2 entry into host epithelial cells. The SARS-CoV-2 spike (S) glycoprotein binds to ACE2 via the receptor-binding domain (RBD) [11]. Upon receptor involvement, several host serine proteases, including TMPRSS2, TMPRSS4, furin, and endosomal cathepsins, cleave the SARS-CoV-2 S protein at the junction between the S1 and S2 fragments, allowing fusion of host and viral membranes and delivery of the viral genome to the cytosol. In addition to ACE2, many other host molecules reportedly support the binding of SARS-CoV-2 to cells and act as entry factors, including CD147, neuropilin-1, sialic acid, and heparan sulfate [12,13].

Numerous studies suggest a significant role for several pro-inflammatory factors. These factors are interleukins (IL)-2, -6, -8, -10, -1β, tumor necrosis factors (TNF), interferons (IFNs), and granulocyte-macrophage colony-stimulating factor (GM-CSF) in the course of the disease COVID-19 [14–16]. In addition, chemokines, including CCL2, CCL3, CCL5, CXCL10, and other chemotactic cytokines [17], have been described as essential determinants of mortality during SARS-CoV-2 infection. Most of the co-expression of these genes has been well-described as actively expressed on the mucosal gene expression of antimicrobial peptides in inflammatory conditions caused by proinflammatory diseases, such as inflammatory bowel disease [18] or even cancer [19]. Their expression is also active in stereotyped and specific human responses to bacteria [20]. Interleukin- 1β plays a pivotal role in the induction of cytokine storms due to uncontrolled immune responses in COVID-19 infections [21]. At the same time, IL-6 may even serve as an early biomarker for monitoring inflammatory and immune responses in COVID-19 [10]. In turn, the level of TNF- α is associated directly with the probability of hospitalization and severe COVID-19 [22]. High levels of TNF- α and other proinflammatory interleukins lead to ARDS aggravation and widespread tissue damage [23]. Interferon-inducible chemokines, such as CXCL10 (IP-10), are one of the leading players in the antiviral responses [24], while CXCL8 acts as a trafficking mediator for neutrophils [25]. SARS-CoV-2 infection is closely related to the cytokine storm. This storm is a highly deadly immune disorder characterized by the rapid proliferation and hyperactivation of NK cells, macrophages, and T lymphocytes, as well as the excessive secretion of over 150 chemical mediators and inflammatory cytokines by non-immune and immune cells [26]. This phenomenon plays a crucial role in the progression of SARS-CoV-2 infection and may be a significant cause of multiple organ damage and increased mortality in immunocompromised patients. IL,

IFN, TNF, and GM-CSF are the main cytokines involved in generating cytokine storms in COVID-19 (Figure 1) [27]. Cytokines are generally divided into two categories based on their functionality during infection: pro-inflammatory cytokines/factors (IL-6, IL-12, IL-1β, IFN, and TNF) and anti-inflammatory cytokines/factors (IL-4, IL-7, IL-10, and TGF-β).

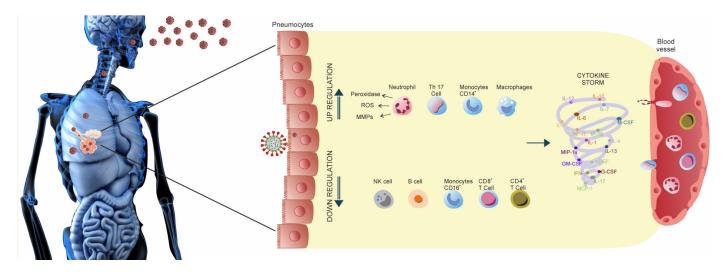


Figure 1. Cytokine storm in COVID-19.

The release of multiple cytokines is associated with various clinical symptoms, such as the over-secretion of IFN- γ , which causes headaches, chills, dizziness, fatigue, and fever [28]. Similar to IFN- γ , TNF- α causes flu-like symptoms, along with fever, fatigue, and malaise, but it can also lead to lung damage, leaky blood vessels, heart failure, and acute-phase protein synthesis [29]. The secretion of IL-6 causes vascular leakage syndrome, activating clotting and complement pathways, leading to clear indications of cytokine release syndrome, i.e., blockage of small blood vessels. Moreover, IL-6 has been associated with the induction of cardiomyopathy by stimulating coronary disease and myocardial dysfunction [30]. Additionally, severe cytokine release syndrome can occur due to the activation of endothelial cells, and endothelial dysfunction can result in hypotension, capillary leakage, and impaired blood clotting. In summary, the inappropriate secretion of various cytokines leads to the activation of a cytokine storm, causing immunopathogenic damage to multiple organs and tissues, even in the presence of a strong, suppressive immune system response.

From the point of view of supporting the treatment of severe COVID-19 cases, it is essential to search for substances that will mitigate the effects of uncontrolled damage in the body and reduce the risk of death of patients. A crucial, vital element of the host immune response during viral infection is the production of reactive oxygen species (ROS) and reactive nitrogen species (RNS), which leads to the destruction of infected cells in the early stages of the immune response [31]. Despite the beneficial effect of producing these molecules in the early stages of infection, their excessive production leads to oxidative stress, an imbalance between pro- and anti-inflammatory cytokines, and damage to tissues and organs (Figure 2). Many anti-inflammatory substances can neutralize the production of free radicals. These include glutathione; carotenoids; polyphenols; vitamins, such as C, D, and E, selenium, and zinc.

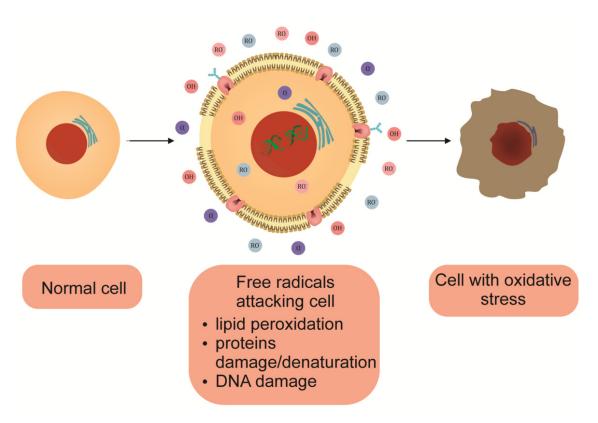


Figure 2. The effect of oxidative stress on the eukaryotic cell.

3. Natural Antioxidants to Treat or Support COVID-19 Patients?

The course of COVID-19 can be mild, moderate, or severe. The treatment regimen depends on the patient's condition. The World Health Organization (WHO) recommends treatment with casirivimab and imdevimab (for those with the highest risk of hospitalization), administering convalescent plasma for the treatment of patients with non-severe COVID-19, and using IL-6 receptor blockers (tocilizumab or sarilumab) [32]. However, the WHO advises against using hydroxychloroquine, chloroquine, or lopinavir/ritonavir to treat COVID-19 [32]. Patients with moderate-to-severe pneumonia may develop (life-threatening) sepsis, followed by septic shock, which is associated with multiple organ failure (MOF) and high mortality [33]. Septic shock during COVID-19 is characterized by the increased production of ROS and RNS [33]. SARS-CoV-2 affects the intracellular level of GSH (glutathione) by reducing intracellular functions [34]. Therefore, an alternative to supporting the treatment of COVID-19 patients may be antioxidant therapy, which has been known since Hippocrates, who used myrth for anti-inflammatory, medicinal purposes [35].

The primary mechanism of antioxidants is to increase intracellular GSH levels and scavenge free radicals, thereby protecting DNA, cytosolic proteins, and membrane lipids [35]. GSH deficiency is associated with a more severe manifestation of COVID-19. Among 4 COVID-19 patients, Polonikov et al. [36] observed a decrease in ROS (in patients with higher GSH levels) and a shorter course of the disease. In contrast, lower GSH levels have been associated with more severe disease symptoms and higher ROS [36]. SARS-CoV-2 has been shown to affect intracellular GSH levels by reducing the function of intracellular NRF2, which plays a crucial role in protecting cells from oxidative damage by regulating GSH production [34]. Horowitz et al. [37] showed that GSH supplementation in 2 COVID-19 patients reduced dyspnea (within 1 h of GSH administration), and this improved with each subsequent dose. Hiedra et al. [38] administered 1 g of VC intravenously (every 8 h for 3 days) to 17 COVID-19 patients. These researchers demonstrated reduced mortality, decreased intubation and mechanical ventilation, a significant decrease in inflammatory markers (ferritin and D-dimer), and a tendency toward a reduced need for FiO₂ [38]. In turn, Colunga Biancatelli et al. [39] recommended a combination of VC and quercetin (for a synergistic antimicrobial effect) in treating COVID-19 patients. Several studies on the impact of VC supplementation are summarized in Table 1.

Other compounds are also involved in antioxidant therapy. Antioxidant therapy may involve a combination of two or more compounds. A study of the effects of vitamin C, vitamin E, N-acetylcysteine (NAC), and pentoxifylline (Px) on 110 COVID-19 patients was conducted by Chavarría et al. [35] The researchers found that antioxidant therapy improved survival rates. The improvement was measured using the SOFA (sequential organ failure assessment) score, the Apache II (Acute Physiology and Chronic Health Evaluation) score, the SAPS II (Simplified Acute Physiology Score), the COVID-GRAM (critical illness prediction tool), and the GCS (Glasgow Coma Scale). In addition, the researchers also took into account the hospitalization for antioxidant therapy, the duration of mechanical ventilation, and the length of stay in the intensive care unit [35].

An example is the combination of NAC + Px in the early stage of the disease. This combination leads to a reduction in inflammation and cell damage [34]. In turn, Chavarría et al. [35] showed that combining Px supplementation with any antioxidant served to decrease IL-6 and CRP levels in COVID-19 patients. Serum zinc content (SZC) influences the progression of COVID-19 in the disease and may constitute a valuable, helpful biomarker [40]. The study data clearly show that many COVID-19 patients were zinc-deficient. More complications and deficiencies were associated with prolonged hospitalization and increased mortality [41].

Study Design	Patients	Dose of VC	Results	References [42]	
Case report	A 74-year-old woman	1 g twice a day (for a total of 10 days)	 fewer days on mechanical ventilation; shorter ICU stay; earlier recovery 		
Case series	17 patients	1 g every 8 h for 3 days	 reduced mortality; reduced number of intubations; decreased inflammatory markers (ferritin and D-dimer); reduced need for FiO₂ 	[38]	
Retrospective case series	12 patients	A median of 162.7 (71.1–328.6) mg/kg (body weight)/day in severe patients, and 178.6 (133.3–350.6) mg/kg/day in critical patients	patients, protein, lymphocyte, and CD4+ T-cell		
Case report	A 29-year-old man	VC treatment together with inhalation therapy	• the patient died of tension pneumothorax	[44]	
Case report	2 patients (a 50-year-old and a 71-year-old man)	200 mg/kg/day, for 96 h	 an adverse effect associated with high-dose intravenous VC administration; ATI and oxalate nephropathy are likely caused by excessive VC 	[45]	
Retrospective study	102 patients	73 patients received supplementation with VC and zinc	 high mortality no change in overall survival of patients 	[46]	
Open-label, randomized, and controlled trial	30 patients with severe COVID-19 infection	1.5 g of IV VC every 6 h for 5 days plus lopinavir/ritonavir and oral hydroxychloroquine vs. lopinavir/ritonavir and oral hydroxychloroquine alone	• no difference in the length of ICU stay, intubation rate, or mortality rate	[47]	
Open-label, randomized, and controlled trial	75 patients	50 g/kg/day	 no significant difference was found in the need for mechanical ventilation or mortality 	[48]	

Table 1. Effect of VC treatment/supplementation among COVID-19 patients.

ICU—intensive care unit; SOFA—Sequential Organ Failure Assessment score; ATI—acute tubular injury; COVID-19—coronavirus disease 2019.

Antioxidant therapy may be beneficial in those patients with comorbidities who take medications each day on a permanent basis. Intracellular GSH levels in RBCs are reduced in patients with type II diabetes [49]. Therefore, supplementation with antioxidants during COVID-19 may be helpful in this group of patients. Sinaci et al. [50] showed that the levels

of vitamin D in pregnant women with moderate/severe courses of COVID-19 were lower (13.69 ng/mL) compared to those of pregnant women with a mild approach (13.69 ng/mL). In addition, Schmitt et al. [51] obtained similar results. They showed that the level of vitamin D in pregnant women (patients and controls) was <20 ng/mL, while in the case of mild COVID-19, the level of vitamin D in pregnant patients was lower (<12 ng/mL) [51].

In contrast, in another study, vitamin D levels were not associated with the level of severity of COVID-19 during pregnancy as similar levels of vitamin D deficiency were found in COVID-19-positive patients and controls [52]. Therefore, studies are ongoing, including clinical ones, on the use of antioxidants to treat COVID-19 patients [53]. A list of the most important antioxidants is presented in Table 2.

Table 2. The role of selected antioxidants in the course of COVID-19.

Antioxidant	Properties	Role in COVID-19	References	
Vitamin C	 a cofactor of many enzymes; protects neutrophils and phagocytes from the damage that occurs after oxidative burst; modulation of nuclear transcription factor kappa B (NFkB); attenuation of pro-inflammatory cytokine production 	 inhibits the production of superoxide (O²⁻) and peroxynitrite (ONOO⁻) by inhibiting O²⁻ NADPH oxidase production and mRNA expression induced nitric oxide synthase (iNOS); in patients with septic shock, decreases MOF, SOFA score, ICU stay time, oxidative stress (OS), and inflammation 	[54,55]	
Vitamin D	 reduces the expression levels of proinflammatory type 1 cytokines, such as IL-12, IL-16, IL-8, TNF-α, and IFN-γ, while increasing type 2 cytokines, such as IL-4, IL-5, IL-10, and regulatory T cells 	• a potential role in the suppression of the cytokine storm during COVID-19	[56,57]	
Vitamin E	 the lipophilic antioxidant in cell membranes; scavenger of O²⁻ and hydroxyl (OH) radicals 	 improvement of the immune response an antioxidant in the acute phase of COVID-19 in patients with septic shock, decreases MOF, SOFA score, ICU stay time, oxidative stress (OS), and inflammation 	[58]	
Melatonin (MT)	 sequesters ROS, thus protecting lipids in cell membranes, proteins in the cytosol, DNA, and mitochondria; stimulates antioxidant enzymes, such as catalase (CAT), superoxide dismutase (SOD) isoforms, GPx, and GR 	 improves sleep habits, reduces anxiety, and boosts immunity; in patients with septic shock, decreases MOF, SOFA score, ICU stay time, oxidative stress (OS), and inflammation decrease; can alleviate septic shock via the NLRP3 pathway; prevents sepsis-induced kidney damage, septic cardiomyopathy, and liver injury 	[59–61]	

Antioxidant	Properties	Role in COVID-19	References	
Pentoxifylline (Px)	 antioxidant and anti-inflammatory effects, such as maintaining GSH levels and mitochondrial viability; inhibition of TNF-α production and maintenance of vascular endothelial function 	• in patients with septic shock, decreases MOF, SOFA score, ICU stay time, oxidative stress (OS), and inflammation	[33,35]	
N-acetylocysteine (NAC)	• reduces the incidence of pneumonia	 binds to Cys-145, the active site of the M protein, and therefore inhibits protease activity and viral replication; a potentially specific, first-line drug for SARS-CoV-2; can help to slow down the aggressive and lethal development of COVID-19 with the use of a moderate dose 	[62,63]	
Zinc	 regulates inflammatory activity; antiviral and antioxidant functions 	 increases the number of cytotoxic T lymphocytes; reduces the number of activated T helper cells; improves cellular immunity, reduces oxidative stress and the production of chronic inflammatory cytokines 	[64]	

Table 2. Cont.

MOF—multi-organ failure; SOFA—Sequential Organ Failure Assessment; ICU—intensive care unit; OS—oxidative stress; SARS-CoV-2—severe acute respiratory syndrome coronavirus 2; COVID-19—coronavirus disease 2019.

In addition to antioxidants, diet can be an essential consideration when COVID-19 is mild. Many ingredients of natural origin can influence the course of the disease. An example is propolis, considered a natural means of supporting the immune system. Propolis promotes the immunoregulation of pro-inflammatory cytokines, reducing IL-6, IL-1, TNF- α , Jak2/STAT3, and NF- κ B. The reduction effects lower the risk of cytokine storm syndrome (Figure 1).

Phytochemicals that show promise for inhibiting human coronaviruses include quercetin, myricetin, and caffeic acid, which are all constituents of propolis [65]. Propolis is used to treat comorbidities, such as diabetes, cancer, and hypertension [66]. Flavonoids also have a positive effect on inhibiting the cytokine storm because they regulate inflammatory mediators; inhibit endothelial activation, NLRP3 inflammasome, Toll-like receptors (TLR), and bromodomain protein 4 (BRD4); and activate Nrf2 [67]. The sources of flavonoids are herbs, fruits, and vegetables. Su et al. [68] showed that baicalein and baicalin inhibited SARS-CoV-2. Additionally, baicalin is an ACE2 inhibitor [69]. Other compounds that inhibit SARS-CoV-2 replication include curcumin, quercetin, kaempferol, capsaicin, sesamine, cyanidins, puerarin, scutellarin, ursolic acid, rutin, cesalamin B, ebsalene, galangin analogues, ellagic acid, and coumarin [70–74]. On the other hand, activity against SARS-CoV-2, due to a high affinity with the S protein of RBD, has been demonstrated for hesperidin, curcumin, braziline, and galangin [75]. Another active ingredient in the food is allicin, the source of which is garlic. Allicin inhibits inflammation by inhibiting TNF- α , which induces the expression of IL-1 β , IL-8, IP-10, and IFN- γ [76]. Thus, consuming garlic may be helpful during a mild course of COVID-19 [57].

Supplementation therapy with antioxidants may contribute to a decrease in mortality among COVID-19 patients. However, caution should be exercised as many oxidant-based treatments have produced negative or inconclusive results [77]. Additionally, data on the possible effects of VC on COVID-19 in children are not yet available.

4. Vitamin C

Ascorbic acid (AA), which is a saccharide derivative produced from D-glucose (by plants and animals) or D-galactose (by plants), was first isolated in 1928 from pepper and adrenal gland extracts [78]. The AA molecule has four stereoisomers: biologically active L(+)-ascorbic acid (VC), D-ascorbic acid and L-isoascorbic acid (inactive forms), and D(-)-isoascorbic acid (which has little biological activity).

VC began to be used in the early 20th century to treat scurvy [79]. The inability to synthesize VC in primates due to a mutation in the gene encoding the L-glucono- γ -lactone oxidase (Gulo) which is involved in the final step of AA biosynthesis makes it necessary to provide it via the dietary route. The enzyme which oxidizes L-gluconolactone to AA is also lacking in some animals—certain species of fish, birds, bats, and guinea pigs, which provides a useful animal model for research [80]. Nowadays, hypovitaminosis C leading to scurvy, although still reported, is rare because of the awareness of the need to supply it via an adequate diet. The Recommended Dietary Allowance (RDA) of VC is an average of 75 mg for women and 90 mg for men. (RDA values may vary depending on the country.) VC deficiency can result from malabsorption disorders, malnutrition, and improper diets that reduce AA absorption, and it is also observed in patients with infections or cancer, and in critically ill patients. Increased supplementation should apply to women with pregnancies (especially multiple pregnancies), lactating women, smokers, people with hypertension or diabetes, and those living under stress [81–83].

Ascorbate is transported into the cell across cell membranes by sodium-dependent VC transporters (SVCT1 and SVCT2). In contrast, in the transport of the oxidized form, dehydroascorbic acid (DHA), which is reduced again in the cell, glucose transporters (GLUT) are involved [84]. In healthy individuals, plasma VC concentrations are approximately 50–100 μ M [85]. Accumulation of AA in organs and tissues varies, with the highest levels found in endocrine cells and neurons, mainly in the adrenal and pituitary glands [86]. The kidneys excrete excessive amounts of VC; therefore, in most cases, there is no danger associated with its consumption, even at a dose that exceeds its daily requirement (1 to 200 g) [87]. Possible adverse side effects of high doses of VC include abdominal bloating and/or transient osmotic diarrhea and polyuria. Controversial, however, is the increased risk of urinary stone formation due to the increased concentration of urinary oxalate under the influence of VC doses exceeding 1 g per day. Therefore, one may be mindful of excluding this supplementation in individuals with kidney stones or renal impairment [88–90].

Ascorbate is a cofactor for a family of biosynthetic and regulatory metalloenzymes, such as hydroxylases (prolyl 4-hydroxylase and lysyl hydroxylase), or oxygenases (coppercontaining monooxygenases, iron (II)-, and 2-oxoglutarate-dependent dioxygenases). Consequently, ascorbate is involved in the stabilization of the tertiary structure of collagen, as well as in the synthesis of carnitine and hormones (noradrenaline/adrenaline and peptide hormones). For this reason, with VC deficiency, the integrity of basement membranes is weakened, and due to disorders in the synthesis of connective tissue, wound healing is impaired, and the risk of osteoporosis and fractures increases [91–93]. AA has also been shown to positively affect the efficiency and accuracy of epigenetic reprogramming by demethylating the lysine 36 of histone H3 (H3K36). This process facilitates the formation of induced pluripotent stem cells (iPSCs). AA also reduces the frequency of the unwanted spontaneous differentiation of iPSCs [94].

An essential role of AA, an electron donor, is the antioxidant activity mentioned earlier—protecting cells from free radicals, such as peroxide radicals, hydrogen peroxide, singlet oxygen, and hydroxyl radicals—formed during metabolism. Ascorbic acid, because of oxidation, converts to an ascorbic anion which, after donating an electron, becomes a nonreactive ascorbic radical. DHA, characterized by the same biological activity as the reduced form, is formed by losing another electron [84,95].

VC plays a vital role in the proper functioning of the immune system. Its concentration in immune cells (lymphocytes, monocytes, and neutrophils) in individuals consuming

 \geq 100 mg ascorbate per day is up to 100 times higher than plasma concentrations [93,96]. AA affects immune cell function in the hypoxic environment in inflammation and cancer because it is required for the optimal hydroxylase activity that regulates the activity of the hypoxia-inducible factors (HIFs) that direct inflammatory and immune responses. The activation of HIFs prolongs neutrophil survival and their antibacterial function. HIFs may also be important in T-cell differentiation, activation, and function [97]. Thus, AA affects lymphocyte development and function. Studies have shown that AA plays a role in the normal maturation of T cells in the thymus, probably through the epigenetic modulation of gene expression. Its involvement in regulating the activity of mature T lymphocytes is also considered [98]. In the case of B lymphocytes, results remain inconclusive in establishing the effect of VC on their proliferation and function [99]. VC, on the other hand, may protect neutrophils from the ROS that they produce and from oxidative damage, as well as increasing the motility and migration of various peripheral blood leukocytes, such as lymphocytes or polymorphonuclear leukocytes, in innate immune responses (Figure 3) [100].

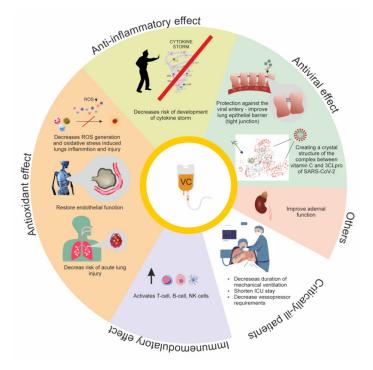


Figure 3. The effect of vitamin C in SARS-CoV-2 infection.

The effect of VC on the immune system has been observed in vivo studies. With age, there is excessive apoptosis and a gradual decrease in the proportion of functional T cells. In older adults, after 3 months of supplementation (500 mg/day), improvements in the immune functions—increased lymphocyte proliferation and interleukin-2 release—were found [101]. In turn, in the studies by Bozonet et al. [102], an increase in VC levels in the neutrophils of individuals with a low VC intake, as influenced by administration in its natural form (kiwi fruits), resulted in a significant increase in neutrophil chemotaxis and superoxide generation. The role of antioxidants in altering the expression of adhesion molecules and inhibiting monocyte adhesion to endothelial cells (ECs) in blood vessels observed in early atherosclerotic lesions has also been suggested [103]. This conclusion is supported by the study of Woollard et al. [104], which demonstrated the normalization of monocyte adhesion under the influence of VC supplementation in people showing reduced VC levels. It is difficult to assess the role of AA in NK cells. Studies indicate that it may have a positive effect by increasing the cytotoxicity of impaired NK cells, while no AA effect on properly functioning NK cells has been observed [99,105,106].

Brain cells, particularly susceptible to oxidative damage, show increased sensitivity to VC deficiency. Due to the synaptic activity and intense neuronal oxygen metabolism

of central nervous system cells and the associated increased production of free radicals, a sufficiently high level of antioxidants is required. In the case of glial cells, such an oxidant is glutathione, and in nerve cells, it is mainly AA [107,108]. Thus, there is a possible link between ascorbate deficiency and neurodegenerative diseases, including Alzheimer, Parkinson, and Huntington disease, as well multiple sclerosis [109].

Currently, much research is focused on using VC as a supplement in treating many diseases (such as cancer) and infections, including sepsis and septic shock [110]. In the case of cancer, attention has been drawn to the contribution of VC in reducing the toxicity of chemotherapy. The positive effect of using VC in preventing the development of malignant tumors, such as those caused by colorectal adenocarcinoma, lung cancer, and endometrial cancer, has also been described. Moreover, its synergistic effect with many chemotherapeutic agents (including carboplatin, arsenic trioxide, and paclitaxel) used in the treatment of various cancers has been indicated [87,111–113]. This effect is probably related to the fact that VC, in addition to its antioxidant properties, may exhibit pro-oxidant effects. If catalytic metals, especially iron and copper, are present in the environment, AA reduces them (Fe³⁺ \rightarrow Fe²⁺, Cu²⁺ \rightarrow Cu⁺). Their new forms can react with oxygen, forming superoxide or hydroxyl radicals [84,114]. The effect of AA cytotoxicity on inhibiting the growth rate of many cancer cell types was demonstrated in vitro and in vivo studies by Chen et al. [115]. Parenteral administration of ascorbate in mice bearing glioblastoma xenografts at pharmacological doses (>0.2 mM) resulted in sustained ascorbate radical and hydrogen peroxide formation in the extracellular space, followed by the autophagy of cancer cells, while having no toxic effects on normal tissues. The process is associated with the inhibited activity of enzymes that neutralize oxidative stress in cancer cells, leading to a reduced ability to metabolize H₂O₂ compared to normal cells [116]. Elevated free-radical levels can lead to the depletion of antioxidant mechanisms and the death of cancer cells. Apoptosis of breast cancer cells induced by AA at physiological concentrations (100 μ M) was observed by Hong et al. [117], while Tronci et al. [118] demonstrated the antitumor effect of VC against papillary thyroid carcinoma cells. The oral intake of VC by patients with malignant tumors likely has no impact because, at lower concentrations, VC exhibits antioxidant, rather than pro-oxidant, activity [119]. However, more clinical studies are needed to confirm the positive effects of VC in oncology patients. One should also consider the results of research conducted on leukemia and lymphoma cell lines by Heaney et al. [120], which indicated that VC could inhibit the effects of anticancer drugs, such as doxorubicin, which produce reactive oxygen species.

Since VC can affect the cytokines secreted by immune cells, its beneficial effects have been shown in many studies using animals infected with viral and bacterial infections or Candida albicans. In studies, decreased or increased production of the pro-inflammatory cytokines TNF- α , IFN- γ , IL-6, and IL-1 β , and increased production of anti-inflammatory IL-10 and IL-4, depending on the cell type, as well as inflammation stimulating factor, have been observed under its influence [121–125]. VC deficiency increases the risk of infection, which is why pneumonia was one of the most common complications of scurvy found in the 20th century [126]. At the same time, due to an overactive inflammatory response resulting in increased metabolism, increased demand for VC is observed in the course of infection. This demand increases significantly when the disease is more severe [127]. A study by Fisher et al. [128] found that, in mice treated with lethal doses of lipopolysaccharide, parenteral administration of AA attenuated proinflammatory states and reduced microvascular thrombosis. The attenuation prevented pulmonary vascular damage in the course of sepsis. Another paper [129], which demonstrated in a mouse model the enhancement of the alveolar-bronchial epithelial barrier function, confirms the potency of VC in preventing multiple organ dysfunction syndromes and treating sepsis. However, the results of clinical trials are not as conclusive. Shrestha et al. [130] described that using VC improved critically ill patients and significantly reduced their intensive care unit (ICU) stay. Hemilä and Chalker [131] reached similar conclusions with a meta-analysis that evaluated the effect of VC on the length of stay and the duration of mechanical ventilation

of patients in the ICU. Fowler et al. [132], assessing the impact of 96-h intravenous VC infusion in patients with sepsis and acute respiratory distress syndrome, found no reduction in vascular damage, changes in organ failure, or C-reactive protein levels. However, there was a significant reduction in 28-day patient mortality compared with the placebo group. In contrast, different results (no improvement in mortality) were shown in patients with severe sepsis or septic shock in the following two studies [133,134]. Therefore, further randomized controlled trials are needed to evaluate the role of VC in the treatment of patients with sepsis.

As previously mentioned, it has also been suggested that VC may be used in the treatment of various viral infections. A daily dose of 1 g of VC is widely believed to reduce the risk of respiratory tract infections. However, not much research supports this assumption [135–138]. In contrast, supplementation at a dose of 0.25–1.0 g/day is recommended for athletes undergoing intensive training and/or during periods of increased risk of upper respiratory tract infections so as to reduce the incidence or shorten the duration of the common cold [139,140]. VC probably does not have a direct virucidal effect, and the impact of reducing viral loads in infected cells is due to its immunomodulatory properties, as described previously [141]. In a study using Gulo knockout (-/-) mice as the study model, it was found that their infection with the H3N2 or H1N1 influenza viruses resulted in more significant lung damage and higher mortality compared to wild-type mice. The effect was associated with an inadequate immune response, including reduced IFN- α/β production in VC-deficient mice [142,143].

During infection, an oxidative imbalance in the host cells occurs. The imbalance results in the overproduction of cytokines and contributes to a severe inflammatory response which, in respiratory infections, leads to lung tissue damage [144]. Furthermore, viruses exploit changes in redox homeostasis for their replication by activating various cellular pathways. VC, through its properties, reduces the oxidative stress that occurs during infection and inflammation due to the activation of phagocytes and ROS release. The VC-mediated reduction of ROS prevents ROS-induced lung damage in the course of viral infections [125,145]. Increased survival associated with AA treatment was observed by Valero et al. [146] in mice infected with the Venezuelan equine encephalitis virus and by Cai et al. [147] in restraint-stressed mice infected with the H1N1 virus.

In contrast, a study conducted on a group of patients with severe viral pneumonia with respiratory failure did not show a reduction in their mortality from receiving VC as compared to a control group [148]. In contrast, the evaluation of the effect of VC on the treatment of patients with herpes zoster has shown a reduction in the incidence of postherpetic neuralgia, as the antioxidant activity may play an essential role in reducing the pain caused by herpes zoster [149]. However, due to the limitations of the research, including the lack of the measurement of plasma VC concentration and the heterogeneity of the groups compared, the authors suggest conducting a multicenter, randomized study.

4.1. Variability of Genes and Their Expression Related to VC Metabolism

4.1.1. Genetic Background of VC Carriers and Related Genes

It could be noticed by tracking the interactions described in the scientific literature dedicated to VC by web interface for generating hypotheses about gene function, analyzing gene lists, and prioritizing genes for functional assays [150] and these VC carriers (Solute Carrier Family 23 members—sodium-dependent SLC23a genes), most of the genes described as involved in a response to COVID-19 were noticed in the mucosal gene expression of antimicrobial peptides in bowel disease [18], the expression of genes engaged in liver cancers [151], and in drug-induced liver injuries [152]. These genes were also involved in expression patterns in primary human liver tissue connected to diabetes, drug response, lipid levels, prostate cancer [153], tumor gene expression [154], a drug-cell interaction in multiple myeloma patients [155], and oncogenic pathway signatures [19], but also in a pluripotent state of stem cells [156], genes characteristic for central nervous system distinct

sites [157], and, finally, distinctive gene expression patterns in human mammary epithelial cells and breast cancers [158].

A list of genes involved in the above interactions, and related to the solute carrier family 23 (*slc23a1*, *a2*, and *a3*) and solute carrier family 2 (*slc2A1* and *A3*) were input into the ConsensusPathDB [159] and analyzed for grouping into pathways related to COVID-19. This procedure enriched transcription factor target sets with calculated *p*-values and q-values. Then, all statistically significant, selected groups were rechecked for any relation to the VC data in scientific literature. Surprisingly the presence of the slc23 subfamily and/or slc2 subfamily was annotated in the miRNA sets, miR-223-3p, miR-203a-3p, and miR-122b-5p. The miR-223-3p set was mentioned as being involved in regulating the SARS-CoV-2-induced inflammatory pathology [160]. According to the authors, the negative fold of the expression of miR-223-3p suggested contribution in vivo to limiting the excessive lung inflammation induced by SARS-CoV infection, and the inhibition of the miRNA-223 increases the mRNA expression levels of NLRP3 inflammasome and pro-inflammatory chemokines CXCL10, CXCL2, and IL-1ß. In this set, only *slc2a4* (*glut4*) was present. Still, according to Shaghagi et al. [161], it is postulated that VC accumulation in cells occurs partly through DHA transport by the carriers of the SLC2A family. Another set, miR-122b-5p, is much more interesting, despite its low *p*-value (p < 0.01). That set of miRNAs differed depending on whether study participants were healthy or had moderate-to-severe COVID-19. This differentiation suggests that the presence of long and short RNAs in human cells is integral to the post-transcriptional gene regulation of the gene set involved in the response to infection and the relevant to RNA of SARS-CoV-2.

Polymorphisms and the pathological relevance of genes *slc23a1* and *slc23a2* were examined for population differences in population differentiation by Eck et al. [162], and the authors found differences between alleles of 2 analyzed genes, depending on the origins of the sample donors: self-described African Americans differed from people self-described as Caucasian. A genetic variant (rs33972313 C/A/G/T) was found in exon 8 of the SLC23A1 locus. It is suggested that this missense causes a conformational change or protein failure responsible for lower concentrations of circulating ascorbic acid ($-5.98 \mu mol/L$) in the general population [163]. Moreover, Zanon-Moreno et al. [164] found that the presence/absence of rs1279386 (A>G) SNP is related to the different risks of primary open-angle glaucoma. They also demonstrated that the SLC23A2 genotype modifies the serum levels of VC [164].

A similar problem concerns slc2a1 gene transporting, with the exception of glucose and dehydroascorbic acid. The polymorphism of that gene, especially SNP rs1385129 (A/G), is known to be related to diabetic cardiovascular disease (CVD) with diabetes mellitus type 2 (T2DM) [165] and diabetic microvascular complications [166,167]. In turn, the slc2a2 gene's polymorphisms (rs5438, rs8192675) may play a role in influencing high-density lipoproteins, and thus the metabolic risk of cardiovascular disease [168] in European Americans. Moreover, the decreased expression of GLUT1 (slc2A1) is critically involved in the disease progression of SARS-CoV-2 infection [169].

Another gene, *ace2* also known to be polymorphic, did not exhibit a statistically significant effect on COVID-19. According to Ivanov et al. [170], who performed experiments in vitro on SAEC and HMEC cell cultures, it is possible that AA can directly modulate the expression of the *ace2* gene and indirectly modulate individual susceptibility to COVID-19. This fact can support the assumption about the crucial role of VC in preventing infection and limiting the multiplication of the virus in the early stages of the disease.

4.1.2. Absorption of VC and the Microbiome

Not only genetic variation can affect the level of VC in organisms. Heskett et al. [171] proved that the presence of enteropathogenic *E. coli* (EPEC) in gut microbiota could affect the uptake of ascorbic acid via the dysregulation of its transporters' (SVCT1 and SVCT2 encoded by the *slc23a2* and *slc23a1* genes) expression.

Among many factors affecting the absorption of VC in the gastrointestinal tract, the intestinal microbiome is one which has the most influence. Commensal microorganisms affect both the synthesis of specific vitamins and the modulation of their absorption [172,173]. Subramanian et al. investigated the effects of bacterially derived lipopolysaccharide (LPS) on AA homeostasis in enterocytes. They showed that treating cell lines (Caco-2 line) affects the reduction of SVCT1 and SVCT2 proteins, mRNA, and hnRNA expression. The reduction of the expression of these elements, in turn, inhibits the absorption of ascorbic acid. Downregulating the expression of ascorbic acid transporters and the transcriptional regulation of SLC23A1 and SLC23A2 genes were the main reason for the inhibition [174].

On the other hand, studies show that supplementation with high doses of VC may impact changes in the microbiome's composition. Research conducted by Otten et al. [175] indicated that, in the case that patients were supplemented with VC in doses of 1000 mg, decreases in the relative number of *Lachnospiraceae*, *Bacteroidetes*, *Enterococcus*, and *Gemmiger formicilis* were observed [175]. The latest information about COVID-19 patients shows that the virus is present in gut cells and can affect their condition [176]. This imbalance can also affect the relation of the gut microbiome's regulating role with the absorption of ascorbic acid.

4.2. High-Dose Intravenous VC Administration

The intravenous usage of antioxidants in severe cases of infection is due to their ability to inhibit cytokine storms [177]. VC has been proven in animal and tissue models to modulate the immunological system by enhancing interferon production, suppressing oxidative stress and thrombosis, inhibiting cytokine storms, and lessening alveolar damage, and it has many other effects on the human body [178]. It is also known to enhance the maturation and proliferation of lymphocytes. VC also has a proven suppressive impact on the cellular expression of ACE2. The cellular expression is the crucial receptor for the SARS-CoV-2 virus to enter the human cells. Thus, VC may hinder the infection process [170]. Based on their research results and available references, Patterson et al. [179] suggested that, in SARS-CoV-2 infected patients, SVCT1 and SVCT2 AA transporters are downregulated, which might be an essential factor related to the severity of COVID-19.

The hypothesis regarding the benefits related to VC administration in severe cases of infection is based on the observation that, in this group of patients, levels of ascorbic acid during the recovery period are very low and, in many cases, undetectable [180,181]. In healthy people, ascorbic acid serum levels are about 0.4–2.0 mg/dL [180]. There is evidence that the majority of critically ill COVID-19 patients has low-to-undetectable VC levels. Tomasa-Irriguible and Bielsa-Berrocal [180] found that, in 82% of patients, within 24 h of ICU admission, the VC was below the normal range, with a mean value of 0.14 mg/dL (+/- SD 0.05 mg/dL). Similar results were observed by Chiscano-Camón et al. [181]—undetectable ascorbic acid levels were found in 94.4% of ICU patients with SARS-CoV-2-associated ARDS. Intravenous AA administration could lead to as much as a 100-fold increase in this vitamin level in the serum within 24 h [182].

Zhao B. et al. [183] found a correlation between patients suffering from multiple organ failure and low serum VC levels. Based on observations conducted during the pandemic, they deducted that high-dose intravenous VC (HDIVC) may prevent disease aggravation in moderate-type COVID-19 and serve as a rescue therapy for severe and/or critical cases of the disease, and they presented a flowchart of a HDIVC protocol proposed by the Shanghai COVID-19 Clinical Treatment Expert Group. The protocol differentiates the severity of the disease. However, it suggests relatively high doses for long-term periods (\geq 100 mg/kg for 7 days). In 2021, Zhao et al. [183] included SARS-CoV-2-infected patients in the study. They conducted 110 observations before and after COVID-19 therapy, over the course of 7 days. The researchers had two groups of patients: HDIVC-treated patients (n = 55) and the control group (n = 55). They observed differences in both of the examined groups of patients in forms of infection which worsened from moderate to severe (HDIVC: 7.3%; control: 21.8%; p = 0.03). In the HDIVC group, the patients exhibited shorter durations of

systemic inflammatory response syndrome (SIRS) (p = 0.0004) during the first week and lower SIRS occurrence (9.5% vs. 45.5%, p = 0.0086) on day 7 (6–7 days after admission) in comparison to the control group. Additionally, C-reactive protein serum concentrations were lower on the seventh day of observation in HDIVC patients than in the control group [184]. The relationship between VC and CRP in healthy people who have been exposed to some inflammatory factors is known [185,186], but the responsible mechanisms are still being examined.

The results from a retrospective cohort study [187] confirmed the information mentioned above, that HDIVC significantly lowers the levels of serum inflammatory markers, such as hs-CRP, IL-6, and TNF- α . (High-dose VC was intravenously administered over a 6-day course in dosages in excess of 100 mg/kg of body weight. VC was diluted in 50 mL of saline solution and infused within 30 min every 6 h on day 1, and 100 mg VC/kg of body weight was diluted in 50 mL of saline solution and infused within 30 min every 12 h for the next 5 d.) The researchers stated that hyperinflammation is linked to the severity of COVID-19 disease. The patients received over 2 g of VC per day for 6 days. In another study, Xia G et al. [188] found that, besides the lower levels of inflammatory markers after HDIVC (100 mg/kg every 6 h for 1 day, followed by 100 mg/kg every 12 h for an additional 5 days during hospitalization), the positive influence on myocardial damage in consecutively severe and critically ill COVID-19 patients was highlighted [188]. After 21 days of hospital therapy, levels of inflammatory markers (CRP, IL-6, IL-8, and TNF- α) significantly decreased in HDIVC patients.

In addition, during HDIVC therapy, the signs and symptoms in COVID-19 patients have improved. CT scans conducted by Tehrani et al. [189] in 2020 on a group of 54 patients showed that HDIVC (2 g/6 h for 5 days) might improve the rate of pulmonary involvement in moderate-to-critical cases. In addition, VC therapy improved the oxygen saturation and lowered the respiratory rate [189].

Despite the above-mentioned benefits, a meta-analysis by Rawat et al. [190], based on results from 572 cases, states that there is no significant decrease in the mortality rate, length of stay in the ICU, duration of hospital stay, or invasive mechanical ventilation after HDIVC therapy. The number of included papers was six. Three were also evaluated in a meta-analysis by Kwak et al. [191] In four analyses, VC was administrated via HDIVC; in two, the administration was oral. The results of the meta-analyses of the efficacy of using VC in the treatment of COVID-19 is summarized in Table 3.

Reference	Month, Year	No. of Studies	PICO	PRISMA	RCTs Only	Number of Patients	VC Dose	Adverse Effects	Conclusion
[190]	November– December 2021	6	YES	YES	YES	572	≥ 1 g daily, IV or oral	ND	No significant benefit
[192]	February 2022	7	NO	YES	NO (3 RCT, 4 retrospec- tive studies)	807	≥2 g daily HDIVC	ND	No significance in mortality rate or disease severity
[193]	January 2022, Beran A. et al.	9	YES	YES	NO (4 RCT, 5 retrospec- tive studies)	1488	≥ 1 g daily, IV or oral	ND	No significance in mortality, intubation rate, or hospitalization length
[191]	March 2022	5	YES	YES	NO (3 RCT, 2 retrospec- tive studies)	374	≥2 g daily HDIVC	No significance	No significance in mortality rate or hospitalization length
[194]	February 2022	11	NO	YES	NO (4 RCT, 7 retrospec- tive studies)	1807	≥1 g daily, IV or oral	ND	No significance in mortality, a longer length of stay for VC group

Table 3. Meta-analyses conducted on the topic of VC use in COVID-19 patients.

Those conclusions were confirmed in a randomized, open-label clinical trial. In Iran, 2 groups of ICU patients (n = 60) were examined: an HDIVC case group (1.5 g VC every 6 h for 5 days) and a control group [47]. All patients were treated with lopinavir/ritonavir twice daily, with a single dose of hydroxychloroquine (400 mg) on the first day of hospitalization. On the third day of treatment, the case group had significantly higher peripheral oxygen saturation (SpO₂: 90.5% vs. 88.0%; p = 0.014) levels and lower body temperatures (36.73 vs. 37.24; p = 0.001) than the control group, but the median length of stay in the hospital was longer in the case group than in the control group (8.5 and 6.5, respectively, p = 0.028). The JamaliMoghadamSiahkali [47] trials found no other benefits from the COVID-19 treatment, including mortality and the length of ICU stay. In this study, all patients were treated with hydroxychloroquine. For this reason, the results of this analysis pertain to a combined therapy, and the findings might be related to the combined effect of the hydroxychloroquine and ascorbic acid, rather than being related to AA alone.

Additionally, in a retrospective study from Wuhan conducted by Zheng et al. [195], there was no improvement observed in the clinical outcome, and no association between HDIVC use (2-4 g/day for 1 to 30 days) and the risk of death in severe cases of COVID-19 [195].

Over the last two years, there have been few randomized control trials (RCTs) considering high doses (over 2 g/day, over 5 to 10 days) of intravenous VC treatment in severe COVID-19 patients (Table 4). A meta-analysis from December 2021 suggests that no significant reduction in mortality rate or length of hospital stay was determined. Kwak et al. [191] observed an insignificant reduction in the in-hospital mortality rate among HDIVC patients as compared with the control group. There was also a lower occurrence of SIRS in this group of patients. The final results and the comparison of the effects of VC administration found in RCTs do not support the assumption that VC plays an essential role in treating COVID-19 patients, with the exception of the HDIVC treatment proposed by different authors who treated patients with high doses of VC. However, reports from studies on the long-term results of HDIVC treatment have included the reduction of inflammatory agents in COVID-19 patients [183,184,187,188], limiting the risk of thrombosis and inflammatory conditions affecting the functioning of multiple organs.

Reference	Study Design	No. of Patients	VC Age	VC Sex, Female (%)	C Age	C Sex	Patient	Disease Severity	Usage of VC	Conclusion
[48]	RCT	150	52 ± 11	ND	53 ± 12	ND	Inpatients	Severe	50 mg/kg/day IV	No impact on mortality or need for mechanical ventilation; difference in length of hospitalization
[196]	RCT	214	46 ± 15	33(69)	42 ± 15	31 (62)	Outpatients	Mild/ moderate	8000 mg/ day OR	No significant decrease in the duration of symptoms compared with standard of care
[47]	RCT	60	$\begin{array}{c} 58 \pm \\ 18 \end{array}$	15(50)	61 ± 16	15 (50)	Inpatients	Severe	4500 mg/day IV	No significantly better outcomes in the IV VC group
[197]	RCT	56	66 ± 11	12(44.4)	67 ± 14	7 (24)	Inpatients	Severe	24,000 mg/day IV	No significant difference in invasive mechanical ventilation-free days/28 days between groups
	Pilot ran- domized trial	20		59 ± 7(35			Inpatients	Severe	6000 mg/day IV	No considerable improvement in the clinical status of patients
[199]	RCT	72	36 ± ND	14(37)	$37 \pm \text{ND}$	12 (35)	Inpatients	Moderate (VAP excluded)	1000 mg/day OR	Vitamin C and vitamin E failed as an adjunctive treatment for COVID-19 patients
[200]	RCT	60	$\begin{array}{c} 51 \pm \\ 17 \end{array}$	15(50)	53 ± 7	14 (47)	Inpatients	Severe	2000 mg/day ND	No significant difference in length of hospitalization
[201]	RCT	120	$^{59\pm}_{15}$	12(39)	64 ± 16	28 (42)	Inpatients	Severe	500 mg/day OR	Higher mean survival duration compared to control group

Table 4. RCTs conducted on the topic of VC use in COVID-19 patients.

RCT—Randomized controlled trial; VC Age—age of people taking vitamin C; VC Sex—sex of people taking vitamin C; C Age—age of people in the control group not taking vitamin C; C Sex—sex of people in the control group not taking vitamin C; VC—vitamin C; ND—no data; VAP—ventilator-associated pneumonia; IV—intravenous; OR—oral; COVID-19 coronavirus disease 2019.

4.3. Ascorbic Acid Orally Supplementation in COVID-19

There is evidence from randomized clinical trials that VC uptake may shorten the duration of SARS-CoV-2 infection. Hemilä et al. [202] reanalyzed the results obtained by Thomas et al. [196]. They found that ascorbic acid (8000 mg divided over 2–3 times per day, with meals) increased the recovery rate by 70% (95% CI 6.8% to 170%, p = 0.025) in ambulatory SARS-CoV-2 infected patients.

A RCT performed by Majidi et al. [201] in December 2021 found that the ICU study group, taking 500 mg of enteral VC per day over 14 days, had a statistically significant higher survival rate. Additionally, the survival duration of patients positively correlated with the time of supplementation [201]. Scientists have also stated that a daily dose of greater than 500 mg drastically lowers its bioavailability in healthy persons. A retrospective clinical trial of 739 critically ill patients hospitalized with UTIs at a governmental tertiary hospital in Saudi Arabia found that low oral doses (1000 mg per day) of VC supplementation do not reduce mortality [201]. Al Sulaiman et al. [203] found that low amounts of VC in a study minimize the incidence of thrombosis in patients, which is the standard of care after COVID-19 infection. This might be related to the decrease in cytokine storms during VC therapy. This effect was observed by Abulmeat et al. [204]. They found that oral supplementation of antioxidants reduces cytokine storm characteristics in an early phase of SARS-CoV-2 infection. The reduction of thrombosis might be associated with the same pathway as found in cardiovascular diseases. Zhu et al. [205] concluded that the protective role of VC in cardiovascular diseases is related to its anti-inflammatory properties. They suspected that the most enriched pathways of VC are JAK-STAT, PD-1, EGFR, FoxO, and the chemokine-signaling pathways.

Vitamin supplementation during care after ICU discharge might be essential [205]. After COVID-19, patients are malnourished, and most experience weight loss (about 25%, or more than 8 kg). Patients are also burdened with psychological disorders, such as depression or anxiety. VC, among other micro- and macroelements, may help them recover faster by diminishing the malnutrition and the scarcity of vitamins and microelements [206].

All of the mentioned reports share one problem concerning the inequality of VC administration programs for patients (see Table 4), limiting the possibility of comparisons between them. The routes of supplementation and the frequency of doses are constantly changing, making comparisons very difficult. In the early stages of COVID-19, the oral administration of 500–1000 mg of VC does not decrease mortality, but it can minimize the risk of thrombosis and the chance of the cytokine storm. Definitively, the administration of VC does not affect the level of mortality. It is possible that, in future, we will know more about the role of VC in avoiding complications, such as the risk of thrombosis after severe COVID-19.

4.4. Ascorbic Acid Supplementation in Combined Pharmacological COVID-19 Treatment

There have been studies on the possible use of VC in combination with other medications in patients with COVID-19. Multiple doses of VC (1.5 g, 4 times per day) were used in the research conducted with hydrocortisone and thiamine, reducing organ failure and mortality from 40.4% to 8.5%. It has also been proven that in a critically ill COVID-19 patient, intravenous sodium ascorbate (60 g) restored arterial pressure, improved renal function, and increased arterial blood oxygen levels. Those findings suggest that megadoses of VC should be trialed as a treatment for sepsis and COVID-19 [207]. It has been shown that their combination can prevent acute kidney damage and reduce mortality in patients with septic shock or sepsis [208]. There is evidence that VC and quercetin co-administration exert synergistic, antiviral action due to overlapping antiviral and immunomodulatory properties. It has been noted that ascorbate's ability to recycle quercetin increases its efficacy. Quercetin displays a broad range of antiviral properties, for example, virus entry and replication. The co-administration of VC can augment these therapeutic effects. Quercetin and VC may disrupt virus entry and replication, concurrently fortifying the immune response, promoting early IFN production, modulating interleukins, promoting T-cell maturation, and promoting phagocytic activity. This therapeutic option was proposed in addition to the promising treatment options, such as Remdesivir or convalescent plasma [39], for COVID-19 patients. Other combinations which include VC have also been studied.

For example, preliminary results of a clinical trial showed that the treatment of severe COVID-19 with a mixture of MB (methylene blue), VC, and N-acetyl Cysteine is safe and feasible. The study used methylene blue (1 mg/kg) with VC (1500 mg/kg) and N-acetyl Cysteine (1500 mg/kg) orally or intravenously in ICU COVID-19 patients. The immediate effect was to increase the $SPO_2\%$ by reducing met-Hb. (All patients had been receiving 100% oxygen). The delayed effects were the acceleration of the typically slow NADPH-methemoglobin reductase, the improvement of inflammatory marker levels (CRP and LDH), and the decrease in disease severity, which may also have been caused by antimicrobial effects [209]. Other studies suggest the addition of thiamine to a corticosteroid (methylprednisolone), VC, and the anticoagulant, heparin (enoxaparin). The proposed dosing is as follows: methylprednisolone 80 mg loading dose, followed by 40 mg q 12 hourly for at least 7 days, and in patients with an increasing CRP or a worsening clinical status, increase the dose to 80 mg q 12 hourly (and then 120 mg if required), AA 3 g IV q 6 hourly for at least 7 days, thiamine 200 mg IV q 12 hourly, heparin depending on contraindications and whether the patient is at risk, and supplemental oxygen ox. This treatment acts synergistically to restore microvascular function in patients with COVID-19. It has been suggested that the following supplements should also be administered: melatonin (6–12 mg at night), famotidine (40 mg) daily (20 mg in case of renal impairment), vitamin D (2000-4000 u PO daily), elemental zinc (50-75 mg daily), magnesium, atorvastatin (80 mg/daily), and thiamine, which is essential for cellular energy production. Thiamine, along with AA and glucocorticoids, has been shown to reduce delirium in critically ill patients with COVID-19 [210]. Another study hypothesized that combining VC and glycyrrhizic acid is a potential option for treating COVID-19 patients. VC and glycyrrhizic acid individually have potent pharmacological efficacy against viral pneumonia. The research used a strategy of network pharmacology and bioinformatics (including GO and KEGG) to uncover the integrative pharmacological mechanism of VC and glycyrrhizic acid against COVID-19. Findings suggest that VC and glycyrrhizic acid may be able to suppress COVID-19 through their combined antioxidative, antiviral, and anti-inflammatory effects, along with activating the immune system, specifically, T-cell activation and leukocyte adhesion.

Moreover, according to KEGG pathway analysis, the pharmacological mechanisms of VC and glycyrrhizic acid against COVID-19 involve specific modulations of immune responses [211]. Findings suggest that a combination of VC, curcumin, and glycyrrhizic acid may help regulate the immune response to combat COVID-19 infections and inhibit excessive inflammatory responses to prevent the onset of the cytokine storm [212]. Based on recent advanced research, a novel combination of VC, curcumin, and glycyrrhizic acid, which has potential against COVID-19 infection, has been developed. Specifically, it has the potential to regulate the innate immune response by acting on the NOD-like and Toll-like signaling pathways. They promote interferon production and activate and balance T cells. They also regulate the inflammatory response by inhibiting the PI3K/AKT, NF- κ B, and MAPK signaling pathways.

The combination of VC and other widely used agents with antioxidant properties, i.e., quercetin, methylene blue, or thiamine, results in increased efficacy through additive effect. The following advantages have been noted: an increased saturation rate, renal function and vascular barrier improvement, and an increased anti-inflammatory effect, requisite to lipid and carbon metabolism regulation. In COVID-19 prevention and/or in mild cases of infection, vitamin C supplementation possibly benefits patients.

5. Conclusions

The facts that the polymorphism of solute carrier family genes affects the uptake of VC according to the phylogenic origin of patients; that it affects their health conditions, such as T2DM, which increases the risk of hospitalization and death in COVID-19; and

that there are interactions between the solute carrier family and a broad spectrum of cardiological diseases all suggest an increased susceptibility of patients to complications and risk of death.

Surprisingly, despite the best efforts, only a few publications have focused directly on the role of VC transporters and their modulation as factors related to the severity of COVID-19. Although there are many publications concerning the role of VC in treating patients with COVID-19, no paper has simultaneously considered the nuances of VC uptake, transport, and genetic variability. The genetic variability related to SNPs is significant in differentiating populations of people across the world. The potential role of SNPs in the susceptibility to a spectrum of diseases with differing severities seems to be well-known, and, according to collected data, the comparison between allele frequency, an abundance of bacteria species in gut microbiota, and a correlation with the plasma levels of VC should be developed as soon as possible, especially since indirect sources indicate that, for example, the abundance of *E. coli* in the gut microbiota of patients with severe symptoms of COVID-19 is higher than in the gut microbiota of healthy patients.

Undoubtedly, studies of VC's influence on the prevention of infection and treatment during COVID-19 should include an analysis of the *slc23* and *slc2* family of genes' polymorphism and expression as the essential factors affecting the uptake and regulation of ascorbic acid levels in organisms. Additionally, studies devoted to research on the gut microbiome could affect the conclusions reached by analyzing glycyrrhizic acid, VC, and curcumin. These compounds affect gut microbiota and can suppress or induce the expression of many genes associated with the regulation of the cell cycle, apoptosis, cell adhesion, phosphatases, and kinases, but they also act as epigenetic modulators.

On the other hand, there are many mutually cancelling publications concerning the role of VC in minimizing the symptoms of COVID-19. The influence of VC could be cleared up if it is related to genetic background. Undoubtedly, the role of VC in human organisms has been efficiently clarified, but many unexplored areas still require intensive basic and clinical research. The most puzzling phenomena are the broad spectrum of VC's activities and the equally wide spectrum of results obtained with different modes of administration and different dosages. Finally, knowing that VC is a most-welcomed supplement in the human organism, and even above-average dosages of that vitamin are not so prone to risk, it could be confirmed that even rare cases of a positive impact could be considered as positive evidence of a supportive effect for VC.

The positive influence of VC has been mentioned as improving the immune system [101], regulating the immune system [121–125], decreasing ROS [35,125,144,149], and reducing inflammatory markers [38,43]. The positive influence of VC has been described in the apoptosis of cancer tumors [116,119]. In viral infections, including COVID-19, the positive effect of VC administration has also been reported [135–138,203,209]. The neutral effect of VC has been mentioned in the case of patients with COVID-19 in a few publications [47,48,190–194,196,198–200], and the adverse effects of the administration of high doses of VC have been mentioned as an aspect of urinary and kidney stones [88–90] and nephropathy [45]. These last effects were found to be neutral. The negatives mainly concern problems with the continuous and extended administration of the vitamin, rather than being basically adverse effects. In light of aforementioned information, it can be concluded that the balance between the positive and negative effects should be considered only when there is evidence of antagonism between vitamin C and other therapeutic agents. VC does not harm, so using that vitamin as an additive to the primary treatment will not be wrong because there is always a chance that HDIVC will improve the effects.

This leads to the conclusion that VC helps, rather than harms, but it should not be considered a supplement that plays an essential role in COVID-19 treatment.

Author Contributions: Conceptualization, K.S. and K.G.-B.; formal analysis, K.S. and A.K.; data curation, K.G.-B., N.W.-K. and J.K.-P.; writing—original draft preparation, K.G.-B., N.W.-K., A.B., J.K.-P., J.P. and D.S.; writing—review and editing, K.G.-B., K.S., N.W.-K., A.B., J.K.-P. and A.K.; visualization, K.G.-B., K.S. and A.K.; supervision, K.G.-B., K.S. and E.G.-K.; funding acquisition, K.S. and E.G.-K. All authors have read and agreed to the published version of the manuscript.

Funding: This article was financed with funds from a contest held by the "Excellent Initiative Research University" program at Nicolaus Copernicus University in Toruń.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

- 1. Wu, F.; Zhao, S.; Yu, B.; Chen, Y.M.; Wang, W.; Song, Z.G.; Hu, Y.; Tao, Z.W.; Tian, Y.; Pei, Y.Y.; et al. A new coronavirus associated with human respiratory disease in China. *Nature* **2020**, *579*, 265–269. [CrossRef] [PubMed]
- World Health Organisation (WHO). WHO Coronavirus (COVID-19) Dashboard. 2022. Available online: https://covid19.who.int/ (accessed on 4 February 2022).
- 3. European Centre for Disease Prevention and Control (ECDC). Variants of Concern. Available online: https://www.ecdc.europa. eu/en/covid-19/variants-concern (accessed on 16 May 2022).
- Mehta, P.; Porter, J.; Manson, J.; Isaacs, J.; Openshaw, P.; McInnes, I.; Summers, C.; Chambers, R. Therapeutic blockade of granulocyte macrophage colony-stimulating factor in COVID-19-associated hyperinflammation: Challenges and opportunities. *Lancet Respir. Med.* 2020, *8*, 822–830. [CrossRef]
- Lee, J.; Park, S.S.; Kim, T.Y.; Lee, D.G.; Kim, D.W. Lymphopenia as a Biological Predictor of Outcomes in COVID-19 Patients: A Nationwide Cohort Study. *Cancers* 2021, 13, 471. [CrossRef]
- Gibson, P.G.; Qin, L.; Puah, S.H. COVID-19 acute respiratory distress syndrome (ARDS): Clinical features and differences from typical pre-COVID-19 ARDS. *Med. J. Aust.* 2020, 213, 54–56. [CrossRef] [PubMed]
- Rabaan, A.A.; Al-Ahmed, S.H.; Muhammad, J.; Khan, A.; Sule, A.A.; Tirupathi, R.; Mutair, A.A.; Alhumaid, S.; Al-Omari, A.; Dhawan, M.; et al. Role of Inflammatory Cytokines in COVID-19 Patients: A Review on Molecular Mechanisms, Immune Functions, Immunopathology and Immunomodulatory Drugs to Counter Cytokine Storm. *Vaccines* 2021, 9, 436. [CrossRef]
- 8. Yang, L.; Xie, X.; Tu, Z.; Fu, J.; Xu, D.; Zhou, Y. The signal pathways and treatment of cytokine storm in COVID-19. *Signal Transduct. Target. Ther.* **2021**, *6*, 255. [CrossRef]
- 9. Deana, C.; Vetrugno, L.; Fabris, M.; Curcio, F.; Sozio, E.; Tascini, C.; Bassi, F. Pericardial Cytokine "Storm" in a COVID-19 Patient: The Confirmation of a Hypothesis. *Inflammation* **2022**, *45*, 1–5. [CrossRef]
- Jing, X.; Xu, M.; Song, D.; Yue, T.; Wang, Y.; Zhang, P.; Zhong, Y.; Zhang, M.; Tsan-Yuk Lam, T.; Faria, N.R.; et al. Association between inflammatory cytokines and anti-SARS-CoV-2 antibodies in hospitalized patients with COVID-19. *Immun. Ageing* 2022, 19, 12. [CrossRef]
- 11. Ou, X.; Liu, Y.; Lei, X.; Li, P.; Mi, D.; Ren, L.; Guo, L.; Guo, R.; Chen, T.; Hu, J.; et al. Characterization of spike glycoprotein of SARS-CoV-2 on virus entry and its immune cross-reactivity with SARS-CoV. *Nat. Commun.* **2020**, *11*, 1620. [CrossRef]
- Beyerstedt, S.; Casaro, E.B.; Rangel, É.B. COVID-19: Angiotensin-converting enzyme 2 (ACE2) expression and tissue susceptibility to SARS-CoV-2 infection. *Eur. J. Clin. Microbiol. Infect. Dis.* 2021, 40, 905–919. [CrossRef]
- 13. Wang, K.; Chen, W.; Zhang, Z.; Deng, Y.; Lian, J.Q.; Du, D.; Wei, D.; Zhang, Y.; Sun, X.; Gong, L.; et al. CD147-spike protein is a novel route for SARS-CoV-2 infection to host cells. *Signal. Transduct. Target Ther.* **2020**, *5*, 283. [CrossRef] [PubMed]
- Ma, A.; Zhang, L.; Ye, X.; Chen, J.; Yu, J.; Zhuang, L.; Weng, C.; Petersen, F.; Wang, Z.; Yu, X. High Levels of Circulating IL-8 and Soluble IL-2R Are Associated with Prolonged Illness in Patients with Severe COVID-19. *Front. Immunol.* 2021, 12, 626235. [CrossRef] [PubMed]
- Mortaz, E.; Tabarsi, P.; Jamaati, H.; Dalil Roofchayee, N.; Dezfuli, N.K.; Hashemian, S.M.; Moniri, A.; Malekmohammad, M.; Mansouri, D.; Varahram, M.; et al. Increased Serum Levels of Soluble TNF-α Receptor Is Associated with ICU Mortality in COVID-19 Patients. *Front. Immunol.* 2021, 12, 592727. [CrossRef] [PubMed]
- 16. Mu, X.; Liu, K.; Li, H.; Wang, F.S.; Xu, R. Granulocyte-macrophage colony-stimulating factor: An immunotarget for sepsis and COVID-19. *Cell Mol. Immunol.* **2021**, *18*, 2057–2058. [CrossRef] [PubMed]
- 17. Khalil, B.A.; Elemam, N.M.; Maghazachi, A.A. Chemokines and chemokine receptors during COVID-19 infection. *Comput. Struct. Biotechnol. J.* **2021**, *19*, 976–988. [CrossRef] [PubMed]
- Arijs, I.; De Hertogh, G.; Lemaire, K.; Quintens, R.; Van Lommel, L.; Van Steen, K.; Leemans, P.; Cleynen, I.; Van Assche, G.; Vermeire, S.; et al. Mucosal gene expression of antimicrobial peptides in inflammatory bowel disease before and after first infliximab treatment. *PLoS ONE* 2009, *4*, e7984. [CrossRef] [PubMed]

- 19. Bild, A.H.; Yao, G.; Chang, J.T.; Wang, Q.; Potti, A.; Chasse, D.; Joshi, M.B.; Harpole, D.; Lancaster, J.M.; Berchuck, A.; et al. Oncogenic pathway signatures in human cancers as a guide to targeted therapies. *Nature* **2006**, *19*, 353–357. [CrossRef]
- Boldrick, J.C.; Alizadeh, A.A.; Diehn, M.; Dudoit, S.; Liu, C.L.; Belcher, C.E.; Botstein, D.; Staudt, L.M.; Brown, P.O.; Relman, D.A. Stereotyped and specific gene expression programs in human innate immune responses to bacteria. *Proc. Natl. Acad. Sci. USA* 2002, 22, 972–977. [CrossRef]
- Mardi, A.; Meidaninikjeh, S.; Nikfarjam, S.; Majidi; Zolbanin, N.; Jafari, R. Interleukin-1 in COVID-19 Infection: Immunopathogenesis and Possible Therapeutic Perspective. *Viral Immunol.* 2021, 34, 679–688. [CrossRef]
- Kokkotis, G.; Kitsou, K.; Xynogalas, I.; Spoulou, V.; Magiorkinis, G.; Trontzas, I.; Trontzas, P.; Poulakou, G.; Syrigos, K.; Bamias, G. Systematic review with meta-analysis: COVID-19 outcomes in patients receiving anti-TNF treatments. *Aliment. Pharmacol. Ther.* 2022, 55, 154–167. [CrossRef]
- Ragab, D.; Salah, E.H.; Taeimah, M.; Khattab, R.; Salem, R. The COVID-19 Cytokine Storm; What We Know So Far. Front. Immunol. 2020, 16, 1446. [CrossRef] [PubMed]
- Hayney, M.S.; Henriquez, K.M.; Barnet, J.H.; Ewers, T.; Champion, H.M.; Flannery, S.; Barrett, B. Serum IFN-γ-induced protein 10 (IP-10) as a biomarker for severity of acute respiratory infection in healthy adults. *J. Clin. Virol.* 2017, 90, 32–37. [CrossRef] [PubMed]
- Henriquez, K.M.; Hayney, M.S.; Xie, Y.; Zhang, Z.; Barrett, B. Association of interleukin-8 and neutrophils with nasal symptom severity during acute respiratory infection. *J. Med. Virol.* 2015, *87*, 330–337. [CrossRef] [PubMed]
- Hojyo, S.; Uchida, M.; Tanaka, K.; Hasebe, R.; Tanaka, Y.; Murakami, M.; Hirano, T. How COVID-19 induces cytokine storm with high mortality. *Inflamm. Regen.* 2020, 40, 37. [CrossRef]
- Blot, M.; Bour, J.B.; Quenot, J.P.; Bourredjem, A.; Nguyen, M.; Guy, J.; Monier, S.; Georges, M.; Large, A.; Dargent, A.; et al. The dysregulated innate immune response in severe COVID-19 pneumonia that could drive poorer outcome. *J. Transl. Med.* 2020, 18, 457; Erratum in *J. Transl. Med.* 2021, 19, 100. [CrossRef]
- Abdel-Hamed, E.F.; Ibrahim, M.N.; Mostafa, N.E.; Moawad, H.S.F.; Elgammal, N.E.; Darwiesh, E.M.; El-Rafey, D.S.; ElBadawy, N.E.; Al-Khoufi, E.A.; Hindawi, S.I. Role of interferon gamma in SARS-CoV-2-positive patients with parasitic infections. *Gut Pathog.* 2021, 13, 29. [CrossRef]
- Guo, Y.; Hu, K.; Li, Y.; Lu, C.; Ling, K.; Cai, C.; Wang, W.; Ye, D. Targeting TNF-α for COVID-19: Recent Advanced and Controversies. *Front. Public Health* 2022, 10, 833967. [CrossRef]
- Fontes, J.A.; Rose, N.R.; Čiháková, D. The varying faces of IL-6: From cardiac protection to cardiac failure. *Cytokine* 2015, 74, 62–68. [CrossRef]
- 31. Gostner, J.M.; Becker, K.; Fuchs, D.; Sucher, R. Redox regulation of the immune response. Redox Rep. 2013, 18, 88–94. [CrossRef]
- 32. World Health Organization (WHO). Therapeutics and COVID-19. Living Guideline. 2021. Available online: https://www.who. int/publications/i/item/WHO-2019-nCoV-therapeutics-2022.3 (accessed on 7 December 2021).
- Aisa-Alvarez, A.; Soto, M.E.; Guarner-Lans, V.; Camarena-Alejo, G.; Franco-Granillo, J.; Martinez-Rodríguez, E.A.; Ávila, G.R.; Pech, L.M.; Pérez-Torres, I. Usefulness of antioxidants as adjuvant therapy for septic shock: A randomized clinical trial. *Medicina* 2020, 56, 619. [CrossRef]
- 34. Guloyan, V.; Oganesian, B.; Baghdasaryan, N.; Yeh, C.; Singh, M.; Guilford, F.; Ting, Y.S.; Venketaraman, V. Glutathione Supplementation as an Adjunctive Therapy in COVID-19. *Antioxidants* **2020**, *9*, 914. [CrossRef] [PubMed]
- 35. Chavarría, A.P.; Vázquez, R.R.V.; Cherit, J.G.D.; Bello, H.H.; Suastegui, H.C.; Moreno-Castañeda, L.; Alanís Estrada, G.; Hernández, F.; González-Marcos, O.; Saucedo-Orozco, H.; et al. Antioxidants and pentoxifylline as coadjuvant measures to standard therapy to improve prognosis of patients with pneumonia by COVID-19. *Comput. Struct. Biotechnol. J.* 2021, 19, 1379–1390. [CrossRef] [PubMed]
- Polonikov, A. Endogenous deficiency of glutathione as the most likely cause of serious manifestations and death in patients with the novel coronavirus infection (COVID-19): A hypothesis based on literature data and own observations. ACS Infect. Dis. 2020, 6, 1558–1562. [CrossRef]
- Horowitz, R.I.; Freeman, P.R.; Bruzzese, J. Ecacy of glutathione therapy in relieving dyspnea associated with COVID-19 pneumonia: A report of 2 cases. *Respir. Med. Case Rep.* 2020, 30, 101063. [PubMed]
- Hiedra, R.; Lo, K.B.; Elbashabsheh, M.; Gul, F.; Wright, R.M.; Albano, J.; Azmaiparashvili, Z.; Patarroyo Aponte, G. The use of IV vitamin C for patients with COVID-19: A case series. *Expert Rev. Anti-Infect. Ther.* 2020, 18, 1259–1261. [CrossRef]
- Colunga Biancatelli, R.M.L.; Berrill, M.; Catravas, J.D.; Marik, P.E. Quercetin and Vitamin C: An Experimental, Synergistic Therapy for the Prevention and Treatment of SARS-CoV-2 Related Disease (COVID-19). Front. Immunol. 2020, 11, 1451. [CrossRef]
- Vogel-González, M.; Talló-Parra, M.; Herrera-Fernández, V.; Pérez-Vilaró, G.; Chillón, M.; Nogués, X.; Gómez-Zorrilla, S.; López-Montesinos, I.; Villar, J.; Sorli-Redó, M.L.; et al. Low zinc levels at clinical admission associates with poor outcomes in COVID-19. Nutrients 2021, 13, 562. [CrossRef]
- Jothimani, D.; Kailasam, E.; Danielraj, S.; Nallathambi, B.; Ramachandran, H.; Sekar, P.; Manoharan, S.; Ramani, V.; Narasimhan, G.; Kaliamoorthy, I.; et al. COVID-19: Poor outcomes in patients with zinc deficiency. *Int. J. Infect. Dis.* 2020, 100, 343–349. [CrossRef]
- 42. Waqas Khan, H.M.; Parikh, N.; Megala, S.M.; Predeteanu, G.S. Unusual Early Recovery of a Critical COVID-19 Patient after Administration of Intravenous Vitamin C. *Am. J. Case Rep.* 2020, *21*, e925521. [CrossRef]

- 43. Zhao, B.; Ling, Y.; Li, J.; Peng, Y.; Huang, J.; Wang, Y.; Qu, H.; Gao, Y.; Li, Y.; Hu, B.; et al. Beneficial aspects of high dose intravenous vitamin C on patients with COVID-19 pneumonia in severe condition: A retrospective case series study. *Ann. Palliat. Med.* **2021**, *10*, 1599–1609. [CrossRef]
- Deliwala, S.S.; Ponnapalli, A.; Seedahmed, E.; Berrou, M.; Bachuwa, G.; Chandran, A. A 29-Year-Old Male with a Fatal Case of COVID-19 Acute Respiratory Distress Syndrome (CARDS) and Ventilator-Induced Lung Injury (VILI). *Am. J. Case Rep.* 2020, 21, e926136. [CrossRef] [PubMed]
- 45. Fontana, F.; Cazzato, S.; Giovanella, S.; Ballestri, M.; Leonelli, M.; Mori, G.; Alfano, G.; Ligabue, G.; Magistroni, R.; Cenacchi, G.; et al. Oxalate Nephropathy Caused by Excessive Vitamin C Administration in 2 Patients with COVID-19. *Kidney Int. Rep.* 2020, *5*, 1815–1822. [CrossRef] [PubMed]
- Capone, S.; Abramyan, S.; Ross, B.; Rosenberg, J.; Zeibeq, J.; Vasudevan, V.; Samad, R.; Gerolemou, L.; Pinelis, E.; Gasperino, J.; et al. Characterization of Critically Ill COVID-19 Patients at a Brooklyn Safety-Net Hospital. *Cureus* 2020, 12, e9809. [CrossRef] [PubMed]
- JamaliMoghadamSiahkali, S.; Zarezade, B.; Koolaji, S.; SeyedAlinaghi, S.; Zendehdel, A.; Tabarestani, M.; Sekhavati Moghadam, E.; Abbasian, L.; Dehghan Manshadi, S.A.; Salehi, M.; et al. Safety and effectiveness of high-dose vitamin C in patients with COVID-19: A randomized open-label clinical trial. *Eur. J. Med. Res.* 2021, 26, 20. [CrossRef]
- 48. Kumari, P.; Dembra, S.; Dembra, P.; Bhawna, F.; Gul, A.; Ali, B.; Sohail, H.; Kumar, B.; Memon, M.K.; Rizwan, A. The Role of Vitamin C as Adjuvant Therapy in COVID-19. *Cureus* 2020, *12*, 10–13. [CrossRef]
- Lutchmansingh, F.K.; Hsu, J.W.; Bennett, F.I.; Badaloo, A.V.; McFarlane-Anderson, N.; Gordon-Strachan, G.M.; Wright-Pascoe, R.A.; Jahoor, F.; Boyne, M.S. Glutathione metabolism in type 2 diabetes and its relationship with microvascular complications and glycemia. *PLoS ONE* 2018, 13, e0198626. [CrossRef]
- 50. Sinaci, S.; Ocal, D.F.; Yetiskin, D.F.Y. Impact of vitamin D on the course of COVID-19 during pregnancy: A case control study. J. Steroid Biochem. Mol. Biol. 2021, 213, 105964. [CrossRef]
- 51. Schmitt, G.; Labdouni, S.; Soulimani, R.; Delamare, C.; Bouayed, J. Oxidative stress status and vitamin D levels of asymptomatic to mild symptomatic COVID-19 infections during the third trimester of pregnancy: A retrospective study in Metz, France. *J. Med. Virol.* **2022**, *94*, 2167–2173. [CrossRef]
- 52. Tekin, A.B.; Yassa, M.; Birol, P.; Unlu, S.N.; Bura, A.M.; Ayanogu, E.; Tug, N. Vitamin D status is not associated with clinical severity of COVID-19 in pregnant women. *Eur. J. Nutr.* **2021**, *61*, 1035–1041. [CrossRef]
- 53. ClinicalTrials.gov. Available online: www.clinicaltrials.gov (accessed on 14 February 2022).
- 54. Smirnoff, N. Ascorbic acid metabolism and functions: A comparison of plants and mammals. *Free Radic. Biol. Med.* **2018**, 122, 116–129. [CrossRef]
- Pérez-Torres, I.; Manzano-Pech, L.; Rubio-Ruíz, M.E.; Soto, M.E.; Guarner-Lans, V. Nitrosative stress and its association with cardiometabolic disorders. *Molecules* 2020, 25, 2555. [CrossRef] [PubMed]
- Jeffery, L.E.; Burke, F.; Mura, M.; Zheng, Y.; Qureshi, O.S.; Hewison, M.; Walker, L.S.K.; Lammas, D.A.; Raza, K.; Sansom, D.M. 1, 25-Dihydroxyvitamin D3 and IL-2 combine to inhibit T cell Production of inflammatory cytokines and promote development of regulatory T cells expressing CTLA-4 and FoxP3. J. Immunol. 2009, 183, 5458–5467. [CrossRef] [PubMed]
- Mrityunjaya, M.; Pavithra, V.; Neelam, R.; Janhavi, P.; Halami, P.M.; Ravindra, P.V. Immune-Boosting, Antioxidant and Antiinflammatory Food Supplements Targeting Pathogenesis of COVID-19. *Front. Immunol.* 2020, *11*, 570122. [CrossRef] [PubMed]
- Pérez-Torres, I.; Guarner-Lans, V.; Rubio-Ruiz, M.E. Reductive stress in inflammation-associated diseases and the pro-effect of antioxidant agents. Int. J. Mol. Sci. 2017, 18, 2098. [CrossRef] [PubMed]
- 59. Sánchez-Barceló, E.J.; Mediavilla, M.D.; Tan, D.X.; Reiter, R.J. Clinical uses of melatonin: Evaluation of human trials. *Curr. Med. Chem.* 2010, *17*, 2070–2095. [CrossRef]
- 60. Galley, H.F.; Lowes, D.A.; Allen, L.; Cameron, G.; Aucott, L.S.; Webster, N.R. Melatonin as a potential therapy for sepsis: A phase I dose escalation study and an ex vivo whole blood model under conditions of sepsis. J. Pineal Res. 2014, 56, 427–438. [CrossRef]
- Shneider, A.; Kudriavtsev, A.; Vakhrusheva, A. Can melatonin reduce the severity of COVID-19 pandemic? *Int. Rev. Immunol.* 2020, 39, 153–162. [CrossRef]
- 62. Guthappa, R. Molecular docking studies of N-acetylcysteine, zincacetylcysteine and niclosamide on SARS-Cov-2 protease and its comparison with hydroxychloroquine. *Chemarxiv* 2020. [CrossRef]
- 63. Zhongcheng, S.; Puyo, C.A. N-Acetylcysteine to combat COVID-19: An evidence review. *Ther. Clin. Risk Manag.* 2020, 16, 1047–1055.
- 64. Read, S.A.; Obeid, S.; Ahlenstiel, C.; Ahlenstiel, G. The role of zinc in antiviral immunity. Adv. Nutr. 2019, 10, 696–710. [CrossRef]
- 65. Mani, J.S.; Johnson, J.B.; Steel, J.C.; Broszczak, D.A.; Neilsen, P.M.; Walsh, K.B.; Naiker, M. Natural product-derived phytochemicals as potential agents against coronaviruses: A review. *Virus Res.* **2020**, *284*, 197989. [CrossRef] [PubMed]
- Berretta, A.A.; Silveira, M.A.D.; Cóndor Capcha, J.M.; De Jong, D. Propolis and its potential against SARS-CoV-2 infection mechanisms and COVID-19 disease: Running title: Propolis against SARS-CoV-2 infection and COVID-19. *Biomed. Pharmacother.* 2020, 131, 110622. [CrossRef] [PubMed]
- Liskova, A.; Samec, M.; Koklesova, L.; Samuel, S.M.; Zhai, K.; Al-Ishaq, R.K.; Abotaleb, M.; Nosal, V.; Kajo, K.; Ashrafizadeh, M.; et al. Flavonoids against the SARS-CoV-2 induced inflammatory storm. *Biomed. Pharmacother.* 2021, 138, 111430. [CrossRef] [PubMed]

- 68. Su, H.; Yao, S.; Zhao, W.; Li, M.; Liu, J.; Shang, W.; Xie, H.; Ke, C.; Gao, M.; Yu, K.; et al. Discovery of baicalin and baicalein as novel, natural product inhibitors of SARS-CoV-2 3CL protease in vitro. *bioRxiv* 2020. [CrossRef]
- 69. Da Silva Antonio, A.; Wiedemann, L.S.M.; Veiga-Junior, V.F. Natural products' role against COVID-19. *RSC Adv.* 2020, 10, 23379–23393. [CrossRef]
- Bhardwaj, V.K.; Singh, R.; Sharma, J.; Rajendran, V.; Purohit, R.; Kumar, S. Identification of bioactive molecules from tea plant as SARS-CoV-2 main protease inhibitors. J. Biomol. Struct. Dyn. 2020, 39, 3449–3458. [CrossRef]
- 71. Huynh, T.; Wang, H.; Luan, B. In Silico Exploration of the Molecular Mechanism of Clinically Oriented Drugs for Possibly Inhibiting SARS-CoV-2's Main Protease. J. Phys. Chem. Lett. 2020, 11, 4413–4420. [CrossRef]
- Enmozhi, S.K.; Raja, K.; Sebastine, I.; Joseph, J. Andrographolide as a potential inhibitor of SARS-CoV-2 main protease: An in silico approach. J. Biomol. Struct. Dyn. 2021, 39, 3092–3098. [CrossRef]
- 73. Kodchakorn, K.; Poovorawan, Y.; Suwannakarn, K.; Kongtawelert, P. Molecular modelling investigation for drugs and nutraceuticals against protease of SARS-CoV-2. *J. Mol. Graph. Model.* **2020**, *101*, 107717. [CrossRef]
- Pandey, A.K.; Verma, S. An in-silico evaluation of dietary components for structural inhibition of SARS-Cov-2 main protease. J. Biomol. Struct. Dyn. 2020, 40, 136–142. [CrossRef]
- 75. Utomo, R.Y.; Ikawati, M.; Meiyanto, E. Revealing the Potency of Citrus and Galangal Constituents to Halt SARS-CoV-2 Infection. *Preprints* **2020**, 2020030214. [CrossRef]
- Lang, A.; Lahav, M.; Sakhnini, E.; Barshack, I.; Fidder, H.H.; Avidan, B.; Bardan, E.; Hershkoviz, R.; Bar-Meir, S.; Chowers, Y. Allicin inhibits spontaneous and TNF-alpha induced secretion of proinflammatory cytokines and chemokines from intestinal epithelial cells. *Clin. Nutr. Edinb. Scotl.* 2004, 23, 1199–1208. [CrossRef] [PubMed]
- 77. Firuzi, O.; Miri, R.; Tavakkoli, M.; Saso, L. Antioxidant Therapy: Current Status and Future Prospects. *Curr. Med. Chem.* 2011, 18, 3871–3888. [CrossRef]
- 78. King, C.G.; Waugh, W.A. The chemical nature of vitamin C. Science 1932, 75, 357–358. [CrossRef]
- 79. Carpenter, K.J. The History of Scurvy and Vitamin C; Cambridge University Press: Cambridge, UK, 1986.
- Chatterjee, I.B.; Kar, N.C.; Ghosh, N.C.; Guha, B.C. Biosynthesis of L-ascorbic acid: Missing steps in animals incapable of synthesizing the vitamin. *Nature* 1961, 192, 163–164. [CrossRef] [PubMed]
- 81. Amrein, K.; Oudemans-van Straaten, H.M.; Berger, M.M. Vitamin therapy in critically ill patients: Focus on thiamine, vitamin C., and vitamin D. *Intensive Care Med.* **2018**, *44*, 1940–1944. [CrossRef] [PubMed]
- McEvoy, C.T.; Shorey-Kendrick, L.E.; Milner, K.; Schilling, D.; Tiller, C.; Vuylsteke, B.; Scherman, A.; Jackson, K.; Haas, D.M.; Harris, J.; et al. Oral Vitamin C (500 mg/d) to Pregnant Smokers Improves Infant Airway Function at 3 Months (VCSIP). A Randomized Trial. *Am. J. Respir. Crit. Care Med.* 2019, 199, 1139–1147. [CrossRef]
- Cerullo, G.; Negro, M.; Parimbelli, M.; Pecoraro, M.; Perna, S.; Liguori, G.; Rondanelli, M.; Cena, H.; D'Antona, G. The Long History of Vitamin C: From Prevention of the Common Cold to Potential Aid in the Treatment of COVID-19. *Front. Immunol.* 2020, 11, 574029. [CrossRef]
- 84. Kaźmierczak-Barańska, J.; Boguszewska, K.; Adamus-Grabicka, A.; Karwowski, B.T. Two Faces of Vitamin C-Antioxidative and Pro-Oxidative Agent. *Nutrients* **2020**, *12*, 1501. [CrossRef]
- Moyad, M.A.; Combs, M.A.; Vrablic, A.S.; Velasquez, J.; Turner, B.; Bernal, S. Vitamin C metabolites, independent of smoking status, significantly enhance leukocyte, but not plasma ascorbate concentrations. *Adv. Ther.* 2008, 25, 995–1009. [CrossRef]
- Kim, H.; Bae, S.; Yu, Y.; Kim, Y.; Kim, H.R.; Hwang, Y.I.; Kang, J.S.; Lee, W.J. The analysis of vitamin C concentration in organs of gulo(-/-) mice upon vitamin C withdrawal. *Immune Netw.* 2012, 12, 18–26. [CrossRef] [PubMed]
- Ngo, B.; Van Riper, J.M.; Cantley, L.C.; Yun, J. Targeting cancer vulnerabilities with high-dose vitamin C. *Nat. Rev. Cancer* 2019, 19, 271–282. [CrossRef] [PubMed]
- 88. Taylor, E.N.; Stampfer, M.J.; Curhan, G.C. Dietary factors and the risk of incident kidney stones in men: New insights after 14 years of follow-up. *J. Am. Soc. Nephrol.* 2004, 15, 3225–3232. [CrossRef] [PubMed]
- 89. Jiang, K.; Tang, K.; Liu, H.; Xu, H.; Ye, Z.; Chen, Z. Ascorbic Acid Supplements and Kidney Stones Incidence Among Men and Women: A systematic review and meta-analysis. *Urol. J.* **2019**, *16*, 115–120. [CrossRef] [PubMed]
- Doseděl, M.; Jirkovský, E.; Macáková, K.; Krčmová, L.K.; Javorská, L.; Pourová, J.; Mercolini, L.; Remião, F.; Nováková, L.; Mladěnka, P.; et al. Vitamin C-Sources, Physiological Role, Kinetics, Deficiency, Use, Toxicity, and Determination. *Nutrients* 2021, 13, 615. [CrossRef]
- 91. Moores, J. Vitamin C: A wound healing perspective. Br. J. Community Nurs. 2013, 18, S6–S11. [CrossRef]
- 92. Aghajanian, P.; Hall, S.; Wongworawat, M.D.; Mohan, S. The Roles and Mechanisms of Actions of Vitamin C in Bone: New Developments. J. Bone Miner. Res. 2015, 30, 1945–1955. [CrossRef]
- Mousavi, S.; Bereswill, S.; Heimesaat, M.M. Immunomodulatory and Antimicrobial Effects of Vitamin C. Eur. J. Microbiol. Immunol. 2019, 9, 73–79. [CrossRef]
- 94. Eid, W.; Abdel-Rehim, W. Vitamin C promotes pluripotency of human induced pluripotent stem cells via the histone demethylase JARID1A. *Biol. Chem.* **2016**, *397*, 1205–1213. [CrossRef]
- 95. Englard, S.; Seifter, S. The biochemical functions of ascorbic acid. Annu. Rev. Nutr. 1986, 6, 365–406. [CrossRef]
- 96. Evans, R.M.; Currie, L.; Campbell, A. The distribution of ascorbic acid between various cellular components of blood, in normal individuals, and its relation to the plasma concentration. *Br. J. Nutr.* **1982**, *47*, 473–482. [CrossRef] [PubMed]

- 97. Ang, A.; Pullar, J.M.; Currie, M.J.; Vissers, M. Vitamin C and immune cell function in inflammation and cancer. *Biochem. Soc. Trans.* **2018**, *46*, 1147–1159. [CrossRef] [PubMed]
- 98. Manning, J.; Mitchell, B.; Appadurai, D.A.; Shakya, A.; Pierce, L.J.; Wang, H.; Nganga, V.; Swanson, P.C.; May, J.M.; Tantin, D.; et al. Vitamin C promotes maturation of T-cells. *Antioxid. Redox Signal.* **2013**, *19*, 2054–2067. [CrossRef] [PubMed]
- 99. Van Gorkom, G.; Klein Wolterink, R.; Van Elssen, C.; Wieten, L.; Germeraad, W.; Bos, G. Influence of Vitamin C on Lymphocytes: An Overview. *Antioxidants* **2018**, *7*, 41. [CrossRef]
- Schwager, J.; Bompard, A.; Weber, P.; Raederstorff, D. Ascorbic acid modulates cell migration in differentiated HL-60 cells and peripheral blood leukocytes. *Mol. Nutr. Food Res.* 2015, 59, 1513–1523. [CrossRef]
- 101. De la Fuente, M.; Sánchez, C.; Vallejo, C.; Díaz-Del Cerro, E.; Arnalich, F.; Hernanz, Á. Vitamin C and vitamin C plus E improve the immune function in the elderly. *Exp. Gerontol.* **2020**, *142*, 11118. [CrossRef]
- Bozonet, S.M.; Carr, A.C.; Pullar, J.M.; Vissers, M.C. Enhanced human neutrophil vitamin C status, chemotaxis and oxidant generation following dietary supplementation with vitamin C-rich SunGold kiwifruit. *Nutrients* 2015, 7, 2574–2588. [CrossRef]
- Marchioli, R.; Schweiger, C.; Levantesi, G.; Tavazzi, L.; Valagussa, F. Antioxidant vitamins and prevention of cardiovascular disease: Epidemiological and clinical trial data. *Lipids* 2001, *36* (Suppl. S1), 53–63. [CrossRef]
- Woollard, K.J.; Phillips, D.C.; Griffiths, H.R. Direct modulatory effect of C-reactive protein on primary human monocyte adhesion to human endothelial cells. *Clin. Exp. Immunol.* 2002, 130, 256–262. [CrossRef]
- Atasever, B.; Ertan, N.Z.; Erdem-Kuruca, S.; Karakas, Z. In vitro effects of vitamin C and selenium on NK activity of patients with beta-thalassemia major. *Pediatr. Hematol. Oncol.* 2006, 23, 187–197. [CrossRef]
- Kim, J.E.; Cho, H.S.; Yang, H.S.; Jung, D.J.; Hong, S.W.; Hung, C.F.; Lee, W.J.; Kim, D. Depletion of ascorbic acid impairs NK cell activity against ovarian cancer in a mouse model. *Immunobiology* 2012, 217, 873–881. [CrossRef] [PubMed]
- 107. Rice, M.E. Ascorbate regulation and its neuroprotective role in the brain. Trends Neurosci. 2000, 23, 209–216. [CrossRef]
- 108. Harrison, F.E.; May, J.M. Vitamin C function in the brain: Vital role of the ascorbate transporter SVCT2. *Free Radic. Biol. Med.* 2009, 46, 719–730. [CrossRef] [PubMed]
- Moretti, M.; Fraga, D.B.; Rodrigues, A. Preventive and therapeutic potential of ascorbic acid in neurodegenerative diseases. CNS Neurosci. Ther. 2017, 23, 921–929. [CrossRef] [PubMed]
- Lv, S.J.; Zhang, G.H.; Xia, J.M.; Yu, H.; Zhao, F. Early use of high-dose vitamin C is beneficial in treatment of sepsis. *Ir. J. Med. Sci.* 2021, 190, 1183–1188. [CrossRef]
- 111. Kuhn, S.O.; Meissner, K.; Mayes, L.M.; Bartels, K. Vitamin C in sepsis. Curr. Opin. Anaesthesiol. 2018, 31, 55-60. [CrossRef]
- 112. Blaszczak, W.; Barczak, W.; Masternak, J.; Kopczyński, P.; Zhitkovich, A.; Rubiś, B. Vitamin C as a Modulator of the Response to Cancer Therapy. *Molecules* **2019**, *24*, 453. [CrossRef]
- 113. Böttger, F.; Vallés-Martí, A.; Cahn, L.; Jimenez, C.R. High-dose intravenous vitamin C, a promising multi-targeting agent in the treatment of cancer. *J. Exp. Clin. Cancer Res.* 2021, 40, 343. [CrossRef]
- 114. Meščić Macan, A.; Gazivoda Kraljević, T.; Raić-Malić, S. Therapeutic Perspective of Vitamin C and Its Derivatives. *Antioxidants* **2019**, *8*, 247. [CrossRef]
- 115. Chen, Q.; Espey, M.G.; Sun, A.Y.; Pooput, C.; Kirk, K.L.; Krishna, M.C.; Khosh, D.B.; Drisko, J.; Levine, M. Pharmacologic doses of ascorbate act as a prooxidant and decrease growth of aggressive tumor xenografts in mice. *Proc. Natl. Acad. Sci. USA* 2008, 105, 11105–11109. [CrossRef]
- 116. Doskey, C.M.; Buranasudja, V.; Wagner, B.A.; Wilkes, J.G.; Du, J.; Cullen, J.J.; Buettner, G.R. Tumor cells have decreased ability to metabolize H₂O₂: Implications for pharmacological ascorbate in cancer therapy. *Redox Biol.* **2016**, *10*, 274–284. [CrossRef] [PubMed]
- 117. Hong, S.W.; Jin, D.H.; Hahm, E.S.; Yim, S.H.; Lim, J.S.; Kim, K.I.; Yang, Y.; Lee, S.S.; Kang, J.S.; Lee, W.J.; et al. Ascorbate (vitamin C) induces cell death through the apoptosis-inducing factor in human breast cancer cells. Oncol. Rep. 2007, 18, 811–815. [CrossRef] [PubMed]
- 118. Tronci, L.; Serreli, G.; Piras, C.; Frau, D.V.; Dettori, T.; Deiana, M.; Murgia, F.; Santoru, M.L.; Spada, M.; Leoni, V.P.; et al. Vitamin C Cytotoxicity and Its Effects in Redox Homeostasis and Energetic Metabolism in Papillary Thyroid Carcinoma Cell Lines. *Antioxidants* 2021, 10, 809. [CrossRef]
- 119. Hoppe, C.; Freuding, M.; Büntzel, J.; Münstedt, K.; Hübner, J. Clinical efficacy and safety of oral and intravenous vitamin C use in patients with malignant diseases. *J. Cancer Res. Clin. Oncol.* **2021**, 147, 3025–3042. [CrossRef] [PubMed]
- 120. Heaney, M.L.; Gardner, J.R.; Karasavvas, N.; Golde, D.W.; Scheinberg, D.A.; Smith, E.A.; O'Connor, O.A. Vitamin C antagonizes the cytotoxic effects of antineoplastic drugs. *Cancer Res.* 2008, *68*, 8031–8038. [CrossRef] [PubMed]
- 121. Jeng, K.C.; Yang, C.S.; Siu, W.Y.; Tsai, Y.S.; Liao, W.J.; Kuo, J.S. Supplementation with vitamins C and E enhances cytokine production by peripheral blood mononuclear cells in healthy adults. *Am. J. Clin. Nutr.* **1996**, *64*, 960–965. [CrossRef] [PubMed]
- 122. Mohammed, B.M.; Fisher, B.J.; Kraskauskas, D.; Farkas, D.; Brophy, D.F.; Fowler, A.A., 3rd; Natarajan, R. Vitamin C: A novel regulator of neutrophil extracellular trap formation. *Nutrients* **2013**, *5*, 3131–3151. [CrossRef] [PubMed]
- 123. Molina, N.; Morandi, A.C.; Bolin, A.P.; Otton, R. Comparative effect of fucoxanthin and vitamin C on oxidative and functional parameters of human lymphocytes. *Int. Immunopharmacol.* **2014**, 22, 41–50. [CrossRef]
- 124. Gao, Y.L.; Lu, B.; Zhai, J.H.; Liu, Y.C.; Qi, H.X.; Yao, Y.; Chai, Y.F.; Shou, S.T. The Parenteral Vitamin C Improves Sepsis and Sepsis-Induced Multiple Organ Dysfunction Syndrome via Preventing Cellular Immunosuppression. *Mediat. Inflamm.* 2017, 2017, 4024672. [CrossRef]

- 125. Hemilä, H. Vitamin C and Infections. Nutrients 2017, 9, 339. [CrossRef]
- 126. Hemilä, H.; Louhiala, P. Vitamin C may affect lung infections. J. R. Soc. Med. 2007, 100, 495–498. [CrossRef] [PubMed]
- 127. Carr, A.C. Vitamin C in Pneumonia and Sepsis. In *Vitamin C: New Biochemical and Functional Insights;* Chen, Q., Ed.; CRC Press: Boca Raton, FL, USA, 2020; pp. 115–135.
- 128. Fisher, B.J.; Seropian, I.M.; Kraskauskas, D.; Thakkar, J.N.; Voelkel, N.F.; Fowler, A.A., 3rd; Natarajan, R. Ascorbic acid attenuates lipopolysaccharide-induced acute lung injury. *Crit. Care Med.* **2011**, *39*, 1454–1460. [CrossRef] [PubMed]
- 129. Fisher, B.J.; Kraskauskas, D.; Martin, E.J.; Farkas, D.; Wegelin, J.A.; Brophy, D.; Ward, K.R.; Voelkel, N.F.; Fowler, A.A., 3rd; Natarajan, R. Mechanisms of attenuation of abdominal sepsis induced acute lung injury by ascorbic acid. *Am. J. Physiol. Lung Cell. Mol. Physiol.* 2012, 303, 20–32. [CrossRef] [PubMed]
- 130. Shrestha, D.B.; Budhathoki, P.; Sedhai, Y.R.; Mandal, S.K.; Shikhrakar, S.; Karki, S.; Baniya, R.K.; Kashiouris, M.G.; Qiao, X.; Fowler, A.A. Vitamin C in Critically Ill Patients: An Updated Systematic Review and Meta-Analysis. *Nutrients* 2021, 13, 3564. [CrossRef]
- 131. Hemilä, H.; Chalker, E. Vitamin C Can Shorten the Length of Stay in the ICU: A Meta-Analysis. Nutrients 2019, 11, 708. [CrossRef]
- 132. Fowler, A.A., 3rd; Truwit, J.D.; Hite, R.D.; Morris, P.E.; DeWilde, C.; Priday, A.; Fisher, B.; Thacker, L.R., 2nd; Natarajan, R.; Brophy, D.F.; et al. Effect of Vitamin C Infusion on Organ Failure and Biomarkers of Inflammation and Vascular Injury in Patients with Sepsis and Severe Acute Respiratory Failure: The CITRIS-ALI Randomized Clinical Trial. JAMA 2019, 322, 1261–1270. [CrossRef]
- Ahn, J.H.; Oh, D.K.; Huh, J.W.; Lim, C.M.; Koh, Y.; Hong, S.B. Vitamin C alone does not improve treatment outcomes in mechanically ventilated patients with severe sepsis or septic shock: A retrospective cohort study. *J. Thorac. Dis.* 2019, 11, 1562–1570. [CrossRef]
- 134. Shin, T.G.; Kim, Y.J.; Ryoo, S.M.; Hwang, S.Y.; Jo, I.J.; Chung, S.P.; Choi, S.H.; Suh, G.J.; Kim, W.Y. Early Vitamin C and Thiamine Administration to Patients with Septic Shock in Emergency Departments: Propensity Score-Based Analysis of a Before-and-After Cohort Study. J. Clin. Med. 2019, 8, 102. [CrossRef]
- Allan, G.M.; Arroll, B. Prevention and treatment of the common cold: Making sense of the evidence. CMAJ Can. Med. Assoc. J. 2014, 186, 190–199. [CrossRef]
- 136. Douglas, R.M.; Hemilä, H.; Chalker, E.; Treacy, B. Vitamin C for preventing and treating the common cold. *Cochrane Database Syst. Rev.* **2007**, *3*, CD000980. [CrossRef]
- 137. Gómez, E.; Quidel, S.; Bravo-Soto, G.; Ortigoza, Á. Does vitamin C prevent the common cold? *Medwave* **2018**, *18*, e7235. [CrossRef] [PubMed]
- 138. Heimer, K.A.; Hart, A.M.; Martin, L.G.; Rubio-Wallace, S. Examining the evidence for the use of vitamin C in the prophylaxis and treatment of the common cold. *J. Am. Acad. Nurse Pract.* **2009**, *21*, 295–300. [CrossRef] [PubMed]
- Kerksick, C.M.; Wilborn, C.D.; Roberts, M.D.; Smith-Ryan, A.; Kleiner, S.M.; Jäger, R.; Collins, R.; Cooke, M.; Davis, J.N.; Galvan, E.; et al. ISSN exercise & sports nutrition review update: Research & recommendations. *J. Int. Soc. Sports Nutr.* 2018, 15, 38. [CrossRef] [PubMed]
- Righi, N.C.; Schuch, F.B.; De Nardi, A.T.; Pippi, C.M.; de Almeida Righi, G.; Puntel, G.O.; da Silva, A.; Signori, L.U. Effects of vitamin C on oxidative stress, inflammation, muscle soreness, and strength following acute exercise: Meta-analyses of randomized clinical trials. *Eur. J. Nutr.* 2020, *59*, 2827–2839. [CrossRef] [PubMed]
- 141. Colunga Biancatelli, R.; Berrill, M.; Marik, P.E. The antiviral properties of vitamin C. *Expert Rev. Anti-Infect. Ther.* **2020**, *18*, 99–101. [CrossRef]
- Li, W.; Maeda, N.; Beck, M.A. Vitamin C deficiency increases the lung pathology of influenza virus-infected gulo-/- mice. J. Nutr. 2006, 136, 2611–2616. [CrossRef]
- 143. Kim, Y.; Kim, H.; Bae, S.; Choi, J.; Lim, S.Y.; Lee, N.; Kong, J.M.; Hwang, Y.I.; Kang, J.S.; Lee, W.J. Vitamin C Is an Essential Factor on the Anti-viral Immune Responses through the Production of Interferon-α/β at the Initial Stage of Influenza A Virus (H3N2) Infection. *Immune Netw.* **2013**, *13*, 70–74. [CrossRef]
- 144. Huang, K.J.; Su, I.J.; Theron, M.; Wu, Y.C.; Lai, S.K.; Liu, C.C.; Lei, H.Y. An interferon-gamma-related cytokine storm in SARS patients. *J. Med. Virol.* 2005, 75, 185–194. [CrossRef]
- 145. Teafatiller, T.; Agrawal, S.; De Robles, G.; Rahmatpanah, F.; Subramanian, V.S.; Agrawal, A. Vitamin C Enhances Antiviral Functions of Lung Epithelial Cells. *Biomolecules* **2021**, *11*, 1148. [CrossRef]
- 146. Valero, N.; Mosquera, J.; Alcocer, S.; Bonilla, E.; Salazar, J.; Álvarez-Mon, M. Melatonin, minocycline and ascorbic acid reduce oxidative stress and viral titers and increase survival rate in experimental Venezuelan equine encephalitis. *Brain Res.* 2015, 1622, 368–376. [CrossRef]
- 147. Cai, Y.; Li, Y.F.; Tang, L.P.; Tsoi, B.; Chen, M.; Chen, H.; Chen, X.M.; Tan, R.R.; Kurihara, H.; He, R.R. A new mechanism of vitamin C effects on A/FM/1/47(H1N1) virus-induced pneumonia in restraint-stressed mice. *BioMed Res. Int.* 2015, 2015, 675149. [CrossRef] [PubMed]
- 148. Lee, S.I.; Lim, C.M.; Koh, Y.; Huh, J.W.; Lee, J.S.; Hong, S.B. The effectiveness of vitamin C for patients with severe viral pneumonia in respiratory failure. *J. Thorac. Dis.* **2021**, *13*, 632–641. [CrossRef] [PubMed]
- 149. Kim, M.S.; Kim, D.J.; Na, C.H.; Shin, B.S. A Study of Intravenous Administration of Vitamin C in the Treatment of Acute Herpetic Pain and Postherpetic Neuralgia. *Ann. Dermatol.* **2016**, *28*, 677–683. [CrossRef] [PubMed]

- Warde-Farley, D.; Donaldson, S.L.; Comes, O.; Zuberi, K.; Badrawi, R.; Chao, P.; Franz, M.; Grouios, C.; Kazi, F.; Lopes, C.T.; et al. The GeneMANIA prediction server: Biological network integration for gene prioritization and predicting gene function. *Nucleic Acids Res.* 2010, *38*, 214–220. [CrossRef] [PubMed]
- 151. Chen, X.; Cheung, S.T.; So, S.; Fan, S.T.; Barry, C.; Higgins, J.; Lai, K.M.; Ji, J.; Dudoit, S.; Ng, I.O.; et al. Gene expression patterns in human liver cancers. *Mol. Biol. Cell* **2002**, *13*, 1929–1939. [CrossRef]
- 152. Jiang, J.; Mathijs, K.; Timmermans, L.; Claessen, S.M.; Hecka, A.; Weusten, J.; Peters, R.; van Delft, J.H.; Kleinjans, J.C.S.; Jennen, D.G.J.; et al. Omics-based identification of the combined effects of idiosyncratic drugs and inflammatory cytokines on the development of drug-induced liver injury. *Toxicol. Appl. Pharmacol.* 2017, 332, 100–108. [CrossRef]
- 153. Innocenti, F.; Cooper, G.M.; Stanaway, I.B.; Gamazon, E.R.; Smith, J.D.; Mirkov, S.; Ramirez, J.; Liu, W.; Lin, Y.S.; Moloney, C.; et al. Identification, replication, and functional fine-mapping of expression quantitative trait loci in primary human liver tissue. *PLoS Genet.* **2011**, *7*, e1002078. [CrossRef]
- 154. Ramaswamy, S.; Tamayo, P.; Rifkin, R.; Mukherjee, S.; Yeang, C.H.; Angelo, M.; Ladd, C.; Reich, M.; Latulippe, E.; Mesirov, J.P.; et al. Multiclass cancer diagnosis using tumor gene expression signatures. *Proc. Natl. Acad. Sci. USA* 2001, *98*, 15149–15154. [CrossRef]
- 155. Burington, B.; Barlogie, B.; Zhan, F.; Crowley, J.; Shaughnessy, J.D., Jr. Tumor cell gene expression changes following short-term in vivo exposure to single agent chemotherapeutics are related to survival in multiple myeloma. *Clin. Cancer Res.* **2008**, *14*, 4821–4829. [CrossRef]
- 156. Mallon, B.S.; Chenoweth, J.G.; Johnson, K.R.; Hamilton, R.S.; Tesar, P.J.; Yavatkar, A.S.; Tyson, L.J.; Park, K.; Chen, K.G.; Fann, Y.C.; et al. StemCellDB: The human pluripotent stem cell database at the National Institutes of Health. *Stem Cell Res.* 2013, 10, 57–66. [CrossRef]
- 157. Roth, R.B.; Hevezi, P.; Lee, J.; Willhite, D.; Lechner, S.M.; Foster, A.C.; Zlotnik, A. Gene expression analyses reveal molecular relationships among 20 regions of the human CNS. *Neurogenetics* **2006**, *7*, 67–80. [CrossRef] [PubMed]
- 158. Perou, C.M.; Jeffrey, S.S.; van de Rijn, M.; Rees, C.A.; Eisen, M.B.; Ross, D.T.; Pergamenschikov, A.; Williams, C.F.; Zhu, S.X.; Lee, J.C.; et al. Distinctive gene expression patterns in human mammary epithelial cells and breast cancers. *Proc. Natl. Acad. Sci. USA* 1999, 96, 9212–9217. [CrossRef] [PubMed]
- 159. Kamburov, A.; Pentchev, K.; Galicka, H.; Wierling, C.; Lehrach, H.; Herwig, R. ConsensusPathDB: Toward a more complete picture of cell biology. *Nucleic Acids Res.* **2011**, *39*, D712–D717. [CrossRef] [PubMed]
- Morales, L.; Oliveros, J.C.; Enjuanes, L.; Sola, I. Contribution of Host miRNA-223-3p to SARS-CoV-Induced Lung Inflammatory Pathology. *Mbio* 2022, 13, e0313521. [CrossRef] [PubMed]
- 161. Shaghaghi, M.A.; Kloss, O.; Eck, P. Genetic Variation in Human Vitamin C Transporter Genes in Common Complex Diseases. *Adv. Nutr.* **2016**, *7*, 287–298. [CrossRef]
- Eck, P.; Erichsen, H.C.; Taylor, J.G.; Yeager, M.; Hughes, A.L.; Levine, M.; Chanock, S. Comparison of the genomic structure and variation in the two human sodium-dependent vitamin C transporters, SLC23A1 and SLC23A2. *Hum. Genet.* 2004, 115, 285–294. [CrossRef]
- 163. Timpson, N.J.; Forouhi, N.G.; Brion, M.J.; Harbord, R.M.; Cook, D.G.; Johnson, P.; McConnachie, A.; Morris, R.W.; Rodriguez, S.; Luan, J.; et al. Genetic variation at the SLC23A1 locus is associated with circulating concentrations of L-ascorbic acid (vitamin C): Evidence from 5 independent studies with >15,000 participants. Am. J. Clin. Nutr. 2010, 92, 375–382. [CrossRef]
- 164. Zanon-Moreno, V.; Ciancotti-Olivares, L.; Asencio, J.; Sanz, P.; Ortega-Azorin, C.; Pinazo-Duran, M.D.; Corella, D. Association between a SLC23A2 gene variation, plasma vitamin C levels, and risk of glaucoma in a Mediterranean population. *Mol. Vis.* 2011, 17, 2997–3004.
- 165. Lee, D.H.; Won, G.W.; Lee, Y.H.; Shin, J.S.; Ku, E.J.; Oh, T.K.; Jeon, H.J. Polymorphism in the HaeIII single nucleotide polymorphism of the SLC2A1 gene and cardiovascular disease in the early type 2 diabetes mellitus. *Diabetes Vasc. Dis. Res.* 2021, 18, 14791641211041225. [CrossRef]
- 166. Amini, S.; Javanmardi, M.; Mokarizadeh, A.; Maroofi, F.; Jalali, C.; Azadi, N.A.; Mohammadi, H.; Abdi, M. Association of HaeIII single nucleotide polymorphisms in the SLC2A1 gene with risk of diabetic nephropathy; evidence from Kurdish patients with type 2 diabetes mellitus. QJM 2016, 109, 399–404. [CrossRef]
- 167. Siokas, V.; Fotiadou, A.; Dardiotis, E.; Kotoula, M.G.; Tachmitzi, S.V.; Chatzoulis, D.Z.; Zintzaras, E.; Stefanidis, I.; Tsironi, E.E. SLC2A1 Tag SNPs in Greek Patients with Diabetic Retinopathy and Nephropathy. *Ophthalmic Res.* 2019, 61, 26–35. [CrossRef] [PubMed]
- 168. Le, M.T.; Lobmeyer, M.T.; Campbell, M.; Cheng, J.; Wang, Z.; Turner, S.T.; Chapman, A.B.; Boerwinkle, E.; Gums, J.G.; Gong, Y.; et al. Impact of genetic polymorphisms of SLC2A2, SLC2A5, and KHK on metabolic phenotypes in hypertensive individuals. *PLoS ONE* 2013, 8, e52062. [CrossRef] [PubMed]
- Mustroph, J.; Hupf, J.; Hanses, F.; Evert, K.; Baier, M.J.; Evert, M.; Meindl, C.; Wagner, S.; Hubauer, U.; Pietrzyk, G.; et al. Decreased GLUT1/NHE1 RNA expression in whole blood predicts disease severity in patients with COVID-19. *ESC Heart Fail*. 2021, *8*, 309–316. [CrossRef] [PubMed]
- 170. Ivanov, V.; Goc, A.; Ivanova, S.; Niedzwiecki, A.; Rath, M. Inhibition of ACE2 Expression by Ascorbic Acid Alone and its Combinations with Other Natural Compounds. *Infect. Dis.* **2021**, *14*, 1178633721994605. [CrossRef]

- Heskett, C.W.; Teafatiller, T.; Hennessey, C.; Gareau, M.G.; Marchant, J.S.; Said, H.M.; Subramanian, V.S. Enteropathogenic Escherichia coli Infection Inhibits Intestinal Ascorbic Acid Uptake via Dysregulation of Its Transporter Expression. *Dig. Dis. Sci.* 2021, 66, 2250–2260. [CrossRef]
- 172. Barone, M.; D'Amico, F.; Brigidi, P.; Turroni, S. Gut microbiome-micronutrient interaction: The key to controlling the bioavailability of minerals and vitamins? *Biofactors* 2022, 48, 307–314. [CrossRef]
- 173. Steinert, R.E.; Lee, Y.K.; Sybesma, W. Vitamins for the Gut Microbiome. Trends Mol. Med. 2020, 26, 137–140. [CrossRef]
- 174. Subramanian, V.S.; Sabui, S.; Moradi, H.; Marchant, J.S.; Said, H.M. Inhibition of intestinal ascorbic acid uptake by lipopolysaccharide is mediated via transcriptional mechanisms. *Biochim. Biophys. Acta Biomembr.* **2018**, *1860*, 556–565. [CrossRef]
- 175. Otten, A.T.; Bourgonje, A.R.; Peters, V.; Alizadeh, B.Z.; Dijkstra, G.; Harmsen, H.J.M. Vitamin C Supplementation in Healthy Individuals Leads to Shifts of Bacterial Populations in the Gut—A Pilot Study. *Antioxidants* **2021**, *10*, 1278. [CrossRef]
- 176. Sun, Z.; Song, Z.G.; Liu, C.; Tan, S.; Lin, S.; Zhu, J.; Dai, F.; Gao, J.; She, J.; Mei, Z.; et al. Gut microbiome alterations and gut barrier dysfunction are associated with host immune homeostasis in COVID-19 patients. *BMC Med.* 2022, 20, 24. [CrossRef]
- 177. Feyaerts, A.F.; Luyten, W. Vitamin C as prophylaxis and adjunctive medical treatment for COVID-19? *Nutrition* **2020**, 79–80, 110948. [CrossRef] [PubMed]
- 178. Miranda-Massari, J.R.; Toro, A.P.; Loh, D.; Rodriguez, J.R.; Borges, R.M.; Marcial-Vega, V.; Olalde, J.; Berdiel, M.J.; Riordan, N.H.; Martinez, J.M.; et al. The Effects of Vitamin C on the Multiple Pathophysiological Stages of COVID-19. *Life* **2021**, *11*, 1341. [CrossRef] [PubMed]
- Patterson, T.; Isales, C.M.; Fulzele, S. Low level of Vitamin C and dysregulation of Vitamin C transporter might be involved in the severity of COVID-19 Infection. *Aging Dis.* 2021, 12, 14–26. [CrossRef] [PubMed]
- 180. Tomasa-Irriguible, T.M.; Bielsa-Berrocal, L. COVID-19: Up to 82% critically ill patients had low Vitamin C values. *Nutr. J.* 2021, 20, 66. [CrossRef] [PubMed]
- Chiscano-Camón, L.; Ruiz-Rodriguez, J.; Ruiz-Sanmartin, A.; Roca, O.; Ferrer, R.; Care, C. Vitamin C levels in patients with SARS-CoV-2-associated acute respiratory distress syndrome. *Crit. Care* 2020, 24, 522. [CrossRef]
- 182. Marik, P.E. Vitamin C for the treatment of sepsis: The scientific rationale. Pharmacol. Ther. 2018, 189, 63–70. [CrossRef]
- 183. Zhao, B.; Li, M.; Ling, Y.; Peng, Y.; Huang, J.; Qu, H.; Gao, Y.; Li, Y.; Hu, B.; Lu, S.; et al. Potential benefit of high-dose intravenous vitamin C for coronavirus disease 2019 pneumonia. *Chin. Med. J.* **2021**, *135*, 23–25. [CrossRef]
- 184. Zhao, B.; Liu, M.; Liu, P.; Peng, Y.; Huang, J.; Li, M.; Wang, Y.; Xu, L.; Sun, S.; Qi, X.; et al. High Dose Intravenous Vitamin C for Preventing the Disease Aggravation of Moderate COVID-19 Pneumonia. A Retrospective Propensity Matched Before-After Study. *Front. Pharmacol.* 2021, 12, 638556. [CrossRef]
- 185. Block, G.; Jensen, C.D.; Dalvi, T.B.; Norkus, E.P.; Hudes, M.; Crawford, P.B.; Holland, N.; Fung, E.B.; Schumacher, L.; Harmatz, P. Vitamin C treatment reduces elevated C-reactive protein. *Free Radic. Biol. Med.* **2009**, *46*, 70–77. [CrossRef]
- 186. Safabakhsh, M.; Emami, M.R.; Khosroshahi, M.Z.; Asbaghi, O.; Khodayari, S.; Khorshidi, M.; Alizadeh, S.; Viri, E.H. Vitamin C supplementation and C-reactive protein levels: Findings from a systematic review and meta-analysis of clinical trials. J. Complement. Integr. Med. 2020, 17, 20190151. [CrossRef]
- 187. Xia, G.; Qin, B.; Ma, C.; Zhu, Y.; Zheng, Q. High-dose vitamin C ameliorates cardiac injury in COVID-19 pandemic: A retrospective cohort study. *Aging* **2021**, *13*, 20906–20914. [CrossRef]
- 188. Xia, G.; Fan, D.; He, Y.; Zhu, Y.; Zheng, Q. High-dose intravenous vitamin C attenuates hyperinflammation in severe coronavirus disease 2019. *Nutrition* **2021**, *91*, 111405. [CrossRef]
- 189. Tehrani, S.; Yadegarynia, D.; Abrishami, A.; Moradi, H.; Gharaei, B.; Rauofi, M.; Maghsoudi Nejad, F.; Sali, S.; Khabiri, N.; Abolghasemi, S. An investigation into the Effects of Intravenous Vitamin C on Pulmonary CT Findings and Clinical Outcomes of Patients with COVID 19 Pneumonia A Randomized Clinical Trial. Urol. J. 2021, 18, 6863. [CrossRef]
- 190. Rawat, D.; Roy, A.; Maitra, S.; Gulati, A.; Khanna, P.; Baidya, D.K. Vitamin C and COVID-19 treatment: A systematic review and meta-analysis of randomized controlled trials. *Diabetes Metab. Syndr. Clin. Res. Rev.* 2021, 15, 102324. [CrossRef]
- 191. Kwak, S.G.; Choo, Y.J.; Chang, M.C. The effectiveness of high-dose intravenous vitamin C for patients with coronavirus disease 2019: A systematic review and meta-analysis. *Complement. Ther. Med.* **2022**, *64*, 102797. [CrossRef]
- 192. Ao, G.; Li, J.; Yuan, Y.; Wang, Y.; Nasr, B.; Bao, M.; Gao, M.; Qi, X. Intravenous vitamin C use and risk of severity and mortality in COVID-19: A systematic review and meta-analysis. *Nutr. Clin. Pract.* **2022**, *37*, 274–281. [CrossRef]
- 193. Beran, A.; Mhanna, M.; Srour, O.; Ayesh, H.; Stewart, J.M.; Hjouj, M.; Khokher, W.; Mhanna, A.S.; Ghazaleh, D.; Khader, Y.; et al. Clinical significance of micronutrient supplements in patients with coronavirus disease 2019: A comprehensive systematic review and meta-analysis. *Clin. Nutr. ESPEN* **2022**, *48*, 167–177. [CrossRef]
- 194. Gavrielatou, E.; Xourgia, E.; Xixi, N.A.; Mantelou, A.G.; Ischaki, E.; Kanavou, A.; Zervakis, D.; Routsi, C.; Kotanidou, A.; Siempos, I.I. Effect of Vitamin C on Clinical Outcomes of Critically Ill Patients With COVID-19: An Observational Study and Subsequent Meta-Analysis. Front. Med. 2022, 11, 814587. [CrossRef]
- 195. Zheng, S.; Chen, Q.; Jiang, H.; Guo, C.; Luo, J.; Li, S.; Wang, H.; Li, H.; Zheng, X.; Weng, Z. No significant benefit of moderate-dose vitamin C on severe COVID-19 cases. *Open Med.* **2021**, *16*, 1403–1414. [CrossRef]
- 196. Thomas, S.; Patel, D.; Bittel, B.; Wolski, K.; Wang, Q.; Kumar, A.; Il'Giovine, Z.; Mehra, R.; McWilliams, C.; Nissen, S.E.; et al. Effect of High-Dose Zinc and Ascorbic Acid Supplementation vs Usual Care on Symptom Length and Reduction among Ambulatory Patients with SARS-CoV-2 Infection: The COVID A to Z Randomized Clinical Trial. JAMA Netw. Open 2021, 4, e210369. [CrossRef]

- 197. Zhang, J.; Rao, X.; Li, Y.; Zhu, Y.; Liu, F.; Guo, G.; Luo, G.; Meng, Z.; De Backer, D.; Xiang, H.; et al. Pilot trial of high-dose vitamin C in critically ill COVID-19 patients. *Ann. Intensive Care* **2021**, *11*, 5. [CrossRef] [PubMed]
- 198. Darban, M.; Malek, F.; Memarian, M.; Gohari, A.; Kiani, A.; Emadi, A.; Lavvaf, S.; Bagheri, B. Efficacy of High Dose Vitamin C, Melatonin and Zinc in Iranian Patients with Acute Respiratory Syndrome due to Coronavirus Infection: A Pilot Randomized Trial. J. Cell. Mol. Anesth. 2020, 6, 164–167. [CrossRef]
- 199. Hakamifard, A.; Soltani, R.; Maghsoudi, A.; Rismanbaf, A.; Aalinezhad, M.; Tarrahi, M.; Mashayekhbakhsh, S.; Dolatshahi, K. The effect of vitamin E and vitamin C in patients with COVID-19 pneumonia; a randomized controlled clinical trial. *Immunopathol. Persa* **2021**, *8*, e08.
- Beigmohammadi, M.T.; Bitarafan, S.; Hoseindokht, A.; Abdollahi, A.; Amoozadeh, L.; Soltani, D. The effect of supplementation with vitamins A, B, C, D, and E on disease severity and inflammatory responses in patients with COVID-19: A randomized clinical trial. *Trials* 2021, 22, 802. [CrossRef]
- 201. Majidi, N.; Rabbani, F.; Gholami, S.; Gholamalizadeh, M.; BourBour, F.; Rastgoo, S.; Hajipour, A.; Shadnoosh, M.; Akbari, M.E.; Bahar, B.; et al. The Effect of Vitamin C on Pathological Parameters and Survival Duration of Critically Ill Coronavirus Disease 2019 Patients: A Randomized Clinical Trial. *Front. Immunol.* **2021**, *12*, 717816. [CrossRef]
- 202. Hemilä, H.; Carr, A.; Chalker, E. Vitamin C May Increase the Recovery Rate of Outpatient Cases of SARS-CoV-2 Infection by 70%: Reanalysis of the COVID A to Z Randomized Clinical Trial. *Front. Immunol.* **2021**, *12*, 674681. [CrossRef]
- 203. Al Sulaiman, K.; Aljuhani, O.; Saleh, K.B.; Badreldin, H.A.; Al Harthi, A.; Alenazi, M.; Alharbi, A.; Algarni, R.; Al Harbi, S.; Alhammad, A.M.; et al. Ascorbic acid as an adjunctive therapy in critically ill patients with COVID-19: A propensity score matched study. *Sci. Rep.* 2021, *11*, 17648. [CrossRef]
- 204. Abulmeaty, M.M.A.; Aljuraiban, G.S.; Shaikh, S.M.; ALEid, N.E.; Mazrou, L.R.A.; Turjoman, A.A.; Aldosari, M.S.; Razak, S.; El-Sayed, M.M.; Areabi, T.M.; et al. The Efficacy of Antioxidant Oral Supplements on the Progression of COVID-19 in Non-Critically Ill Patients: A Randomized Controlled Trial. *Antioxidants* 2021, 10, 804. [CrossRef]
- Zhu, N.; Huang, B.; Jiang, W. Targets of Vitamin C withTherapeutic Potential for Cardiovascular Disease and Underlying Mechanisms: A Study of Network Pharmacology. *Front. Pharmacol.* 2021, 11, 591337. [CrossRef]
- 206. Chadli, A.; Haraj, N.E.; El Aziz, S.; Laidi, S.; Mounir, A.; Bensbaa, S.; Mjabber, A.; Barrou, L.; Hamidi, C.E.K.E.; Nsiri, A.; et al. COVID-19: Patient care after discharge from the Intensive Care Unit. *Int. J. Clin. Pract.* 2021, 75, e14270. [CrossRef]
- 207. May, C.N.; Bellomo, R.; Lankadeva, Y.R. Therapeutic potential of megadose vitamin C to reverse organ dysfunction in sepsis and COVID-19. *Br. J. Pharmacol.* **2021**, *178*, 3864–3868. [CrossRef] [PubMed]
- Marik, P.E.; Khangoora, V.; Rivera, R.; Hooper, M.H.; Catravas, J. Hydrocortisone, Vitamin C, and Thiamine for the Treatment of Severe Sepsis and Septic Shock: A Retrospective Before-After Study. *Chest* 2017, 151, 1229–1238. [CrossRef] [PubMed]
- 209. Alamdari, D.H.; Moghaddam, A.B.; Amini, S.; Keramati, M.R.; Zarmehri, A.M.; Alamdari, A.H.; Damsaz, M.; Banpour, H.; Yarahmadi, A.; Koliakos, G. Application of methylene blue -vitamin C -N-acetyl cysteine for treatment of critically ill COVID-19 patients, report of a phase-I clinical trial. *Eur. J. Pharmacol.* 2020, 15, 173494. [CrossRef] [PubMed]
- Marik, P.E.; Kory, P.; Varon, J.; Iglesias, J.; Meduri, G.U. MATH+ protocol for the treatment of SARS-CoV-2 infection: The scientific rationale. *Expert Rev. Anti-Infect. Ther.* 2021, 19, 129–135. [CrossRef]
- Li, R.; Wu, K.; Li, Y.; Liang, X.; Lai, K.P.; Chen, J. Integrative pharmacological mechanism of vitamin C combined with glycyrrhizic acid against COVID-19: Findings of bioinformatics analyses. *Brief Bioinform.* 2021, 22, 1161–1174. [CrossRef]
- Chen, L.; Hu, C.; Hood, M.; Zhang, X.; Zhang, L.; Kan, J. A Novel Combination of Vitamin C, Curcumin and Glycyrrhizic Acid Potentially Regulates Immune and Inflammatory Response Associated with Coronavirus Infections: A Perspective from System Biology Analysis. *Nutrients* 2020, *12*, 1193. [CrossRef]