

Review

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Mechanisms by Which Atopic Dermatitis Predisposes to Food Allergy and the Atopic March

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ABSTRACT

The Atopic march denotes the progression from atopic dermatitis (AD) to the development of other allergic disorders such as immunoglobulin (Ig) E-mediated food allergy, allergic rhinitis and asthma in later childhood. There is increasing evidence from prospective birth cohort studies that early-onset AD is a risk factor for other allergic diseases or is found in strong association with them. Animal studies now provide mechanistic insights into the pathways that may be responsible for triggering the progression from the skin barrier dysfunction seen in AD to epicutaneous sensitization, food allergy and allergic airway disorders. Recent large randomized controlled trials have demonstrated the efficacy of early interventions targeted at AD and food allergy prevention. These show great promise for research into future strategies aimed at prevention of the atopic march.

Keywords: Atopic march; atopic dermatitis; allergic rhinitis, asthma, food allergy

INTRODUCTION

The increasing prevalence of allergic disorders worldwide imposes a significant socioeconomic burden on society. The International Study of Asthma and Allergies in Childhood (ISAAC) Phase IIIwhich surveyed around 1.2 million children across 233 centers in 98 countries found the prevalence of atopic dermatitis (AD) to be 7.9%, asthma 11.7%, and rhinoconjunctivitis 8.5% in 6- to 7-year old children, and 7.3% (AD), 14.1% (asthma) and 7.3% (rhinoconjunctivitis) in 13- to 14-year olds.¹ Food allergy affects up to 8% of children and its prevalence has been increasing over the past 2 decades.²

The term "atopic march" was coined to describe the progression from AD in early infancy to other allergic diseases such as food allergy, asthma and allergic rhinitis in later childhood (**Fig. 1**).^{3,4} AD is a chronic inflammatory skin disorder which is marked by skin barrier dysfunction, frequent skin infections and impaired quality of life. In many children, AD is the first manifestation of an atopic phenotype which begins in early infancy. A recent study suggested that the complete manifestations of the atopic march occurred in less than 10% of AD patients⁵; however, approximately 40% of AD patients have food allergy. Severity of AD is a factor determining subsequent development of allergic disease.

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Fig. 1. The atopic march. The figure shows the age at diagnosis of each allergic disease. Reproduced with permission from *Ann Allergy Asthma Immunol* 2018;120:131-7.⁴

Several prospective birth cohorts have shown associations between early-onset AD and development of asthma and allergic rhinitis in school age.^{6,7} The risks appear to be even greater in children with the early-onset persistent AD phenotype.⁸ The Canadian Healthy Infant Longitudinal Development Study found that children with AD and allergic sensitization had a greatly increased risk of food allergy, asthma and allergic rhinitis compared to non-sensitized children with AD.⁹

The combined study by the high-risk Melbourne Atopy Cohort Study (MACS) and the population-based Influences of Lifestyle-Related Factors on the Immune System and the Development of Allergies in Childhood plus Air Pollution and Genetics (LISAplus) cohort found that food sensitization in the first 2 years of life was associated with increased risks of asthma and allergic rhinitis by age 10 years.¹⁰ A meta-analysis of 13 cohorts studying the atopic march also demonstrated that early food sensitization was associated with increased risks of wheeze/asthma and allergic rhinitis at 4 to 8 years of age.¹¹ Hill *et al.*¹² likewise observed that children with food allergy, particularly peanut, milk and egg allergy, were at increased risk of developing asthma and rhinitis in later childhood.

This review explores the evidence supporting the concept that impaired epithelial barrier in AD initiates progression to epicutaneous sensitization, food allergy and allergic airway disease in later life as well as intervention strategies aimed at preventing progression through the atopic march.

GENETIC PREDISPOSITION TO SKIN BARRIER DYSFUNCTION AND AD

The dual-allergen exposure hypothesis postulates that exposure to environmental food allergens induces sensitization and ensuing food allergy through an impaired skin barrier as in AD. Immune tolerance can, however, be achieved through early oral exposure to food allergens.¹³

Intact healthy skin serves as an important physical and immunological barrier against the entry of environmental allergens and microbes into the body. The epidermis is comprised of terminally



differentiated keratinocytes, which form a cornified layer of keratin filaments aggregated by filaggrin (FLG), involucrin and loricrin. Epidermal lipids, such as ceramides, are produced by keratinocytes and surround the keratin filaments, forming an impermeable skin barrier.¹⁴ Skin barrier defects may arise from multiple factors, including defects in terminal epithelial differentiation such as lack of FLG, deficiency of antimicrobial peptides (AMPs), altered stratum corneum intercellular lipids, skin microbiome dysbiosis and immune dysregulation.¹⁵

Genetic defects encoding skin barrier proteins and abnormalities in lipid production or tight junctions contribute to epithelial barrier dysfunction that characterizes AD. The *FLG* gene encodes filaggrin, an essential epidermal protein expressed in the outer layers of the epidermis, playing an important role in maintaining skin barrier integrity. *FLG* loss-of-function mutations are associated with an earlier onset as well as a more persistent phenotype of AD.¹⁶ Polymorphisms in the thymic stromal lymphopoietin (*TSLP*) gene and its receptor as well as dysregulation of genes involved in epidermal lipid metabolism have likewise been reported to be associated with increased risks of AD and its persistence.¹⁷⁴⁹ Other skin barrier mutations involving the *SPINK5* gene²⁰ and corneodesmosin²¹ as well as genome-wide association study (GWAS)-identified genetic variants²² have also been linked to AD and the development of food allergy.

Kelleher *et al.*²³ found that neonates with increased transepidermal water loss (TEWL) in the first week of life had an increased risk of developing AD at 12 months of age. The strong genetic basis for AD described above supports the view that skin barrier impairment may be evident very early in life, even before the onset of clinical AD.

SKIN BARRIER DYSFUNCTION PREDISPOSES TO EPICUTANEOUS SENSITIZATION AND FOOD ALLERGY

There is now a growing body of evidence supporting the notion that epicutaneous allergen sensitization occurs more readily through an impaired skin barrier.^{15,24} Murine models demonstrated that the application of ovalbumin (OVA) or peanut allergens on abraded mice skin resulted in significant systemic egg and peanut-specific immunoglobulin (Ig) E responses.^{25,26} In a mouse model of *FLG* deficiency, flaky tailed mice with a 1-bp deletion mutation (5303delA) within the murine *FLG* gene were treated with topical applications of OVA. These mice displayed increased cutaneous inflammatory infiltrates and enhanced cutaneous allergen priming with raised specific IgE to OVA compared to wild-type mice, even without prior abrasion of the skin. This suggests that *FLG* mutation-induced skin barrier dysfunction allowed for cutaneous penetration of allergens and the development of systemic allergic responses.²⁷

Several murine models have also described the immunological pathways involved in the progression from epicutaneous sensitization to food allergy. Mice which were epicutaneously sensitized with topical applications of OVA on tape-stripped skin (to mimick scratching) exhibited IgE-dependent intestinal mast cell expansion, increased serum interleukin (IL)-4 levels and systemic anaphylaxis following oral OVA challenge.²⁸ In another study, mice epicutaneously sensitized with OVA or peanut on an AD-like skin lesion demonstrated *TSLP*-dependent basophil expansion in the skin, which promoted antigen-specific Th2 cytokine responses, increased antigen-specific serum IgE levels and accumulation of mast cells in the intestine akin to IgE-mediated food allergy.²⁹ Conversely, disruption of the *TSLP*-basophil axis



abrogated this allergic response. A follow-up study showed infiltration of IL-4-competent basophils and eosinophils into the skin of these epicutaneously OVA-sensitized mice, indicating that *TSLP*-dependent basophils mediate Th2-allergic sensitization to food antigens through the production of IL-4 and that these mechanisms likely play a key role in the pathogenesis of epicutaneously induced intestinal food allergy.³⁰

The role of IL-33 in mediating oral anaphylaxis through epicutaneous sensitization is now also emerging. In another murine model of epicutaneous OVA-sensitization, Galand *et al.*³¹ showed that mechanical skin damage induced by tape stripping induced local and systemic IL-33 release, which then enhanced IgE-mediated mast cell degranulation and induced anaphylaxis after oral antigen challenge. Disruption of IL-33 signalling through blockade of ST2 (the IL-33 receptor) resulted in the inhibition of oral anaphylaxis.

Findings from clinical studies also support the concept of the atopic march from early skin barrier dysfunction to the development of food sensitization and clinical food allergy. Increased TEWL at 2 days of life was found to be predictive of food allergy development at 2 years of age.³² A birth cohort study found that children with chronic low-dose topical exposure to peanut allergens, in the form of arachis oil, on inflamed skin had an increased risk of peanut allergy at age 5 years.³³ Another study found that 32% of children with AD who used oat-based creams had positive patch tests to oat compared to 0% in those who did not.³⁴ The MACS study found that children with AD at age 6 months had increased risks of developing new-onset food and inhalant sensitization by 1 and 2 years of age, respectively.³⁵ In the HealthNuts cohort, infants with AD in the first year of life had an 11-fold risk of developing peanut allergy and 6-fold risk of egg allergy by age 12 months.³⁶ Up to 50% of the infants who had moderate to severe AD in the first 3 months of age requiring topical corticosteroid use subsequently developed challenge-proven food allergy.

Skin microbiome dysbiosis also contributes to skin barrier dysfunction and modulates epicutaneous allergen sensitization. Up to 90% of AD patients are colonized with *Staphylococcus aureus*. Disease severity and exacerbations are linked to increased *S. aureus* abundance and reduced local bacterial diversity, which are evident even at non-lesional sites.³⁷ *S. aureus* perpetuates the pathogenesis of AD through various mechanisms, including disruption of epidermal integrity through protease activity, down-regulation of terminal differentiation markers in the skin and production of virulence factors such as cytolysins, protein A and superantigens.³⁸ Staphylococcal-derived superantigens, such as staphylococcal enterotoxin B (SEB), have been shown to enhance Th2 inflammation in the skin in response to peanut allergens and drive the development of peanut allergy.³⁹ Jones *et al.*⁴⁰ also observed that colonization with *S. aureus*, particularly methicillin-resistant *S. aureus* (MRSA) was associated with a higher risk of food allergy.

THE MARCH FROM AD TO ALLERGIC RHINITIS AND ASTHMA

The link between AD and allergic airway diseases is influenced by the age of AD onset and its severity. High-risk infants in the MACS study with early-onset persistent AD had a 3-fold risk of developing asthma and allergic rhinitis in later childhood, compared to children with late-onset AD that began after 2 years of age.⁴¹ In another Swedish cohort, more than 60% of children with severe AD before 3 years of age developed asthma by age 7, compared to only



20% of those with mild AD.⁴² The presence of AD was also associated with increased asthma severity as well as greater persistence into adulthood.^{43,44}

Inherent genetic susceptibility may also influence one's risk of progression from AD to allergic airway disease. A GWAS study identified shared asthma, hay fever and AD immunerelated gene variants, suggesting that these allergic disorders may co-exist because they share genetic risk loci that result in dysregulation of immune-related genes.⁴⁵ Another AD GWAS study also found genetic loci that overlapped between asthma and AD.⁴⁶ A multi-stage GWAS study on children with infantile AD and childhood asthma identified novel genetic loci (rs9357733 located in EFHC1 on chromosome 6p12.3 and rs993226 between TMTC2 and SLC6A15 on chromosome 12q21.3) which were specific for the AD-to-asthma atopic march phenotype.⁴⁷ A Kinesin family member 3A (KIF3A) genetic variant was also found to be associated with AD-asthma comorbidity in a population cohort from the Greater Cincinnati Pediatric Clinic Repository (GCPCR).⁴⁸

The mechanisms behind the march from AD to allergic airway disease likely arise from initial epicutaneous allergen sensitization inducing robust local and systemic Type 2 immune responses, supporting the view that AD is not merely a disease confined to the skin, but is in fact a systemic disease. Epicutaneous exposure to the *Aspergillus fumigatus* aeroallergen has been reported to induce nasal hyperresponsiveness to methacholine and inflammation upon subsequent intranasal *A. fumigatus* challenge in mice through a STAT6-dependent pathway.⁴⁹ Epicutaneous OVA application to tape-stripped mouse skin, followed by a bronchial OVA challenge, also induced bronchial eosinophilia and airway hyperresponsiveness to methacholine, akin to allergic asthma.²⁶

The inflammatory responses induced by AD are manifested by increased production of Type 2 cytokines such as IL-4, IL-13, IL-25 and IL-33 as well as TSLP. The latter now appears to be a major player in inducing systemic Type 2-immune responses, which are responsible for the pathogenesis of allergic airway diseases. TSLP is an IL-7 related cytokine that is expressed predominantly in skin keratinocytes, pulmonary airways and intestinal epithelium. Increased TSLP expression has been observed in both the skin of AD subjects and asthmatic pulmonary epithelia.⁵⁰ Intradermal administration of TSLP together with OVA, in mice, leads to an AD-like skin inflammation with epicutaneous OVA sensitization, and results in an allergic asthma-like phenotype upon airway OVA challenge.⁵¹ Another murine model demonstrated that overproduction of TSLP by AD skin promoted airway sensitization to house dust mite (HDM) aeroallergens and induced allergic airway responses in sensitized mice.⁵²

EPICUTANEOUS SENSITIZATION WITHOUT AN IMPAIRED SKIN BARRIER

Interestingly, epicutaneous sensitization has been found to cause clinical food allergy or airway allergic disease even without a disrupted skin barrier. These events occur in the presence of an adjuvant or following highly concentrated chronic exposure. Dunkin *et al.*⁵³ found that mice sensitized to the milk allergen, α -lactalbumin (ALA), through the cutaneous route by direct skin application of ALA and cholera toxin, used as an adjuvant, developed systemic anaphylaxis upon oral challenge with ALA. An outbreak of new-onset IgE-mediated wheat protein allergy was also reported in previously healthy Japanese adults after chronic usage of a wheat-containing facial soap on intact facial skin.⁵⁴ HDM allergens



applied epicutaneously to unmanipulated mouse ears or under an occlusive tape induced Th2 sensitization and eosinophilic airway inflammation upon intranasal HDM challenge.⁵⁵ In another study, mice cutaneously sensitized to HDM allergen Der p 2 through intact skin developed significant systemic allergen-specific IgE and IgG1 antibody responses.⁵⁶ These responses were augmented by the addition of lipopolysaccharide (LPS), suggesting that it could act as an adjuvant inducing aeroallergen sensitization through the skin.

Likewise, Walker *et al.*⁵⁷ demonstrated that when neonatal mice heterozygous for skin barrier mutations were cutaneously sensitized to food allergens in the presence of fungal (*Alternaria alternata*) or HDM extracts, they developed food allergen sensitization, foodinduced anaphylaxis and increased skin and intestinal inflammation upon oral food allergen challenge. These events were not observed when mice were sensitized with food allergen alone. In these mice, pre-exposure with oral peanut extract before skin sensitization was able to induce oral peanut tolerance. However, co-exposure to *A. alternata* at the same time as the oral peanut extract reduced the induction of oral tolerance. This suggested that co-exposure to environmental allergens, which alter the skin barrier, may facilitate food allergen sensitization and the development of food allergies early in life in infants with skin barrier mutations.

PREVENTIVE MEASURES AGAINST THE ATOPIC MARCH

The increasing body of knowledge surrounding risk factors for the atopic march has triggered renewed interest in the study of preventive interventions aimed at halting progression through the atopic march. Many of the primary prevention measures have been targeted at reducing the risk of AD development through prophylactic skin barrier protection beginning very early in life in high-risk neonates (Fig. 2). Horimukai et al.58 and Simpson et al.⁵⁹ both demonstrated that regular application of standard emollients from birth reduced the incidence of AD at 8 and 6 months of age, respectively. The latter study also reported that children in the emollient group had a lower skin pH and altered skin microbiome, with increased proportions of Streptococcus salivarius organisms at various skin sites, compared to the control group, suggesting that these mechanisms may contribute to the preventative effects of emollients in these high-risk infants.⁶⁰ The Prevention of AD By a Barrier Lipid Equilibrium Strategy (PEBBLES) study, which utilized topical applications of a trilipidrich ceramide-dominant emollient in high-risk infants from birth to 6 months, showed decreased food sensitization at age 6 and 12 months when emollient therapy was introduced during the first 3 weeks of life.⁶¹ Several larger trials are also currently ongoing which aim to assess the efficacy of regular moisturizing for skin barrier protection in early life against the development of not just AD, but also challenge-proven food allergy and allergic airway diseases.62,63

There is some evidence that probiotic supplementation in pregnancy or infancy may be protective against the development of AD, but no protection against food allergy or other allergic disorders have been proved.⁶⁴ There is still insufficient evidence for the use of other nutritive interventions such as prebiotics, hydrolyzed formulas or vitamin D supplementation in the primary prevention of allergic diseases.

Secondary prevention of allergic disease involves interventions targeted at high-risk infants or children with known disease such as pre-existing AD, or food sensitization without clinical





Fig. 2. Emollient therapy for AD prevention. Early skin barrier protection with prophylactic emollient therapy might prevent AD development. Original source: *J Allergy Clin Immunol* 2014;134:818-23, an open access article under the CC BY license (http://creativecommons.org/licenses/by/3.0/).⁵⁹ AD, atopic dermatitis; FLG, filaggrin; TSLP, thymic stromal lymphopoietin.

food allergy, aiming to prevent progression to the next phase of the atopic march. The Japanese Prevention of Allergy via Cutaneous Intervention (PACI) study is one such ongoing randomized controlled trial that plans to measure the effects of aggressive (proactive) treatment of 7- to 13-week old infants with AD versus standard (reactive) AD treatment for the prevention of challenge-proven IgE-mediated hen's egg allergy at 28 weeks of age.⁶⁵

Several randomized controlled trials have now shown that early introduction of allergenic foods such as peanut or egg to high-risk infants with severe AD or pre-existing food sensitization may reduce the risk of developing peanut or egg allergies, respectively. High-risk infants aged 4-11 months with severe AD and/or egg allergy and peanut skin prick test (SPT) responses of 1-4 mm were enrolled into the Learning Early About Peanut Allergy (LEAP) trial. Infants randomized to consume 6 grams of peanut protein at least 3 times a week or 2 grams 3 times a week had an 81.4% lower risk of developing challenge-proven peanut allergy by age 5 years, compared to those who completely avoided peanut products.⁶⁶ The Prevention of Egg allergy with Tiny amount InTake (PETIT) study investigated the effects of early introduction of heated egg powder to high-risk Japanese infants with AD on the development of egg allergy.⁶⁷ The intervention group which received 50 mg of heated whole egg powder between 6-9 months of age, then 250 mg between 9-12 months of age, had 30% lower risk of developing challenge-proven hen's egg allergy at 12 months of age, compared to the placebo group.

Several primary prevention strategies for asthma and allergic rhinitis have also been investigated. These include HDM avoidance in early life and prophylactic sublingual immunotherapy in sensitized children. None, however, have shown sufficient evidence for adoption into routine clinical practice yet.^{68,69}



CONCLUSIONS

There is strong evidence for the link between early-onset AD and the development of other allergic diseases in later life. Several preventive interventions such as early regular emollient use and early introduction of peanut and egg in high-risk infants have shown promising results for the prevention of AD as well as peanut and egg allergy. However, more studies are needed to determine the efficacy of these interventions on prevention of other food allergies and allergic airway disorders. Future research should also focus on identifying biomarkers predictive of risk of progression through the atopic march, in order to develop interventions specific to this high-risk group. There is reason to hope that more options will eventually be available to prevent or halt progression through the atopic march.

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REFERENCES

- Mallol J, Crane J, von Mutius E, Odhiambo J, Keil U, Stewart A, et al. The International Study of Asthma and Allergies in Childhood (ISAAC) phase three: a global synthesis. Allergol Immunopathol (Madr) 2013;41:73-85.
 PUBMED | CROSSREF
- Sicherer SH, Sampson HA. Food allergy: epidemiology, pathogenesis, diagnosis, and treatment. J Allergy Clin Immunol 2014;133:291-307.
 PUBMED | CROSSREF
- Johansson E, Hershey GK. Contribution of an impaired epithelial barrier to the atopic march. Ann Allergy Asthma Immunol 2018;120:118-9.
 PUBMED | CROSSREF
- Hill DA, Spergel JM. The atopic march: critical evidence and clinical relevance. Ann Allergy Asthma Immunol 2018;120:131-7.
 PUBMED | CROSSREF
- Belgrave DC, Granell R, Simpson A, Guiver J, Bishop C, Buchan I, et al. Developmental profiles of eczema, wheeze, and rhinitis: two population-based birth cohort studies. PLoS Med 2014;11:e1001748.
 PUBMED | CROSSREF
- Saunes M, Øien T, Dotterud CK, Romundstad PR, Storrø O, Holmen TL, et al. Early eczema and the risk of childhood asthma: a prospective, population-based study. BMC Pediatr 2012;12:168.
 PUBMED | CROSSREF
- von Kobyletzki LB, Bornehag CG, Hasselgren M, Larsson M, Lindström CB, Svensson Å. Eczema in early childhood is strongly associated with the development of asthma and rhinitis in a prospective cohort. BMC Dermatol 2012;12:11.
 PUBMED I CROSSREF
- Carlsten C, Dimich-Ward H, Ferguson A, Watson W, Rousseau R, Dybuncio A, et al. Atopic dermatitis in a high-risk cohort: natural history, associated allergic outcomes, and risk factors. Ann Allergy Asthma Immunol 2013;110:24-8.
 PUBMED I CROSSREF
- 9. Tran MM, Lefebvre DL, Dharma C, Dai D, Lou WY, Subbarao P, et al. Predicting the atopic march: results from the Canadian Healthy Infant Longitudinal Development Study. J Allergy Clin Immunol 2018;141:601-607.e8.

PUBMED | CROSSREF



- 10. Alduraywish SA, Standl M, Lodge CJ, Abramson MJ, Allen KJ, Erbas B, et al. Is there a march from early food sensitization to later childhood allergic airway disease? Results from two prospective birth cohort studies. Pediatr Allergy Immunol 2017;28:30-7. PUBMED | CROSSREF
- 11. Alduravwish SA, Lodge CJ, Campbell B, Allen KJ, Erbas B, Lowe AJ, et al. The march from early life food sensitization to allergic disease: a systematic review and meta-analyses of birth cohort studies. Allergy 2016:71:77-89. PUBMED | CROSSREF
- 12. Hill DA, Grundmeier RW, Ram G, Spergel JM. The epidemiologic characteristics of healthcare providerdiagnosed eczema, asthma, allergic rhinitis, and food allergy in children: a retrospective cohort study. BMC Pediatr 2016;16:133. PUBMED | CROSSREF
- 13. Lack G. Epidemiologic risks for food allergy. J Allergy Clin Immunol 2008;121:1331-6. PUBMED | CROSSREE
- Berdyshev E, Goleva E, Bronova I, Dyjack N, Rios C, Jung J, et al. Lipid abnormalities in atopic skin are 14. driven by type 2 cytokines. JCI Insight 2018;3:98006. PUBMED | CROSSREE
- 15. Kim BE, Leung DY. Significance of skin barrier dysfunction in atopic dermatitis. Allergy Asthma Immunol Res 2018;10:207-15. PUBMED | CROSSREF
- Barker JN, Palmer CN, Zhao Y, Liao H, Hull PR, Lee SP, et al. Null mutations in the filaggrin gene (FLG) 16. determine major susceptibility to early-onset atopic dermatitis that persists into adulthood. J Invest Dermatol 2007;127:564-7.
 - PUBMED | CROSSREF
- 17. Margolis DJ, Kim B, Apter AJ, Gupta J, Hoffstad O, Papadopoulos M, et al. Thymic stromal lymphopoietin variation, filaggrin loss of function, and the persistence of atopic dermatitis. JAMA Dermatol 2014:150:254-9. PUBMED | CROSSREF
- 18. Cole C, Kroboth K, Schurch NJ, Sandilands A, Sherstnev A, O'Regan GM, et al. Filaggrin-stratified transcriptomic analysis of pediatric skin identifies mechanistic pathways in patients with atopic dermatitis. J Allergy Clin Immunol 2014;134:82-91. PUBMED | CROSSREF
- 19. Ewald DA, Malajian D, Krueger JG, Workman CT, Wang T, Tian S, et al. Meta-analysis derived atopic dermatitis (MADAD) transcriptome defines a robust AD signature highlighting the involvement of atherosclerosis and lipid metabolism pathways. BMC Med Genomics 2015;8:60. PUBMED | CROSSREF
- 20. Ashley SE, Tan HT, Vuillermin P, Dharmage SC, Tang ML, Koplin J, et al. The skin barrier function gene SPINK5 is associated with challenge-proven IgE-mediated food allergy in infants. Allergy 2017;72:1356-64. PUBMED | CROSSREF
- 21. Marenholz I, Esparza-Gordillo J, Lee YA. The genetics of the skin barrier in eczema and other allergic disorders. Curr Opin Allergy Clin Immunol 2015;15:426-34. PUBMED | CROSSREE
- 22. Hirota T, Nakayama T, Sato S, Yanagida N, Matsui T, Sugiura S, et al. Association study of childhood food allergy with genome-wide association studies-discovered loci of atopic dermatitis and eosinophilic esophagitis. J Allergy Clin Immunol 2017;140:1713-6. PUBMED | CROSSREF
- 23. Kelleher M, Dunn-Galvin A, Hourihane JO, Murray D, Campbell LE, McLean WH, et al. Skin barrier dysfunction measured by transepidermal water loss at 2 days and 2 months predates and predicts atopic dermatitis at 1 year. J Allergy Clin Immunol 2015;135:930-935.e1. PUBMED | CROSSREF
- 24. Lack G. Update on risk factors for food allergy. J Allergy Clin Immunol 2012;129:1187-97. PUBMED | CROSSREF
- 25. Strid J, Hourihane J, Kimber I, Callard R, Strobel S. Disruption of the stratum corneum allows potent epicutaneous immunization with protein antigens resulting in a dominant systemic Th2 response. Eur J Immunol 2004;34:2100-9. PUBMED | CROSSREF
- 26. Spergel JM, Mizoguchi E, Brewer JP, Martin TR, Bhan AK, Geha RS. Epicutaneous sensitization with protein antigen induces localized allergic dermatitis and hyperresponsiveness to methacholine after single exposure to aerosolized antigen in mice. J Clin Invest 1998;101:1614-22. PUBMED | CROSSREF



- Fallon PG, Sasaki T, Sandilands A, Campbell LE, Saunders SP, Mangan NE, et al. A homozygous frameshift mutation in the mouse FLG gene facilitates enhanced percutaneous allergen priming. Nat Genet 2009;41:602-8.
 PUBMED | CROSSREF
- Bartnikas LM, Gurish MF, Burton OT, Leisten S, Janssen E, Oettgen HC, et al. Epicutaneous sensitization results in IgE-dependent intestinal mast cell expansion and food-induced anaphylaxis. J Allergy Clin Immunol 2013;131:451-460.e1-6.
 PUBMED | CROSSREF
- Noti M, Kim BS, Siracusa MC, Rak GD, Kubo M, Moghaddam AE, et al. Exposure to food allergens through inflamed skin promotes intestinal food allergy through the thymic stromal lymphopoietinbasophil axis. J Allergy Clin Immunol 2014;133:1390-9, 1399.e1-6.
 PUBMED | CROSSREF
- Hussain M, Borcard L, Walsh KP, Pena Rodriguez M, Mueller C, Kim BS, et al. Basophil-derived IL-4 promotes epicutaneous antigen sensitization concomitant with the development of food allergy. J Allergy Clin Immunol 2018;141:223-234.e5.
- Galand C, Leyva-Castillo JM, Yoon J, Han A, Lee MS, McKenzie AN, et al. IL-33 promotes food anaphylaxis in epicutaneously sensitized mice by targeting mast cells. J Allergy Clin Immunol 2016;138:1356-66.
 PUBMED | CROSSREF
- 32. Kelleher MM, Dunn-Galvin A, Gray C, Murray DM, Kiely M, Kenny L, et al. Skin barrier impairment at birth predicts food allergy at 2 years of age. J Allergy Clin Immunol 2016;137:1111-1116.e8.
 PUBMED | CROSSREF
- 33. Lack G, Fox D, Northstone K, Golding JAvon Longitudinal Study of Parents and Children Study Team. Factors associated with the development of peanut allergy in childhood. N Engl J Med 2003;348:977-85. PUBMED | CROSSREF
- Boussault P, Léauté-Labrèze C, Saubusse E, Maurice-Tison S, Perromat M, Roul S, et al. Oat sensitization in children with atopic dermatitis: prevalence, risks and associated factors. Allergy 2007;62:1251-6.
 PUBMED | CROSSREF
- 35. Lowe AJ, Abramson MJ, Hosking CS, Carlin JB, Bennett CM, Dharmage SC, et al. The temporal sequence of allergic sensitization and onset of infantile eczema. Clin Exp Allergy 2007;37:536-42.
 PUBMED | CROSSREF
- 36. Martin PE, Eckert JK, Koplin JJ, Lowe AJ, Gurrin LC, Dharmage SC, et al. Which infants with eczema are at risk of food allergy? Results from a population-based cohort. Clin Exp Allergy 2015;45:255-64. PUBMED | CROSSREF
- Tauber M, Balica S, Hsu CY, Jean-Decoster C, Lauze C, Redoules D, et al. *Staphylococcus aureus* density on lesional and nonlesional skin is strongly associated with disease severity in atopic dermatitis. J Allergy Clin Immunol 2016;137:1272-1274.e3.
 PUBMED | CROSSREF
- Czarnowicki T, Krueger JG, Guttman-Yassky E. Skin barrier and immune dysregulation in atopic dermatitis: an evolving story with important clinical implications. J Allergy Clin Immunol Pract 2014;2:371-9.
 PUBMED | CROSSREF
- Forbes-Blom E, Camberis M, Prout M, Tang SC, Le Gros G. Staphylococcal-derived superantigen enhances peanut induced Th2 responses in the skin. Clin Exp Allergy 2012;42:305-14.
 PUBMED | CROSSREF
- Jones AL, Curran-Everett D, Leung DY. Food allergy is associated with *Staphylococcus aureus* colonization in children with atopic dermatitis. J Allergy Clin Immunol 2016;137:1247-1248.e3.
 PUBMED | CROSSREF
- Lowe AJ, Angelica B, Su J, Lodge CJ, Hill DJ, Erbas B, et al. Age at onset and persistence of eczema are related to subsequent risk of asthma and hay fever from birth to 18 years of age. Pediatr Allergy Immunol 2017;28:384-90.
 PUBMED | CROSSREF
- Gustafsson D, Sjöberg O, Foucard T. Development of allergies and asthma in infants and young children with atopic dermatitis--a prospective follow-up to 7 years of age. Allergy 2000;55:240-5.
 PUBMED | CROSSREF
- 43. Strachan DP, Butland BK, Anderson HR. Incidence and prognosis of asthma and wheezing illness from early childhood to age 33 in a national British cohort. BMJ 1996;312:1195-9.
 PUBMED | CROSSREF



- 44. Martinez FD, Wright AL, Taussig LM, Holberg CJ, Halonen M, Morgan WJ, et al. Asthma and wheezing in the first six years of life. N Engl J Med 1995;332:133-8.
 PUBMED | CROSSREF
- 45. Ferreira MA, Vonk JM, Baurecht H, Marenholz I, Tian C, Hoffman JD, et al. Shared genetic origin of asthma, hay fever and eczema elucidates allergic disease biology. Nat Genet 2017;49:1752-7. PUBMED | CROSSREF
- 46. Weidinger S, Willis-Owen SA, Kamatani Y, Baurecht H, Morar N, Liang L, et al. A genome-wide association study of atopic dermatitis identifies loci with overlapping effects on asthma and psoriasis. Hum Mol Genet 2013;22:4841-56.
 PURMED L CROSSREF
- Marenholz I, Esparza-Gordillo J, Rüschendorf F, Bauerfeind A, Strachan DP, Spycher BD, et al. Metaanalysis identifies seven susceptibility loci involved in the atopic march. Nat Commun 2015;6:8804.
 PUBMED | CROSSREF
- Johansson E, Biagini Myers JM, Martin LJ, He H, Pilipenko V, Mersha T, et al. KIF3A genetic variation is associated with pediatric asthma in the presence of eczema independent of allergic rhinitis. J Allergy Clin Immunol 2017;140:595-598.e5.
- Akei HS, Brandt EB, Mishra A, Strait RT, Finkelman FD, Warrier MR, et al. Epicutaneous aeroallergen exposure induces systemic TH2 immunity that predisposes to allergic nasal responses. J Allergy Clin Immunol 2006;118:62-9.
 PUBMED | CROSSREF
- Cianferoni A, Spergel J. The importance of TSLP in allergic disease and its role as a potential therapeutic target. Expert Rev Clin Immunol 2014;10:1463-74.
- Han H, Xu W, Headley MB, Jessup HK, Lee KS, Omori M, et al. Thymic stromal lymphopoietin (TSLP)mediated dermal inflammation aggravates experimental asthma. Mucosal Immunol 2012;5:342-51.
 PUBMED | CROSSREF
- 52. Jiang H, Hener P, Li J, Li M. Skin thymic stromal lymphopoietin promotes airway sensitization to inhalant house dust mites leading to allergic asthma in mice. Allergy 2012;67:1078-82.
 PUBMED | CROSSREF
- Dunkin D, Berin MC, Mayer L. Allergic sensitization can be induced via multiple physiologic routes in an adjuvant-dependent manner. J Allergy Clin Immunol 2011;128:1251-1258.e2.
 PUBMED | CROSSREF
- Yagami A, Aihara M, Ikezawa Z, Hide M, Kishikawa R, Morita E, et al. Outbreak of immediate-type hydrolyzed wheat protein allergy due to a facial soap in Japan. J Allergy Clin Immunol 2017;140:879-881.e7.
 PUBMED | CROSSREF
- 55. Deckers J, Sichien D, Plantinga M, Van Moorleghem J, Vanheerswynghels M, Hoste E, et al. Epicutaneous sensitization to house dust mite allergen requires interferon regulatory factor 4-dependent dermal dendritic cells. J Allergy Clin Immunol 2017;140:1364-1377.e2.
 PUBMED | CROSSREF
- 56. Stremnitzer C, Manzano-Szalai K, Starkl P, Willensdorfer A, Schrom S, Singer J, et al. Epicutaneously applied Der p 2 induces a strong TH 2-biased antibody response in C57BL/6 mice, independent of functional TLR4. Allergy 2014;69:741-51.
 PUBMED | CROSSREF
- Walker MT, Green JE, Ferrie RP, Queener AM, Kaplan MH, Cook-Mills JM. Mechanism for initiation of food allergy: dependence on skin barrier mutations and environmental allergen costimulation. J Allergy Clin Immunol 2018;141:1711-1725.e9.
 PUBMED | CROSSREF
- Horimukai K, Morita K, Narita M, Kondo M, Kitazawa H, Nozaki M, et al. Application of moisturizer to neonates prevents development of atopic dermatitis. J Allergy Clin Immunol 2014;134:824-830.e6.
 PUBMED | CROSSREF
- Simpson EL, Chalmers JR, Hanifin JM, Thomas KS, Cork MJ, McLean WH, et al. Emollient enhancement of the skin barrier from birth offers effective atopic dermatitis prevention. J Allergy Clin Immunol 2014;134:818-23.
 PUBMED | CROSSREF
- Glatz M, Jo JH, Kennedy EA, Polley EC, Segre JA, Simpson EL, et al. Emollient use alters skin barrier and microbes in infants at risk for developing atopic dermatitis. PLoS One 2018;13:e0192443.
 PUBMED | CROSSREF



- Lowe AJ, Su JC, Allen KJ, Abramson MJ, Cranswick N, Robertson CF, et al. A randomized trial of a barrier lipid replacement strategy for the prevention of atopic dermatitis and allergic sensitization: the PEBBLES pilot study. Br J Dermatol 2018;178:e19-21.
 PUBMED | CROSSREF
- 62. Chalmers JR, Haines RH, Mitchell EJ, Thomas KS, Brown SJ, Ridd M, et al. Effectiveness and costeffectiveness of daily all-over-body application of emollient during the first year of life for preventing atopic eczema in high-risk children (The BEEP trial): protocol for a randomised controlled trial. Trials 2017;18:343.
 PUBMED | CROSSREF
- Lowe AJ, Leung DY, Tang ML, Su JC, Allen KJ. The skin as a target for prevention of the atopic march. Ann Allergy Asthma Immunol 2018;120:145-51.
 PUBMED | CROSSREF
- 64. Fiocchi A, Pawankar R, Cuello-Garcia C, Ahn K, Al-Hammadi S, Agarwal A, et al. World Allergy Organization-McMaster University Guidelines for Allergic Disease Prevention (GLAD-P): probiotics. World Allergy Organ J 2015;8:4.
 PUBMED | CROSSREF
- 65. Ohya Y. Prevention of Allergy via Cutaneous Intervention (PACI) study. Trial registration ID: UMIN000028043 [Internet]. Tokyo: National Center for Child Health and Development; 2017 [cited 2018 Jan 7] Available from: http://paci-study.jp/english.html.
- 66. Du Toit G, Roberts G, Sayre PH, Bahnson HT, Radulovic S, Santos AF, et al. Randomized trial of peanut consumption in infants at risk for peanut allergy. N Engl J Med 2015;372:803-13.
 PUBMED | CROSSREF
- Natsume O, Kabashima S, Nakazato J, Yamamoto-Hanada K, Narita M, Kondo M, et al. Two-step egg introduction for prevention of egg allergy in high-risk infants with eczema (PETIT): a randomised, double-blind, placebo-controlled trial. Lancet 2017;389:276-86.
- Beasley R, Semprini A, Mitchell EA. Risk factors for asthma: is prevention possible? Lancet 2015;386:1075-85.
 PUBMED | CROSSREF
- 69. Tsilochristou OA, Douladiris N, Makris M, Papadopoulos NG. Pediatric allergic rhinitis and asthma: can the march be halted? Paediatr Drugs 2013;15:431-40.
 PUBMED | CROSSREF