Filgrastim fails to improve haemopoietic reconstitution following myeloablative chemotherapy and peripheral blood stem cell rescue

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Summary The morbidity of high-dose chemotherapy has been considerably reduced by the use of autologous peripheral blood progenitor cell reinfusion. Most studies have used myeloid colony-stimulating factors after stem cell reinfusion, making it difficult to determine the relative contribution of each of these variables to the early recovery of blood cells. The financial implications of colony-stimulating factor use are an area of concern as dose intensification in chemosensitive malignancies is increasingly employed. We have studied 19 consecutive patients receiving high-dose chemotherapy with and without filgrastim (Amgen, granulocyte colony-stimulating factor, G-CSF) after stem cell infusion to examine its effect on the kinetics of blood cell recovery, the complications of myelosuppression and the associated costs. Analysis of the two treatment groups reveals that administration of filgrastim $10 \,\mu g \, kg^{-1} \, day^{-1}$ following stem cell reinfusion does not further accelerate haemopoietic recovery, fails to reduce the incidence of neutropenic fever or antibiotic usage and significantly increases the cost of the procedure. The results of this study do not support the routine use of filgrastim after high-dose chemotherapy and peripheral blood stem cell reinfusion.

The reinfusion of peripheral blood stem cells (PBSCs) following high-dose chemotherapy to accelerate haemopoietic reconstitution is now standard practice in many centres (Gianni *et al.*, 1990; Kessinger & Armitage, 1991; Sheridan *et al.*, 1992). Several groups have reported reduced durations of thrombocytopenia with PBSC rescue compared with autologous bone marrow transplantation. There have, however, been some reports of patients with acute myeloid leukaemia (AML) receiving purged bone marrow having better diseasefree survival than those receiving peripheral stem cell autografts (Korbling *et al.*, 1991). It is important to administer sufficient stem cells for engraftment and most centres now recognise a threshold number of reinfused colony-forming unit-granulocyte/macrophage (CFU-GM) cells kg^{-1} to ensure haemopoietic reconstitution (To *et al.*, 1986; Reiffers

et al., 1988). Myeloid colony-stimulating factors (CSFs), alone or in combination with chemotherapy, are extensively used to aid harvesting of adequate numbers of PBSCs (Duhrsen et al., 1988; Siena et al., 1989; Socinski et al., 1988). Although most centres continue CSF administration after PBSC infusion, there remains considerable doubt about the value of myeloid CSFs in this setting in terms of further accelerating haemopoietic recovery and thus reducing the morbidity and mortality of the procedure. We have recently audited a group of 19 consecutive patients undergoing highdose chemotherapy between 1991 and 1993 and retrospectively compared the clinical characteristics of patients who received filgrastim (G-CSF, Amgen) after PBSC reinfusion with those who did not.

Table I Characteristics of 19 patients receiving high-dose chemotherapy and peripheral blood stem cell reinfusion

Patient	Age	Sex	Diagnosis	Conditioning regimen	Bone marrow reinfusion	CD34 × 10 ⁶ kg ⁻¹ reinfused	CFU-GM × 10 ^s kg ⁻ reinfused
Filgrast	im afte	er PBS	SC reinfusion				
1	22	Μ	Hodgkin's	EAM	Yes	2.5	3.8
2	38	F	Non-Hodgkin's	EAM	Yes	2.7	4.0
3	23	Μ	Non-Hodgkin's	TBI/CTX	Yes	4.3	3.0
4	23	Μ	Non-Hodgkin's	TBI/CTX	Yes	5.0	5.9
5	33	F	Non-Hodgkin's	TBI/CTX	Yes	2.6	2.3
6	39	F	Teratoma	CEC	Yes	6.5	6.6
7	31	Μ	Teratoma	CEC	Yes	6.7	Contaminated
8	29	Μ	Teratoma	CEC	Yes	5.1	3.2
9	44	Μ	Non-Hodgkin's	BEAM	Yes	10.0	10.0
10	16	Μ	Sarcoma	TBI/M	No	9.1	10.4
No filgr	astim d	after P	BSC reinfusion				
11	31	M	Non-Hodgkin's	TBI/CTX	No	2.4	3.1
12	53	Μ	Non-Hodgkin's	TBI/CTX	No	2.4	8.0
13	44	Μ	Non-Hodgkin's	TBI/CTX	No	6.2	9.0
14	62	F	Non-Hodgkin's	EAM	No	10.1	13.1
15	27	Μ	Teratoma	CEC	No	9.9	20.0
16	45	Μ	Non-Hodgkin's	EAM	Yes	26.7	17.0
17	39	F	Non-Hodgkin's	EAM	No	8.1	17.0
18	52	F	Non-Hodgkin's	EAM	No	11.3	13.0
19	29	Μ	Teratoma	CEC	No	6.4	20.9

Conditioning regimens: BEAM, BCNU 300 mg m⁻², etoposide 1,200 mg m⁻², cytosine arabinoside 800 mg m⁻², melphalan 140 mg m⁻²; EAM, etoposide 1,200 mg m⁻², cytosine arabinoside 800 mg m⁻², melphalan 140 mg m⁻²; CEC, carboplatin AUC × 20, etoposide 1,600 mg m⁻², cyclophosphamide 60 mg kg⁻¹. TBI/CTX, total body irradiation 14.4 Gy, cyclosphosphamide 3.6 g m⁻²; TBI/M, total body irradiation 14.4 Gy, melphalan 140 mg m⁻².

Patients and methods

The patients selected for high-dose consolidation chemotherapy had testicular teratoma (5), non-Hodgkin and Hodgkin's lymphoma (12), myeloma (1) and sarcoma (1), and had received conventional dose induction chemotherapy prior to high-dose chemotherapy. Patient details are listed in Table I. There were 14 males and five females, who ranged in age from 16 to 62 years. A number of patients also received autologous bone marrow reinfusion (ABMR). Different conditioning regimens were used as appropriate for different tumour types. PBSC harvests were carried out on a Cobe Spectra following patient priming with conventional chemotherapy and filgrastim $5 \mu g k g^{-1} da y^{-1}$ subcutaneously from the time of the white cell nadir for 5 days. On the 2 days of leucopheresis they received filgrastim 10 µg kg⁻¹ day⁻¹. Following myeloablative chemotherapy all patients received PBSC reinfusion containing haemopoietic progenitors in excess of our recognised threshold values for marrow rescue $(2 \times 10^5 \text{ GFU-GM cells } \text{kg}^{-1} \text{ and } 1 \times 10^6 \text{ CD34}^+ \text{ cells } \text{kg}^{-1})$ (Table I). Ten patients received filgrastim $10 \,\mu\text{g kg}^{-1} \,\text{day}^{-1}$ subcutaneously or intravenously from day 5 following stem cell reinfusion until the neutrophil count was $> 1.0 \times 9^{-1}$.

End point analysis included days of neutrophils $<0.5 \times 10^{9} \, l^{-1}$, days of neutrophils $<1.0 \times 10^{9} \, l^{-1}$, days of platelets $<15 \times 10^{9} \, l^{-1}$, platelet transfusions, febrile days (temperature $<38^{\circ}$ C on two separate occasions in a 24 h period), days on intravenous antibiotics, cost of intravenous antibiotics, number of in-patient days and overall cost of the procedure. Statistical methods applied to the results included the Mann–Whitney U-test to compare median parameters in each group and calculation of the Spearman rank correlation coefficient.

Results

The initial ten patients received autologous bone marrow in addition to PBSC, but as confidence in PBSC rescue increased bone marrow harvests were discontinued. The addition of autologous bone marrow reinfusion to PBSC did not lead to more rapid haemopoietic reconstitution than PBSC reinfusion alone (95% CI for the difference between median number of days to engraftment -3.0 to 9.0 days, Mann-Whitney U-test, P-value 0.285). No correlation between dose of CFU-GM cells kg^{-1} and rate of engraftment was estab-lished (Spearman rank correlation coefficient -0.208). Ten patients (mean age 29.8 years) received filgrastim 10 µg kg⁻¹ day⁻¹ s.c. or i.v. daily from day 5 after progenitor cell reinfusion until peripheral blood neutrophil counts were $> 1.0 \times 10^9 l^{-1}$. Nine patients (mean age 42.2 years) received peripheral blood stem cell reinfusion alone (Table I). Patients in the cohort receiving filgrastim were on average reinfused fewer CFU-GM cells kg^{-1} (5.46 × 10⁵ cells kg^{-1} vs 13.4 × 10⁵ cells kg^{-1} reinfused, 95% CI for difference between median number of CFU-GM kg⁻¹ infused -3.00 to -13.5, Mann-Whitney U-test, P-value 0.0062). There was no significant difference between mean numbers of CD34⁺ cells kg^{-1} reinfused in each treatment group (9.32 ± 2.1 × 10⁶ cells kg⁻¹ vs 9.27 ± 7.2 × 10⁶ cells kg⁻¹). There were no significant differences between the two treatment groups in terms of days of neutropenia, days of thrombocytopenia, febrile days or days on antibiotics (Table II). On average, the patients who received filgrastim stayed in hospital longer than those who did not, but this difference did not reach statistical significance (mean 20.6 days vs 17.1 days, 95% CI for difference between median number of in-patient days -3.00 to 9.00, Mann-Whitney U-test, P-value 0.58). The administration of filgrastim approximately doubled the average overall cost of the procedure in those patients receiving filgrastim following PBSC transfusion.

Table II	Data of 19 patients receiving high-dose chemotherapy and peripheral blood stem cell reinfusion; details of morbidity and engraftment. All parameters are expressed as	
	II Da	

	Days to ree	Days to recover after PBSC transfusion	transfusion						
	ANC ^a <0.5 × 10°/1-1	$4NC^{\circ}$ $4NC^{\circ}$ $4NC$ $<0.5 \times 10^{\circ}/1^{-1}$ $<1.0 \times 10^{\circ}/1^{-1}$	$Platelets = 10^{\circ} 1^{-1}$	Platelets Units of platelets No. of days <15 × 10° t ⁻¹ transfused febrile	No. of days febrile	No. of days on i.v. Abs	In-patient days	Cost of i.v. Abs (£)	No. of days on In-patient days Cost of i.v. Abs Cost of procedure i.v. Abs (E)
+ Filgrastim	11.2 (8 16)	11.2 (8 16) 13.7 (10-24)	21.9 (10-42)	45 (15-20)	6.4 (2-12)	13.7 (9–25)	13.7 (9-25) 20.6 (12-36)	1798.60	7817.30
– Filgrastim	12.9 (6 20)	12.9 (6 20) 14.6 (7-23)	16.8 (2 - 45)	29 (5-80)	3.5 (1–9)	10.3 (4 20)	17.1 (12–25)	(530 -6,615) 799.25	(4,299 – 16,815) 3449.89
								(151–1,390)	(2,551–6,175)
*ANC (absolu stay × £200). Th methods section	*ANC (absolute neutrophil count per 10° cells 1^{-1} stay × £200). The cost of using filgrastim after PBSC methods section. In our centre 600 µg of filgrastim	t per 10° cells l ⁻¹). astim after PBSC ¹ µg of filgrastim c). ^b Cost of proced creinfusion was ba costs £172.00.	*ANC (absolute neutrophil count per 10° cells 1 ⁻¹). ^b Cost of procedure based on cost of conditioning drugs + antibiotics + cost of filgrastim + cost of in-patient stay (Days of stay × £200). The cost of using filgrastim after PBSC reinfusion was based on the administration of 600 µg per patient per day for the number of days specified in the Patients and methods section. In our centre 600 µg of filgrastim costs £172.00.	f conditioning (ration of 600 µg	drugs + antibiotics I per patient per da	+ cost of filgrastin y for the number	m + cost of in-pat of days specified i	ient stay (Days of n the Patients and

Discussion

Although this is a small study, the results suggest that the administration of filgrastim after PBSC transfusion fails to further accelerate haemopoietic reconstitution, reduce the morbidity of the procedure or reduce the duration of hospitalisation. Importantly, its use significantly increases the cost of the procedure. The failure of filgrastim to significantly accelerate neutrophil recovery contradicts the results of Spitzer et al. (1993), who studied a similar number of patients but found that a combination of G-CSF and GM-CSF encouraged earlier neutrophil recovery. However, the authors found that administration of growth factors after PBSC infusion made no difference to the duration of hospital stay or fever, results that concur with the findings in our study. The authors make no comment on the use of antibiotics or cost of the procedure. So far there have been no published randomised studies comparing haemopoietic reconstitution following high-dose chemotherapy and PBSC rescue, with and without myeloid growth factors. Although a relatively small number of patients were included in our analysis, important conclusions can be drawn. The failure of filgrastim to further accelerate myeloid reconstitution after high-dose chemotherapy and PBSC reinfusion resulted in similar morbidity, number of febrile days and antibiotic use

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in the two groups of patients (Table II). Although most patients receiving filgrastim received fewer CFU-GM cells, all patients received numbers in excess of threshold values which produce optimal times to haemopoietic reconstitution. Our data failed to establish any correlation between numbers of CFU-GM cells reinfused above this level and the rate of subsequent haemopoietic reconstitution.

There are obvious differences between the cost of the procedure in the two treatment cohorts. The overall cost of the procedure was calculated by adding costs of antibiotics, conditioning chemotherapy and cost of filgrastim to the bed cost per day of the in-patient stay after PBSC reinfusion. The major difference in cost of the procedure was due to the administration of filgrastim, which doubled procedural expense in the group receiving filgrastim after PBSC reinfusion. The period of hospitalisation and intensive support after myeloablative therapy was not influenced by the administration of CSFs. This study fails to support the value of routine administration of myeloid colony-stimulating factors after such therapy and has significant financial implications for centres carrying out high-dose chemotherapy.

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