

1 **Research Letter**

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3 **Decreasing ELK3 expression improves Bone Morphogenetic Protein Receptor 2 signaling**
4 **and pulmonary vascular cell function in PAH**

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32 **Author contribution:** MKA and ES conceptualised the study design. MKA performed the
33 experiments and data analysis. All authors contributed to data collection, data interpretation,
34 writing, and editing of the manuscript. ES: fund acquisition and supervision.

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37 **Abstract:**

38 ELK3 is upregulated in blood and pulmonary vascular cells of PAH patients and may play a
39 significant role in PAH potentially through modulating BMPR2 signaling.

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41 **Introduction:**

42 Pulmonary arterial hypertension (PAH) is a rare but complex, severe, life-threatening condition
43 of the small pulmonary arteries. PAH is characterized by pulmonary arterial remodeling and
44 increased pulmonary vascular resistance; the consequent elevated pulmonary arterial pressures
45 causes right ventricular afterload and, ultimately, failure. The disease remains incurable despite
46 intense efforts to identify new therapies to treat PAH patients. In order to identify new therapies,
47 it is crucial to understand the cause and exact molecular mechanisms of the disease. Although the
48 exact cause of PAH is unclear, many genetic, epigenetic, and environmental factors have been
49 shown to contribute to the development and progression of the disease. For example,
50 heterozygous loss of function mutation in bone morphogenic protein receptor 2 (BMPR2) occurs
51 in 53-86% of familial PAH patients and 14-35% in sporadic idiopathic PAH patients(1).
52 However, the disease penetrance rate of the mutation carriers is low, indicating that other
53 unidentified factors may contribute to the disease development in addition to the gene mutation.
54 Significantly, BMPR2 signaling is thought to be impaired in PAH patients regardless of the
55 etiology of PAH, making the signaling a master switch in the disease. Previously, we and others
56 have shown that targeting BMPR2 signaling with repurposed drugs (FK506, Enzastaurin) or
57 rebalancing the pathway with BMP9 and Sotatercept improved PAH in animal models and pilot
58 studies in patients (1).

59

60 To identify BMPR2 signaling modifier genes, our team previously performed an siRNA-
61 mediated high throughput screening (HTS) of ~22,000 genes in a BRE-ID1 incorporated mouse
62 myoblastoma reporter cell line, which yielded two important novel BMPR2 modifier genes

63 (FHIT, LCK) in PAH (2). In the HTS data set, we also found that E26 transformation-specific
64 transcription factor (ELK3) knockdown decreased Id1 levels. Surprisingly, our validation
65 experiments in human pulmonary arterial endothelial cells (PAECs) and pulmonary arterial
66 smooth muscle cells (PASMCs) showed that ELK3 modulated BMPR2 signaling in the opposite
67 direction compared to the HTS experiments using the mouse myoblastoma cell line. ELK3
68 knockdown increased BMPR2 signaling. ELK3, also known as NET/SAP-2/ERP, is a
69 transcription factor which can form a ternary complex with serum response factor and DNA.
70 While ELK3 generally acts as a transcriptional repressor, it can also work as a transcriptional
71 activator when phosphorylated by the Ras/mitogen-activated protein kinase signaling pathway.
72 ELK3 regulates various biological processes in health and disease, such as proliferation,
73 apoptosis, migration, and angiogenesis. Elevated expression of ELK3 plays a significant role in
74 accelerating the progression and metastasis of different cancers, such as prostate, breast, bladder,
75 gastric, and liver cancer. A significant upregulation of ELK3 expression was also observed in rat
76 carotid arteries following balloon-injury and in human plaques (3). ELK3 was also found to
77 attenuate angiogenesis in VEGF-induced angiogenesis assays *in vitro* and *in vivo* (4). While the
78 role of ELK3 has been studied in different cancers and cardiovascular diseases, its role in PAH is
79 unknown.

80

81 **Results and Discussion:**

82 We first checked whether ELK3 expression is dysregulated in the whole blood of PAH patients
83 using a large RNA sequencing (RNAseq) data set comprising 359 patients with idiopathic,
84 heritable, and drug-induced PAH as well as 72 age- and sex-matched healthy (5). We found a
85 significant upregulation of ELK3 in the blood of PAH patients compared to healthy controls
86 (**Figure 1A**). We next tried to determine whether the upregulation of ELK3 in peripheral blood is
87 mirrored by an ELK3 upregulation in the PAH pulmonary vasculature. Thus, we re-analyzed
88 publicly available RNAseq data sets generated from two critical pulmonary vascular cell types in
89 PAH, PAECs and PASMCs of PAH patients and healthy controls. We found a significant
90 increase in ELK3 expression in the PASMCs of 4 idiopathic (IPAH) patients compared to 4
91 healthy control PASMCs (**Figure 1B**, GSE144274). We also measured the expression of ELK3
92 in PASMCs of a different cohort of 4 healthy controls and 3 PAH patients by quantitative reverse
93 transcription PCR (qRT-PCR) but did not observe a significant change in ELK3 expression

94 between the two groups (**Figure 1C**). Possible confounding factors are low ELK3 mRNA
95 expression, low sample size and different PAH etiologies, which warrants further validation in a
96 larger cohort. Previously, several studies showed increased expression of ELK3 in PAECs of
97 patients. In a single-cell RNAseq analysis study of lung tissues from 6 healthy controls and 4
98 IPAH patients, ELK3 expression was shown to be up-regulated in endothelial cells of IPAH
99 patients compared to healthy controls(6). As another example, Reyes-Palomares and colleagues
100 identified that the transcriptional activity of ELK3 was more active in PAH patients than healthy
101 controls (7). We therefore analyzed ELK3 expression in PAECs of PAH patients using a publicly
102 available RNAseq data set comprising 9 healthy controls and 8 PAH patients (GSE0126262). We
103 did not observe a significant change in ELK3 expression in PAECs of PAH patients in this
104 RNAseq dataset (**Figure 1D**). The increased detection of ELK3 in blood of PAH patients could
105 not be attributed to PASCs and PAECs in our relatively small sample size, given that these
106 cells could be the source of ELK3 in blood but that was not demonstrated in this study.

107
108 ELK3 is predicted to be connected with the inhibitor of DNA (ID) signaling pathway
109 (wikipathways). Furthermore, homozygous deficiency in ELK3 was shown to up-regulate
110 expression of SERPINE1 (Serpin family E member 1), also called plasminogen activator
111 inhibitor 1 (PAI-1) in prostate cancer cells(8). PAI-1 is a known downstream target of the BMP
112 signaling pathway and is strongly linked to PAH(9). A recent integrated bioinformatic analysis
113 revealed SERPINE1 (PAI-1) as one of the most significant markers in PAH(10). We therefore
114 hypothesized that the observed increase in ELK3 might downregulate BMPR2 signaling and
115 thereby be involved in PAH pathogenesis. We further hypothesized that decreasing ELK3 might
116 do the opposite – increase BMPR2 signaling – and thereby might be beneficial in PAH. We
117 conducted experiments in PAECs subjected to ELK3 or non-target siRNA with and without
118 BMP9 stimulation. We demonstrated that ELK3 silencing further increased BMP9-induced
119 phospho-SMAD1/5/9 levels measured by western blot (**Figures 2A-B**). ELK3 protein
120 concentration is very low in PSMCs, as seen in **Figure 2C**, and therefore we used qRT-PCR to
121 assess the effect of ELK3 knockdown on BMPR2 signaling. We found a significant increase in
122 BMPR2 as well as ID1 expression, a downstream target of the BMPR2 signaling, following the
123 knockdown of ELK3 with siRNA in PSMCs, while BMPR2 knockdown did not change the
124 ELK3 expression (**Figures 2D-G**), suggesting ELK3 to be upstream of BMPR2. We also

125 observed that inhibition of ELK3 decreased PASMCs proliferation (**Figure 2H**), which is in line
126 with the proposed model that ELK3 inhibition improves BMPR2 signaling and function in
127 PAEC and PASMC (**Figure 2I**).

128

129 Previous studies suggested that ELK3 is downregulated during hypoxia, releasing repression of
130 several genes and leading to increased expression of Egr1 and VEGF, as well as PHD2, PHD3,
131 and Siah2 destabilizing HIF1 α (6). Therefore, its enhanced expression in PAH is surprising and
132 might be expected to inhibit angiogenesis. Further studies need to explore the exact role and
133 molecular mechanisms of ELK3 in PAH.

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135 This study has several limitations. *First*, we were not able to validate the RNAseq expression of
136 ELK3 by qRT-PCR. *Second*, while ELK3 inhibition increases BMPR2 signaling, it is critical to
137 explore whether overexpression of ELK3 does the opposite, that is, inhibits BMPR2 expression
138 and signaling and induces PAH. *Third*, the molecular mechanisms of how ELK3 regulates
139 BMPR2 signaling, and PAH still need to be clarified.

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141 **Conclusion:**

142 In summary, through preliminary experimental and clinical sample analysis, we uncovered
143 ELK3 as a clinically meaningful BMPR2 signaling modulator that influences pulmonary
144 vascular cell function. Further studies are required to fully elucidate the role and molecular
145 mechanisms of ELK3 expression in the pathogenesis of PAH.

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147 **Conflict of Interest statement:**

148 The authors declared no conflict of interest exists.

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155 in the whole blood of PAH patients and healthy controls by RNAseq.

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157 **Author contribution:**

158 MKA and ES conceptualised the study design. MKA performed the experiments and data
159 analysis. All authors contributed to data collection, data interpretation, writing, and editing the
160 manuscript. ES: fund acquisition and supervision.

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163 **References:**

- 164 1. Ali MK, Ichimura K, Spiekerkoetter E. Promising therapeutic approaches in pulmonary
165 arterial hypertension. *Curr Opin Pharmacol*. 2021;59:127-39.
- 166 2. Dannewitz Prosseda S, Tian X, Kuramoto K, Boehm M, Sudheendra D, Miyagawa K, et
167 al. FHIT, a Novel Modifier Gene in Pulmonary Arterial Hypertension. *Am J Respir Crit Care*
168 *Med*. 2019;199(1):83-98.
- 169 3. Krawczyk KK, Skovsted GF, Perisic L, Dreier R, Berg JO, Hedin U, et al. Expression of
170 endothelin type B receptors (EDNRB) on smooth muscle cells is controlled by MKL2, ternary
171 complex factors, and actin dynamics. *Am J Physiol Cell Physiol*. 2018;315(6):C873-C84.
- 172 4. Heo SH, Cho JY. ELK3 suppresses angiogenesis by inhibiting the transcriptional activity
173 of ETS-1 on MT1-MMP. *Int J Biol Sci*. 2014;10(4):438-47.
- 174 5. Rhodes CJ, Otero-Nunez P, Wharton J, Swietlik EM, Kariotis S, Harbaum L, et al.
175 Whole-Blood RNA Profiles Associated with Pulmonary Arterial Hypertension and Clinical
176 Outcome. *Am J Respir Crit Care Med*. 2020;202(4):586-94.
- 177 6. Saygin D, Tabib T, Bittar HET, Valenzi E, Sembrat J, Chan SY, et al. Transcriptional
178 profiling of lung cell populations in idiopathic pulmonary arterial hypertension. *Pulm Circ*.
179 2020;10(1).
- 180 7. Reyes-Palomares A, Gu M, Grubert F, Berest I, Sa S, Kasowski M, et al. Remodeling of
181 active endothelial enhancers is associated with aberrant gene-regulatory networks in pulmonary
182 arterial hypertension. *Nat Commun*. 2020;11(1):1673.
- 183 8. Mao Y, Li W, Hua B, Gu X, Pan W, Chen Q, et al. Silencing of ELK3 Induces S-M
184 Phase Arrest and Apoptosis and Upregulates SERPINE1 Expression Reducing Migration in
185 Prostate Cancer Cells. *Biomed Res Int*. 2020;2020:2406159.
- 186 9. Chen T, Huang JB, Dai J, Zhou Q, Raj JU, Zhou G. PAI-1 is a novel component of the
187 miR-17~92 signaling that regulates pulmonary artery smooth muscle cell phenotypes. *Am J*
188 *Physiol Lung Cell Mol Physiol*. 2018;315(2):L149-L61.
- 189 10. Tang S, Liu Y, Liu B. Integrated bioinformatics analysis reveals marker genes and
190 immune infiltration for pulmonary arterial hypertension. *Sci Rep*. 2022;12(1):10154.

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193 **Figure legends**

194 **Figure 1. ELK3 is upregulated in the blood and PASMCs of PAH patients.** A) RNAseq
195 analysis of the whole blood collected from the 72 healthy controls (HC) and 359 patients with

196 IPAH, APH or HPAH showed a significant increase in ELK3 expression in PAH compared to
197 HC. For detail subject characteristics and hemodynamic data, please see (5). B) ELK3 expression
198 was found to be upregulated in PSMCs of IPAH patients in a publicly available RNAseq data
199 seta comprising 4 HC and 4 IPAH patients (GSE144274). C) ELK3 expression was not altered in
200 PSMCs of a small cohort of PAH patients by qRT-PCR. D) ELK3 expression was not changed
201 in PAECs of a small cohort of PAH patients (GSE126262). Wilcoxon rank sum test with
202 continuity correction was performed to compare ELK3 expression in the whole blood RNAseq
203 data. Student t-test was used to compare ELK3 expression in the PSMCs of PAH RNAseq
204 data. **P<0.01. TPM, transcripts per million.

205

206 **Figure 2. ELK3 is involved in regulating BMPR2 signaling and PSMCs proliferation.** A-
207 B) Inhibition of ELK3 with siRNA increased BMP9-induced pSMAD1/5/9 in PAECs, as
208 measured by western blot, 48h knock down, 2h BMP9 stimulation. C) Western blot verification
209 of ELK3 knockdown with siRNA in PSMCs. D-G) siRNA-mediated knockdown of ELK3
210 increased ID1 levels in PSMCs. BMPR2, ID1 and ELK3 levels were measured by qRT-PCR
211 following 72 hours of ELK3 or BMPR2 knockdown with 60nM siRNA and 3ul RNAimax. Data
212 are represented as mean +/- standard error mean (n=6/group). Student t test, **P<0.01. H)
213 Silencing of ELK3 decreases hPSMCs proliferation as measured by MTT assay. Student t test,
214 ****P<0.0001. I) Proposed model for how ELK3 regulates BMPR2 signaling and PAH.

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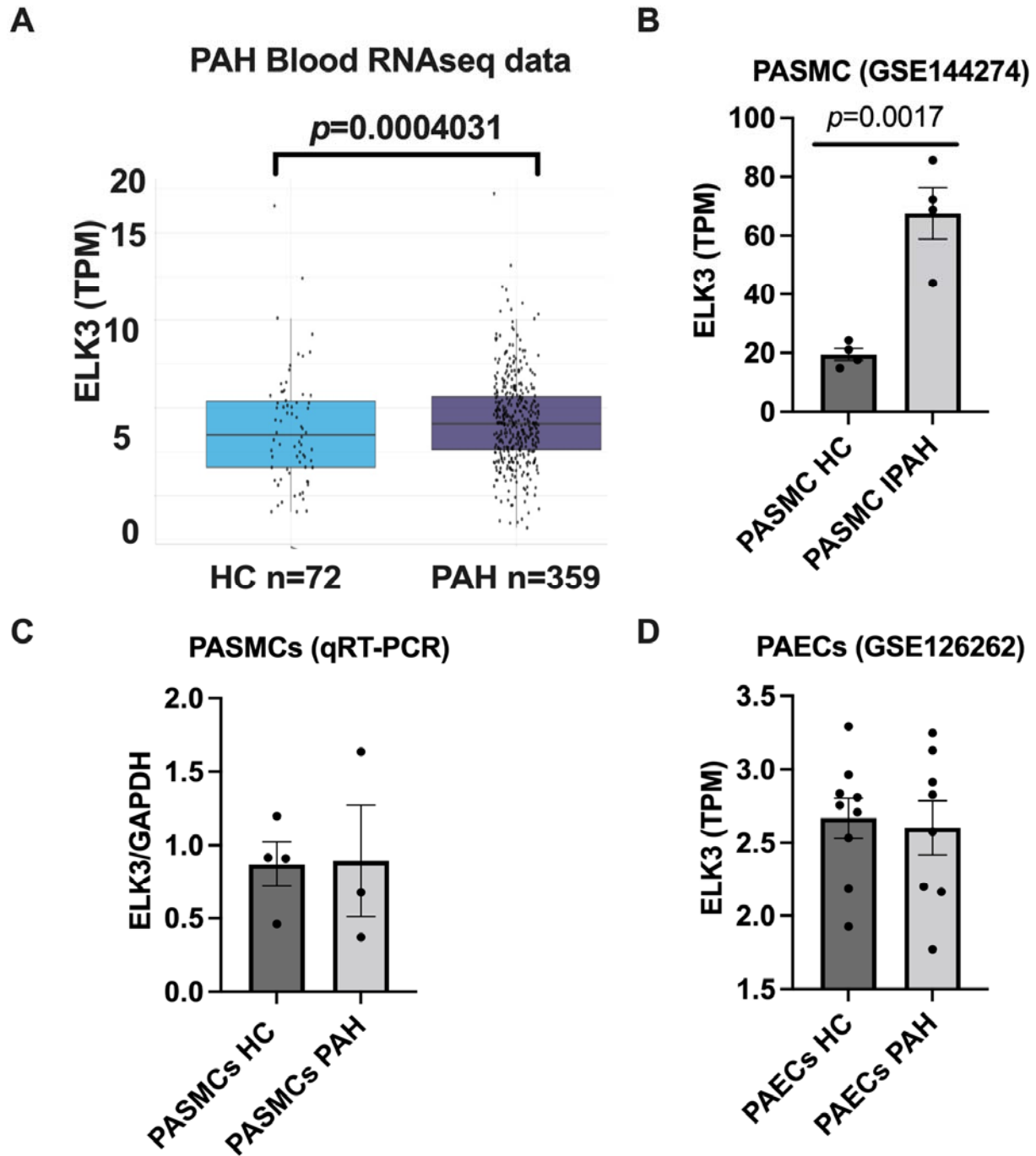
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Figure 1



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235 **Figure 2**

