

# LRP1 Functions as an Atheroprotective Integrator of TGF $\beta$ and PDGF Signals in the Vascular Wall: Implications for Marfan Syndrome

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**Background.** The multifunctional receptor LRP1 controls expression, activity and trafficking of the PDGF receptor- $\beta$  in vascular smooth muscle cells (VSMC). LRP1 is also a receptor for TGF $\beta$ 1 and is required for TGF $\beta$  mediated inhibition of cell proliferation. **Methods and Principal Findings.** We show that loss of LRP1 in VSMC (smLRP<sup>-</sup>) in vivo results in a Marfan-like syndrome with nuclear accumulation of phosphorylated Smad2/3, disruption of elastic layers, tortuous aorta, and increased expression of the TGF $\beta$  target genes thrombospondin-1 (TSP1) and PDGFR $\beta$  in the vascular wall. Treatment of smLRP1<sup>-</sup> animals with the PPAR $\gamma$  agonist rosiglitazone abolished nuclear pSmad accumulation, reversed the Marfan-like phenotype, and markedly reduced smooth muscle proliferation, fibrosis and atherosclerosis independent of plasma cholesterol levels. **Conclusions and Significance.** Our findings are consistent with an activation of TGF $\beta$  signals in the LRP1-deficient vascular wall. LRP1 may function as an integrator of proliferative and anti-proliferative signals that control physiological mechanisms common to the pathogenesis of Marfan syndrome and atherosclerosis, and this is essential for maintaining vascular wall integrity.

Citation: Boucher P, Li W-P, Matz RL, Takayama Y, Auwerx J, et al (2007) LRP1 Functions as an Atheroprotective Integrator of TGF $\beta$  and PDGF Signals in the Vascular Wall: Implications for Marfan Syndrome. PLoS ONE 2(5): e448. doi:10.1371/journal.pone.0000448

## INTRODUCTION

Cholesterol-induced atherosclerosis is a major cause of morbidity and mortality in Western societies. In the liver, the low-density lipoprotein (LDL) receptor functions in concert with another LDL receptor family member, the LDL receptor-related protein 1 (LRP1), in removing cholesterol carrying lipoprotein particles from the bloodstream. However, in the smooth muscle cells of the artery wall LRP1 acts through a different, cholesterol-independent mechanism in the maintenance of vascular wall integrity and atherosclerosis prevention, which relies in part on the ability of LRP1 to regulate the activity and subcellular trafficking of the PDGFR $\beta$  [1–3]. SmLRP<sup>-</sup> mice that lack LRP1 in vascular smooth muscle cells show thickening of the muscular layer and greatly increased susceptibility to atherosclerotic lesion development, even at low plasma cholesterol levels. On a high cholesterol diet, lesion development and progression is dramatically accelerated, resulting in the occlusion of major arteries within 2 months of cholesterol feeding. Pharmacologic inhibition of PDGFR $\beta$  signaling reduced both lesion progression and smooth muscle over-proliferation, suggesting that the abnormal activation of PDGFR $\beta$  is at least in part responsible for the accelerated lesion development and progression [1].

On the other hand, the characteristic disruption of elastic layers, elongation of the aortas and aneurysms are hallmarks of Marfan syndrome (MFS) [4,5], an autosomal dominant genetic disease. MFS is caused by loss of function mutations in fibrillin-1, a microfibrillar protein that coats the surface of elastic fibers where it binds and immobilizes the TGF $\beta$  large latency complex (LLC) [6]. Haploinsufficiency of fibrillin-1 [7,8] markedly changes the extracellular targeting, sequestration and turnover of latent TGF $\beta$ , resulting in pronounced activation of the growth factor and the signaling pathway [4,5,9]. A phenotypically similar Marfan-like syndrome is caused by loss of function mutations in TGF $\beta$  receptors 1 or 2 [10–12], which nevertheless result in a paradoxical

activation of the pathway and nuclear accumulation of pSmad2 [10]. Thus, abnormal activation of TGF $\beta$  signaling appears to be a common feature of Marfan and Marfan-like disorders.

Recently, LRP1 was reported to be identical to TGF $\beta$ R(V), which is coexpressed together with TGF $\beta$  receptors I, II and III. LRP1/TGF $\beta$ R(V) is required for mediating the growth inhibitory response of TGF $\beta$ , in conjunction with Smad2/3 signaling through TGF $\beta$ R(I) and (II) [13,14]. TGF $\beta$ 1 can induce the expression of connective tissue growth factor (CTGF), PDGF-B and PDGF receptors [15–19]. Activation of a PDGF/PDGFR autocrine loop by TGF $\beta$  mediates epithelial-mesenchymal transition and constitutes a crucial mechanism that enhances tumor metastasis [20] and the malignancy of gliomas [21]. All of these proteins are not only ligands for LRP1, they also promote smooth muscle cell proliferation [16,22]. Furthermore, TSP1, another ligand for LRP1 and a TGF $\beta$  target gene [23], directly mediates

**Academic Editor:** Mariko Hatakeyama, RIKEN Genomic Sciences Center, Japan

**Received** January 26, 2007; **Accepted** April 23, 2007; **Published** May 16, 2007

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**Funding:** This work was supported by grants from, Fondation pour la Recherche Médicale, Fondation de France, Conseil Scientifique de l'Université Louis Pasteur de Strasbourg, the National Institutes of Health (HL20948, HL63762, NS43408), and the Perot Family Foundation. J.H. is an Established Investigator of the American Heart Association and Parke-Davis and is the recipient of a Wolfgang-Paul Award from the Humboldt Foundation. None of the sponsors has placed any restrictions on the work or its publication.

**Competing Interests:** The authors have declared that no competing interests exist.

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the conversion of latent TGF $\beta$  to the active form [24,25] and TSP1 binding to an LRP1/calreticulin complex regulates cell adhesion [26]. Thus, LRP1 is tightly involved in the control of growth regulating signaling processes that involve TGF $\beta$ , PDGF, and their receptors. This prompted us to investigate, whether abnormal activation of TGF $\beta$  signaling could be responsible for the striking disruption of the elastic fiber network, fibrosis, and accelerated atherosclerotic lesion development in the vascular wall of smLRP $^{-}$  mice [1].

## RESULTS

### Activation of TGF $\beta$ signaling in LRP1-deficient VSMC

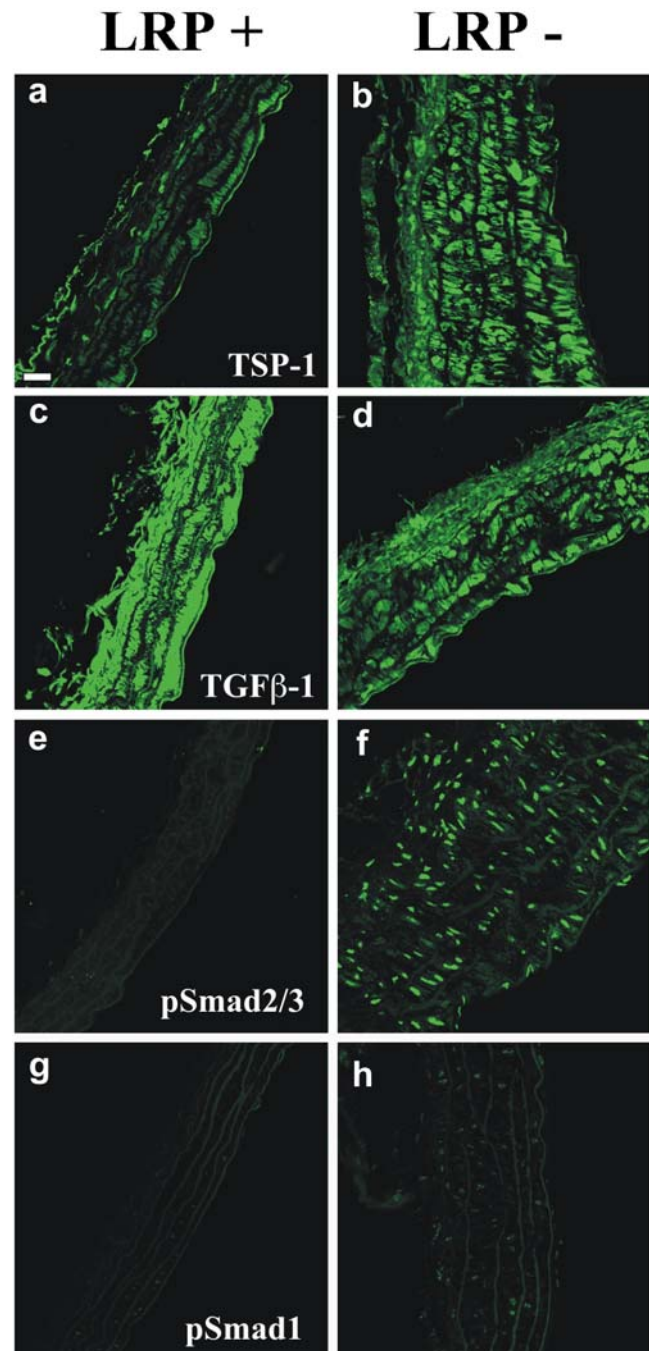
Vascular wall fibrosis, disruption of elastic layers, and tortuosity of the aorta were suggestive of abnormal TGF $\beta$  signaling in smLRP-deficient mice. To obtain further evidence for the presence of increased TGF $\beta$  activity, we used immunohistochemistry to visualize the levels of TGF $\beta$ 1 and TSP1, as well as the phosphorylated forms of Smad-1 and Smad-2, the intracellular mediators of TGF $\beta$  family signals, in aortas from cholesterol-fed, LDL receptor deficient mice expressing (LRP $^{+}$ ) or not expressing LRP1 (LRP $^{-}$ ) in their VSMC (Figure 1). Expression of TSP-1, a direct TGF $\beta$ 1 target gene, was strikingly increased in LRP $^{-}$  compared to LRP $^{+}$  aortas (Panels a and b), whereas total TGF $\beta$ 1 immunoreactivity was slightly reduced (Panels c and d), consistent with an increased activation and turnover of matrix-associated TGF $\beta$ .

TGF $\beta$ 1 and other TGF $\beta$  family members, as well as angiotensin II are prominent inducers of Smad2-dependent target gene transcription, resulting in collagen deposition and disruption of the elastic layers in the vascular wall [27,28]. Prominent accumulation of nuclear phosphorylated Smad2/3 (pSmad2/3) was found in VSMC in the absence of LRP1 (Panels e and f). This effect was specific for Smad2, which is the target of TGF $\beta$  signals, but not for Smad1, which responds to signals elicited by bone morphogenic proteins (BMPs). pSmad1 was not significantly elevated in the absence of LRP1 (Panels g and h). These findings, taken together with the molecular properties of LRP1, make it likely that TGF $\beta$  signaling in VSMC is indeed selectively increased in the absence of LRP1.

### Rosiglitazone reduces TGF $\beta$ signals in vitro and in vivo

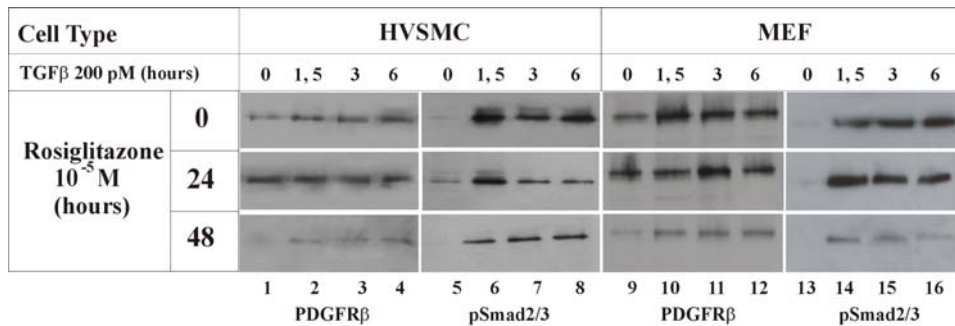
PPAR $\gamma$  agonists of the thiazolidinedione class have been reported to inhibit TGF $\beta$  signaling, CTGF expression and fibrosis [29,30], and they also can reduce atherosclerosis [31]. We therefore decided to test, whether activation of PPAR $\gamma$  by rosiglitazone, a compound of this class that is in clinical use, might result in reduced nuclear accumulation of pSmad2 and suppression of atherosclerosis in mice lacking LRP1. We first examined the effect of rosiglitazone on PDGFR $\beta$  and pSmad2/3 levels in cultured cells in vitro. While TGF $\beta$ 1 only moderately elevated PDGFR $\beta$  expression in vitro, it markedly increased pSmad2/3 levels in cultured human VSMC and in murine fibroblasts. This increase was significantly blunted when the cells were pretreated with rosiglitazone (Figure 2).

To test, whether rosiglitazone was able to correct the abnormal accumulation of pSmad2/3 levels and activation of PDGFR $\beta$  signaling in smLRP deficient animals in vivo, we fed smLRP $^{+}$  and smLRP $^{-}$  animals a cholesterol-rich diet that either contained or did not contain rosiglitazone for 5 weeks (Figure 3). Aortic extracts were prepared as described earlier [1] and analyzed by immunoblotting (Panel A). Rosiglitazone treatment (lane 3) significantly reduced the elevated pSmad2/3, PDGFR $\beta$  and pErk1/2 in smLRP $^{-}$  aortas (lane 2) compared to smLRP $^{+}$  controls (lane 1). We verified this result by immunofluorescence staining of aortas (Panel B). Rosiglitazone treatment (Panels c, f, i)



**Figure 1. Increased pSmad2/3 expression and activation of TGF $\beta$  signaling in LRP $^{-}$  mouse aorta.** Longitudinal sections of abdominal aorta from SM22Cre $^{+}$ ;LRP $^{flox/flox}$ ;LDLR $^{-/-}$  (LRP $^{-}$ ) and LRP $^{flox/flox}$ ;LDLR $^{-/-}$  (LRP $^{+}$ ) mice were stained with anti-TSP1, anti-TGF $\beta$ 1, anti-pSmad2/3 and anti-pSmad1 antibodies. Reduced LRP1 expression results in greatly enhanced expression of pSmad2/3 and its target gene, TSP1. By contrast, TGF $\beta$ 1 levels were slightly reduced, pSmad1 levels did not change. Bar in a indicates 20  $\mu$ m.  
doi:10.1371/journal.pone.0000448.g001

abolished the increased nuclear pSmad2/3, overexpression of PDGFR $\beta$ , and the increased pErk1/2 levels in the smLRP $^{-}$  animals (Panels b, e, h). As had been shown previously using the tyrosine kinase inhibitor Gleevec [1], rosiglitazone treatment also considerably improved the integrity of the elastic layers (Panel C).



**Figure 2. Rosiglitazone treatment decreases expression of PDGFRβ and activation of Smad2/3 (pSmad2/3) *in vitro*.** Western blot analysis of activated Smad2/3 and PDGFRβ in whole cell lysates from human VSMC and MEF LRP<sup>+/+</sup> cells pretreated 0, 24 or 48 hours with rosiglitazone at 10<sup>-5</sup>M and then stimulated with human TGFβ1 (200 pM) for 0, 1.5, 3 or 6 hours.  
doi:10.1371/journal.pone.0000448.g002

### Rosiglitazone prevents atherosclerosis in smLRP<sup>-</sup> mice

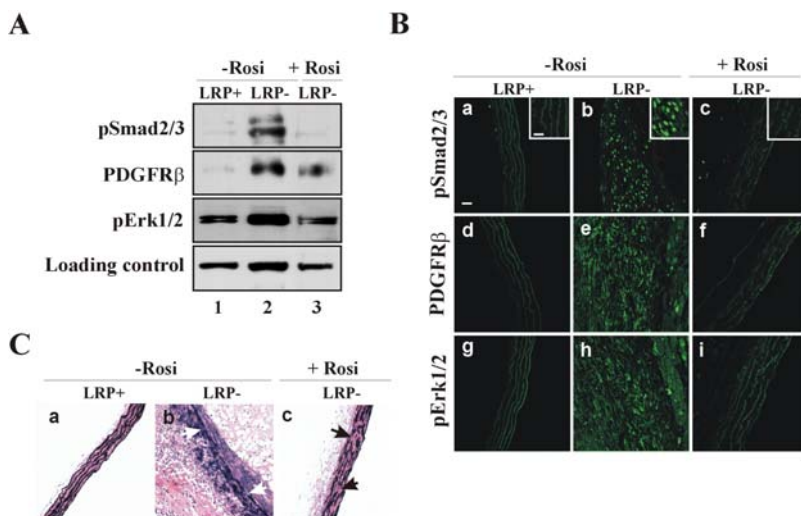
Thiazolidinediones have been reported to protect male LDLR-deficient mice from developing atherosclerotic lesions [31], coinciding with reductions in gelatinase and TNFα expression. To test whether blockade of the increased Smad activation by rosiglitazone would also reduce the marked atherosclerosis in the smLRP<sup>-</sup> mice, we quantitatively examined the aortas from the atherogenic diet-fed mice. Atherosclerotic lesion areas were significantly reduced in the presence of the PPARγ agonist (Figure 4A and C), and the striking thickening and fibrosis of the LRP deficient vessel wall was almost completely abolished (Figure 4B). Importantly, these changes were not caused by or secondary to any effect of rosiglitazone on the plasma lipoprotein profile, triglyceride or cholesterol levels (Figure 4D–F).

### DISCUSSION

The findings we have presented here reveal a novel mechanism by which the multifunctional receptor LRP1 functions as an

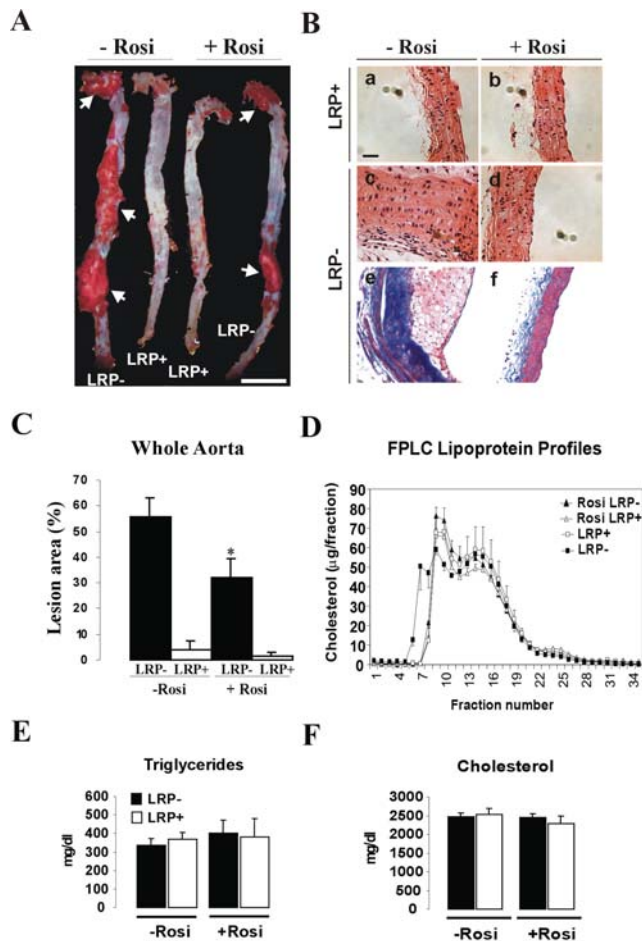
integrator of two distinct cellular signaling pathways in the vascular wall. They also suggest a functional relationship between the molecular events that underlie MFS, the formation of aneurysms and atherosclerosis. Our results show that LRP1 controls Smad2/3-as well as PDGF-dependent signaling in VSMC in a coordinated fashion and by distinct mechanisms.

LRP1 is a target of PDGFRβ-dependent Src kinase activation [2,32] and we showed earlier that loss of LRP1 expression in VSMCs results in increased PDGFRβ expression and activation [1,3]. In the present study we have demonstrated, that loss of LRP1 expression also results in increased Smad2/3-dependent signaling in VSMCs. A potent inducer of Smad2 phosphorylation and nuclear translocation is TGFβ [33]. Moreover, induction of PDGF as well as PDGFRβ expression in response to TGFβ stimulation has been recognized as an important component of epithelial-mesenchymal transition, a process that underlies metastasis of some tumors [20] and increases the aggressiveness of others [21]. Our results show that increased pSmad2/3 signaling in the LRP1-deficient vascular wall results in increased PDGFRβ expression and activation, making it likely that TGFβ is directly involved.



**Figure 3. Activation of TGFβ and PDGF signaling in LRP<sup>-</sup> mouse aortas are both prevented upon rosiglitazone treatment.** Mice had been cholesterol-fed for 5 weeks in the absence (-Rosi) or presence (+Rosi) of rosiglitazone (GlaxoSmithKline, 25 mg/kg/day) before analysis. Mouse aortas expressing (LRP<sup>+</sup>) or not expressing (LRP<sup>-</sup>) LRP in VSMC were analyzed by western blot (Panel A) and immunohistochemistry (Panel B) for expression of PDGFRβ (d–f), and for activation of Smad2/3 (pSmad2/3, a–c), and Erk1/2 (pErk1/2, g–i). Panel C shows elastic staining of corresponding sections and gaps in elastic fiber continuity (arrows). Bar indicates 40 μm, insert scale bar in B, a indicates 10 μm.  
doi:10.1371/journal.pone.0000448.g003





**Figure 4. Quantitative analysis of atherosclerotic lesion size in aortas from cholesterol-fed mice with or without rosiglitazone treatment.** (A) Aortas from 20-week-old mice that express (LRP<sup>+</sup>) or lack (LRP<sup>-</sup>) LRP in VSMC (n=6 mice per group). Mice had been cholesterol-fed for 5 weeks in the absence (-Rosig) or presence (+Rosig) of rosiglitazone (GlaxoSmithKline, 25 mg/kg/day) before analysis. Aortae were stained en face with Sudan IV and arrows indicate lipid-laden (Sudan-positive) atherosclerotic lesions. Scale bar, 1.2 cm. (B) Histological analysis of thoracic aortas from animals cholesterol-fed in the absence or presence of rosiglitazone. Hematoxylin and eosin (a and b, LRP<sup>+</sup>; c and d, LRP<sup>-</sup>), and trichrome staining (e and f, LRP<sup>-</sup>) of longitudinal sections. Scale bar in a, 15  $\mu$ m. (C) Quantitative analysis of atherosclerotic lesion size in aortas from cholesterol-fed LRP<sup>-</sup> and control (LRP<sup>+</sup>) mice (n=5 mice per group) with and without rosiglitazone treatment. Values are expressed as mean $\pm$ s.e.m. \*, p<0.05 for LRP<sup>-</sup> treated versus untreated. (D) FPLC profile of plasma lipoproteins from untreated LRP<sup>-</sup> (filled squares) and LRP<sup>+</sup> (open squares) and rosiglitazone treated LRP<sup>-</sup> (filled triangles) and LRP<sup>+</sup> (open triangles). (E) Plasma triglycerides and (F) cholesterol from untreated and rosiglitazone treated LRP<sup>-</sup> and LRP<sup>+</sup> mice. Values are expressed as mean $\pm$ S.E.M. (n=10 mice per group). doi:10.1371/journal.pone.0000448.g004

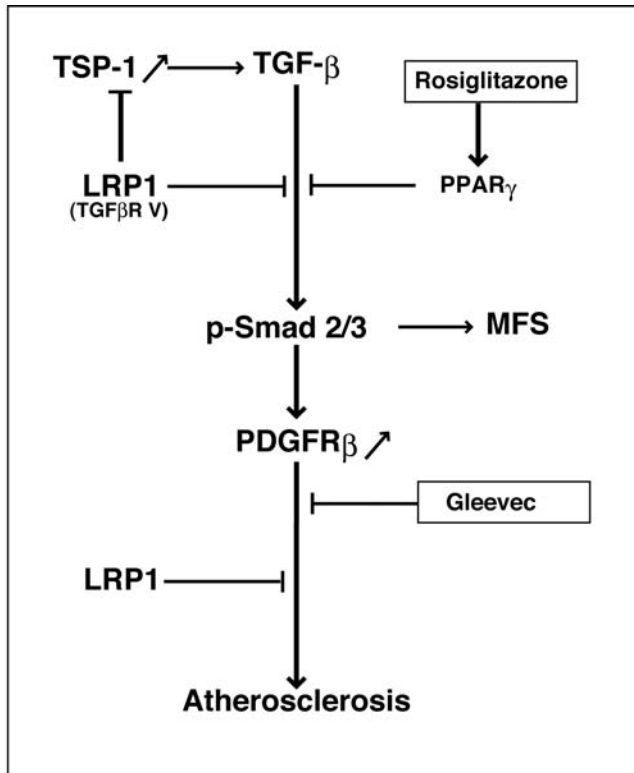
Furthermore, LRP1/TGF $\beta$ R(V), can directly bind TGF $\beta$ 1 [13], as well as enter into a complex with TGF $\beta$ R(I) [34]. By binding and endocytosis of the active growth factor and also by sequestering TGF $\beta$ R(I) away from TGF $\beta$ R(II), LRP1 is in a central position where it can suppress the normal mode of TGF $\beta$  signaling and Smad2/3 activation through the TGF $\beta$ R(I)/R(II) complex. In addition, LRP1 mediates the endocytosis of TSP1 as well as several matrix metalloproteinases (MMP) from the extracellular space [35–37], and TSP1 as well as active MMP2 and MMP9 levels are increased in smLRP deficient aortas (Figure 1, and [1]).

TSP1 and MMPs potently and independently activate TGF $\beta$ 1 [24,25,38]. Loss of smLRP would thus be predicted to activate TGF $\beta$ /Smad signaling in VSMC by multiple independent mechanisms, and this is consistent with the dramatically increased nuclear pSmad2/3 levels in smLRP<sup>-</sup> aortas (Figure 1). The resulting disruption of elastic layers, vascular fibrosis, elongation and distension of the aorta, and high incidence of aortic aneurysms directly support the pivotal role of TGF $\beta$  signaling in the pathogenesis of MFS and reveal LRP1 as a critical regulator of this pathway in vivo. As such, LRP1 could conceivably contribute as a genetic modifier to the variable severity of expression of MFS in humans.

We have further demonstrated the importance of TGF $\beta$  signaling through pharmacological intervention and reversal of the smLRP<sup>-</sup> phenotype with the PPAR $\gamma$  agonist rosiglitazone. Activation of PPAR $\gamma$  with thiazolidinediones potently inhibits TGF $\beta$  signaling in vitro and in vivo [29,39,40]. The striking effect of rosiglitazone on the nuclear accumulation of pSmad2/3 and the near complete reversal of the aortic wall thickening and fibrosis, and prevention of atherosclerotic lesion progression confirm the central importance of TGF $\beta$  pathway activation for this phenotype. PPAR $\gamma$  agonists might thus complement and enhance the potential therapeutic use of the angiotensin II type 1 receptor blocker Losartan in the treatment of MFS [27].

Increased susceptibility to atherosclerosis is not a part of the MFS complex, but a striking feature of smLRP<sup>-</sup> mice [1]. Likewise, PPAR $\gamma$  agonists are only partially effective against diet-induced atherosclerosis in LDLR deficient mice when LRP1 is functional [31]. By contrast, we have shown that rosiglitazone (this study) as well as the PDGFR tyrosine kinase inhibitor Gleevec [1] are highly potent in protecting smLRP<sup>-</sup> mice from atherosclerosis. Our findings thus suggest that abnormal activation of PDGFR $\beta$  signaling, which has so far not been reported in MFS, is the major mediator of the increased atherosclerosis in the smLRP<sup>-</sup> mice, and that increased PDGFR $\beta$  activity is brought about by two independent and synergistic mechanisms. First, activation of Smad/TGF $\beta$  signals in the absence of LRP1 results in the disruption of elastic layers and a tortuous aorta [1], as well as increased expression of PDGF receptors (Figure 2 and 3) [15,20,41]. However, since LRP1 also controls PDGFR $\beta$  activity and recycling at the level of the plasma membrane [3], loss of LRP1 in smooth muscle cells also results in a greatly increased mitogenic response, VSMC proliferation and the promotion of atherosclerotic lesion formation. This second event is prevented in MFS by the presence of LRP1, which suppresses the abnormal PDGFR $\beta$  activation through a posttranslational mechanism [3]. Thus, rosiglitazone (as a Smad/TGF $\beta$  antagonist) and Gleevec (as a PDGFR $\beta$  inhibitor) act at different steps in a linear sequence of events (Figure 5) that result in a Marfan-like syndrome and culminate in extensive atherosclerosis in smLRP<sup>-</sup> artery walls.

Nevertheless, blockade of PDGFR $\beta$  signaling alone by administration of Gleevec was sufficient to reduce atherosclerotic lesion progression, as well as vascular wall thickness, in smLRP<sup>-</sup> mice [1], suggesting that signaling through this pathway is necessary for elastic layer disruption and wall thickening. This raises the possibility that PDGFR $\beta$  signaling may be activated to some extent in the Marfan vessel wall, or alternatively the mere suppression of its baseline activity may be sufficient to normalize the vessel wall thickening that is initiated by increased TGF $\beta$  activity. Consistent with an important role of PDGFR $\beta$  in vascular thickening is a recent report that showed transphosphorylation of a truncated PDGFR $\beta$  induced by the Angiotensin II type 1 receptor (AT1) [42], which can also activate Smad2-dependent signals independent of TGF $\beta$  [28]. AT1 is inhibited by Losartan,



**Figure 5. Control of TGF $\beta$  and PDGF signaling and protection of vascular wall integrity by LRP1.** Absence of LRP1 results in increased activation of TGF $\beta$  signaling. This is accompanied by disruption of elastic layers, tortuous aortas and increased fibrosis similar to what is observed in Marfan and Marfan-like syndromes in which the genes for fibrillin-1 or TGF $\beta$  receptors are defective. Loss of LRP1 expression leads to increased expression of PDGF receptors. LRP1 also controls PDGFR $\beta$  signaling and trafficking through an independent mechanism, and absence of LRP1 promotes VSMC proliferation and severe atherosclerosis. Rosiglitazone blocks TGF $\beta$  signaling upstream of PDGFR, which is inhibited by Gleevec. Both drugs are thus effective in reducing arterial wall thickening and atherosclerosis, which are induced by increased PDGFR signaling.

doi:10.1371/journal.pone.0000448.g005

a drug that also reduced wall thickening in a mouse model of Marfan syndrome [27]. Taken together, these findings indicate a critical role for Smad2- and AT1-dependent signaling mechanisms in Marfan and Marfan-like syndromes and identify PDGFR $\beta$  as another important component of the pathogenic mechanism. Our findings also suggest that rosiglitazone, a clinically approved antidiabetic drug, may be useful for the treatment of vasculopathies that are caused by increased TGF $\beta$  signaling, e.g. Marfan syndrome, as well as for the supportive treatment of certain malignancies, such as malignant glioblastomas [43].

## MATERIALS AND METHODS

### Transgenic mice and diets

Homozygous LDL receptor (LDLR $^{-/-}$ ) animals LRP deficient in vascular smooth muscle cells (SM22-Cre $^{+}$ ;LRP $^{lox/lox}$ ;LDLR $^{-/-}$ , referred to as smLRP $^{-}$  or LRP $^{-}$ ) have been described previously [1] and were maintained on Teklad 6% (w/w) Mouse/Rat Diet 7002 from Harlan Teklad Premier Laboratory Diets (Madison, WI). Six weeks prior to sacrifice, 3–6 months old LRP $^{-}$  and littermate controls (LRP $^{lox/lox}$ ;LDLR $^{-/-}$ , noted LRP $^{+}$ ) were placed on a high-fat/high-cholesterol (Paigen) diet containing

(21% (w/w) milk fat, 1.25% (w/w) cholesterol, 0.5% (w/w) cholic acid) and where indicated, the PPAR $\gamma$  agonist rosiglitazone (Advantia $^{\circ}$ , GlaxoSmithKline, 25mg/kg/day) was mixed into the diet. All animals used in the experiments were age and sex matched as closely as possible. All animal experiments were conducted according to procedures approved by the institutional Animal Care and Use Committee (IACUC) at UT Southwestern.

### Whole Aorta staining

After 4% paraformaldehyde cardiac perfusion, the whole aortas of animals from the different treated groups were carefully dissected from the heart to the iliac bifurcation. The vessels were rinsed in PBS and dissected to remove connective tissue and attached fat. The vessels were opened longitudinally and stained for lipids with Sudan IV. Atherosclerosis in the aortas was measured by quantitation of Sudan IV-positive lesions as described [44]. Whole mount preparations of the vessels were photographed and digitized. Sudan IV-positive lesions were delineated and analyzed using NIH Image software (NIH, Bethesda, Maryland, USA).

### Histology

Aortic tissue was fixed over night with 4% paraformaldehyde and embedded in paraffin. Longitudinal and transverse sections (10  $\mu$ m thick) were stained with hematoxylin-eosin under standard conditions.

### Immunofluorescence microscopy

Anesthetized wt and LRP KO mice were perfused through the left cardiac ventricle with warmed Hank's balanced salt solution (20 mM HEPES, pH 7.3) followed by the same solution containing 4% (w/v) paraformaldehyde. Aortas were removed and divided into small pieces. The tissue was immersion-fixed for an additional hour followed by treatment with a mixture of 60% (v/v) methanol, 10% (v/v) glacial acetic acid, 30% (v/v) inhibisol (1,1,1-trichloroethane) for 24 hours. Tissues were then embedded in paraffin. Paraffin sections (5  $\mu$ m thick) were dewaxed in three changes of xylene (10 minutes each) and rehydrated in PBS. Sections were washed with 50 mM NH $_4$ Cl in PBS for 30 minutes and blocked by incubation for 1 h in TBS (10 mM Tris-HCl, pH 9.0, 150 mM NaCl) containing 10% (v/v) normal goat serum and 1% (w/v) bovine serum albumin. Samples were then incubated overnight at 4 $^{\circ}$ C with rabbit antiserum raised against pSmad2 (1:200 dilution), PDGFR $\beta$  (1:100 dilution), phospho-Erk (1:100 dilution), or preimmune rabbit serum (1:100 dilution). Sections were washed three times in TBS containing 0.1% bovine serum albumin, and bound primary antibody was detected by incubation for 2 h with Alexa-Fluor 488-conjugated goat anti-rabbit IgG (10  $\mu$ g/ml, Molecular Probes, Eugene, OR). The tissues slides were washed three times in TBS containing 0.1% bovine serum albumin, rinsed with water, and mounted using Fluorescence Mounting Medium (DakoCytomation). Images were taken using a Leica TCS SP confocal microscope.

### Lipid and Lipoprotein analyses

At sacrifice, blood was collected by cardiac puncture and plasma was analyzed by FPLC on a Superose 6 column (Sigma Chemical Co.). Plasma and liver cholesterol content was determined spectrofluorimetrically as described previously [45].

### Western blotting

MEF LRP deficient cells were seeded in 100 mm dishes (300 000 cells/dish) and grown to confluency in 10 ml DMEM supple-

mented with 10% (v/v) fetal calf serum and antibiotics. MEF LRP deficient cells were either left untreated or treated for the indicated times with the PPAR $\gamma$  agonist rosiglitazone under reduced serum conditions (0.2% fetal calf serum), followed by stimulation with recombinant human TGF $\beta$ 1 (200 pM) for 0, 1.5, 3 or 6 hours. Whole cell lysates were then prepared as described previously [1], subjected to SDS polyacrylamide gel electrophoresis, transferred to poly(vinylidene) fluoride (PVDF) membranes (Millipore), and blotted for phospho-Smad2/3 or PDGF receptor- $\beta$  using specific antibodies directed against PDGF receptor- $\beta$  and phospho-Smad2/3 (S465/467), respectively (all from Upstate Biotechnology Incorporated, Lake Placid, NY). Proteins were detected using

enhanced chemiluminescence (ECL; Amersham Pharmacia Biotech Inc, Piscataway, NJ).

## ACKNOWLEDGMENTS

We thank Bob Hammer and Sally Comerford for helpful discussions and suggestions and Megan Davenport for excellent technical assistance.

## Author Contributions

Conceived and designed the experiments: JH PB. Performed the experiments: PB WL YT RM. Analyzed the data: RA JH PB JA. Contributed reagents/materials/analysis tools: RA WL YT RM JA. Wrote the paper: JH PB.

## REFERENCES

- Boucher P, Gotthardt M, Li WP, Anderson RG, Herz J (2003) LRP: role in vascular wall integrity and protection from atherosclerosis. *Science* 300: 329–332.
- Boucher P, Liu P, Gotthardt M, Hiesberger T, Anderson RG, et al. (2002) Platelet-derived growth factor mediates tyrosine phosphorylation of the cytoplasmic domain of the low Density lipoprotein receptor-related protein in caveolae. *J Biol Chem* 277: 15507–15513.
- Takayama Y, May P, Anderson RG, Herz J (2005) Low density lipoprotein receptor-related protein 1 (LRP1) controls endocytosis and c-CBL-mediated ubiquitination of the platelet-derived growth factor receptor beta (PDGFR beta). *J Biol Chem* 280: 18504–18510.
- Boileau C, Jondeau G, Mizuguchi T, Matsumoto N (2005) Molecular genetics of Marfan syndrome. *Curr Opin Cardiol* 20: 194–200.
- Dietz HC, Loeys B, Carta L, Ramirez F (2005) Recent progress towards a molecular understanding of Marfan syndrome. *Am J Med Genet C Semin Med Genet* 139: 4–9.
- Kaartinen V, Warburton D (2003) Fibrillin controls TGF-beta activation. *Nat Genet* 33: 331–332.
- Dietz HC, Cutting GR, Pyeritz RE, Maslen CL, Sakai LY, et al. (1991) Marfan syndrome caused by a recurrent de novo missense mutation in the fibrillin gene. *Nature* 352: 337–339.
- Pereira L, Lee SY, Gayraud B, Andrikopoulos K, Shapiro SD, et al. (1999) Pathogenic sequence for aneurysm revealed in mice underexpressing fibrillin-1. *Proc Natl Acad Sci U S A* 96: 3819–3823.
- Neptune ER, Frischmeyer PA, Arking DE, Myers L, Bunton TE, et al. (2003) Dysregulation of TGF-beta activation contributes to pathogenesis in Marfan syndrome. *Nat Genet* 33: 407–411.
- Loeys BL, Chen J, Neptune ER, Judge DP, Podowski M, et al. (2005) A syndrome of altered cardiovascular, craniofacial, neurocognitive and skeletal development caused by mutations in TGFBR1 or TGFBR2. *Nat Genet* 37: 275–281.
- Mizuguchi T, Collod-Beroud G, Akiyama T, Abifadel M, Harada N, et al. (2004) Heterozygous TGFBR2 mutations in Marfan syndrome. *Nat Genet* 36: 855–860.
- Pannu H, Fadulu VT, Chang J, Lafont A, Hasham SN, et al. (2005) Mutations in transforming growth factor-beta receptor type II cause familial thoracic aortic aneurysms and dissections. *Circulation* 112: 513–520.
- Huang SS, Ling TY, Tseng WF, Huang YH, Tang FM, et al. (2003) Cellular growth inhibition by IGFBP-3 and TGF-beta1 requires LRP-1. *Faseb J* 17: 2068–2081.
- Tseng WF, Huang SS, Huang JS (2004) LRP-1/TbetaR-V mediates TGF-beta1-induced growth inhibition in CHO cells. *FEBS Lett* 562: 71–78.
- Battagay EJ, Raines EW, Seifert RA, Bowen-Pope DF, Ross R (1990) TGF-beta induces bimodal proliferation of connective tissue cells via complex control of an autocrine PDGF loop. *Cell* 63: 515–524.
- Igarashi A, Okochi H, Bradham DM, Grotendorst GR (1993) Regulation of connective tissue growth factor gene expression in human skin fibroblasts and during wound repair. *Mol Biol Cell* 4: 637–645.
- Ihn H (2002) Pathogenesis of fibrosis: role of TGF-beta and CTGF. *Curr Opin Rheumatol* 14: 681–685.
- Ikuno Y, Kazlauskas A (2002) TGFbeta1-dependent contraction of fibroblasts is mediated by the PDGFalpha receptor. *Invest Ophthalmol Vis Sci* 43: 41–46.
- Taylor LM, Khachigian LM (2000) Induction of platelet-derived growth factor B-chain expression by transforming growth factor-beta involves transactivation by Smads. *J Biol Chem* 275: 16709–16716.
- Jechlinger M, Sommer A, Moriggl R, Seither P, Kraut N, et al. (2006) Autocrine PDGFR signaling promotes mammary cancer metastasis. *J Clin Invest* 116: 1561–1570.
- Bruna A, Darken RS, Rojo F, Ocana A, Penuelas S, et al. (2007) High TGFbeta-Smad activity confers poor prognosis in glioma patients and promotes cell proliferation depending on the methylation of the PDGF-B gene. *Cancer Cell* 11: 147–160.
- Fan WH, Pech M, Karnovsky MJ (2000) Connective tissue growth factor (CTGF) stimulates vascular smooth muscle cell growth and migration in vitro. *Eur J Cell Biol* 79: 915–923.
- Takekawa M, Tatebayashi K, Itoh F, Adachi M, Imai K, et al. (2002) Smad-dependent GADD45beta expression mediates delayed activation of p38 MAP kinase by TGF-beta. *Embo J* 21: 6473–6482.
- Crawford SE, Stellmach V, Murphy-Ullrich JE, Ribeiro SM, Lawler J, et al. (1998) Thrombospondin-1 is a major activator of TGF-beta1 in vivo. *Cell* 93: 1159–1170.
- Ribeiro SM, Poczatek M, Schultz-Cherry S, Villain M, Murphy-Ullrich JE (1999) The activation sequence of thrombospondin-1 interacts with the latency-associated peptide to regulate activation of latent transforming growth factor-beta. *J Biol Chem* 274: 13586–13593.
- Orr AW, Pedraza CE, Pallero MA, Elzie CA, Goicoechea S, et al. (2003) Low density lipoprotein receptor-related protein is a calreticulin coreceptor that signals focal adhesion disassembly. *J Cell Biol* 161: 1179–1189.
- Habashi JP, Judge DP, Holm TM, Cohn RD, Loeys BL, et al. (2006) Losartan, an AT1 antagonist, prevents aortic aneurysm in a mouse model of Marfan syndrome. *Science* 312: 117–121.
- Ruiz-Ortega M, Rodriguez-Vita J, Sanchez-Lopez E, Carvajal G, Egido J (2007) TGF-beta signaling in vascular fibrosis. *Cardiovasc Res* 74: 196–206.
- Fu M, Zhang J, Zhu X, Myles DE, Willson TM, et al. (2001) Peroxisome proliferator-activated receptor gamma inhibits transforming growth factor beta-induced connective tissue growth factor expression in human aortic smooth muscle cells by interfering with Smad3. *J Biol Chem* 276: 45888–45894.
- Ghosh AK, Bhattacharyya S, Lakos G, Chen SJ, Mori Y, et al. (2004) Disruption of transforming growth factor beta signaling and profibrotic responses in normal skin fibroblasts by peroxisome proliferator-activated receptor gamma. *Arthritis Rheum* 50: 1305–1318.
- Li AC, Brown KK, Silvestre MJ, Willson TM, Palinski W, et al. (2000) Peroxisome proliferator-activated receptor {gamma} ligands inhibit development of atherosclerosis in LDL receptor-deficient mice. *J Clin Invest* 106: 523–531.
- Loukinova E, Ranganathan S, Kuznetsov S, Gorlatova N, Migliorini MM, et al. (2002) Platelet-derived growth factor (PDGF)-induced tyrosine phosphorylation of the low density lipoprotein receptor-related protein (LRP). Evidence for integrated co-receptor function between LRP and the PDGF. *J Biol Chem* 277: 15499–15506.
- Kretzschmar M, Massague J (1998) SMADs: mediators and regulators of TGF-beta signaling. *Curr Opin Genet Dev* 8: 103–111.
- Liu Q, Huang SS, Huang JS (1997) Function of the type V transforming growth factor beta receptor in transforming growth factor beta-induced growth inhibition of mink lung epithelial cells. *J Biol Chem* 272: 18891–18895.
- Emonard H, Bellon G, Troeberg L, Berton A, Robinet A, et al. (2004) Low Density Lipoprotein Receptor-related Protein Mediates Endocytic Clearance of Pro-MMP-2 (middle dot) TIMP-2 Complex through a Thrombospondin-independent Mechanism. *J Biol Chem* 279: 54944–54951.
- Hahn-Dantona E, Ruiz JF, Bornstein P, Strickland DK (2001) The Low Density Lipoprotein Receptor-related Protein Modulates Levels of Matrix Metalloproteinase 9 (MMP-9) by Mediating Its Cellular Catabolism. *J Biol Chem* 276: 15498–15503.
- Herz J, Strickland DK (2001) LRP: a multifunctional scavenger and signaling receptor. *J Clin Invest* 108: 779–784.
- Yu Q, Stamenkovic I (2000) Cell surface-localized matrix metalloproteinase-9 proteolytically activates TGF-beta and promotes tumor invasion and angiogenesis. *Genes Dev* 14: 163–176.
- Law RE, Mehan WP, Xi XP, Graf K, Wuthrich DA, et al. (1996) Troglitazone inhibits vascular smooth muscle cell growth and intimal hyperplasia. *J Clin Invest* 98: 1897–1905.
- Zirlik A, Leugers A, Lohrmann J, Ernst S, Sobel BE, et al. (2004) Direct attenuation of plasminogen activator inhibitor type-1 expression in human adipose tissue by thiazolidinediones. *Thromb Haemost* 91: 674–682.

41. Gotzmann J, Fischer AN, Zojer M, Mikula M, Proell V, et al. (2006) A crucial function of PDGF in TGF-beta-mediated cancer progression of hepatocytes. *Oncogene* 25: 3170–3185.
42. Gao BB, Hansen H, Chen HC, Feener EP (2006) Angiotensin II stimulates phosphorylation of an ectodomain-truncated platelet-derived growth factor receptor-beta and its binding to class IA PI3K in vascular smooth muscle cells. *Biochem J* 397: 337–344.
43. Morosetti R, Servidei T, Mirabella M, Rutella S, Mangiola A, et al. (2004) The PPARgamma ligands PGJ2 and rosiglitazone show a differential ability to inhibit proliferation and to induce apoptosis and differentiation of human glioblastoma cell lines. *Int J Oncol* 25: 493–502.
44. Paigen B, Morrow A, Holmes PA, Mitchell D, Williams RA (1987) Quantitative assessment of atherosclerotic lesions in mice. *Atherosclerosis* 68: 231–240.
45. Ishibashi S, Brown MS, Goldstein JL, Gerard RD, Hammer RE, et al. (1993) Hypercholesterolemia in low density lipoprotein receptor knockout mice and its reversal by adenovirus-mediated gene delivery. *J Clin Invest* 92: 883–893.