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Design, synthesis, antiviral and cytotoxic evaluation of novel acyclic phosphonate nucleotide analogues with a 5,6-dihydro-1*H*-[1,2,3]triazolo[4,5-*d*]pyridazine-4,7-dione system

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Abstract A series of diethyl 2-(4,5-dimethoxycarbonyl-1H-1,2,3-triazol-1-yl)alkylphosphonates was synthesised from ω -azidoalkylphosphonates and dimethyl acetylenedicarboxylate and was further transformed into the respective diamides, dihydrazides, and 5,6-dihydro-1H-[1,2,3]triazolo[4,5-d]pyridazine-4,7-diones as phosphonate analogues of acyclic nucleosides having nucleobases replaced with substituted 1,2,3-triazoles. All compounds containing P-C-C-triazole or P-C-C-CH₂-triazole moieties exist in single conformations in which the diethoxyphosphoryl and substituted 1,2,3-triazolyl or substituted (1,2,3-triazolyl)methyl groups are oriented anti. All phosphonates were evaluated in vitro for activity against a variety of DNA and RNA viruses. None of the compounds were endowed with antiviral activity. They were not cytostatic at 100 µM.

Keywords Cycloadditions · Cyclizations · Heterocycles · NMR spectroscopy · Conformation

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Introduction

In the past two decades acyclic nucleoside phosphonates (ANPs) have become one of the most important classes of antiviral drugs [1]. Three of them (adefovir, cidofovir, tenofovir; Fig. 1) have been marketed for treatment of viral infection caused by HIV, HBV, HSV and other DNA viruses [2–5]. The concept of acyclic nucleosides is based on the assumption that an acyclic moiety most often bearing an oxygen atom mimics the furanose ring at least partially. Acyclic nucleoside phosphonates require conversion in vivo to their triphosphate metabolites to become active [2, 6]. The replacement of the natural phosphate moiety by a phosphonate group makes analogues less susceptible to enzymatic hydrolysis [7, 8].

Further studies in this field brought a new generation of nucleotide/nucleoside analogues in which natural nucleobases were modified as exemplified by the 2,4diaminopyrimidine framework present in antiviral HPMPO-DAPy and PMPO-DAPy [9–11] and the 1,2,4triazole ring in ribavirin (Fig. 2) [12].

The antiviral activity of ribavirin stimulated interest in replacing the 1,2,4-triazole system with an isomeric 1,2,3-triazole ring because several 1,2,3-triazoles exhibit antibacterial [13–15], antifungal [15–17], anticancer [18–20], anti-inflammatory [21, 22] and antiviral [23–26] properties. It was found that carbocyclic analogues 1 and phosphonocarbocyclic analogues 2 of ribavirin displayed antiviral activity against HIV-1 (Fig. 3) [27].

Furthermore, preliminary structure–activity relationship evaluation of 1,2,3-triazole nucleoside phosphonates **3** and **4** suggested that this scaffold could be further optimised to afford selective inhibitors of HCV replication (Fig. 4) [28].

On the other hand, it was reported that nucleoside analogues 5 and 6 containing the 5,6-dihydro-1*H*-imidazo[4,5-





 $(HO)_{2}(O)P \longrightarrow O R = CH_{2}OH Ribavirin PMPO-DAPy R = CH_{3}$

Fig. 2 Structures of HPMPO-DAPy, PMPO-DAPy and ribavirin



Fig. 3 Antiviral analogues of ribavirin having the 1,2,3-triazole ring



Fig. 4 1,2,3-Triazole nucleoside phosphonates as potential inhibitors of HCV replication



Fig. 5 Cyclic nucleoside analogues based on the 5,6-dihydro-1*H*-imidazo[4,5-*d*]pyridazine-4,7-dione framework

d]pyridazine-4,7-dione ring instead of the natural purine framework exhibited antiviral activity by inhibition of a viral helicase from West Nile Virus (WNV) and HCV (Fig. 5). These observations may be useful in designing a lead structure for the development of new classes of antiviral agents [29].

Based on the active compounds already discussed, a novel series of phosphonate analogues **11** having the 5,6dihydro-1*H*-[1,2,3]triazolo[4,5-*d*]pyridazine-4,7-dione system was designed as potential antiviral agents (Scheme 1). Furthermore, because their immediate precursor dihydrazides **10** as well as diamides **9** share several common structural features with ribavirin, they also may show antiviral activity. The key step of our synthetic plan involves the 1,3-dipolar cycloaddition of dimethyl acetylenedicarboxylate and ω -azidoalkylphosphonates **7** which contain structurally diversified alkyl chains to provide the intermediate diesters **8**.

Results and discussion

Chemistry

The 1,2,3-triazoles **8a–8d** and **8f** were synthesised in 64–98 % yield employing the Huisgen 1,3-dipolar cycloaddition of the corresponding azidophosphonates **7** and dimethyl acetylenedicarboxylate at 110 °C in the same way as the known compounds **8e** and **8g** [30, 31]. They were finally purified either by chromatography on a silica gel column or by crystallisation (Scheme 1).

The required azidoalkylphosphonates 7a-7e and 7g have already been described in the literature [30, 32–36]. Azidophosphonate 7f was obtained in the Abramov reaction [37, 38] from 3-azidopropanal [39] and diethyl phosphite in 34 % yield (Scheme 2). It was found that 3-azidopropanal is unstable in the presence of triethylamine used as a catalyst in this reaction. For this reason only 0.4 equiv. of diethyl phosphite was applied to avoid tedious separation of 7f from the reaction mixture.

The diamides **9a–9g** were obtained from diesters **8a–8g** by ammonolysis according to a standard protocol (Scheme 1) [6, 40]. The crude products were subjected to column chromatography on silica gel and finally purified



Reagents and conditions: a. toluene, 110 °C, 4 h; b. aqueous ammonia, EtOH, rt, 24 h; c. hydrazine hydrate, EtOH, reflux, 2 h; d. 10% HCl, 90 °C, 2.5 h.





Fig. 6 Compounds 12a and 13a

by crystallisation to give 9a-9g in 37–61 % yields. The ¹H NMR spectra of diamides 9a-9g in chloroform-*d* confirmed the nonequivalence of protons from the carbamoyl groups because in the 6.0–10.8 ppm region four broad singlets were always observed.

The diesters **8a–8g** were also converted to the corresponding dihydrazides **10a–10g** with hydrazine hydrate (Scheme 1). Preliminary attempts at synthesising dihydrazide **10a** followed the literature procedure [41] and showed that refluxing phosphonate **8a** and hydrazine hydrate in ethanolic solution for 5 h led to the formation of several products. The ³¹P NMR spectrum of the reaction mixture revealed the presence of the expected phosphonate **10a** (76 %), bicyclic 1,2,3-triazolopyridazinedione **11a** (11 %) and their monodealkylated counterparts **12a** and **13a** (10 and 3 %, respectively) (Fig. 6). When hydrazinolysis of the diester **8a** was conducted for 2 h only phosphonates **10a** (79 %) and **11a** (21 %) were produced. For this reason syntheses of dihydrazides **10b–10g** were performed under

the same conditions. However, it appeared that for compounds **8c** and **8g** significant dealkylation still occurred. In both cases the respective crude reaction mixtures contained almost 50 % of by-products. Purifications on silica gel columns and crystallisations gave dihydrazides **10a–10g** in 28–66 % yields.

Finally, heating dihydrazides **10a–10g** with 10 % hydrochloric acid for 2.5 h gave [1,2,3]triazolo[4,5-*d*]pyridazine-4,7-diones **11a–11g** (Scheme 1) in 30–66 % yields [41].

Conformational analysis

Detailed analyses of ¹H and ¹³C NMR spectral data revealed conformational preferences of phosphonates described in this paper. Compounds **8a–11a** contain an 1,2substituted ethylene fragment which does not freely rotate around a C–C bond because their ¹H NMR spectra display AA'XX'P patterns. A similar spectrum was also noticed for 2-azidoethylphosphonate [31]. Antiperiplanar disposition of the diethoxyphosphoryl groups and substituted 1,2,3triazoles **14a** (Fig. 7) was proved by the presence of two identical ³*J*(P–H_X) = 10.5 Hz couplings which were calculated from the ¹H{³¹P} NMR spectrum of **8a**.

In the ¹H NMR spectra of compounds **8g**, **10g**, and **11g** three hydrogen atoms attached to the two-carbon linker appeared as deceptively simple but very similar ABXP spectral patterns. However, the relevant ³J(H1–H2a) and ³J(H1–H2b) coupling constants were precisely calculated (10.2 and 3.6 Hz, respectively) from the ¹H NMR spectrum of diamide **9g**. These values [42] together with small couplings for ³J(P–H2a) and ³J(P–H2b) [43, 44] allowed us to unequivocally establish **14g** (Fig. 7) as the preferred conformation of phosphonate **9g** although other phosphonates from this series very likely adopt the same *anti* conformation.

in this study



Large values (16.9–19.2 Hz) of ${}^{3}J(P-CC-C3)$ [44–46] observed in the ¹³C NMR spectra of phosphonates 8-11 containing a three-carbon fragment between the phosphorus atom and the 1,2,3-triazole ring (series **b**, **e**, and **f**) evidenced the preference of antiperiplanar conformations 14b, 14e, and 14f (Fig. 7) for these compounds. This conclusion was further supported by vicinal H1-H2 couplings calculated for 2- and 1-hydroxyphosphonates (series e and f) which clearly indicated gauche (3.0-3.6 Hz) and anti (9.6-10.8 Hz) arrangements of the respective H-C1C2-H protons.

Although values (9.3–12.1 Hz) of ${}^{3}J(P-C-O-C)$ were easily extracted from the ¹³C NMR spectra of phosphonates 8-11 (series c) they could not be unequivocally applied in the estimation of the extent to which rotation around the PC-OC bond is hindered because the angular dependence of ${}^{3}J(P-C-O-C)$ has not been established so far. However, the rotation around the OC-CN bond is not restricted because vicinal H₂C-CH₂ coupling constants observed for phosphonates 8c-11c fall in the 4.8-5.2 Hz range. On the other hand, on the basis of the values of ³J(HC–CH) found for PH₂C–CH₂O and OH₂C–CH₂N units (7.2-7.8 and 5.1-5.7 Hz, respectively), full conformational freedom within a five-atom linker in phosphonates 8-11 (series d) is anticipated.

Antiviral activity evaluation

The synthesised compounds 9a-9g, 10a-10g, and 11a-11g were evaluated for their antiviral activities against a wide variety of DNA and RNA viruses using the following cellbased assays: (a) human embryonic lung (HEL) cell: herpes simplex virus-1 (KOS), herpes simplex virus-2 (G), herpes simplex virus-1 (TK⁻ ACV^r KOS), vaccinia virus, vesicular stomatitis virus, varicella-zoster virus (TK⁺ VZV strain OKA and TK VZV strain 07-1) and cytomegalovirus (CMV) (strain AD-169 and Davis); (b) CEM cell cultures: human immunodeficiency virus-1 (HIV-1) and HIV-2; (c) Vero cell cultures: para-influenza-3 virus, reovirus-1, Sindbis virus, Coxsackie virus B4, Punta Toro virus; (d) HeLa cell cultures: vesicular stomatitis virus, Coxackie virus B4 and respiratory syncytial virus; (e) Crandel-Rees feline kidney (CRFK) cell cultures: feline corona virus (FIPV) and feline herpes virus (FHV) and (f) Madin Darby Canine kidney (MDCK) cell culture: influenza A virus H1N1 subtype A/PR/8, influenza H3N2 subtype A/HK/7/87 and influenza B virus (B/HK/5/72). Ganciclovir, cidofovir, acvclovir, brivudin, (S)-9-(2,3-dihydroxypropyl)adenine [(S)-DHPA], ribavirin, oseltamivir carboxylate, amantadine and rimantadine were used as the reference compounds. The antiviral activity was expressed as the EC_{50} : the compound concentration required to reduce virus plaques formation (VZV, CMV) by 50 % or to reduce virus-induced cytopathogenicity by 50 % (other viruses). None of the compounds showed appreciable antiviral activity at subtoxic concentrations.

Evaluation of cytotoxicity

The cytotoxicity of the tested compounds towards the uninfected host cells was defined as the minimum cytotoxic concentration (MMC) that causes a microscopically detectable alteration of normal cell morphology. The 50 % cytotoxic concentration (CC_{50}), i.e. causing a 50 % decrease in cell viability, was determined using a colorimetric 3-(4,5-dimethylthiazol-2yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2Htetrazolium (MTS) assay system. None of the tested compounds affected the cell morphology of Vero, HEL, HeLa or CRFK cells (MCC or CC_{50}) at compound concentrations up to 100 µM. They were also not cytostatic against murine leukemia and human CEM and HeLa cells at 100 µM.

Conclusions

Phosphonylated 1,2,3-triazoles 8a-8g were obtained in good to excellent yields by 1,3-dipolar cycloaddition between ω -azidophosphonates 7a–7g and dimethyl acetylenedicarboxylate. New series of diamides 9a-9g, dihydrazides 10a-10g and 1,2,3-triazolopyridazinediones 11a-11g were efficiently synthesised as phosphonate analogues of acyclic nucleosides in which nucleobases were replaced by substituted 1,2,3-triazoles.

Compounds 8–11 (series \mathbf{a} , \mathbf{b} , \mathbf{e} , \mathbf{f} , \mathbf{g}) exist in single conformations in which diethoxyphosphoryl and substituted 1,2,3-triazolyl (series \mathbf{a} and \mathbf{b}) or (1,2,3-triazolyl)methyl groups (series \mathbf{e} , \mathbf{f} , \mathbf{g}) prefer the *anti* orientation.

All synthesised compounds were evaluated for their antiviral activity against DNA and RNA viruses and were inactive. None of the compounds were cytotoxic (Vero, HEL, HeLa) or cytostatic (L1210, CEM, HeLa) at a concentration up to 100μ M.

Experimental

The ¹H NMR spectra were recorded in CDCl₃, CD₃OD or D₂O on the following spectrometers: Varian Mercury-300 and Bruker Avance III (600 MHz) with TMS as an internal standard; chemical shifts δ in ppm with respect to TMS; coupling constants *J* in Hz. The ¹³C NMR spectra were recorded for CDCl₃, CD₃OD or D₂O solutions on Varian Mercury-300 and Bruker Avance III (600 MHz) machines at 75.5 and 150.5 MHz, respectively. The ³¹P NMR spectra were recorded in CDCl₃, CD₃OD or D₂O on Varian Mercury-300 and Bruker Avance III (600 MHz) spectra were recorded in CDCl₃, CD₃OD or D₂O on Varian Mercury-300 and Bruker Avance III (600 MHz) spectra were recorded in CDCl₃, CD₃OD or D₂O on Varian Mercury-300 and Bruker Avance III (600 MHz) spectrometers at 121.5 and 243 MHz, respectively.

IR spectra were measured on an Infinity MI-60 FT-IR spectrometer. Melting points were determined on the Boetius apparatus. Elemental analyses were performed by the microanalytical laboratory of the host institution on Perkin Elmer PE 2400 CHNS analyzer and the results were found to be in good agreement ($\pm 0.3 \%$) with the calculated values.

The following absorbents were used: column chromatography, Merck silica gel 60 (70–230 mesh); analytical TLC, Merck TLC plastic sheets, silica gel 60 F_{254} . TLC plates were developed in chloroform–methanol solvent systems. Visualization of spots was effected with iodine vapours. All solvents were dried according to standard literature methods.

Diethyl 3-azido-1-hydroxypropylphosphonate (**7f**, C₇H₁₆N₃O₄P)

A mixture of 5.77 g 3-azidopropanal [39] (0.0582 mol), 3.0 cm³ diethyl phosphite (0.023 mol) and 0.81 cm³ triethylamine (0.0058 mol) was stirred at 5 °C for 24 h. The crude product was subjected to chromatography on silica gel with chloroform/methanol (100:1 and 50:1, v/v) to give a yellow oil (4.671 g, 34 %). IR (film): $\bar{V} = 3,272, 2,985, 2,934,$ 2,911, 2,874, 2,102, 1,227, 1,028 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz): $\delta = 1.35$ (t, J = 7.1 Hz, 6H, 2 × POCH₂CH₃), 1.87–2.06 (m, 3H, CH₂, OH), 3.55 (t, J = 6.3 Hz, 2H, CH₂N₃), 4.01 (dt, J = 9.8, 4.6 Hz, 1H, H-1), 4.12–4.24 (m, 4H, 2 × POCH₂CH₃) ppm; ¹³C NMR (CDCl₃, 75.5 MHz): $\delta = 16.6$ and 16.7 (2 d, J = 5.4 Hz, POCC), 30.8 (d, J = 2.6 Hz, C-2), 47.7 (d, J = 15.5 Hz, C-3), 62.9 and 63.2 (2 d, J = 7.2 Hz, POC), 64.6 (d, J = 164.6 Hz, C-1) ppm; ³¹P NMR (CDCl₃, 121.5 MHz): $\delta = 25.28$ ppm.

Synthesis of 1,2,3-triazoles 8a-8g (general procedure)

A solution of azidophosphonate **7a–7g** (1.00 mmol) and dimethyl acetylenedicarboxylate **13** (1.00 mmol) in 4 cm³ toluene was refluxed for 4 h. The reaction mixtures were concentrated to dryness to leave yellow oils which were purified on silica gel columns with chloroform/methanol (100:1, v/v) or were crystallised to give 1,2,3-triazoles **8a–8g**.

Diethyl 2-(4,5-dimethoxycarbonyl-1H-1,2,3-triazol-1-yl)ethylphosphonate (**8a**, C₁₂H₂₀N₃O₇P)

From 0.450 g azidophosphonate **7a** (2.17 mmol) and 0.309 g dimethyl acetylenedicarboxylate **13** (2.17 mmol) phosphonate **8a** was obtained as a yellowish oil (0.674 g, 89 %) after chromatography on a silica gel column with chloroform/methanol (100:1, v/v). IR (film): $\bar{V} = 3,462$, 2,985, 2,958, 1,736, 1,468, 1,447, 1,225, 1,140, 1,060, 1,025, 957 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz): $\delta = 1.33$ (t, J = 7.1 Hz, 6H, CH_3CH_2OP), 2.40–2.52 (m, 2H, PCH₂), 3.98 (s, 3H, OCH₃), 4.02 (s, 3H, OCH₃), 4.08–4.18 (m, 4H, CH₃CH₂OP), 4.82–4.91 (m, 2H, CH₂N) ppm; ¹³C NMR (CDCl₃, 75 MHz): $\delta = 16.6$ (d, J = 5.7 Hz, *CCOP*), 27.2 (d, J = 140.6 Hz, PC), 45.4 (s, PCCN), 52.9 (s, OCH₃), 53.7 (s, OCH₃), 62.4 (d, J = 6.3 Hz, *CCOP*), 129.8 and 140.2 (2 s, C=C), 158.6 and 160.4 (2 s, C=O) ppm; ³¹P NMR (CDCl₃, 121 MHz): $\delta = 25.9$ ppm.

Diethyl 3-(4,5-dimethoxycarbonyl-1H-1,2,3-triazol-1-yl)propylphosphonate (**8b**, C₁₃H₂₂N₃O₇P)

From 0.785 g azidophosphonate 7b (3.565 mmol) and 0.507 g dimethyl acetylenedicarboxylate 13 (3.565 mmol) phosphonate 8b was obtained as a yellowish oil (1.067 g, 83 %) after chromatography on a silica gel column with chloroform/methanol (100:1, v/v). IR (film): $\overline{V} = 2.953$, 2,836, 1,736, 1,463, 1,226, 1,030 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz): $\delta = 1.34$ (t, J = 7.1 Hz, 6H, CH₃CH₂OP), 1.72–1.84 (m, 2H, PCH₂CH₂), 2.16–2.31 (m, 2H, PCH₂), 4.00 (s, 3H, OCH₃), 4.03 (s, 3H, OCH₃), 4.05–4.18 (m, 4H, CH₃CH₂OP), 4.71 (t, J = 7.1 Hz, 2H, CH₂N) ppm; ¹³C NMR (CDCl₃, 75 MHz): $\delta = 16.5$ (d, J = 6.0 Hz, POCC), 22.6 (d, J = 143.1 Hz, PC), 23.5 (d, J = 4.3 Hz, PCC), 50.4 (d, J = 16.9 Hz, PCCC), 52.7 (s, OCH₃), 53.2 (s, OCH_3), 61.8 (d, J = 6.3 Hz, POC), 129.7 and 139.8 (2 s, C=C), 158.6 and 160.3 (2 s, C=O) ppm; ³¹P NMR (CDCl₃, 121 MHz): $\delta = 30.7$ ppm.

Diethyl 2-(4,5-dimethoxycarbonyl-1H-1,2,3-triazol-1-yl)ethoxymethylphosphonate (**8c**, C₁₃H₂₂N₃O₈P)

From 1.027 g azidophosphonate **7c** (4.330 mmol) and 0.615 g dimethyl acetylenedicarboxylate **13** (4.330 mmol) phosphonate **8c** was obtained as a yellowish oil (1.538 g, 93 %) after chromatography on a silica gel column with chloroform/

methanol (100:1, v/v). IR (film): $\bar{V} = 3,459, 2,983, 2,957, 2,908, 1,732, 1,462, 1,225, 1,117, 1,060, 1,027, 960 cm⁻¹; ¹H NMR (CDCl₃, 600 MHz): <math>\delta = 1.31$ (t, J = 7.0 Hz, 6H, CH₃CH₂OP), 3.74 (d, J = 7.9 Hz, 2H, PCH₂), 3.98 (s, 3H, OCH₃), 4.02 (s, 3H, OCH₃), 4.02 (t, J = 5.2 Hz, 2H, OCH₂CH₂N), 4.08–4.13 (m, 4H, CH₃CH₂OP), 4.86 (t, J = 5.2 Hz, 2H, OCH₂CH₂N), 4.08–4.13 (m, 4H, CH₃CH₂OP), 4.86 (t, J = 5.2 Hz, 2H, OCH₂CH₂N) ppm; ¹³C NMR (CDCl₃, 75 MHz): $\delta = 16.5$ (d, J = 5.7 Hz, POCC), 49.8 (s, CN), 52.8 (s, OCH₃), 53.5 (s, OCH₃), 62.6 (d, J = 6.5 Hz, POCC), 65.2 (d, J = 164.7 Hz, PC), 71.0 (d, J = 9.3 Hz, PCOC), 131.0 and 139.6 (2 s, C=C), 158.8 and 160.3 (2 s, C=O) ppm; ³¹P NMR (CDCl₃, 243 MHz): $\delta = 20.04$ ppm.

Diethyl 2-[2-(4,5-dimethoxycarbonyl-1H-1,2,3-triazol-1-yl)ethoxy]ethylphosphonate (**8d**, C₁₄H₂₄N₃O₈P)

From 0.647 g azidophosphonate 7d (2.68 mmol) and 0.382 g dimethyl acetylenedicarboxylate 13 (2.68 mmol) phosphonate 8d (1.030 g, 98 %) was obtained as a vellowish oil. The crude product was sufficiently pure and was used in the next step without further purification. IR (film): $\overline{V} = 3,459, 2,983, 2,957, 2,909, 1,736, 1,466$, 1,229, 1,118, 1,063, 1,027, 963 cm⁻¹; ¹H NMR (CDCl₃, 600 MHz): $\delta = 1.32$ (t, J = 7.1 Hz, 6H, CH_3CH_2OP), 2.00 (dt, J = 15.3, 7.7 Hz, 2H, PCH₂), 3.63 (dt, J = 10.4, 7.7 Hz, 2H, PCH₂CH₂O), 3.82 (t, J = 5.2 Hz, 2H, OCH₂CH₂N), 4.00 (s, 3H, OCH₃), 4.01 (s, 3H, OCH₃), 4.05–4.12 (m, 4H, CH₃CH₂OP), 4.82 (t, J = 5.2 Hz, 2H, OCH_2CH_2N) ppm; ¹³C NMR (CDCl₃, 150 MHz): $\delta = 16.3$ (d, J = 5.7 Hz, POCC), 26.7 (d, J = 139.3 Hz, PC), 49.8 (s, OCCN), 52.6 (s, OCH₃), 53.3 (s, OCH₃), 61.6 (d, J = 6.4 Hz, CCOP), 65.3 (s, PCC), 68.7 (s, OCCN), 131.4 and 139.5 (2 s, C=C), 159.1 and 160.3 (2 s, C=O) ppm; ³¹P NMR (CDCl₃, 243 MHz): $\delta = 27.58$ ppm.

Diethyl 1-hydroxy-3-(4,5-dimethoxycarbonyl-1H-1,2,3-triazol-1-yl)propylphosphonate (**8f**, C₁₃H₂₂N₃O₈P)

From 0.560 g azidophosphonate 7f (2.36 mmol) and 0.335 g dimethyl acetylenedicarboxylate 13 (2.36 mmol) phosphonate 8f (0.850 g, 95 %) was obtained as a white amorphous solid after chromatography on a silica gel column with chloroform/methanol (100:1, v/v). M.p.: 107-109 °C; IR (film): $\bar{V} = 3,420, 3,218, 3,081, 2,985, 2,878,$ 1,675, 1,451, 1,280, 1,223, 1,030, 970 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz): $\delta = 1.33$ and 1.34 (2 t, J = 7.2 Hz, 6H, CH₃CH₂OP), 2.28–2.47 (m, 2H, PCCH₂), 3.70 (dd, J = 6.0, 5.1 Hz, 1H, OH), 3.84 (dddd, J = 10.5, 6.3, 6.0,3.3 Hz, 1H, PCH), 3.98 (s, OCH₃), 4.00 (s, OCH₃), 4.10-4.22 (m, 4H, CH₃CH₂OP), 4.75-4.90 (m, 2H, CH₂N) ppm; ¹³C NMR (CDCl₃, 75 MHz): $\delta = 16.6$ and 16.7 (2 d, J = 5.4 Hz, CCOP), 31.9 (d, J = 3.2 Hz, PCC), 47.2 (d, J = 16.3 Hz, PCCC), 52.8 (s, OCH₃), 53.6 (s, OCH₃), 63.0 and 63.3 (2 d, J = 7.3 Hz, CCOP), 64.6 (d, J = 164.2 Hz, PC), 130.3 and 139.7 (2 s, C=C), 158.8 and 160.4 (2 s, C=O) ppm; ³¹P NMR (CDCl₃, 121 MHz): $\delta = 23.5$ ppm.

Synthesis of diamides **9a–9g** (general procedure)

To a solution of the diester **8a–8g** (1.00 mmol) in 14 cm³ ethanol 16 cm³ was added aqueous ammonia. The reaction mixtures were stirred at room temperature for 24 h. Ethanol and excess ammonia were evaporated in vacuo. Diamides **9a–9g** were purified on silica gel columns with chloroform/methanol or by crystallisation.

$Diethyl = 2-(4,5-dicarbamoyl-1H-1,2,3-triazol-1-yl)ethyl-phosphonate (9a, C_{10}H_{18}N_5O_5P)$

From 0.165 g diester **8a** (0.472 mmol) diamide **9a** was obtained as a white amorphous solid (0.065 g, 42 %) after chromatography on a silica gel column with chloroform/ methanol (50:1, 20:1, v/v) followed by crystallisation from ethanol/diethyl ether. M.p.: 172–173 °C; IR (KBr): $\bar{V} = 3,428, 3,228, 3,113, 2,986, 2,932, 1,687, 1,451, 1,221, 1,023 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz): <math>\delta = 1.34$ (t, J = 6.9 Hz, 6H, CH_3CH_2OP), 2.41–2.53 (m, 2H, PCH₂), 4.09–4.21 (m, 4H, CH₃CH₂OP), 5.08–5.17 (m, 2H, CH₂N), 6.01 (s, 2H, NH₂), 7.58 (s, 1H, NH), 10.77 (s, 1H, NH) ppm; ¹³C NMR (CD₃OD, 75 MHz): $\delta = 16.8$ (d, J = 6.0 Hz, CCOP), 27.3 (d, J = 140.3 Hz, PC), 47.2 (s, PCCN), 63.8 (d, J = 6.6 Hz, CCOP), 132.1 and 140.4 (2 s, C=C), 160.3 and 165.2 (2 s, C=O) ppm; ³¹P NMR (CDCl₃, 121 MHz): $\delta = 26.5$ ppm.

Diethyl 3-(4,5-*dicarbamoyl*-1H-1,2,3-*triazol*-1-*yl*)*propylphosphonate* (**9b**, C₁₁H₂₀N₅O₅P)

From 0.114 g diester 8b (0.314 mmol) diamide 9b was obtained as a white amorphous solid (0.070 g, 61 %) after chromatography on a silica gel column with chloroform/ methanol (50:1, v/v) and crystallisation from ethanol. M.p.: 143–144 °C; IR (KBr): $\bar{V} = 3,455, 3,270, 2,984, 2,909,$ 1,694, 1,629, 1,592, 1,454, 1,205, 1,018, 961 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz): $\delta = 1.31$ (t, J = 6.9 Hz, 6H, CH₃CH₂OP), 1.74–1.86 (m, 2H, PCH₂CH₂), 2.18–2.32 (m, 2H, PCH₂), 4.04–4.16 (m, 4H, CH₃CH₂OP), 4.98 (t, J = 6.9 Hz, 2H, CH₂N), 6.13 (s, 1H, NH), 6.25 (s, 1H, NH), 7.62 (s, 1H, NH), 10.79 (s, 1H, NH) ppm; ¹³C NMR $(CDCl_3, 75 \text{ MHz}): \delta = 16.7 \text{ (d, } J = 6.6 \text{ Hz}, CCOP), 23.0$ (d, J = 141.9 Hz, PC), 23.8 (d, J = 4.3 Hz, PCC), 51.8 (d, J = 4.3 Hz, PCC)J = 19.2 Hz, PCCC), 62.0 (d, J = 6.6 Hz, CCOP), 130.7 and 138.7 (2 s, C=C), 158.6 and 163.7 (2 s, C=O) ppm; ³¹P NMR (CDCl₃, 121 MHz): $\delta = 31.4$ ppm.

Diethyl [2-(4,5-dicarbamoyl-1H-1,2,3-triazol-1-yl)ethoxy]-methylphosphonate (9c, $C_{11}H_{20}N_5O_6P$)

From 0.280 g diester **8c** (0.738 mmol) diamide **9c** was obtained as a white amorphous solid (0.139 g, 54 %) after chromatography on a silica gel column with chloroform/ methanol (50:1, v/v). M.p.: 100–101 °C; IR (KBr): $\bar{V} = 3,361, 3,259, 3,110, 2,990, 2,959, 2,901, 1,689, 1,613, 1,454, 1,303, 1,219, 1,118, 1,048, 1,019, 967,$

943 cm⁻¹; ¹H NMR (CD₃OD, 600 MHz): $\delta = 1.29$ (t, J = 7.1 Hz, 6H, CH₃CH₂OP), 3.89 (d, J = 8.5 Hz, 2H, PCH₂), 4.06 (t, J = 5.2 Hz, 2H, OCH₂CH₂N), 4.06–4.10 (m, 4H, CH₃CH₂OP), 5.14 (t, J = 5.2 Hz, 2H, CH₂N) ppm; ¹³C NMR (CD₃OD, 150 MHz): $\delta = 15.3$ (d, J = 5.9 Hz, CCOP), 50.3 (s, CN), 62.8 (d, J = 6.5 Hz, CCOP), 64.0 (d, J = 164.7 Hz, PC), 71.1 (d, J = 11.9 Hz, PCOC), 131.0 and 139.0 (2 s, C=C), 159.1 and 163.9 (2 s, C=O) ppm; ³¹P NMR (CD₃OD, 243 MHz): $\delta = 21.3$ ppm.

Diethyl 2-[2-(4,5-dicarbamoyl-1H-1,2,3-triazol-1-yl)ethoxy]ethylphosphonate (**9d**, C₁₂H₂₂N₅O₆P)

From 0.264 g diester 8d (0.671 mmol) diamide 9d was obtained as a white amorphous solid (0.090 g, 37 %) after chromatography on a silica gel column with chloroform/ methanol (50:1, v/v) and crystallisation from ethyl acetate. M.p.: 102–103 °C; IR (KBr): $\overline{V} = 3,400, 3,306, 3,214,$ 2,986, 2,906, 1,672, 1,608, 1,453, 1,245, 1,109, 1,024, 961 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz): $\delta = 1.31$ (t, J = 6.9 Hz, 6H, CH₃CH₂OP), 2.05 (dt, J = 15.3, 7.8 Hz, 2H, PCH₂), 3.69 (dt, J = 11.1, 7.8 Hz, 2H, PCH₂CH₂O), 3.91 (t, J = 5.4 Hz, 2H, OCH₂CH₂N), 4.01–4.12 (m, 4H, CH₃CH₂OP), 5.11 (t, J = 5.4 Hz, 2H, OCH₂CH₂N), 6.00 (s, 1H, NH), 6.07 (s, 1H, NH), 7.60 (s, 1H, NH), 10.77 (s, 1H, NH) ppm; ¹³C NMR (CDCl₃, 75 MHz): $\delta = 16.6$ (d, J = 6.0 Hz, CCOP), 26.9 (d, J = 139.1 Hz, PC), 51.0 (s, OCCN), 61.9 (d, J = 6.3 Hz, CCOP), 65.0 (s, PCCO), 69.1 (s, OCCN), 130.9 and 139.7 (2 s, C=C), 158.7 and 163.7 (2 s, C=O) ppm; ³¹P NMR (CDCl₃, 121 MHz): $\delta = 29.2$ ppm.

Diethyl 3-(4,5-dicarbamoyl-1H-1,2,3-triazol-1-yl)-2-hy-droxypropylphosphonate (**9e**, C₁₁H₂₀N₅O₆P)

From 0.145 g diester 8e (0.382 mmol) diamide 9e was obtained as a white amorphous solid (0.060 g, 45 %) after chromatography on a silica gel column with chloroform/ methanol (50:1, v/v) and crystallisation from methanol. M.p.: 142–143 °C; IR (KBr): $\bar{V} = 3,428, 3,308, 3,034,$ 2,988, 1,686, 1,667, 1,599, 1,443, 1,375, 1,293, 1,113, 1,083, 1,055, 1,029, 960 cm⁻¹; ¹H NMR (CD₃OD, 300 MHz): $\delta = 1.32$ (t, J = 7.2 Hz, 6H, CH_3CH_2OP), 2.06 (ddd, J = 22.8, 15.3, 7.8 Hz, 1H, PCH_aH_b), 2.18 $(ddd, J = 20.4, 15.3, 5.1 \text{ Hz}, 1\text{H}, \text{PCH}_{a}H_{b}), 4.07-4.19 \text{ (m},$ 4H, CH₃CH₂OP), 4.37–4.50 (m, 1H, PCCH), 4.99 (dd, J = 13.5, 8.1 Hz, 1H, PCCC H_aH_b), 5.02 (dd, J = 13.5, 4.2 Hz, 1H, PCCCH_a $H_{\rm b}$) ppm; ¹³C NMR (CD₃OD, 75 MHz): $\delta = 16.9$ (d, J = 6.3 Hz, CCOP), 32.3 (d, J = 141.7 Hz, PC), 58.1 (d, J = 18.0 Hz, PCCC), 63.4 and 63.7 (2 d, J = 6.6 Hz, CCOP), 67.0 (d, J = 4.0 Hz, PCC), 132.6 and 140.4 (2 s, C=C), 160.6 and 165.3 (2 s, ³¹P C=O) ppm; NMR (CD₃OD, 121 MHz): $\delta = 30.1$ ppm.

Diethyl 3-(4,5-dicarbamoyl-1H-1,2,3-triazol-1-yl)-1-hydroxypropylphosphonate (**9f**, C₁₁H₂₀N₅O₆P)

From 0.180 g diester **8f** (0.475 mmol) diamide **9f** was obtained as a white amorphous solid (0.093 g, 56 %) after crystallisation from methanol. M.p.: 179–180 °C; IR (KBr): $\bar{V} = 3,433$, 3,319, 3,212, 2,974, 2,930, 2,874, 1,668, 1,605, 1,455, 1,399, 1,221, 1,167, 1,066, 1,014, 953 cm⁻¹; ¹H NMR (CD₃OD, 300 MHz): $\delta = 1.32$ (t, J = 7.2 Hz, 6H, CH_3CH_2OP), 2.13–2.28 (m, 1H, PCCH_aH_b), 2.31–2.44 (m, 1H, PCCH_aH_b), 3.89 (ddd, J = 10.5, 7.2, 3.0 Hz, 1H, PCH), 4.10–4.21 (m, 4H, CH₃CH₂OP), 4.95–5.14 (m, 2H, CH₂N) ppm; solubility of **9f** in D₂O or CD₃OD was not sufficient to measure the ¹³C NMR spectrum; ³¹P NMR (CD₃OD, 121 MHz): $\delta = 25.4$ ppm.

Diethyl 2-(4,5-*dicarbamoyl*-1H-1,2,3-*triazol*-1-*yl*)-1-*hydroxyethylphosphonate* (**9g**, C₁₀H₁₈N₅O₆P)

From 0.230 g diester **8g** (0.630 mmol) diamide **9g** was obtained as a white amorphous solid (0.112 g, 53 %) after chromatography on a silica gel column with chloroform/ methanol (20:1, v/v) and crystallisation from methanol. M.p.: 200–202 °C; IR (KBr): $\bar{V} = 3,431, 3,223, 3,081, 2,990, 2,847, 1,671, 1,599, 1,452, 1,393, 1,289, 1,225, 1,031, 979 cm⁻¹; ¹H NMR (CD₃OD, 600 MHz): <math>\delta = 1.39$ and 1.40 (2 t, J = 7.2 Hz, 6H, CH_3CH_2OP), 4.22-4.29 (m, 4H, CH_3CH_2OP), 4.55 (ddd, J = 10.2, 9.9 Hz, 3.6 Hz, 1H, PCH), 5.10 (ddd, J = 13.5, 10.2, 8.7 Hz, 1H, PCCH_aH_b), 5.22 (ddd, J = 13.5, 3.6, 3.1 Hz, 1H, PCCH_aH_b) ppm; solubility of **9g** in D₂O and CD₃OD was not sufficient to measure the ¹³C NMR spectrum; ³¹P NMR (CD₃OD, 243 MHz): $\delta = 21.2$ ppm.

Synthesis of dihydrazides 10a–10g (general procedure)

A solution of the diester 8a-8g (0.73 mmol) and 0.530 cm³ hydrazine hydrate (10.8 mmol) in 6 cm³ ethanol was refluxed for 2 h. The reaction mixtures were concentrated to give yellow oils or solids which were subjected to chromatography on a silica gel column with chloroform/ methanol or crystallisation to obtain dihydrazides 10a-10g.

Diethyl 2-[4,5-bis(hydrazinocarbonyl)-1H-1,2,3-triazol-1yl]ethylphosphonate (**10a**, $C_{10}H_{20}N_7O_5P$)

From 0.115 g diester **8a** (0.330 mmol) dihydrazide **10a** was obtained (0.070 g, 61 %) after chromatography on a silica gel column with chloroform/methanol (50:1, 20:1, 10:1, v/v) as a white amorphous solid. M.p.: 84–86 °C; IR (KBr): $\bar{V} = 3,335, 3,282, 2,984, 2,932, 1,661, 1,606, 1,551, 1,491, 1,262, 1,223, 1,056, 1,020, 963 cm⁻¹; ¹H NMR (CD₃OD, 300 MHz): <math>\delta = 1.32$ (t, J = 6.6 Hz, 6H, CH₃CH₂O), 2.48–2.59 (m, 2H, PCH₂), 4.07–4.18 (m, 4H, CH₃CH₂O), 5.03–5.14 (m, 2H, CH₂N) ppm; ¹³C NMR

(CD₃OD, 75 MHz): $\delta = 16.9$ (d, J = 6.0 Hz, CCOP), 27.4 (d, J = 140.0 Hz, PC), 47.1 (s, PCCN), 63.8 (d, J = 6.3 Hz, CCOP × 2), 130.8 and 139.3 (2 s, C=C), 157.6 and 161.8 (2 s, C=O) ppm; ³¹P NMR (CD₃OD, 121 MHz): $\delta = 28.2$ ppm.

Diethyl 3-[4,5-bis(hydrazinocarbonyl)-1H-1,2,3-triazol-1yl]propylphosphonate (**10b**, C₁₁H₂₂N₇O₅P)

From 0.170 g diester **8b** (0.468 mmol) dihydrazide **10b** was obtained as a white amorphous solid (0.113 g, 66 %) after chromatography on a silica gel column with chloro-form/methanol (50:1, 20:1, 10:1, v/v). M.p.: 73–75 °C; IR (KBr): $\bar{V} = 3,300, 3,192, 2,983, 2,931, 1,656, 1,551, 1,209, 1,024, 969 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz): <math>\delta = 1.28$ (t, J = 6.9 Hz, 6H, CH_3CH_2OP), 1.71–1.82 (m, 2H, PCH₂CH₂), 2.13–2.27 (m, 2H, PCH₂), 4.00–4.13 (m, 4H, CH₃CH₂OP), 4.94 (t, J = 7.2 Hz, 2H, CH₂N), 5.15–6.03 (brs, 6H, NHNH₂) ppm; ¹³C NMR (CDCl₃, 75 MHz): $\delta = 16.7$ (d, J = 6.0 Hz, CCOP), 22.9 (d, J = 139.9 Hz, PCC), 62.0 (d, J = 6.6 Hz, CCOP), 129.5 and 137.6 (2 s, C=C), 156.3 and 161.3 (2 s, C=O) ppm; ³¹P NMR (CDCl₃, 121 MHz): $\delta = 31.5$ ppm.

Diethyl [2-[4,5-bis(hydrazinocarbonyl)-1H-1,2,3-triazol-1yl]ethoxy]methylphosphonate (**10c**, C₁₁H₂₂N₇O₆P)

From 0.430 g diester **8c** (1.13 mmol) dihydrazide **10c** was obtained as a colourless oil (0.139 g, 32 %) after chromatography on a silica gel column with chloroform/methanol (50:1, 20:1, 10:1, v/v). IR (film): $\bar{V} = 3,326, 3,248, 2,985, 2,918, 2,890, 1,666, 1,544, 1,440, 1,300, 1,238, 1,117, 1,024, 968 cm⁻¹; ¹H NMR (CD₃OD, 600 MHz): <math>\delta = 1.28$ (t, J = 7.0 Hz, 6H, CH_3CH_2OP), 3.89 (d, J = 8.5 Hz, 2H, PCH₂), 4.05–4.10 (m, 6H, CH₃CH₂OP and OCH₂CH₂N), 5.14 (t, J = 4.8 Hz, 2H, OCH₂CH₂N) ppm; ¹³C NMR (CD₃OD, 150 MHz): $\delta = 15.3$ (d, J = 5.5 Hz, CCOP), 50.2 (s, CN), 62.8 (d, J = 6.5 Hz, CCOP), 64.0 (d, J = 164.9 Hz, PC), 71.1 (d, J = 11.9 Hz, PCOC), 129.9 and 137.9 (2 s, C=C), 156.5 and 160.5 (2 s, C=O) ppm; ³¹P NMR (CD₃OD, 243 MHz): $\delta = 21.4$ ppm.

Diethyl 2-[2-[4,5-bis(hydrazinocarbonyl)-1H-1,2,3-triazol-1-yl]ethoxy]ethylphosphonate (**10d**, C₁₂H₂₄N₇O₆P)

From 0.216 g diester **8d** (0.549 mmol) dihydrazide **10d** was obtained as a white amorphous solid (0.128 g, 59 %) after chromatography on a silica gel column with chloroform/methanol (50:1, 20:1, v/v) and crystallisation from ethyl acetate. M.p: 95–96 °C; IR (KBr): $\bar{V} = 3,300, 3,232, 2,983, 2,913, 2,827, 1,667, 1,545, 1,550, 1,252, 1,023, 966, 918 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz): <math>\delta = 1.31$ (t, J = 7.2 Hz, 6H, CH_3CH_2OP), 2.03 (dt, J = 15.0, 7.5 Hz, 2H, PCH₂), 3.69 (dt, J = 11.1, 7.5 Hz, 2H, PCH₂CH₂O), 3.91 (t, J = 5.7 Hz, 2H, OCH₂CH₂N), 4.01–4.13 (m, 4H, CH₃CH₂OP), 4.20 (brs, 4H, NH₂), 5.13 (t, J = 5.7 Hz, 2H,

OCH₂CH₂N), 8.81 (s, 1H, NH), 12.0 (s, 1H, NH) ppm; ¹³C NMR (CDCl₃, 75 MHz): δ = 16.6 (d, J = 6.0 Hz, CCOP), 27.0 (d, J = 138.5 Hz, PC), 51.0 (s, OCCN), 61.9 (d, J = 6.3 Hz, CCOP), 65.0 (s, PCCO), 69.1 (s, OCCN), 129.9 and 137.5 (2 s, C=C), 156.5 and 161.3 (2 s, C=O) ppm; ³¹P NMR (CDCl₃, 121 MHz): δ = 28.5 ppm.

Diethyl 3-[4,5-bis(hydrazinocarbonyl)-1H-1,2,3-triazol-1yl]-2-hydroxypropylphosphonate (**10e**, C₁₁H₂₂N₇O₆P)

From 0.290 g diester 8e (0.764 mmol) dihydrazide 10e was obtained as a white amorphous solid (0.140 g, 48 %) after chromatography on a silica gel column with chloroform/ methanol (50:1, 20:1, v/v) and crystallisation from ethanol. M.p.: 140–142 °C; IR (KBr): $\bar{V} = 3,336, 3,275, 2,980,$ 2,930, 1,668, 1,578, 1,549, 1,443,1,260, 1,217, 1,067, 1,027, 957 cm⁻¹; ¹H NMR (CD₃OD, 600 MHz): $\delta = 1.34$ and 1.35 (2 t, J = 7.1 Hz, 6H, CH₃CH₂OP), 2.12 (ddd, J = 23.6, 15.4, 8.2 Hz, 1H, PCH₂H_b), 2.18 (ddd, J = 20.1,15.4, 4.7 Hz, 1H, PCH_aH_b , 4.11–4.19 (m, 4H. CH₃CH₂OP), 4.44–4.47 (m, 1H, PCCH), 4.96 (dd, J = 13.4, 8.4 Hz, 1H, PCCC H_aH_b), 5.05 (dd, J = 13.4, 4.1 Hz, 1H, PCCCH_a H_b) ppm; ¹³C NMR (CD₃OD, 150 MHz): $\delta = 15.3$ (d, J = 5.8 Hz, CCOP), 30.8 (d, J = 141.6 Hz, PC), 56.4 (d, J = 17.3 Hz, PCCC), 61.9 and 62.2 (2 d, J = 6.5 Hz, CCOP), 65.6 (d, J = 3.6 Hz, PCC), 130.1 and 137.9 (2 s, C=C), 156.7 and 160.6 (2 s, C=O) ppm; ³¹P NMR (CD₃OD, 243 MHz): $\delta = 28.9$ ppm.

Diethyl 3-[4,5-bis(hydrazinocarbonyl)-1H-1,2,3-triazol-1yl]-1-hydroxypropylphosphonate (**10f**, $C_{11}H_{22}N_7O_6P$) From 0.262 g diester **8f** (0.691 mmol) dihydrazide **10f** was

From 0.262 g diester 8f (0.691 mmol) dihydrazide 10f was obtained as a white amorphous solid (0.155 g, 60 %) after chromatography on a silica gel column with chloroform/ methanol (50:1, 30:1, v/v). M.p.: 119-120 °C; IR (KBr): $\bar{V} = 3,316, 3,266, 3,160, 2,976, 2,930, 1,659, 1,572, 1,532,$ 1,249, 1,210, 1,072, 1,022, 955 cm⁻¹; ¹H NMR (CD₃OD, 300 MHz): $\delta = 1.31$ (t, J = 7.5 Hz, 6H, CH_3CH_2OP), 2.13-2.29 (m, 1H, PCCH_aH_b), 2.31–2.44 (m, 1H, $PCCH_aH_b$), 3.91 (ddd, J = 10.8, 7.8, 3.3 Hz, 1H, PCH), 4.09-4.21 (m, 4H, CH₃CH₂OP), 4.96-5.15 (m, 2H, CH₂N) ppm; ¹³C NMR (CD₃OD, 75 MHz): $\delta = 17.0$ (d, J = 5.4 Hz, CCOP), 33.4 (s, PCC), 48.4 (d, J = 18.9 Hz, PCCC), 64.2 and 64.5 (d, J = 7.1 Hz, CCOP), 65.6 (d, J = 166.7 Hz, PC), 130.9 and 139.2 (2 s, C=C), 157.7 and ³¹P 161.8 (2 s, C=O \times 2) ppm; NMR (CD₃OD, 121 MHz): $\delta = 25.5$ ppm.

Diethyl 2-[4,5-bis(hydrazinocarbonyl)-1H-1,2,3-triazol-1yl]-1-hydroxyethylphosphonate (10g, $C_{10}H_{20}N_7O_6P$)

From 0.450 g diester **8g** (1.23 mmol) dihydrazide **10g** was obtained as a white amorphous solid (0.128 g, 28 %) after chromatography on a silica gel column with chloroform/ methanol (50:1, 30:1, v/v). M.p.: 149–152 °C; IR (KBr): $\bar{V} = 3,334, 3,207, 2,980, 2,930, 2,870, 1,668, 1,578, 1,449,$

1,217, 1,067, 1,028, 953 cm⁻¹; ¹H NMR (D₂O, 600 MHz): $\delta = 1.31$ (t, J = 7.0 Hz, 6H, CH₃CH₂OP), 4.18–4.24 (m, 4H, CH₃CH₂OP), 4.53–4.58 (m, 1H, PCH), 4.94–5.01 (m, 1H, PCCH_aH_b), 5.11–5.16 (m, 1H, PCCH_aH_b) ppm; ¹³C NMR (D₂O, 150 MHz): $\delta = 15.7$ (d, J = 4.6 Hz, CCOP), 51.9 (d, J = 11.2 Hz, PCCN), 64.7 and 64.8 (2 d, J = 7.4 Hz, CCOP), 65.5 (d, J = 165.8 Hz, PC), 130.4 and 137.5 (2 s, C=C), 156.5 and 160.2 (2 s, C=O) ppm; ³¹P NMR (D₂O, 243 MHz): $\delta = 21.8$ ppm.

Synthesis of 1,2,3-triazolopyridazinediones **11a–11g** (general procedure)

A mixture of the dihydrazide 10a-10g (0.50 mmol) and 5 cm³ 10 % hydrochloric acid was heated at 90 °C for 2.5 h. After concentration in vacuo, crude products were purified on a silica gel column with chloroform/methanol or crystallised from the appropriate solvent.

Diethyl 2-(4,7-dioxo-5,6-dihydro-1H-1,2,3-triazolo[4,5-d]pyridazin-1-y]ethylphosphonate (**11a**, C₁₀H₁₆N₅O₅P)

From 0.167 g dihydrazide **10a** (0.478 mmol) compound **11a** was obtained as a white amorphous solid (0.045 g, 30 %) after chromatography on a silica gel column with chloroform/methanol (50:1, 20:1, 10:1, v/v) followed by crystallisation from ethanol. M.p.: 198–200 °C; IR (KBr): $\bar{V} = 3,385, 2,922, 2,672, 1,687, 1,582, 1,460, 1,262, 1,214,$ 1,014, 1,019, 980 cm⁻¹; ¹H NMR (CD₃OD, 300 MHz): $\delta = 1.28$ (t, J = 6.6 Hz, 6H, CH_3CH_2OP), 2.58 (dt, J = 14.7 Hz, J = 7.5 Hz, PCH₂), 4.03–4.13 (m, 4H, CH₃CH₂OP), 5.12 (dt, J = 12.6 Hz, J = 7.5 Hz, CH₂N) ppm; ¹³C NMR (CD₃OD, 150 MHz): $\delta = 17.5$ (d, J = 5.7 Hz, CCOP), 28.2 (d, J = 141.0 Hz, PC), 46.5 (d, J = 3.0 Hz, PCCN), 64.7 (d, J = 6.6 Hz, CCOP), 131.3 and 141.8 (2 s, C=C), 152.0 and 154.6 (2 s, C=O) ppm; ³¹P NMR (CD₃OD, 121 MHz): $\delta = 27.7$ ppm.

Diethyl 3-(4,7-dioxo-5,6-dihydro-1H-1,2,3-triazolo[4,5d]pyridazin-1-yl)propylphosphonate

$(11b, C_{11}H_{18}N_5O_5P)$

From 0.055 g dihydrazide **10b** (0.151 mmol) compound **11b** was obtained as a white amorphous solid (0.033 g, 66 %) after chromatography on a silica gel column with chloroform/methanol (50:1, 20:1, 10:1, v/v). M.p.: 148– 150 °C; IR (KBr): $\bar{V} = 3,446, 3,074, 2,991, 2,960, 1,662,$ 1,555, 1,462, 1,294, 1,218, 1,032, 972, 805 cm⁻¹; ¹H NMR (CD₃OD, 300 MHz): $\delta = 1.31$ (t, J = 6.9 Hz, 6H, *CH*₃CH₂OP), 1.84–1.95 (m, 2H, PCH₂CH₂), 2.24–2.38 (m, 2H, PCH₂), 4.02–4.15 (m, 4H, CH₃CH₂OP), 4.93 (t, J = 6.9 Hz, 2H, CH₂N) ppm; ¹³C NMR (CD₃OD, 75 MHz): $\delta = 16.7$ (d, J = 5.7 Hz, *C*COP), 22.7 (d, J = 141.9 Hz, PC), 23.9 (s, PCC), 50.5 (d, J = 18.5 Hz, PCCC), 62.6 (d, J = 6.5 Hz, *CCOP*), 129.1 and 139.7 (2 s, C=C), 150.5 and 152.7 (2 s, C=O) ppm; ³¹P NMR (CD₃OD, 121 MHz): δ = 32.6 ppm.

Diethyl [2-(4,7-dioxo-5,6-dihydro-1H-1,2,3-triazolo-[4,5-d]pyridazin-1-yl)ethoxy]methylphosphonate (**11c**, C₁₁H₁₈N₅O₆P)

From 0.090 g dihydrazide **10c** (0.237 mmol) compound **11c** was obtained as a white amorphous solid (0.043 g, 52 %) after chromatography on a silica gel column with chloroform/methanol (50:1, 20:1, v/v). M.p.: 170–172 °C; IR (KBr): $\bar{V} = 3,348, 2,985, 2,930, 2,918, 1,658, 1,552,$ 1,420, 1,260, 1,070, 1,028, 975 cm⁻¹; ¹H NMR (CD₃OD, 600 MHz): $\delta = 1.25$ (t, J = 7.0 Hz, 6H, CH_3CH_2OP), 3.89 (d, J = 8.5 Hz, 2H, PCH₂), 4.01–4.04 (m, 4H, CH₃CH₂OP), 4.16 (t, J = 5.1 Hz, 2H, OCH₂CH₂N), 5.09 (t, J = 5.1 Hz, 2H, OCH₂CH₂N) ppm; ¹³C NMR (CD₃OD, 150 MHz): $\delta = 15.2$ (d, J = 5.7 Hz, CCOP), 49.2 (s, CN), 62.7 (d, J = 6.6 Hz, CCOP), 63.9 (d, J = 164.8 Hz, PC), 70.9 (d, J = 12.1 Hz, PCOC), 129.2 and 139.3 (2 s, C=C), 149.6 and 152.3 (2 s, C=O) ppm; ³¹P NMR (CD₃OD, 243 MHz): $\delta = 21.2$ ppm.

Diethyl 2-[2-(4,7-dioxo-5,6-dihydro-1H-1,2,3-triazolo[4,5-d]pyridazin-1-yl)ethoxy]ethylphosphonate (11d, $C_{12}H_{20}N_5O_6P$)

From 0.165 g dihydrazide 10d (0.419 mmol) compound 11d was obtained as a white amorphous solid (0.087 g, 58 %) after chromatography on a silica gel column with chloroform/methanol (50:1, 20:1, v/v) and crystallisation from ethyl acetate/chloroform. M.p.: 134-135 °C; IR (KBr): $\overline{V} = 3,433, 2,974, 2,928, 2,872, 1,715, 1,645,$ 1,206, 1,113, 1,065, 1,026, 982, 952 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz): $\delta = 1.26$ (t, J = 7.2 Hz, 6H, CH_3CH_2OP), 2.07 (dt, J = 15.0, 7.2 Hz, 2H, PCH₂), 3.73 $(dt, J = 13.5, 7.5 Hz, 2H, PCH_2CH_2O), 3.96-4.10 (m, 6H, 6H)$ CH₃CH₂OP and OCH₂CH₂N), 5.01 (t, J = 5.1 Hz, 2H, OCH₂CH₂N), 10.49 (brs, 2H, NH) ppm; ¹³C NMR (CDCl₃, 75 MHz): $\delta = 16.6$ (d, J = 6.3 Hz, CCOP), 26.8 (d, J = 138.9 Hz, PC), 49.9 (s, OCCN), 62.2 (d, J = 6.5 Hz, CCOP), 65.0 (s, PCCO), 68.8 (s, OCCN), 129.4 and 139.4 (2 s, C=C), 151.1 and 152.8 (2 s, C=O) ppm; ³¹P NMR (CDCl₃, 121 MHz): $\delta = 29.3$ ppm.

Diethyl 3-(4,7-dioxo-5,6-dihydro-1H-1,2,3-triazolo[4,5-d]pyridazin-1-yl)-2-hydroxypropylphosphonate (**11e**, C₁₁H₁₈N₅O₆P)

From 0.270 g dihydrazide **10e** (0.719 mmol) compound **11e** was obtained as a white amorphous solid (0.087 g, 39 %) after chromatography on a silica gel column with chloroform/methanol (20:1, 10:1, v/v) and crystallisation from water. M.p.: 168–170 °C; IR (KBr): $\bar{V} = 3,476$, 2,982, 2,932, 2,867, 1,655, 1,550, 1,427, 1,405, 1,256, 1,222, 1,108, 1,054, 957 cm⁻¹; ¹H NMR (D₂O, 600 MHz): $δ = 1.30 \text{ and } 1.31 (2 \text{ t}, J = 6.1 \text{ Hz}, 6\text{H}, CH_3CH_2OP), 2.25$ (td, J = 16.1, 8.8 Hz, 1H, PCH_aH_b), 2.35 (td, J = 15.8, 3.2 Hz, 1H, PCH_aH_b), 4.10–4.17 (m, 4H, CH₃CH₂OP), 4.52–4.59 (m, 1H, PCCH), 4.91 (dd, J = 12.8, 9.2 Hz, 1H, PCCCH_aH_b), 4.98 (dd, J = 12.8, 2.6 Hz, 1H, PCCCH_aH_b) ppm; ¹³C NMR (D₂O, 150 MHz): δ = 15.6 (d, J = 5.7 Hz, CCOP), 30.0 (d, J = 139.9 Hz, PC), 55.7 (d, J = 17.8 Hz, PCCC), 63.5 and 63.6 (2 d, J = 6.4 Hz, CCOP), 65.4 (d, J = 3.8 Hz, PCC), 130.1 and 140.1 (2 s, C=C), 149.8 and 153.4 (2 s, C=O) ppm; ³¹P NMR (D₂O, 243 MHz): δ = 30.3 ppm.

Diethyl 3-(4,7-dioxo-5,6-dihydro-1H-1,2,3-triazolo-[4,5-d]pyridazin-1-yl)-1-hydroxypropylphosphonate (**11f**, C₁₁H₁₈N₅O₆P)

From 0.163 g dihydrazide 10f (0.430 mmol) compound 11f was obtained as a white amorphous solid (0.086 g, 58 %) after chromatography on a silica gel column with chloroform/methanol (50:1, 20:1, 10:1, v/v). M.p.: 66-68 °C; IR (KBr): $\overline{V} = 3,362, 2,987, 2,920, 1,666, 1,580,$ 1,449, 1,212, 1,050, 1,021, 970 cm⁻¹; ¹H NMR (CD₃OD, 300 MHz): $\delta = 1.32$ (t, J = 7.2 Hz, 6H, CH_3CH_2OP), 2.19–2.28 (m, 1H, PCCH_aH_b), 2.31–2.44 (m, 1H, $PCCH_aH_b$), 3.89 (ddd, J = 10.5, 7.2, 3.0 Hz, 1H, PCH), 4.10-4.21 (m, 4H, CH₃CH₂OP), 4.95-5.12 (m, 2H, CH₂N) ppm; ¹³C NMR (CD₃OD, 75 MHz): $\delta = 17.0$ (d, J = 5.4 Hz, CCOP), 33.3 (d, J = 4.0 Hz, PCC), 48.1 (d, J = 16.2 Hz, PCCC), 64.2 and 64.5 (d, J = 6.9 Hz, CCOP), 65.4 (d, J = 167.3 Hz, PC), 130.2 and 140.8 (2 s, C=C), 148.9 and 152.2 (2 s, C=O) ppm; ³¹P NMR (CD₃OD, 121 MHz): $\delta = 25.3$ ppm.

Diethyl 2-(4,7-dioxo-5,6-dihydro-1H-1,2,3-triazolo-[4,5-d]pyridazin-1-yl)-1-hydroxyethylphosphonate (**11g**, C₁₀H₁₆N₅O₆P)

From 0.100 g dihydrazide **10g** (0.274 mmol) compound **11g** was obtained as a colourless oil (0.036 g, 40 %) after chromatography on a silica gel column with chloroform/ methanol (50:1, 10:1, v/v). IR (film): $\bar{V} = 3,362, 2,988,$ 2,971, 1,667, 1,580, 1,212, 1,021 cm⁻¹; ¹H NMR (CD₃OD, 600 MHz): $\delta = 1.38$ and 1.39 (2 t, J = 7.0 Hz, 6H, CH₃CH₂OP), 4.23–4.29 (m, 4H, CH₃CH₂OP), 4.64–4.69 (m, 1H, PCH), 5.09–5.15 (m, 2H, CH₂N) ppm; ¹³C NMR (CD₃OD, 150 MHz): $\delta = 15.3$ and 15.4 (d, J = 5.2 Hz, CCOP), 51.3 (d, J = 11.6 Hz, PCCN), 63.3 and 63.5 (2 d, J = 6.8 Hz, COP), 66.4 (d, J = 166.2 Hz, PC), 129.2 and 139.4 (2 s, C=C), 150.1 and 152.4 (2 s, C=O) ppm; ³¹P NMR (CD₃OD, 243 MHz): $\delta = 20.5$ ppm.

Assays for antiviral activity other than HIV

The compounds were evaluated against the following viruses: herpes simplex virus type 1 (HSV-1) strain KOS, thymidine kinase-deficient (TK^-) HSV-1 KOS strain

resistant to ACV (ACV^r), herpes simplex virus type 2 (HSV-2) strain G, cytomegalovirus (strains AD-169 and Davis), varicella-zoster virus (VZV) (strains OKA and YS), vaccinia virus Lederle strain, respiratory syncytial virus (RSV) strain Long, vesicular stomatitis virus (VSV), coxsackie b4 virus, parainfluenza 3, influenza virus A (subtypes H1N1, H3N2), influenza virus B, reovirus-1, Sindbis virus and Punta Toro virus. The antiviral assays were based on inhibition of virus-induced cytopathicity or plaque formation in human embryonic lung (HEL) fibroblasts, African green monkey cells (Vero), human epithelial cervix carcinoma cells (HeLa) or MDCK. Confluent cell cultures in microtiter 96-well plates were inoculated with 100 CCID₅₀ of virus (1 CCID₅₀ being the virus dose to infect 50 % of the cell cultures) or 20 or 100 plaque forming units (PFU) (for VZV and CMV, respectively) in the presence of varying concentrations of the test compounds. Viral cytopathicity or plaque formation was recorded as soon as it reached completion in the control virus-infected cell cultures that were not treated with the test compounds. Antiviral activity was expressed as the EC_{50} or compound concentration required to reduce virus-induced cytopathogenicity or viral plaque formation by 50 %.

Anti-HIV activity assays

Inhibition of HIV-1(III_B)- and HIV-2(ROD)-induced cytopathicity in CEM cell cultures was measured in microtiter 96-well plates containing 3×10^5 CEM cells/ cm³ infected with 100 CCID50 of HIV per cm³ and containing appropriate dilutions of the test compounds. After 4–5 days of incubation at 37 °C in a CO₂-controlled humidified atmosphere, CEM giant cell (syncytium) formation was examined microscopically. The EC₅₀ (50 % effective concentration) was defined as the compound concentration required to inhibit HIV-induced giant cell formation by 50 %.

Cytostatic activity assays

All assays were performed in 96-well microtiter plates. To each well were added $(5-7.5) \times 10^4$ tumour cells and a given amount of the test compound. The cells were allowed to proliferate for 48 h (murine leukemia L1210 cells) or 72 h (human lymphocytic CEM and human cervix carcinoma HeLa cells) at 37 °C in a humidified CO₂-controlled atmosphere. At the end of the incubation period, the cells were counted in a Coulter counter. The IC₅₀ (50 % inhibitory concentration) was defined as the concentration of the compound that inhibited cell proliferation by 50 %. Acknowledgments The authors are grateful to Leen Ingels, Leentje Persoons, Frieda De Meyer, Lizette van Berckelaer, Anita Camps, Steven Carmans and Lies Van den Heurck for excellent technical assistance. The work was supported by the KU Leuven (GOA 10/14) and by the Medical University of Lodz (502-34-009 and 503/3-014-01/503-1).

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