



# Article Carbonic Anhydrase IX as a Marker of Disease Severity in Obstructive Sleep Apnea

Ayşegül Altıntop Geçkil <sup>1,\*</sup>, Tuğba Raika Kıran <sup>2</sup>, Nurcan Kırıcı Berber <sup>1</sup>, Önder Otlu <sup>2</sup>, Mehmet Erdem <sup>2</sup> and Erdal İn <sup>1</sup>

- <sup>1</sup> Department of Chest Diseases, Malatya Turgut Özal University, Malatya 44210, Turkey
- <sup>2</sup> Department of Biochemistry, Malatya Turgut Özal University, Malatya 44210, Turkey

Correspondence: aysegul.altintop@gmail.com; Tel.: +90-042-2502-8001

Abstract: Background and Objectives: Carbonic anhydrase (CA) enzymes are a family of metalloenzymes that contain a zinc ion in their active sites. CA enzymes have been implied in important situations such as CO<sub>2</sub> transport, pH regulation, and oncogenesis. CA-IX is a transmembrane glycoprotein and stimulates the expression of hypoxia-inducible factor-1 (HIF-1) CA-IX. This study aimed to determine serum CA-IX levels in OSA patients in whom intermittent hypoxia is important and to investigate the relationship between serum CA-IX levels and disease severity. Materials and Methods: The study included 88 people who applied to Malatya Turgut Özal University Training and Research Hospital Sleep Disorders Center without a history of respiratory disease, malignancy, and smoking. Patients were divided into three groups: control (AHI < 5, n = 31), mild–moderate OSA (AHI = 5–30, n = 27) and severe OSA (AHI > 30, n = 30). The analysis of the data included in the research was carried out with the SPSS (IBM Statistics 25, NY, USA). The Shapiro-Wilk Test was used to check whether the data included in the study had a normal distribution. Comparisons were made with ANOVA in multivariate groups and the t-test in bivariate groups. ANCOVA was applied to determine the effect of the CA-IX parameter for OSA by controlling the effect of independent variables. The differentiation in CA-IX and OSA groups was analyzed regardless of BMI, age, gender, and laboratory variables. ROC analysis was applied to determine the parameter cut-off point. Sensitivity, specificity, and cut-off were calculated, and the area under the curve (AUC) value was calculated. *Results:* Serum CA-IX levels were  $126.3 \pm 24.5$  pg/mL in the control group,  $184.6 \pm 59.1$  pg/mL in the mild–moderate OSA group, and  $332.0 \pm 39.7$  pg/mL in the severe OSA group. Serum CA-IX levels were found to be higher in the severe OSA group compared to the mild-moderate OSA group and control group and higher in the mild–moderate OSA group compared to the control group (p < 0.001, p < 0.001, p < 0.001, respectively). In addition, a negative correlation between CA-IX and minimum  $SaO_2$  and mean  $SaO_2$  (r = -0.371, p = 0.004; r = -0.319, p = 0.017, respectively). A positive correlation between CA-IX and desaturation index (CT90) was found (r = 0.369, p = 0.005). A positive correlation was found between CA-IX and CRP (r = 0.340, p = 0.010). When evaluated by ROC curve analysis, the area under the curve (AUC) value was determined as 0.940 (95% CI 0.322–0.557; p < 0.001). When the cut-off value for CA-IX was taken as 254.5 pg/mL, it was found to have 96.7% sensitivity and 94.8% specificity in demonstrating severe OSA. Conclusions: Our study found that serum CA-IX value was higher in OSA patients than in control patients, and this elevation was associated with hypoxemia and inflammation. CA-IX value can be a fast, precise, and useful biomarker to predict OSA.

Keywords: CA-IX; intermittent hypoxia; OSA

# 1. Introduction

Obstructive sleep apnea (OSA) is a condition characterized by the narrowing of the upper airway during sleep [1]. In OSA, the reduction in airflow in the airway may be partial (hypopnea) or complete (apnea). As a result of all these events, hypoxemia and acidosis occur at the tissue level due to pathophysiological hypoxia–ischemia, expressed



Citation: Geçkil, A.A.; Kıran, T.R.; Berber, N.K.; Otlu, Ö.; Erdem, M.; İn, E. Carbonic Anhydrase IX as a Marker of Disease Severity in Obstructive Sleep Apnea. *Medicina* 2022, *58*, 1643. https://doi.org/ 10.3390/medicina58111643

Academic Editor: Salim Surani

Received: 10 October 2022 Accepted: 28 October 2022 Published: 14 November 2022

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**Copyright:** © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). as "asphyxia". Intermittent hypoxia is very important in the pathophysiology of OSA [2]. Transcription factors such as hypoxia-inducible factor-1 (HIF-1) are very important in OSA pathology [3,4]. It is known that inflammation occurs in the upper respiratory tract in OSA. It is known that various cytokines and acute phase proteins that show systemic inflammation, especially in advanced stages, increase in OSA, and inflammation increases as hypoxemia increases [5].

OSA affects 5–15% of the general population, and its prevalence is increasing day by day [2]. Only about 10% of individuals with OSA are diagnosed and treated. The low rate causes increased complications related to OSA [6,7]. Polysomnography is used as the gold standard for the diagnosis and classification of OSA. Today, there is a need for biomarkers to help determine OSA [8].

Carbonic anhydrase (CA) enzymes are a metalloenzyme containing a zinc ion in their active sites. It catalyzes the reactions of hydration of carbon dioxide (CO<sub>2</sub>) and dehydration of bicarbonate ( $HCO_3^-$ ) alternately. CA enzyme has important physiological roles such as regulating intra and extracellular CO<sub>2</sub>, H<sup>+</sup>, and HCO<sub>3</sub><sup>-</sup> concentrations, CO<sub>2</sub> transport, ion release, pH regulation, calcification, and oncogenesis [9,10]. CA has 16 isoenzymes. These isoforms may differ in catalytic activity, tissue distribution, and subcellular localization. CA has cytosolic, mitochondrial, secretory, and membrane-bound isoforms [11]. CA activity and the membrane-bound enzymes CA-IX and CA-XII increase under environmental stress conditions such as a high degree of hypoxia [12].CA-IX is a transmembrane glycoprotein and stimulates the expression of HIF-1 CA-IX [13].

Previous studies have reported that CA-IX has an important role in cancer angiogenesis, prognosis, and metastasis, and its serum levels increase in various cancer types [14]. CA activity in OSA has been investigated in treatment studies [15]. This study aimed to evaluate the relationship between CA-IX and OSA severity by examining the levels of CA-IX associated with intermittent hypoxia in patients with OSA, and to reveal the contribution of CA-IX to the pathogenesis of OSA.

#### 2. Materials and Methods

## 2.1. Design of the Study and the Subjects

A total of 88 people, who applied to Malatya Turgut Özal University Training and Research Hospital Sleep Disorders Center between June and July 2022, had no respiratory disease, no history of malignancy, no smoking history, and who did not receive OSA treatment were included in the study. All patients underwent polysomnographic evaluation. Patients with OSA as a result of polysomnographic evaluation were included in the study group, and others were included in the control group. Clinical and polysomnographic measurements of the study participants were recorded. Hemoglobin, hematocrit, leukocyte count, platelet count, mean erythrocyte volume (MCV), and mean platelet volume (MPV) values of the patients' hemogram blood parameters (SYSMEX, Automated Hematology, JAPAN) were recorded. Alanine aminotransferase (ALT), acetyl aminotransferase (AST), low-density lipoprotein (LDL), high-density lipoprotein (HDL), triglyceride, thyroidstimulating hormone (TSH), thyrotoxin (T4), triiodothyronine (T3) and C-reactive protein (CRP) values of the patients' biochemistry blood parameters (ARCHITECT, Toshiba, Abbott Park, IL, USA) were recorded. Blood samples were taken from patients with OSA after completing the anamnesis and physical examination.

#### 2.2. Polysomnographic Evaluation

Patients were evaluated using a 55-channel polysomnography (Alice 6<sup>®</sup> Sleepware, Philips Respironics, PA, USA) system. Polysomnography recordings were analyzed according to the American Academy of Sleep Medicine (AASM) criteria version 2.6 [6]. Apnea was defined as a drop in peak thermal sensor excursion by  $\geq$ 90% of baseline for  $\geq$ 10 s. Hypopnea was defined as a drop in the nasal pressure signal by  $\geq$ 30% of baseline for  $\geq$ 10 s, causing a  $\geq$ 3% decrease in the oxygen saturation (SaO<sub>2</sub>) pre-event baseline or arousal. The desaturation index (CT90) was defined as the number of oxygen desaturations per hour during sleep. The minimum SaO<sub>2</sub> was defined as the lowest SaO<sub>2</sub> value recorded during the night. Mean SaO<sub>2</sub> was defined as the average SaO<sub>2</sub> value recorded during the night. The apnea–hypopnea index (AHI) was defined as the mean number of apneic and hypopneic episodes per hour of sleep. An obstructive AHI > 5/h was considered diagnostic of OSA. The patients were categorized into mild (AHI, 5–15), moderate (AHI, 15–30), and severe OSA (AHI, >30) groups. Patients with AHI < 5 were considered as the control group.

# 2.3. Determination of CA-IX Levels in Serum Samples

Venous blood samples taken from the cubital vein for the study were centrifuged at 2000 RPM and stored at -80 °C. The CA-IX value was analyzed according to kit procedures using the commercial enzyme-linked immune assay [Enzyme-Linked Immuno Sorbent Assay (ELISA)] kit (Cloud-Clone Corp., Cat. No: SED076Hu, Wuhan, China). The absorbance of the samples was determined with a microplate reader adjusted to 450 nm wavelength. The measurement range was 7.81–500 pg/mL, while the minimum measurable level was <2.9 pg/mL.

## 2.4. Statistical Analysis

The analysis of the data included in the research was carried out with the SPSS (IBM Statistics 25, New York, NY, USA). The Shapiro–Wilk Test was used to check whether the data included in the study had a normal distribution. Comparisons were made with ANOVA in multivariate groups and the t-test in bivariate groups. ANCOVA was applied to determine the effect of the CA-IX parameter for OSA by controlling the effect of independent variables. The differentiation in CA-IX and OSA groups was analyzed regardless of BMI, age, gender, and laboratory variables. ROC analysis was applied to determine the parameter cut-off point. Sensitivity, specificity, and cut-off were calculated, and the area under the curve (AUC) value was calculated.

## 3. Results

#### 3.1. Basic Demographic and Polysomnographic Data

Patients were divided into three groups: control (AHI < 5, n = 31), mild–moderate OSA (AHI = 5–30, n = 27), and severe OSA (AHI > 30, n = 30). Thirty-five (39.8%) of the patients were female, and 53 (60.2%) were male, and there was a significant difference between the groups in terms of gender (p < 0.001). The mean age of the patients was 46.38 ± 11.79 years, and there was a significant difference between the groups in terms of age (p = 0.03). The mean BMI of the patients was 28.75 ± 5.79, and there was a significant difference between the groups in terms of BMI (p < 0.001). While there was a history of hypertension in 7 people in the mild–moderate OSA group and 17 people in the severe OSA group, there was no hypertension patient in the control group. The patients did not have a regular history of antihypertensive drug use.

The mean AHI value in the polysomnography evaluation was  $41.03 \pm 24.91$ . The mean SaO<sub>2</sub> value of the patients was  $90.58 \pm 3.16$ , and there was a significant difference between the groups (p = 0.002). The mean minimum SaO<sub>2</sub> was  $77.14 \pm 11.67$ , and there was a significant difference between the groups (p < 0.001). The mean CT90 was  $25.15 \pm 29.80$ , and there was a significant difference between the groups (p < 0.001). The mean CT90 loss  $25.15 \pm 29.80$ , and there was a significant difference between the groups (p < 0.001).

Variable	OSA Group	Mean	Std. Deviation	F	р	Difference
Age (year)	Control <sup>1</sup>	41.06	9.40			
	Mild-moderate <sup>2</sup>	47.78	11.62	8.577	< 0.001	1 and 3
	Severe <sup>3</sup>	53.57	13.98			
	Control <sup>1</sup>	24.63	2.33			
BMI (kg/m <sup>2)</sup>	Mild-moderate <sup>2</sup>	29.41	5.51	20.634	< 0.001	1-2, 1-3, 2-3
	Severe <sup>3</sup>	32.53	5.92			
AHI (/h)	Control <sup>1</sup>	3.52	3.24			
	Mild-moderate <sup>2</sup>	19.71	7.22	58.576	< 0.001	1-2, 1-3, 2-3
	Severe <sup>3</sup>	60.70	18.52			
Minimum SPO <sub>2</sub> (%)	Control <sup>1</sup>	92.00	5.65			
	Mild-moderate <sup>2</sup>	83.33	3.88	10.118	< 0.001	1-2, 1-3, 2-3
	Severe <sup>3</sup>	71.20	13.57			
Mean SPO <sub>2</sub> (%)	Control <sup>1</sup>	93.00	6.52			
	Mild-moderate <sup>2</sup>	92.00	1.41	7.222	0.002	1-2, 1-3, 2-3
	Severe <sup>3</sup>	89.14	3.75			
CT90 (/h)	Control <sup>1</sup>	0.30	0.24			
	Mild-moderate <sup>2</sup>	9.81	15.60	10.189	< 0.001	1-2, 1-3, 2-3
	Severe <sup>3</sup>	40.30	32.60			

Table 1. Evaluation of basic demographic and polysomnographic data.

ANOVA Test value, p < 0.05; There is a difference between groups, <sup>1</sup> AHI < 5; <sup>2</sup> AHI, 5–30; <sup>3</sup> AHI > 30.

## 3.2. Evaluation of Laboratory Levels and CA-IX Levels

Serum CA-IX levels were  $126.3 \pm 24.5$  pg/mL in the control group,  $184.6 \pm 59.1$  pg/mL in the mild–moderate OSA group, and  $332.0 \pm 39.7$  pg/mL in the severe OSA group. Serum CA-IX levels were significantly higher in the severe OSA group compared to the mild–moderate OSA group and the control group (p < 0.001, p < 0.001, respectively). Serum CA-IX levels were also found to be statistically significantly higher in the mild–moderate OSA group compared to the control group (p < 0.001) (Table 1, Figure 1). A significant difference was found between the control group and severe OSA regarding hemoglobin and hematocrit values, which are hemogram parameters (p = 0.05, p = 0.02). There was no difference between the groups regarding leukocyte count, platelet count, MPV, and MCV values. There was no significant difference between the control group and severe OSA in terms of LDL and triglyceride values, which are biochemistry parameters (p = 0.023, p = 0.026). A significant difference was found between the control group and severe OSA regarding CRP value (p = 0.022). There was no difference between the groups regarding AST, ALT, HDL, TSH, T3, and T4 values (Table 2). CA-IX values did not differ in OSA groups according to the presence of hypertension (p > 0.05).

When the effects of BMI, age, gender, hemoglobin, hematocrit, LDL, triglyceride, and CRP variables were fixed using ANCOVA, the CA-IX variable did not differ in the OSA groups. It was observed that 63.1% ( $\eta^2 = 0.631$ , p = 0.001) of the change in OSA groups was caused by CA-IX. When the intragroup CA-IX differences were examined, a statistically significant difference was found between the control group and severe OSA and between the mild–moderate OSA and the severe OSA group (p < 0.05). There was no significant difference between the control group and the mild–moderate OSA group (p > 0.05) (Table 3).



Figure 1. The comparison of serum CA IX levels in OSA groups.

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Variable	OSA Group	Mean	Std. Deviation	F	р	Difference
CA-IX (pg/mL)	Control <sup>1</sup>	126.58	24.93			
	Mild-moderate <sup>2</sup>	184.60	59.07	149.271	0.000 *	1-2, 1-3, 2-3
	Severe <sup>3</sup>	324.43	48.69			
	Control <sup>1</sup>	44.54	7.43			
(mg/dI)	Mild-moderate <sup>2</sup>	43.85	9.75	0.178	0.838	none
(IIIg/uL)	Severe <sup>3</sup>	45.79	12.98			
IDI	Control <sup>1</sup>	89.31	24.40			
LDL	Mild-moderate <sup>2</sup>	97.35	31.67	4.052	0.023 *	1 and 3
(ing/uL)	Severe <sup>3</sup>	119.29	34.53			
ТС	Control <sup>1</sup>	128.10	41.77			
(ma/dI)	Mild-moderate <sup>2</sup>	178.00	87.24	3.940	0.026 *	1 and 3
(mg/dL)	Severe <sup>3</sup>	232.08	128.95			
AST (U/L)	Control <sup>1</sup>	14.00	0.5			
	Mild-moderate <sup>2</sup>	19.48	5.05	1.030	0.365	none
	Severe <sup>3</sup>	18.95	5.46			
ALT (U/L)	Control <sup>1</sup>	10.50	2.12			
	Mild-moderate <sup>2</sup>	22.92	7.57	1.314	0.278	none
	Severe <sup>3</sup>	25.29	16.61			
T3 (pg/mL)	Control <sup>1</sup>	3.19	0.72			
	Mild-moderate <sup>2</sup>	3.41	0.38	0.414	0.664	none
	Severe <sup>3</sup>	3.31	0.47			
T4 (ng/mL)	Control <sup>1</sup>	1.05	0.02			
	Mild-moderate <sup>2</sup>	1.24	0.17	1.013	0.371	none
	Severe <sup>3</sup>	1.23	0.18			
TSH (mU/L)	Control <sup>1</sup>	2.30	0.84			
	Mild-moderate <sup>2</sup>	1.70	0.53	0.757	0.475	none
	Severe <sup>3</sup>	2.03	1.35			
Uamaalahir	Control <sup>1</sup>	13.78	2.02			
(a/dL)	Mild-moderate <sup>2</sup>	14.87	1.64	5.617	0.005 *	1 and 3
(g/uL)	Severe <sup>3</sup>	15.33	1.49			

Variable	OSA Group	Mean	Std. Deviation	F	p	Difference
Hematocrit (%)	Control <sup>1</sup>	40.93	5.48			
	Mild-moderate <sup>2</sup>	44.07	4.51	6.796	0.002 *	1 and 3
	Severe <sup>3</sup>	45.81	4.64			
Leukocyte 10 <sup>3</sup> /L	Control <sup>1</sup>	7.50	1.69			
	Mild-moderate <sup>2</sup>	7.33	1.58	1.559	0.217	none
	Severe <sup>3</sup>	8.15	1.85			
Thrombocyte 10 <sup>3</sup> /L	Control <sup>1</sup>	289.30	63.11			
	Mild-moderate <sup>2</sup>	251.73	67.10	2.107	0.129	none
	Severe <sup>3</sup>	280.45	73.44			
MCV (fL)	Control <sup>1</sup>	85.18	4.81			
	Mild-moderate <sup>2</sup>	85.20	4.90	0.022	0.979	none
	Severe <sup>3</sup>	85.45	5.39			
MPV (fL)	Control <sup>1</sup>	10.28	0.81			
	Mild-moderate <sup>2</sup>	10.21	0.86	0.043	0.958	none
	Severe <sup>3</sup>	10.25	0.94			
CRP (mg/dL)	Control <sup>1</sup>	0.21	0.18			
	Mild-moderate <sup>2</sup>	0.36	0.37	4.100	0.022 *	1 and 3
	Severe <sup>3</sup>	0.71	0.68			

#### Table 2. Cont.

ANOVA Test value; \* p < 0.05; There is a difference between groups, <sup>1</sup> AHI < 5, <sup>2</sup> AHI, 5–30, <sup>3</sup> AHI > 30.

Table 3. Effect of CA-IX on OSA groups when the effect of variables is standardized.

Variable	F	p	$\eta^2$
BMI	0.407	0.528	0.013
Age	0.033	0.857	0.001
Gender	0.464	0.501	0.015
LDL	0.377	0.544	0.012
TG	0.021	0.885	0.001
Hemoglobin	0.095	0.760	0.003
Hematocrit	0.001	0.977	0.001
CRP	0.033	0.857	0.001
OSA Group	25.732	0.001 *	0.632
CA IX	Control <sup>1</sup>	Mild-moderate <sup>2</sup>	Severe <sup>3</sup>
Mean $\pm$ ss	$124.99\pm22.46$	$172.28\pm14.84$	$327.81\pm16.77$
Difference		1–3, 2–3	

CA-IX, dependent variable; ss, standard deviation;  $\eta^2$ , partial Eta squared; F, ANCOVA test value; \* p < 0.05; There is a difference between groups, <sup>1</sup> AHI < 5; <sup>2</sup> AHI, 5–30; <sup>3</sup> AHI > 30.

# 3.3. Correlation Analysis

There was a negative correlation between serum CA-IX levels and minimum SaO<sub>2</sub> and mean SaO<sub>2</sub> (r = -0.371, p = 0.004; r = -0.319, p = 0.017, respectively). There was a positive correlation (r = 0.369, p = 0.005) between serum CA-IX levels and CT90. There was a positive correlation between CRP and CA-IX values (r = 0.340, p = 0.01).

# 3.4. ROC Curve Analysis

When evaluated by ROC curve analysis, the area under the curve (AUC) value was determined as 0.940 (95% CI 0.322–0.557; p < 0.001). When the cut-off value for CA-IX was taken as 254.5 pg/mL, it was found to have 96.7% sensitivity and 94.8% specificity in demonstrating severe OSA (Figure 2).



**Figure 2.** ROC curve analysis of the utility of CA IX to severe OSA. The red line is the reference and the blue line is the ROC curve.

# 4. Discussion

The results of our study conducted to determine serum CA-IX levels in OSA patients show that serum CA-IX levels are significantly higher in both mild–moderate and severe OSA patients compared to control patients. Serum CA-IX levels were negatively correlated with minimum SaO<sub>2</sub> and mean SaO<sub>2</sub> values and positively correlated with CT90. Serum CA-IX levels showed a positive correlation with the CRP value.

CA-IX, a membrane-associated CA strongly induced by hypoxia, is particularly overexpressed in some types of cancer and has been associated with metastasis and poor prognosis [16]. Tumor-associated CA-IX has two main forms. One is the form expressed in gastric mucosa and various types of cancer, and the other is CA-IX, which is released from transmembrane and intracellular arrays by cleavage with proteolytic enzymes [17]. Studies have shown that CA-IX contributes to the acidification of hypoxic tumors by reducing the extracellular pH (<7.28) [18]. Low pH has been associated with extracellular matrix degradation, tumor transformation, and tumor cell migration [19]. In renal cell carcinoma, unlike other tumors, CA-IX overexpression has been associated with a good prognosis and a good response to immunotherapy [20]. Inhibition of CA-IX release in these patients has been shown to reduce tumor proliferation and tumor resistance to conventional anti-cancer therapies [20]. It has also been found that acetazolamide, a CA inhibitor, prevents tumor invasion [16].

CA enzymes are abundant in the human body and have also been associated with cardiovascular diseases [21]. The enzymatic function of CA is essential for maintaining acid–base balance and, therefore, controlling respiration. High CA activity in OSA has been clinically investigated by therapy with drugs with CA-inhibiting properties [15]. Recent studies have shown that pharmacological blockade of the CA enzyme by acetazolamide improves both blood pressure and sleep apnea [22]. It has also been reported that using zonisamide, a CA inhibitor, reduces blood pressure in OSA patients [23]. Acetazolamid has also been used to prevent hypertension in altitude sickness [24].

When the studies on OSA in the literature were examined, Wang et al. found that patients with OSA and concomitant hypertension had higher blood CA activity than

OSA patients without hypertension. It was observed that OSA and blood pressure were controlled, and CA activity decreased by inhibition of CA with acetazolamide [25]. In another study, long-term positive airway pressure (PAP) treatment was found to improve OSA and blood pressure, but it was observed that CA activity did not change [12]. In a prospective study conducted on patients with OSA and hypertension, CPAP treatment was given to one group, and acetazolamide treatment was given to the other group in addition to CPAP treatment, and the results were compared after two weeks. At the end of the study, it was found that there was a significant improvement in AHI, CT90, mean SaO<sub>2</sub>, and minimum SaO<sub>2</sub> in patients who were given additional acetazolamide [23]. The improvement in hypoxia-related parameters can be explained by the reduction of loop gain in OSA by acetazolamide and the decrease in  $HCO_3^-$  levels following acetazolamide treatment [26].

In our study, in which we investigated CA-IX levels, a parameter associated with tissue hypoxemia in OSA patients, we found a correlation between OSA severity and CA-IX. When the cut-off value for CA-IX was taken as 254.5 pg/mL, we found that it had 96.7% sensitivity and 94.8% specificity in demonstrating severe OSA. It showed that these results were independent of hypertension and that hypertension did not affect CA-IX levels among the OSA groups. When the effects of age, gender, BMI, hemogram, and biochemistry parameters were fixed in covariance analysis, we found that the CA-IX value differed between OSA grades. We concluded that this difference between the groups was associated with hypoxemia and inflammation. The negative correlation between CA-IX levels and minimum SaO<sub>2</sub> and mean SaO<sub>2</sub> values and the positive correlation with CT90 also show the relationship with hypoxia. The positive correlation between CA-IX and CRP shows the relationship with inflammation; as hypoxemia increases in OSA, inflammation also increases.

The limitation of our study was the relatively small number of OSA and control patients and the lack of CA-IX evaluation of OSA patients after PAP treatment. CA-IX may be a promising parameter in diagnosing and treating OSA and other respiratory tract diseases in which hypoxemia is at the forefront. More extensive studies are needed to determine the importance of CA-IX and to support our findings.

## 5. Conclusions

In conclusion, this study showed that serum CA-IX levels are increased in severe OSA patients, which is associated with hypoxemia and inflammation. CA-IX can be a fast, precise, and useful biomarker to predict patients at high risk of OSA.

**Author Contributions:** Conceptualization, A.A.G. and T.R.K.; methodology, A.A.G. and T.R.K.; software, M.E. and A.A.G.; formal analysis, A.A.G., T.R.K., Ö.O. and E.İ.; resources, M.E., Ö.O. and N.K.B.; data curation, A.A.G. and N.K.B.; writing-original draft preparation, A.A.G. and M.E.; writing-review and editing, A.A.G., E.İ. and T.R.K.; supervision, E.İ. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

**Institutional Review Board Statement:** Approval was obtained from Malatya Turgut Ozal University Clinical Research Ethics Committee (Approval number: 2022/25, date: 17 June 2022).

**Informed Consent Statement:** Informed consent was obtained from all participants included in the study.

**Data Availability Statement:** The datasets are not publicly available but are available from the corresponding author upon reasonable request.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

- Olson, L.G.; King, M.T.; Hensley, M.J.; Saunders, N.A. A community study of snoring and sleep-disordered breathing prevalence. *Am. J. Respir. Crit. Care Med.* 1995, 152, 711–716. [CrossRef] [PubMed]
- Lévy, P.; Kohler, M.; McNicholas, W.T.; Barbé, F.; McEvoy, R.D.; Somers, V.K.; Lavie, L.; Pépin, J.L. Obstructive sleep apnoea syndrome. *Nat. Rev. Dis. Prim.* 2015, 1, 1–21. [CrossRef] [PubMed]
- Arnaud, C.; Bochaton, T.; Pépin, J.L.; Belaidi, E. Obstructive sleep apnoea and cardiovascular consequences: Pathophysiological mechanisms. Arch. Cardiovasc. Dis. 2020, 113, 350–358. [CrossRef] [PubMed]
- 4. Hunyor, I.; Cook, K.M. Models of intermittent hypoxia and obstructive sleep apnea: Molecular pathways and their contribution to cancer. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **2018**, *315*, 669–687. [CrossRef]
- 5. Cilekar, S.; Beysel, S.; Karatas, S.; Balci, A.; Akaslan, K.; Uncu, A. Circulating sTweak is associated with visceral adiposity and severity in patients with obstructive sleep apnea syndrome. *Sci. Rep.* **2021**, *11*, 22058. [CrossRef] [PubMed]
- Berry, R.B.; Quan, S.F.; Abreu, A.R.; Bibbs, M.L.; DelRosso, L.; Harding, S.M.; Mao, M.M.; Plante, D.T.; Presman, M.R.; Troester, M.M.; et al. *The AASM Manual for the Scoring of Sleep and Associated Events: Rules, Terminology and Technical Specifications*; Version 2.6; American Academy of Sleep Medicine: Darien, IL, USA, 2020.
- Sánchez-de-la-Torre, M.; Campos-Rodriguez, F.; Barbé, F. Obstructive sleep apnoea and cardiovascular disease. *Lancet Respir.* Med. 2013, 1, 61–72. [CrossRef]
- 8. Ekin, S.; Yildiz, H.; Alp, H.H. NOX4, MDA, IMA and oxidative DNA damage: Can these parameters be used to estimate the presence and severity of OSA? *Sleep Breath.* **2021**, *25*, 529–536. [CrossRef] [PubMed]
- 9. Alver, A.; Menteşe, A.; Menteşe, Ü.; Sümer, A.; Uçar, F.; Us Altay, D. Anti-carbonic anhydrase II antibodies in end-stage renal disease patients. *Med. Princ. Pract.* 2014, 23, 331–335. [CrossRef] [PubMed]
- 10. Mentese, A.; Erkut, N.; Demir, S.; Yaman, S.O.; Sumer, A.; Erdem, M.; Alver, A.; Sönmez, M.G. Serum carbonic anhydrase I and II autoantibodies in patients with chronic lymphocytic leukaemia. *Cent. Eur. J. Immunol.* **2018**, *43*, 276–280. [CrossRef] [PubMed]
- Lucarini, L.; Magnelli, L.; Schiavone, N.; Crisci, A.; Innocenti, A.; Puccetti, L.; Cianchi, F.; Peri, S.; Supuran, C.T.; Papucci, L.; et al. Plasmatic carbonic anhydrase IX as a diagnostic marker for clear cell renal cell carcinoma. *J. Enzyme Inhib. Med. Chem.* 2018, 33, 234–240. [CrossRef] [PubMed]
- 12. Hoff, E.; Zou, D.; Schiza, S.; Demir, Y.; Grote, L.; Bouloukaki, I.; Beydemir, Ş.; Eskandari, D.; Stenlöf, K.; Hedner, J. Carbonic anhydrase, obstructive sleep apnea and hypertension: Effects of intervention. J. Sleep Res. 2020, 29, e12956. [CrossRef] [PubMed]
- Mahon, B.P.; Pinard, M.A.; McKenna, R. Targeting carbonic anhydrase IX activity and expression. *Molecules* 2015, 20, 2323–2348. [CrossRef]
- Loncaster, J.A.; Harris, A.L.; Davidson, S.E.; Logue, J.P.; Hunter, R.D.; Wycoff, C.C.; Pastorek, J.; Ratceliffe, P.C.; Stratford, I.C.; West, C.M. Carbonic anhydrase (CA IX) expression, a potential new intrinsic marker of hypoxia: Correlations with tumor oxygen measurements and prognosis in locally advanced carcinoma of the cervix. *Cancer Res.* 2001, *61*, 6394–6399. [PubMed]
- Hedner, J.; Stenlöf, K.; Zou, D.; Hoff, E.; Hansen, C.; Kuhn, K.; Lennartz, P.; Grote, L. A randomized controlled clinical trial exploring safety and tolerability of sulthiame in sleep apnea. *Am. J. Respir. Crit. Care Med.* 2022, 205, 1461–1469. [CrossRef] [PubMed]
- 16. Robertson, N.; Potter, C.; Harris, A.L. Role of carbonic anhydrase IX in human tumor cell growth, survival, and invasion. *Cancer Res.* **2004**, *64*, 6160–6165. [CrossRef]
- 17. Kaluz, S.; Kaluzova, M.; Stanbridge, E.J. Expression of the hypoxia marker carbonic anhydrase IX is critically dependent on SP1 activity. Identification of a novel type of hypoxia-responsive enhancer. *Cancer Res.* **2003**, *63*, 917–922. [PubMed]
- Dubois, L.; Peeters, S.; Lieuwes, N.G.; Geusens, N.; Thiry, A.; Wigfield, S.; Carta, F.; Mcintyre, A.; Scozzafava, A.; Dogné, J.M.; et al. Specific inhibition of carbonic anhydrase IX activity enhances the in vivo therapeutic effect of tumor irradiation. *Radiother. Oncol.* 2011, *99*, 424–431. [CrossRef]
- 19. Gatenby, R.A.; Gillies, R.J. Why do cancers have high aerobic glycolysis? Nat. Rev. Cancer 2009, 4, 891–899. [CrossRef] [PubMed]
- 20. Sandlund, J.; Oosterwijk, E.; Grankvist, K.; Oosterwijk-Wakka, J.; Ljungberg, B.; Rasmuson, T. Prognostic impact of carbonic anhydrase IX expression in human renal cell carcinoma. *BJU Int.* **2007**, *100*, 556–560. [CrossRef] [PubMed]
- Aamand, R.; Dalsgaard, T.; Jensen, F.B.; Simonsen, U.; Roepstorff, A.; Fago, A. Generation of nitric oxide from nitrite by carbonic anhydrase: A possible link between metabolic activity and vasodilation. *Am. J. Physiol. Heart Circ. Physiol.* 2009, 297, 2068–2074. [CrossRef] [PubMed]
- Eskandari, D.; Zou, D.; Grote, L.; Hoff, E.; Hedner, J. Acetazolamide reduces blood pressure and sleep-disordered breathing in patients with hypertension and obstructive sleep apnea: A randomized controlled trial. *J. Clin. Sleep Med.* 2018, 14, 309–317. [CrossRef] [PubMed]
- 23. Eskandari, D.; Zou, D.; Karimi, M.; Stenlof, K.; Grote, L.; Hedner, J. Zonisamide reduces obstructive sleep apnoea: A randomised placebo-controlled study. *Eur. Respir. J.* 2014, 441, 140–149. [CrossRef]
- Estrada, V.H.N.; Franco, D.M.; Medina, R.D.; Garay, A.G.G.; Martí-Carvajal, A.J.; Arevalo-Rodriguez, I. Interventions for preventing high altitude illness: Part 1. Commonly-used classes of drugs. *Cochrane Database Syst. Rev.* 2017, 6, CD009761.

- 25. Wang, T.; Eskandari, D.; Zou, D.; Grote, L.; Hedner, J. Increased carbonic anhydrase activity is associated with sleep apnea severity and related hypoxemia. *Sleep* **2015**, *38*, 1067–1073. [CrossRef] [PubMed]
- Edwards, B.A.; Sands, S.A.; Eckert, D.J.; White, D.P.; Butler, J.P.; Owens, R.L.; Malhotra, A.; Wellman, A. Acetazolamide improves loop gain but not the other physiological traits causing obstructive sleep apnoea. *J. Physiol.* 2012, 590, 1199–1211. [CrossRef] [PubMed]