REVIEW



Leptomeningeal Disease: Current Approaches and Future Directions

Ugur Sener^{1,2} • Jessica A. Wilcox³ • Adrienne A. Boire^{3,4}

Accepted: 2 March 2025 © The Author(s) 2025

Abstract

Purpose of Review Leptomeningeal disease (LMD), or spread of cancer cells into the pia and arachnoid membranes encasing the brain and spinal cord, is associated with high symptom burden and poor survival at 2 to 5 months. Conventional treatments including photon-based radiation therapy, systemic chemotherapy, and intrathecal chemotherapy demonstrate limited efficacy. Despite significant successes for a range of solid tumors, immunotherapy has not yet demonstrated significant efficacy in management of LMD. Advances in understanding of LMD pathophysiology, improved diagnostics, and novel therapeutics are shifting this paradigm. In this article, we review diagnostic and treatment challenges associated with LMD.

Recent Findings We discuss the use of novel cerebrospinal fluid (CSF) analysis techniques such as circulating tumor cell and CSF cell-free DNA assessment to overcome limitations of conventional diagnostic modalities. We then review advances in treatment including clinical trial data demonstrating efficacy of proton craniospinal radiation to treat the entire neuroaxis. We discuss emerging data regarding targeted therapeutics conferring durable survival benefit.

Summary Novel therapeutics and combinatorial treatment approaches will likely further improve outcomes for patients with LMD.

 $\textbf{Keywords} \ \ Leptomeningeal \ disease \cdot Leptomeningeal \ metastases \cdot Craniospinal \ radiation \cdot Intrathecal \ therapy \cdot Central \ nervous \ system \ metastases \cdot Cerebrospinal \ fluid$

Introduction

Leptomeningeal disease (LMD) describes spread of cancer cells into the cerebrospinal fluid (CSF)-filled membranes encasing the brain and spinal cord [1, 2]. LMD describes a particularly destructive pattern of cancer progression associated with median overall survival (mOS) of 2 to 5 months [3]. Reflecting the diffuse nature of disease location, LMD

Adrienne A. Boire boirea@mskcc.org

Published online: 18 March 2025

- Department of Neurology, Mayo Clinic, Rochester, MN, USA
- Department of Medical Oncology, Mayo Clinic, Rochester, MN, USA
- Department of Neurology, Memorial Sloan Kettering Cancer Center, New York, NY, USA
- Brain Tumor Center, Human Oncology and Pathogenesis Program, Memorial Sloan Kettering Cancer Center, New York, NY, USA

is associated with significant symptom burden including headache, seizures, cranial neuropathies, weakness, and gait difficulty [4]. LMD related to solid tumors most commonly occurs in the setting of breast cancer, lung cancer, and melanoma; though in practice, any tumor may result in LMD [1, 5]. LMD has been reported at initial cancer diagnosis or as part of initial intracranial involvement in 2–12% of cases, but more often occurs as a late complication [6]. Prognosis of LMD varies by primary tumor type and tumor molecular markers [2]. For example, LMD related to breast cancer is generally associated with superior survival than that observed with lung cancers [5–8]. Within breast cancer subtypes, LMD related to human epidermal growth factor receptor 2 (HER2) and hormone receptor positive breast cancer carries a better prognosis compared to triple negative tumors [7]. Treatment approaches for LMD include radiation therapy, molecularly targeted systemic therapy in applicable cases, immunotherapy, systemic chemotherapy, and intrathecal chemotherapy [9]. In this review we will discuss recent advances in pathophysiology, diagnosis, and treatment of LMD.

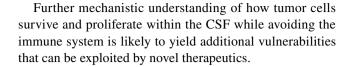


Pathophysiology of Leptomeningeal Disease

The leptomeninges consist of the pia and arachnoid membranes. The arachnoid is laminated to the dural surface, forming the outer limit of the central nervous system. The pia adhere to the cortical surface. Interaction between the pia and astrocytic endfeet leads to generation of extracellular matrix forming the glia limitans. Circulating CSF, generated by the choroid plexuses (and, to a lesser extent, the ependyma) fills the ventricular system and the space between these two membranes. Once cancer cells gain access to this space and successfully grow within the space, LMD results [9]. Mechanisms of tumor cell entry into the CSF include hematogenous dissemination through the choroid plexus or retrograde venous extension [10, 11]. Tumor cells may also directly seed the CSF through parenchymal brain or spinal cord metastases [11]. Upregulation of complement component 3 (C3) has been identified as a critical mechanism through which cancer cells activate the C3a receptor in the choroid plexus, thereby disrupting the blood-CSF barrier [11]. Although C3a is necessary for cancer cell entry through the barrier, it is not sufficient to mediate entry of tumor cells into the space. Mechanism(s) governing cancer cell entry into the leptomeninges represent an active area of basic research.

Within the leptomeningeal space, LMD generates two phenotypes [12]. Tumor cells may adhere to the leptomeninges, creating a plaque-like appearance that can be detected on magnetic resonance imaging (MRI) of the neuroaxis. Alternatively, tumor cells may be free-floating, detected by CSF cytology. LMD related to free-floating tumor cells has been associated with poorer survival compared to plaque-like adherent LMD in the setting of breast and lung cancer [12]. Similarly, in a prior retrospective study of 225 patients with LMD, cytology positive for tumor cells was noted as a negative prognostic factor [3]. In the same study, among patients with cytology negative for tumor cells, presence of nodular disease on MRI was a negative prognostic factor compared to radiographic linear enhancement on MRI [3].

Tumor cells in the leptomeninges are faced with an unfavorable environment with profound nutritional scarcity. One mechanism that has been identified to facilitate cancer cell survival within the space is through expression of iron-scavenging protein and receptor pair, lipocalin-2 (LCN2) and SCL22A17 [13]. Cancer cells secrete LCN2 into the CSF to sequester iron from CSF macrophage, thereby impairing macrophage function and supporting metastatic growth [13]. Following promising results from disruption of this pathway in preclinical models, intrathecal administration of the iron chelator deferoxamine for treatment of LMD is currently under investigation (NCT05184816) [14].



Diagnostic Advances in Leptomeningeal Disease

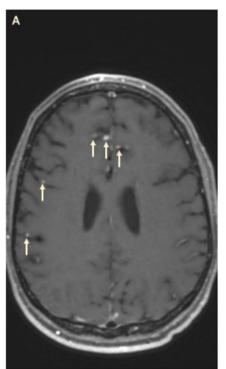
Because LMD encompasses the entire leptomeningeal space, diagnostic and monitoring evaluation of LMD requires imaging of the entire neuroaxis as well as CSF analysis [9]. On conventional MRI, intracranial LMD can appear as contrast enhancement along cranial nerves, surrounding the brainstem, and within the cerebellar folia (Fig. 1) [15]. Along the spinal cord, LMD can appear as enhancement over the cord surface or as clumping and enhancement of the cauda equina or individual nerve roots [15]. However, radiographic findings related to LMD can be subtle, and the differential diagnosis for leptomeningeal enhancement is broad. Moreover, a normal MRI of the neuroaxis does not exclude the diagnosis [2].

Radiographic assessment of LMD also represents a challenge for determination of disease progression and response to treatment. The Response Assessment in Neuro-Oncology (RANO) criteria for LMD have been proposed to address this issue and improve radiographic disease assessment [16]. However, these criteria have not yet been prospectively validated in their entirety. Perhaps due to the complexity of these criteria, their use in routine clinical practice has been limited [17]. More recently, a new imaging scorecard was validated in a small cohort of 20 patients [18]. Joint guidelines from the European Association of Neuro-Oncology (EANO) and European Society for Medical Oncology (ESMO) are also available for follow up and disease assessment [3, 19, 20].

The CSF biochemical and cytological profile is abnormal in the setting of LMD, with lymphocytic pleocytosis, elevated protein, and decreased glucose representing the most common findings [21]. Conventional cytology allows for direct visualization of tumor cells within CSF and is the gold standard for diagnosis of LMD. However, yield from a single CSF sample is low and the false negative rate for conventional cytology is high [15]. While sensitivity of CSF cytology is improved with repeated sampling, this can be cumbersome for patients and delay diagnosis [22].

Novel approaches for CSF evaluation include CSF circulating tumor cell (CTC) and cell-free tumor DNA (ctDNA) assessment [2]. CTCs can be detected based on surface protein expression. As an example, epithelial cell adhesion molecule (EpCAM) is expressed on the surface of epithelial cells, including tumor cells of breast or lung cancer origin [23–25]. Thus, EpCAM-based assays can be used for detection of CTCs in the CSF. Similarly, High-Molecular





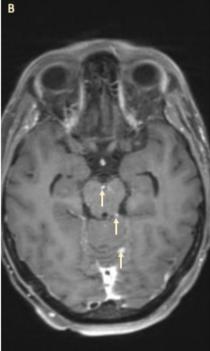




Fig. 1 Intracranial and Spinal Appearance of Leptomeningeal Disease. **A** MRI brain with contrast demonstrating multiple leptomeningeal deposits (arrows). **B** MRI brain with contrast demonstrating leptomeningeal deposits involving the brainstem and cerebellum

(arrows). **C** MRI of the lumbar spine with contrast demonstrating diffuse enhancement along the spinal cord (yellow arrows) and cauda equina nerve roots (red arrows)

Weight-Melanoma-Associated Antigen/Melanoma-associated Chondroitin Sulfate Proteoglycan (HMW-MAA/MCSP) has been explored for detection of CTCs related to melanoma [26]. In a prospective study of patients with breast cancer, CSF CTC detection demonstrated superior sensitivity in comparison to traditional cytology to detect LMD [27]. Similar findings were reported in lung cancer LMD; CTC sensitivity was 94%, cytology 76% [28].

In addition to commercially available cell-surface protein-based assays, flow cytometry can also establish the diagnosis of LMD. In this technique, fluorescently labeled antibodies against cell surface proteins of interest detect of CSF CTCs [29]. Use of flow cytometry for detection of LMD from solid tumors remains extremely limited; surface antigens are less established in these tumors. However, in hematologic malignancies, this technique is standard-of-care [30].

CSF CTC measurement also provides a quantitative measure of LMD burden with potential clinical utility for treatment response assessment and detection of disease progression. As an example, in 15 patients with LMD related to breast cancer, higher CSF CTC enumeration was associated with more rapid disease progression [31]. Importantly, high CSF CTC burden did not necessarily correlate with more pronounced radiographic appearance of LMD, underscoring the ability of this quantitative biomarker assessment

to provide diagnostic, prognostic, and response assessment information [31]. In a subsequent retrospective study, increased CSF CTC count was associated with increased risk of mortality as a continuous variable. Greater than or equal to 61 CSF-CTCs per 3 ml of CSF was identified as the optimal cutoff, at which risk of mortality doubled [32]. However, limitations of CSF CTC quantification include limited availability of commercial assays at most centers, possibility of epithelial-to-mesenchymal transformation theoretically reducing the yield of EpCAM-based assays, and need to develop assays or flow cytometry techniques for surface proteins associated with different malignancies [2].

Tumor cells within the CSF shed DNA, which can be detected by CSF ctDNA analysis [33]. As an example, in a prior study of 43 CSF samples from 22 patients with LMD, cytology was positive for malignant cells in 72% of the samples, compared to 93% positive for CSF ctDNA [34]. CSF ctDNA has successfully been employed for diagnosis and classification of primary CNS tumors [35, 36]. The quantitative nature of CSF ctDNA may have utility in response assessment and prognostication [37]. However, the qualitative and descriptive aspects of CSF ctDNA analysis are of perhaps the greatest value, enabling identification of targetable mutations with potentially actionable targeted therapies or identifying divergent mutational profiles between CNS



and the primary tumor site [33]. For instance, a study of patients with lung cancer harboring epithelial growth factor (EGFR) mutations revealed unique mutations and copy number variations in CSF ctDNA not detected in analysis of the primary tumor [38]. In a more recent study, analysis of CSF ctDNA from patients with LMD secondary to lung cancer captured molecular characteristics and heterogeneity of metastatic disease [39]. Thus, examination of CSF ctDNA may identify resistance mechanisms and suggest therapeutic strategies unique to the CNS compartment [2]. Limitations to the use of CSF ctDNA include lack of standardized processes for DNA extraction, limited availability of the testing for widespread use and the processing time required that may preclude clinical decision-making based on results [33]. Furthermore, ctDNA may be shed into the CSF from both parenchymal brain metastases and LMD. Thus, the presence of CSF ctDNA does not provide diagnostic certainty. Optimized protocols for CSF ctDNA isolation are under development and clinical utility of this test for molecular characterization of LMD is likely to increase [40].

Further advances in advanced diagnostic tools for detection and monitoring of LMD may in time permit earlier detection, improved diagnostic certainty, and more widely available quantitative response assessment. Utilization of CSF next-generation sequencing has the potential to identify resistance mechanisms, informing development of new therapies. Table 1 summarizes emerging CSF diagnostic strategies for LMD.

Therapeutic Advances in Leptomeningeal Disease

Joint EANO and ESMO guidelines provide guidance for management based on tumor type, presence of brain metastases, and extent of systemic disease [19, 20]. Treatment of LMD primarily involves radiation therapy to the symptomatic sites followed by CNS-penetrant systemic therapy. Due to the diffuse nature of LMD, surgical resection is

not possible. In select cases, ventriculoperitoneal shunting (VPS) can be considered for palliation of symptoms related to obstructive hydrocephalus [41]. In a study of 190 patients with LMD who underwent CSF diversion, symptomatic relief was noted among 83% [42]. Complications were uncommon with 5% of patients experiencing a shunt related infection and 6.3% having a subdural hygroma/hematoma [42]. Peritoneal carcinomatosis secondary to VPS is exceedingly rare [43]. Recently, combination of an occludable VPS with IT chemotherapy was retrospectively associated with better survival compared to occludable shunt alone [44].

Radiation Therapy

Historically, LMD has been treated with photon-based whole brain radiation therapy (WBRT) and focal spinal radiation therapy (RT) to symptomatic sites of disease [45]. The disadvantage of this technique is that it does not address disease within the entire CSF compartment, and therefore has not been associated with a survival benefit. Photon-based craniospinal irradiation (CSI) is an option, but this is associated with profound toxicities in adults and is not recommended [46].

Proton CSI has emerged as a more tolerable alternative to photon CSI to treat the entire neuroaxis [47]. Rather than the progressive dose fall-off seen with photon radiation, protons deposit most of their energy within a few millimeters of their end range, thereby eliminating exit radiation dose and limiting off-target toxicities [48]. The resultant efficient and focused delivery of RT to the craniospinal axis largely spares the vertebral bone marrow, reducing risk of myelotoxicity. Anterior torso organ systems receive less radiation with proton compared to photon CSI, resulting in less toxicity to these organs.

Safety and tolerability of proton CSI for patients with LMD from solid tumors was established in a phase I clinical trial [47]. In this study, among 24 patients with LMD treated with proton CSI, median central nervous system (CNS) progression free survival (PFS) was 7 months and mOS was

Table 1 Emerging Cerebrospinal Fluid Analytic Strategies for Leptomeningeal Disease

	CTC Analysis	ctDNA Analysis
Technique Advantages	Tumor cell detection based on expression of surface proteins • Improved sensitivity compared to conventional cytology • Quantitative assessment allows longitudinal monitoring of therapeutic response • Potential prognostic value based on quantity of CTCs	Sequencing of DNA released from tumor cells into the CSF • Potential to detect targetable tumor genetic alterations • Potential to detect mutational divergence between CNS and extracranial tumor cells
Disadvantages	 Limited availability Requires specialized equipment Individual assays specific to individual tumor types 	 Poor specificity: May also originate from parenchymal brain metastases, limiting diagnostic attribution Limited availability Lack of standardization Prolonged processing time



8 months [47]. The most common toxicities were fatigue and self-limiting lymphopenia and thrombocytopenia.

In a subsequent clinical trial, proton CSI was compared directly to involved field photon radiotherapy (IFRT) [49]. IFRT was defined as WBRT and/or focal spinal RT to symptomatic areas. Forty-two patients treated with proton CSI were compared to 21 patients treated with IFRT. A significant benefit in CNS PFS was observed with proton CSI (median 7.5 months; 95% CI, 6.6 months to not reached) compared with IFRT (2.3 months; 95% CI, 1.2 to 5.8 months; P, 0.001). OS for proton CSI was 9.9 months (95% CI, 7.5 months to not reached) compared to 6.0 months for IFRT (95% CI, 3.9 months to not reached; P 5 0.029).

Importantly, the proton CSI trials enrolled a heterogeneous population of patients, some with actionable tumor genetic mutations. In practice, toxicities (particularly highgrade fatigue) are most prominent in patients greater than 60 years of age, or those that are heavily pretreated. Further, limited availability of centers capable of delivering proton therapy limits generalizability of the findings. Nevertheless, the results of the phase 1 and phase 2 trials are promising and represent a significant advancement in treatment of LMD. Proton CSI should be considered for select patients with LMD when possible, particularly for those in whom the goal of treatment is durable disease control. Sequencing CNS-active systemic therapy with proton CSI may further improve patient outcomes [50, 51].

Conventional Systemic Therapies

LMD is often encountered in the setting of progressive extracranial disease [20]. As such, systemic therapies that can target both extracranial and CNS compartments represent an attractive option. However, systemically-administered, cytotoxic chemotherapies have demonstrated limited efficacy in treatment of LMD. High-dose IV methotrexate is among the most frequently employed approaches but is associated with a modest survival benefit [52]. Despite frequent use in other CNS tumors, single agent temozolomide has not demonstrated efficacy for LMD related to solid tumors [53]. Capecitabine alone or in combination with trastuzumab represents one tumor-specific systemic therapy option with modest benefit in treatment of LMD related to breast and esophageal cancer [54, 55].

Intrathecal Therapy

Intrathecal (IT) chemotherapy describes direct administration of chemotherapy into the CSF space by a lumbar puncture or through the use of an intraventricular catheter (e.g., Ommaya reservoir) [2]. To safely receive IT chemotherapy, patients must have non-bulky disease with no CSF flow obstruction [21]. In a retrospective study,

intraventricular therapy via Ommaya catheter was associated with better OS compared to IT chemotherapy administered into the lumbar cistern [56].

Methotrexate, cytarabine, topotecan, pemetrexed, and thiotepa represent the most commonly administered IT chemotherapy agents [2]. The mOS from early studies of IT chemotherapy range from 2-4 months. However, comparative clinical trials are lacking, optimal dosing regimens are unknown, and the patient population from these historical trials do not reflect the modern era of cancer-directed therapies [57–61]. More modern trials of IT chemotherapies are suggestive of better survival outcomes [62–65]. A common pattern of IT chemotherapy administration in clinical practice involves use of methotrexate, cytarabine, topotecan, or thiotepa twice weekly for four weeks [66]. If cytologic response is favorable following the initial four-week period, treatment can continue once weekly for four weeks and then once monthly for maintenance.

IT chemotherapy can be associated with toxicities related to chemical meningitis including fever, headaches, and nausea [66]. IT methotrexate is associated with leukoencephalopathy in certain cases, particularly in the post-RT setting [67]. Very limited data is available for combinatorial IT chemotherapy strategies, such as a small study of cytarabine and methotrexate combination demonstrating improved disease control compared to methotrexate alone [68]. In another study investigating combinatorial therapy, IT liposomal cytarabine combined with systemic chemotherapy was associated with improved survival compared to systemic chemotherapy alone among patients with LMD related to breast cancer [62]. On the other hand, a study of IT methotrexate or cytarabine combined with systemic chemotherapy compared to systemic chemotherapy demonstrated no added benefit from IT chemotherapy among patients with LMD related to breast cancer [61].

IT administration of immunotherapy is a consideration, but published experience remains limited to small studies and case reports. In a retrospective report of 43 patients with LMD from melanoma treated with IT interleukin-2 (IL-2), mOS was 7.8 months but was accompanied by a high rate of symptomatic intracranial hypertension [69]. A subset of patients achieved long-term disease control with 5-year OS rate reported at 13%. These findings suggest a subset of patients with LMD secondary to melanoma may derive significant benefit from IT IL-2, but patient or tumor-specific factors contributing to this prolonged survival remain unclear. Combination of IT immune checkpoint inhibitor (ICI) therapy with intravenous ICI is an emerging treatment option for ICI-sensitive cancers [70]. Interim results from a phase 1 study of IT plus IV nivolumab in patients with melanoma LMD demonstrated a preliminary mOS of 4.9 months with tolerable safety data [70, 71].



Similarly, IT administration of molecularly targeted therapies may be a consideration in select settings. Administration of IT trastuzumab was evaluated in a clinical trial for patients with LMD related to HER2-positive breast cancer [72]. Thirty-four patients with LMD were treated. During the phase 1 portion of the study, mOS for patients treated with the 80 mg dose selected for phase 2 was 8.3 months; mOS during the phase 2 portion of the study was 10.5 months [72]. In another study, 7 patients with HER2-positive breast cancer received IT trastuzumab [73]. mOS was 13.7 months in the IT trastuzumab group, compared to 9.3 months among patients who did not receive IT therapy [73]. IT trastuzumab in the setting of HER2 positive esophageal carcinoma was also evaluated, but data is very limited with one patient experiencing durable disease control and the other experiencing rapid progression and short survival [74]. While these findings were encouraging, a meta-analysis of 45 publications representing 208 patients identified no difference in mOS between IT trastuzumab compared to oral or IV administration of HER2 targeted therapy [75].

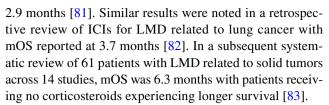
Based on current evidence, use of conventional IT therapy for LMD is best reserved for patients with reasonable performance status, limited CNS-penetrant systemic therapy options, reasonably controlled extracranial disease, and/or in the context of a clinical trial. The potential utility of IT-based immunotherapy for melanoma and HER2 targeted therapy for LMD related to HER2-positive cancer is additionally encouraging. Further studies may better delineate a role for IT therapy for selected subpopulations of patients based on patient and tumor characteristics.

Immunotherapy

Checkpoint regulators such as programmed cell death protein 1 (PD-1) downregulate T cell activation [76]. ICIs such as the anti-PD1 agent pembrolizumab work by blocking these checkpoints with intent to generate a sustained antitumor immune response. ICIs have proven efficacy in numerous systemic malignancies including lung cancer, breast cancer, and melanoma [77]. Despite these successes, the role of ICIs for LMD remain limited.

Two clinical trials have evaluated pembrolizumab in mixed tumor populations of LMD [78, 79]. In the first study, 13 patients with LMD were treated with pembrolizumab with median OS at 4.9 months [78]. In the second study, 20 patients with LMD were treated with pembrolizumab giving an OS of 3.6 months, for a three-month OS of 60% [79].

In a study of melanoma patients with brain metastases, four patients with LMD were treated with anti-PD1 ICI nivolumab monotherapy [80]. No patients responded to treatment. In another study combining anti-cytotoxic T-lymphocyte-associated protein 4 (CTLA-4) ICI ipilimumab and nivolumab for 18 patients with LMD, median OS was



Taken together, these mOS data from small studies with ICI monotherapy for management of LMD are comparable to mOS associated with conventional treatments such as systemic or intrathecal chemotherapy. To date, ICIs have not satisfactorily improved survival from LMD as single agents. Combinatorial strategies such as use of ICIs with proton CSI, use of dual ICI, or combinations of other systemic and IT therapies with ICIs are currently under investigation.

Molecularly Derived Targeted Therapies

Systemic therapies targeting driver mutations have remarkable efficacy in management of a range of malignancies. As an example, EGFR and anaplastic lymphoma kinase (ALK) targeted therapies have dramatically altered management of non-small cell lung cancer (NSCLC). V-raf murine sarcoma viral oncogene homolog B1 (BRAF) and mitogen-activated protein kinase (MEK) inhibitors are used in treatment of melanoma and other BRAF-mutant tumors. These agents have also demonstrated efficacy in LMD related to solid tumors.

EGFR-targeted tyrosine kinase inhibitor (TKI) osimertinib has demonstrated significant efficacy in treatment of LMD related to EGFR-mutant NSCLC [84–86]. In the phase 1 BLOOM study, 41 patients with LMD secondary to EGFR-mutant NSCLC were treated with osimertinib with an mOS of 11 months [84]. Similarly, within the AURA program, 22 patients with LMD secondary to EGFR-mutant NSCLC were treated with osimertinib and the mOS was 11.1 months [85]. In a recent meta-analysis of 243 patients and 282 lines of EGFR-TKI for patients with LMD from NSCLC, median OS was 14.5 months [86]. Osimertinib was associated with improved outcomes compared to other EGFR TKIs [86].

While early-generation ALK inhibitor crizotinib had limited CNS activity, the dual ALK and ROS1 inhibitor lorlatinib has been associated with CNS responses ranging from 8 to 22 months [87]. Among 11 patients with LMD due to NSCLC with ALK (n=9) or ROS1 (n=2) mutations, lorlatinib therapy demonstrated an intracranial objective response rate (ORR) of 45%; survival data was not provided [88]. Responses to the ALK inhibitor alectinib have also been reported in setting of LMD secondary to NSLC [89, 90].

Responses to BRAF/MEK inhibitor combinations dabrafenib/trametinib and encorafenib/binimetinib have been reported [91–93]. While individual case reports have indicated some instances of durable responses, patients often experience progression on these targeted therapeutics at



the time of LMD diagnosis, limiting their clinical utility specifically for management of LMD due to BRAF-mutant tumors [2].

For patients with LMD related to HER2-positive breast cancer, the antibody drug conjugate trastuzumab deruxtecan has been associated with durable responses despite uncertain CSF penetration [94, 95]. In a retrospective study of 8 patients with LMD secondary to HER2-positive breast cancer, mOS was 10.4 months [95]. In the ROSET-BM study, among 19 patients with radiographic LMD, 12-month overall survival rate was 87.1% though cytologic-positivity and responses were not reported [94]. The selective HER2 inhibitor tucatinib in combination with trastuzumab and capecitabine was associated with improved CNS disease control and survival (CNS-PFS = 9.9 mo; mOS = 18.1 mo) compared to trastuzumab and capecitabine without tucatinib (CNS-PFS = 4.2 mo; mOS = 12.0 mo). While large-scale data is lacking, a PFS of 7 months and OS of 10 months was reported for a patient with LMD secondary to HER2 positive breast cancer treated with tucatinib [96]. Furthermore, preliminary results of a phase 2 study of tucatinib, capecitabine, and trastuzumab in newly diagnosed LMD suggest an LM-ORR of 38% and 100% of patients achieving clinical benefit [97]. Tucatinib, like other small molecule inhibitors, has documented CSF penetration [98],

Another potential therapeutic approach may be trastuzumab deruxtecan, an antibody drug conjugate consisting of anti-HER2 antibody trastuzumab linked to the topoisomerase I inhibitor deruxtecan. Among 19 patients with breast cancer related LMD, trastuzumab-deruxtecan was associated with 12-month survival rate of 87.1%, though CSF cytology data was not reported in this study and clinical experience with the antibody–drug conjugate is so far lacking [94]. It remains to be determined whether combined administration of IT trastuzumab with systemic HER2-acting agents such as tucatinib or trastuzumab-deruxtecan is feasible or associated with any additional benefit.

The vascular endothelial growth factor (VEGF) inhibitor bevacizumab is frequently used in treatment of primary CNS tumors as a steroid-sparing agent for management of symptomatic cerebral edema, even though it is not associated with an OS benefit [99]. Among patients with LMD, high levels of VEGF have been negatively correlated with survival [100]. In a prior study, combination of bevacizumab, etoposide, and cisplatin was associated with CNS response and improved OS in setting of LMD related to breast cancer [101]. Given withdrawal of bevacizumab approval for breast cancer, this combination is not frequently used in the clinic [102]. Similarly, possible benefit from combination of erlotinib with bevacizumab was reported for LMD secondary to EGFR-mutant NSCLC [103]. However, given subsequent development of and clear benefit associated with

CNS-penetrant EGFR inhibitor osimertinib, this combination is unlikely to be used clinically.

Clinical experience with molecularly targeted therapies to date indicates clear benefits from CNS-penetrant agents such as osimertinib, lorlatinib, tucatinib, and trastuzumab deruxtecan. Many patients with primary tumors harboring actionable mutations will have already received targeted therapy at the time of LMD diagnosis. Nevertheless, for patients who present with LMD at the time of initial tumor diagnosis or treated with an earlier generation TKI and experiencing progression, targeted therapies represent viable treatment options.

Conclusion

LMD remains a challenging complication of malignancy resulting in significant morbidity and mortality. Significant advances have been made in diagnosis and treatment of LMD over the past decade, suggesting potential for change. Emerging diagnostic tools such as CTCs and CSF ctDNA provide earlier diagnosis, prior to the onset of irreversible neurological damage. These tools also provide novel ways to monitor the course of LMD, permitting earlier detection of treatment response and guiding personalized treatment based on tumor molecular profiles.

Advances in radiation therapy, notably proton CSI, provide a treatment option for the entire CSF compartment for select patients. Combination proton CSI with systemic therapy may further improve outcomes and is an area of active investigation. Available clinical trial data has demonstrated the potential of CNS-penetrant targeted therapies in LMD. With improved understanding of tumor growth mechanisms and LMD pathophysiology, further systemic therapy options are likely to be developed.

Given the wealth of treatment resistance mechanisms already established in solid tumor malignancies, regardless of primary or site of metastasis, it is unlikely that any single treatment modality will sufficiently treat LMD and achieve long-term survival. Combinatorial strategies individualized to individual tumor profiles and adjusted over the disease course have the highest likelihood to improve local disease control, symptom burden, and survival.

Key References

• Lamba N, Cagney DN, Catalano PJ, Elhalawani H, Haas-Kogan DA, Wen PY, Wagle N, Lin NU, Aizer AA, Tanguturi S (2023) Incidence proportion and prognosis of leptomeningeal disease among patients with breast vs. non-breast primaries. Neuro



Oncol 25 (5):973–983. https://doi.org/10.1093/neu-onc/noac249

- O This large single institution review represents a modern epidemiologic study of LMD, highlighting the high incidence of LMD arising from breast cancer and superior survival in patients with breast LMD as opposed to LMD from other solid cancers.
- Chi Y, Remsik J, Kiseliovas V, Derderian C, Sener U, Alghader M, Saadeh F, Nikishina K, Bale T, Iacobuzio-Donahue C, Thomas T, Pe'er D, Mazutis L, Boire A (2020) Cancer cells deploy lipocalin-2 to collect limiting iron in leptomeningeal metastasis. Science 369 (6501):276–282. https://doi.org/10.1126/science.aaz2193
- O This publication highlights an iron-scavenging pathway critical for leptomeningeal cell surivival in the nutrient-sparse CSF. Understanding the unique mechanisms of cancer dissemination and survival within the leptomeningeal space is crucial for the development of novel therapeutics against LMD.
- Le Rhun E, Devos P, Winklhofer S, Lmalem H, Brandsma D, Kumthekar P, Castellano A, Compter A, Dhermain F, Franceschi E, Forsyth P, Furtner J, Galldiks N, Gallego Perez-Larraya J, Gempt J, Hattingen E, Hempel JM, Lukacova S, Minniti G, O'Brien B, Postma TJ, Roth P, Ruda R, Schaefer N, Schmidt NO, Snijders TJ, Thust S, van den Bent M, van der Hoorn A, Vogin G, Smits M, Tonn JC, Jaeckle KA, Preusser M, Glantz M, Wen PY, Bendszus M, Weller M (2022) Prospective validation of a new imaging scorecard to assess leptomeningeal metastasis: A joint EORTC BTG and RANO effort. Neuro Oncol 24 (10):1726–1735. https://doi.org/10.1093/neuonc/noac043
- This publication outlines the validation of an updated RANO-LM imaging scorecard and represents an international collaboration between the EORTC BTG and RANO to improve response monitoring in LMD.
- Yang JT, Wijetunga NA, Pentsova E, Wolden S, Young RJ, Correa D, Zhang Z, Zheng J, Steckler A, Bucwinska W, Bernstein A, Betof Warner A, Yu H, Kris MG, Seidman AD, Wilcox JA, Malani R, Lin A, DeAngelis LM, Lee NY, Powell SN, Boire A (2022) Randomized Phase II Trial of Proton Craniospinal Irradiation Versus Photon Involved-Field Radiotherapy for Patients With Solid Tumor Leptomeningeal Metastasis. J Clin Oncol 40 (33):3858–3867. https://doi.org/10.1200/JCO.22.01148

- This phase II clinical trial of proton CSI versus IFRT is the first to demonstrate a significant survival benefit from a radiation technique in the treatment of LMD.
- Glitza Oliva IC, Ferguson SD, Bassett R, Jr., Foster AP, John I, Hennegan TD, Rohlfs M, Richard J, Iqbal M, Dett T, Lacey C, Jackson N, Rodgers T, Phillips S, Duncan S, Haydu L, Lin R, Amaria RN, Wong MK, Diab A, Yee C, Patel SP, McQuade JL, Fischer GM, McCutcheon IE, O'Brien BJ, Tummala S, Debnam M, Guha-Thakurta N, Wargo JA, Carapeto FCL, Hudgens CW, Huse JT, Tetzlaff MT, Burton EM, Tawbi HA, Davies MA (2023) Concurrent intrathecal and intravenous nivolumab in leptomeningeal disease: phase 1 trial interim results. Nat Med 29 (4):898–905. https://doi.org/10.1038/s41591-022-02170-x
- O The interim results from a phase I study of IV and IT nivolumab detail the emerging use of intrathecal immune checkpoint inhibition in the treatment of LMD.
- Kumthekar PU, Avram MJ, Lassman AB, Lin NU, Lee E, Grimm SA, Schwartz M, Bell Burdett KL, Lukas RV, Dixit K, Perron I, Zhang H, Gradishar WJ, Pentsova EI, Jeyapalan S, Groves MD, Melisko M, Raizer JJ (2023) A phase I/II study of intrathecal trastuzumab in human epidermal growth factor receptor 2-positive (HER2-positive) cancer with leptomeningeal metastases: Safety, efficacy, and cerebrospinal fluid pharmacokinetics. Neuro Oncol 25 (3):557–565. https://doi.org/10.1093/neuonc/noac195
- O This phase I/II study of IT trastuzumab demonstrates the successful conversion of a non-CNS penetrant targeted therapy to an intraventricular formulation for the treatment of LMD.

Acknowledgements • This publication was supported by Grant Number UL1 TR002377 from the National Center for Advancing Translational Sciences (NCATS). Its contents are solely the responsibility of the authors and do not necessarily represent the official views of the NIH.

- This research was funded in part through the NIH/NCI Cancer Center Support Grant P30 CA008748 and 1R01CA245499-01A1.
- We are deeply grateful to our patients who continue to inspire us with their generosity and grace in the face of an uncertain future

Author Contributions J.A.W and A. A. B conceived the manuscript, U.S. prepared Fig. 1. U.S., J.A.W., and A.A.B wrote the manuscript. A.A.B edited and prepared manuscript for publication. All authors reviewed the manuscript.

Data Availability No datasets were generated or analysed during the current study.



Declarations

Statements and Declarations This publication was supported by Grant Number UL1 TR002377 from the National Center for Advancing Translational Sciences (NCATS) to Ugur Sener.

This work was supported by National Institute of Health/National Cancer Institute P30 CA008748 and 1R01CA245499-01A1 to Adrienne A. Boire.

Competing Interests • Ugur Sener reports: Servier (Mayo Clinic Glioma Center of Excellence Advisory Board – paid).

• Adrienne Boire reports: Evren Technologies (scientific advisory board - unpaid), Apellis Pharmaceuticals (consultant), Patents (inventor) via Sloan Kettering Institute..

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

References

- Clarke JL, Perez HR, Jacks LM, Panageas KS, Deangelis LM. Leptomeningeal metastases in the MRI era. Neurology. 2010;74(18):1449-54. https://doi.org/10.1212/WNL.0b013 e3181dc1a69.
- Sener U, Kumthekar P, Boire A. Advances in the diagnosis, evaluation, and management of leptomeningeal disease. Neurooncol Adv. 2021;3(Suppl 5):v86–95. https://doi.org/10.1093/noajnl/vdab108.
- Le Rhun E, Devos P, Weller J, Seystahl K, Mo F, Compter A, Berghoff AS, Jongen JLM, Wolpert F, Ruda R, Brandsma D, van den Bent M, Preusser M, Herrlinger U, Weller M. Prognostic validation and clinical implications of the EANO ESMO classification of leptomeningeal metastasis from solid tumors. Neuro Oncol. 2021;23(7):1100–12. https://doi.org/10.1093/neuonc/ noaa298.
- Pan Z, Yang G, He H, Yuan T, Wang Y, Li Y, Shi W, Gao P, Dong L, Zhao G. Leptomeningeal metastasis from solid tumors: clinical features and its diagnostic implication. Sci Rep. 2018;8(1):10445. https://doi.org/10.1038/s41598-018-28662-w.
- Lamba N, Cagney DN, Catalano PJ, Elhalawani H, Haas-Kogan DA, Wen PY, Wagle N, Lin NU, Aizer AA, Tanguturi S. Incidence proportion and prognosis of leptomeningeal disease among patients with breast vs. non-breast primaries. Neuro Oncol. 2023;25(5):973–83. https://doi.org/10.1093/neuonc/noac249.
- Lamba N, Wen PY, Aizer AA. Epidemiology of brain metastases and leptomeningeal disease. Neuro Oncol. 2021;23(9):1447–56. https://doi.org/10.1093/neuonc/noab101.
- Abouharb S, Ensor J, Loghin ME, Katz R, Moulder SL, Esteva FJ, Smith B, Valero V, Hortobagyi GN, Melhem-Bertrandt A. Leptomeningeal disease and breast cancer: the importance of tumor subtype. Breast Cancer Res Treat. 2014;146(3):477–86. https://doi.org/10.1007/s10549-014-3054-z.

- Lee SJ, Lee JI, Nam DH, Ahn YC, Han JH, Sun JM, Ahn JS, Park K, Ahn MJ. Leptomeningeal carcinomatosis in non-small-cell lung cancer patients: impact on survival and correlated prognostic factors. J Thorac Oncol. 2013;8(2):185–91. https://doi.org/10. 1097/JTO.0b013e3182773f21.
- Wilcox JA, Li MJ, Boire AA. Leptomeningeal Metastases: New Opportunities in the Modern Era. Neurotherapeutics. 2022;19(6):1782-98. https://doi.org/10.1007/s13311-022-01261-4.
- Yao H, Price TT, Cantelli G, Ngo B, Warner MJ, Olivere L, Ridge SM, Jablonski EM, Therrien J, Tannheimer S, McCall CM, Chenn A, Sipkins DA. Leukaemia hijacks a neural mechanism to invade the central nervous system. Nature. 2018;560(7716):55– 60. https://doi.org/10.1038/s41586-018-0342-5.
- Boire A, Zou Y, Shieh J, Macalinao DG, Pentsova E, Massague J. Complement Component 3 Adapts the Cerebrospinal Fluid for Leptomeningeal Metastasis. Cell. 2017;168(6):1101–13.
- Remsik J, Chi Y, Tong X, Sener U, Derderian C, Park A, Saadeh F, Bale T, Boire A. Leptomeningeal metastatic cells adopt two phenotypic states. Cancer Rep (Hoboken). 2022;5(4):e1236. https://doi.org/10.1002/cnr2.1236.
- Chi Y, Remsik J, Kiseliovas V, Derderian C, Sener U, Alghader M, Saadeh F, Nikishina K, Bale T, Iacobuzio-Donahue C, Thomas T, Pe'er D, Mazutis L, Boire A. Cancer cells deploy lipocalin-2 to collect limiting iron in leptomeningeal metastasis. Science. 2020;369(6501):276–82. https://doi.org/10.1126/science.aaz2193.
- Wilcox J, Modelevsky LR, Thomas T, Cremers S, Young RJ, Reiner AS, Panageas K, Yu HA, Boire AA (2022) A phase Ia/ Ib study of intrathecal deferoxamine in patients with leptomeningeal metastases. Journal of Clinical Oncology 40 (16_ suppl):TPS2074-TPS2074. https://doi.org/10.1200/JCO.2022. 40.16_suppl.TPS2074
- Freilich RJ, Krol G, DeAngelis LM. Neuroimaging and cerebrospinal fluid cytology in the diagnosis of leptomeningeal metastasis. Ann Neurol. 1995;38(1):51–7. https://doi.org/10.1002/ana. 410380111.
- Chamberlain M, Junck L, Brandsma D, Soffietti R, Ruda R, Raizer J, Boogerd W, Taillibert S, Groves MD, Le Rhun E, Walker J, van den Bent M, Wen PY, Jaeckle KA. Leptomeningeal metastases: a RANO proposal for response criteria. Neuro Oncol. 2017;19(4):484–92. https://doi.org/10.1093/neuonc/now183.
- Chukwueke UN, Wen PY (2019) Use of the Response Assessment in Neuro-Oncology (RANO) criteria in clinical trials and clinical practice. CNS Oncol 8 (1):CNS28. https://doi.org/10.2217/cns-2018-0007
- 18. Le Rhun E, Devos P, Winklhofer S, Lmalem H, Brandsma D, Kumthekar P, Castellano A, Compter A, Dhermain F, Franceschi E, Forsyth P, Furtner J, Galldiks N, Gallego Perez-Larraya J, Gempt J, Hattingen E, Hempel JM, Lukacova S, Minniti G, O'Brien B, Postma TJ, Roth P, Ruda R, Schaefer N, Schmidt NO, Snijders TJ, Thust S, van den Bent M, van der Hoorn A, Vogin G, Smits M, Tonn JC, Jaeckle KA, Preusser M, Glantz M, Wen PY, Bendszus M, Weller M. Prospective validation of a new imaging scorecard to assess leptomeningeal metastasis: A joint EORTC BTG and RANO effort. Neuro Oncol. 2022;24(10):1726–35. https://doi.org/10.1093/neuonc/noac043.
- Le Rhun E, Weller M, Brandsma D, Van den Bent M, De Azambuja E, Henriksson R, Boulanger T, Peters S, Watts C, Wick W, Wesseling P. EANO–ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up of patients with leptomeningeal metastasis from solid tumours. Annals of oncology. 2017 Jul 1;28:iv84-99. https://doi.org/10.1093/annonc/mdx221
- 20. E Rhun Le M Weller M Bent van den D Brandsma J Furtner R Ruda D Schadendorf J Seoane JC Tonn P Wesseling W Wick G Minniti S Peters G Curigliano M Preusser EG Committee



- clinicalguidelines@esmo.org EGCEa, 2023 Leptomeningeal metastasis from solid tumours: EANO-ESMO Clinical Practice Guideline for diagnosis, treatment and follow-up ESMO Open 8 5 101624https://doi.org/10.1016/j.esmoop.2023.101624
- 21. Nayar G, Ejikeme T, Chongsathidkiet P, Elsamadicy AA, Blackwell KL, Clarke JM, Lad SP, Fecci PE. Leptomeningeal disease: current diagnostic and therapeutic strategies. Oncotarget. 2017;8(42):73312-28. https://doi.org/10.18632/oncotarget. 20272
- 22. Leal T, Chang JE, Mehta M, Robins HI. Leptomeningeal Metastasis: Challenges in Diagnosis and Treatment. Curr Cancer Ther Rev. 2011;7(4):319–27. https://doi.org/10.2174/1573394117 97642597.
- 23. Tu Q, Wu X, Le Rhun E, Blonski M, Wittwer B, Taillandier L, De Carvalho BM, Faure GC. Cell Search technology applied to the detection and quantification of tumor cells in CSF of patients with lung cancer leptomeningeal metastasis. Lung Cancer. 2015;90(2):352-7. https://doi.org/10.1016/j.lungcan.2015. 09.008
- 24. Le Rhun E, Massin F, Tu Q, Bonneterre J, Bittencourt Mde C, Faure GC. Development of a new method for identification and quantification in cerebrospinal fluid of malignant cells from breast carcinoma leptomeningeal metastasis. BMC Clin Pathol. 2012;12:21. https://doi.org/10.1186/1472-6890-12-21.
- 25. van Bussel MTJ, Pluim D, Bol M, Beijnen JH, Schellens JHM, Brandsma D. EpCAM-based assays for epithelial tumor cell detection in cerebrospinal fluid. J Neurooncol. 2018;137(1):1–10. https://doi.org/10.1007/s11060-017-2691-6.
- 26. Le Rhun E, Tu Q, De Carvalho BM, Farre I, Mortier L, Cai H, Kohler C, Faure GC. Detection and quantification of CSF malignant cells by the Cell Search technology in patients with melanoma leptomeningeal metastasis. Med Oncol. 2013;30(2):538. https://doi.org/10.1007/s12032-013-0538-3.
- 27. Darlix A, Cayrefourcq L, Pouderoux S, Menjot de Champfleur N, Bievelez A, Jacot W, Leaha C, Thezenas S, Alix-Panabieres C. Detection of Circulating Tumor Cells in Cerebrospinal Fluid of Patients with Suspected Breast Cancer Leptomeningeal Metastases: A Prospective Study. Clin Chem. 2022;68(10):1311-22. https://doi.org/10.1093/clinchem/hvac127.
- 28. van Bussel MTJ, Pluim D, Milojkovic Kerklaan B, Bol M, Sikorska K, Linders DTC, van den Broek D, Beijnen JH, Schellens JHM, Brandsma D. Circulating epithelial tumor cell analysis in CSF in patients with leptomeningeal metastases. Neurology. 2020;94(5):e521-8. https://doi.org/10.1212/WNL.0000000000
- 29. Subira D, Simo M, Illan J, Serrano C, Castanon S, Gonzalo R, Granizo JJ, Martinez-Garcia M, Navarro M, Pardo J, Bruna J. Diagnostic and prognostic significance of flow cytometry immunophenotyping in patients with leptomeningeal carcinomatosis. Clin Exp Metastasis. 2015;32(4):383–91. https://doi.org/10. 1007/s10585-015-9716-3.
- 30. Schroers R, Baraniskin A, Heute C, Vorgerd M, Brunn A, Kuhnhenn J, Kowoll A, Alekseyev A, Schmiegel W, Schlegel U, Deckert M, Pels H. Diagnosis of leptomeningeal disease in diffuse large B-cell lymphomas of the central nervous system by flow cytometry and cytopathology. Eur J Haematol. 2010;85(6):520-8. https://doi.org/10.1111/j.1600-0609.2010.01516.x.
- 31. Malani R, Fleisher M, Kumthekar P, Lin X, Omuro A, Groves MD, Lin NU, Melisko M, Lassman AB, Jeyapalan S, Seidman A, Skakodub A, Boire A, DeAngelis LM, Rosenblum M, Raizer J, Pentsova E. Cerebrospinal fluid circulating tumor cells as a quantifiable measurement of leptomeningeal metastases in patients with HER2 positive cancer. J Neurooncol. 2020;148(3):599-606. https://doi.org/10.1007/s11060-020-03555-z.
- 32. Diaz M, Singh P, Kotchetkov IS, Skakodub A, Meng A, Tamer C, Young RJ, Reiner AS, Panageas KS, Ramanathan LV, Pentsova

- E. Quantitative assessment of circulating tumor cells in cerebrospinal fluid as a clinical tool to predict survival in leptomeningeal metastases. J Neurooncol. 2022;157(1):81-90. https://doi.org/10. 1007/s11060-022-03949-1.
- 33. Boire A. Brandsma D. Brastianos PK, Le Rhun E. Ahluwalia M, Junck L, Glantz M, Groves MD, Lee EO, Lin N, Raizer J, Ruda R, Weller M, Van den Bent MJ, Vogelbaum MA, Chang S, Wen PY, Soffietti R. Liquid biopsy in central nervous system metastases: a RANO review and proposals for clinical applications. Neuro Oncol. 2019;21(5):571-84. https://doi.org/10.1093/
- 34. White MD, Klein RH, Shaw B, Kim A, Subramanian M, Mora JL, Giobbie-Hurder A, Nagabhushan D, Jain A, Singh M, Kuter BM, Nayyar N, Bertalan MS, Stocking JH, Markson SC, Lastrapes M, Alvarez-Breckenridge C, Cahill DP, Gydush G, Rhoades J, Rotem D, Adalsteinsson VA, Mahar M, Kaplan A, Oh K, Sullivan RJ, Gerstner E, Carter SL, Brastianos PK. Detection of Leptomeningeal Disease Using Cell-Free DNA From Cerebrospinal Fluid. JAMA Netw Open. 2021;4(8):e2120040. https://doi. org/10.1001/jamanetworkopen.2021.20040.
- 35. Valerius AR, Webb MJ, Hammad N, Sener U, Malani R. Cerebrospinal Fluid Liquid Biopsies in the Evaluation of Adult Gliomas. Curr Oncol Rep. 2024;26(4):377-90. https://doi.org/ 10.1007/s11912-024-01517-6.
- 36. Afflerbach AK, Rohrandt C, Brandl B, Sonksen M, Hench J, Frank S, Bornigen D, Alawi M, Mynarek M, Winkler B, Ricklefs F, Synowitz M, Duhrsen L, Rutkowski S, Wefers AK, Muller FJ, Schoof M, Schuller U. Classification of Brain Tumors by Nanopore Sequencing of Cell-Free DNA from Cerebrospinal Fluid. Clin Chem. 2024;70(1):250-60. https://doi.org/10.1093/clinc hem/hvad115.
- 37. Escudero L, Martinez-Ricarte F, Seoane J. ctDNA-Based Liquid Biopsy of Cerebrospinal Fluid in Brain Cancer. Cancers (Basel). 2021;13(9):1989. https://doi.org/10.3390/cancers13091989.
- 38. Li YS, Jiang BY, Yang JJ, Zhang XC, Zhang Z, Ye JY, Zhong WZ, Tu HY, Chen HJ, Wang Z, Xu CR, Wang BC, Du HJ, Chuai S, Han-Zhang H, Su J, Zhou Q, Yang XN, Guo WB, Yan HH, Liu YH, Yan LX, Huang B, Zheng MM, Wu YL. Unique genetic profiles from cerebrospinal fluid cell-free DNA in leptomeningeal metastases of EGFR-mutant non-small-cell lung cancer: a new medium of liquid biopsy. Ann Oncol. 2018;29(4):945–52. https://doi.org/10.1093/annonc/mdy009.
- 39. Liu X, Mei F, Fang M, Jia Y, Zhou Y, Li C, Tian P, Lu C, Li G. Cerebrospinal fluid ctDNA testing shows an advantage over plasma ctDNA testing in advanced non-small cell lung cancer patients with brain metastases. Front Oncol. 2023;13:1322635. https://doi.org/10.3389/fonc.2023.1322635.
- 40. Song HH, Park H, Cho D, Bang HI, Oh HJ, Kim J. Optimization of a Protocol for Isolating Cell-free DNA From Cerebrospinal Fluid. Ann Lab Med. 2024;44(3):294–8. https://doi.org/10.3343/ alm.2023.0267.
- 41. Nigim F, Critchlow JF, Kasper EM. Role of ventriculoperitoneal shunting in patients with neoplasms of the central nervous system: An analysis of 59 cases. Mol Clin Oncol. 2015;3(6):1381–6. https://doi.org/10.3892/mco.2015.627.
- 42. Bander ED, Yuan M, Reiner AS, Garton ALA, Panageas KS, Brennan CW, Tabar V, Moss NS. Cerebrospinal fluid diversion for leptomeningeal metastasis: palliative, procedural and oncologic outcomes. J Neurooncol. 2021;154(3):301–13. https://doi. org/10.1007/s11060-021-03827-2.
- 43. Sener U, Elmore K, Jayaseelan K, Porter J, Marghoob A, Rosenblum MK, Haque S, Khakoo Y. Neurocutaneous melanocytosisassociated malignant melanoma presenting with peritoneal seeding. Pediatr Dermatol. 2021;38(5):1298-301. https://doi.org/10. 1111/pde.14789.



- Huntoon KM, Gasco J, Glitza Oliva IC, Ferguson SD, Majd NK, McCutcheon IE. Ventriculoperitoneal shunting with an on-off valve for patients with leptomeningeal metastases and intracranial hypertension. Neurooncol Pract. 2024;11(1):56–63. https:// doi.org/10.1093/nop/npad056.
- Buszek SM, Chung C. Radiotherapy in Leptomeningeal Disease: A Systematic Review of Randomized and Non-randomized Trials. Front Oncol. 2019;9:1224. https://doi.org/10.3389/fonc. 2019.01224.
- Barney CL, Brown AP, Grosshans DR, McAleer MF, de Groot JF, Puduvalli V, Tucker SL, Crawford CN, Gilbert MR, Brown PD, Mahajan A. Technique, outcomes, and acute toxicities in adults treated with proton beam craniospinal irradiation. Neuro Oncol. 2014;16(2):303–9. https://doi.org/10.1093/neuonc/ not155.
- 47. Yang TJ, Wijetunga NA, Yamada J, Wolden S, Mehallow M, Goldman DA, Zhang Z, Young RJ, Kris MG, Yu HA, Seidman AD, Gavrilovic IT, Lin A, Santomasso B, Grommes C, Piotrowski AF, Schaff L, Stone JB, DeAngelis LM, Boire A, Pentsova E. Clinical trial of proton craniospinal irradiation for leptomeningeal metastases. Neuro Oncol. 2021;23(1):134–43. https://doi.org/10.1093/neuonc/noaa152.
- 48. Song S, Park HJ, Yoon JH, Kim DW, Park J, Shin D, Shin SH, Kang HJ, Kim SK, Phi JH, Kim JY. Proton beam therapy reduces the incidence of acute haematological and gastrointestinal toxicities associated with craniospinal irradiation in pediatric brain tumors. Acta Oncol. 2014;53(9):1158–64. https://doi.org/10.3109/0284186X.2014.887225.
- 49. Yang JT, Wijetunga NA, Pentsova E, Wolden S, Young RJ, Correa D, Zhang Z, Zheng J, Steckler A, Bucwinska W, Bernstein A, Betof Warner A, Yu H, Kris MG, Seidman AD, Wilcox JA, Malani R, Lin A, DeAngelis LM, Lee NY, Powell SN, Boire A. Randomized Phase II Trial of Proton Craniospinal Irradiation Versus Photon Involved-Field Radiotherapy for Patients With Solid Tumor Leptomeningeal Metastasis. J Clin Oncol. 2022;40(33):3858–67. https://doi.org/10.1200/JCO.22.01148.
- Webb MJ, Breen WG, Laack NN, Leventakos K, Campian JL, Sener U (2023) Proton craniospinal irradiation with bevacizumab and pembrolizumab for leptomeningeal disease: a case report. CNS Oncol 12 (3):CNS101. https://doi.org/10.2217/ cns-2023-0005
- 51. Sener U, Webb M, Breen WG, Neth BJ, Laack NN, Routman D, Brown PD, Mahajan A, Frechette K, Dudek AZ, Markovic SN, Block MS, McWilliams RR, Dimou A, Kottschade LA, Montane HN, Kizilbash SH, Campian JL. Proton Craniospinal Irradiation with Immunotherapy in Two Patients with Leptomeningeal Disease from Melanoma. J Immunother Precis Oncol. 2024;7(1):1–6. https://doi.org/10.36401/JIPO-23-20.
- Glantz MJ, Cole BF, Recht L, Akerley W, Mills P, Saris S, Hochberg F, Calabresi P, Egorin MJ. High-dose intravenous methotrexate for patients with nonleukemic leptomeningeal cancer: is intrathecal chemotherapy necessary? J Clin Oncol. 1998;16(4):1561–7. https://doi.org/10.1200/JCO.1998.16.4. 1561.
- Segura PP, Gil M, Balana C, Chacon I, Langa JM, Martin M, Bruna J. Phase II trial of temozolomide for leptomeningeal metastases in patients with solid tumors. J Neuroncol. 2012;109(1):137–42. https://doi.org/10.1007/s11060-012-0879-3.
- Giglio P, Tremont-Lukats IW, Groves MD. Response of neoplastic meningitis from solid tumors to oral capecitabine. J Neurooncol. 2003;65(2):167–72. https://doi.org/10.1023/b:neon.0000003752.89814.ca.
- 55. Shigekawa T, Takeuchi H, Misumi M, Matsuura K, Sano H, Fujiuchi N, Okubo K, Osaki A, Aogi K, Saeki T. Successful treatment of leptomeningeal metastases from breast cancer

- using the combination of trastuzumab and capecitabine: a case report. Breast Cancer. 2009;16(1):88–92. https://doi.org/10.1007/s12282-008-0056-x.
- de Oca Montes, Delgado M, Cacho Diaz B, Santos Zambrano J, Guerrero Juarez V, Lopez Martinez MS, Castro Martinez E, Avendano Mendez-Padilla J, Mejia Perez S, Reyes Moreno I, Gutierrez Aceves A, Gonzalez Aguilar A. The Comparative Treatment of Intraventricular Chemotherapy by Ommaya Reservoir vs. Lumbar Puncture in Patients With Leptomeningeal Carcinomatosis. Front Oncol. 2018;8:509. https://doi.org/10.3389/fonc.2018.00509.
- Gutin PH, Levi JA, Wiernik PH, Walker MD. Treatment of malignant meningeal disease with intrathecal thioTEPA: a phase II study. Cancer Treat Rep. 1977;61(5):885–7.
- Hitchins RN, Bell DR, Woods RL, Levi JA. A prospective randomized trial of single-agent versus combination chemotherapy in meningeal carcinomatosis. J Clin Oncol. 1987;5(10):1655–62. https://doi.org/10.1200/JCO.1987.5.10.1655.
- Orlando L, Curigliano G, Colleoni M, Fazio N, Nole F, Martinelli G, Cinieri S, Graffeo R, Peruzzotti G, Goldhirsch A. Intrathecal chemotherapy in carcinomatous meningitis from breast cancer. Anticancer Res. 2002;22(5):3057–9.
- Groves MD, Glantz MJ, Chamberlain MC, Baumgartner KE, Conrad CA, Hsu S, Wefel JS, Gilbert MR, Ictech S, Hunter KU, Forman AD, Puduvalli VK, Colman H, Hess KR, Yung WK. A multicenter phase II trial of intrathecal topotecan in patients with meningeal malignancies. Neuro Oncol. 2008;10(2):208–15. https://doi.org/10.1215/15228517-2007-059.
- Boogerd W, van den Bent MJ, Koehler PJ, Heimans JJ, van der Sande JJ, Aaronson NK, Hart AA, Benraadt J, Vecht ChJ. The relevance of intraventricular chemotherapy for leptomeningeal metastasis in breast cancer: a randomised study. Eur J Cancer. 2004;40(18):2726–33. https://doi.org/10.1016/j.ejca.2004.08. 012.
- 62. Le Rhun E, Wallet J, Mailliez A, Le Deley MC, Rodrigues I, Boulanger T, Lorgis V, Barriere J, Robin YM, Weller M, Bonneterre J. Intrathecal liposomal cytarabine plus systemic therapy versus systemic chemotherapy alone for newly diagnosed leptomeningeal metastasis from breast cancer. Neuro Oncol. 2020;22(4):524–38. https://doi.org/10.1093/neuonc/noz201.
- 63. Li H, Zheng S, Lin Y, Yu T, Xie Y, Jiang C, Liu X, Qian X, Yin Z. Safety, Pharmacokinetic and Clinical Activity of Intrathecal Chemotherapy With Pemetrexed via the Ommaya Reservoir for Leptomeningeal Metastases From Lung Adenocarcinoma: A Prospective Phase I Study. Clin Lung Cancer. 2023;24(2):e94–104. https://doi.org/10.1016/j.cllc.2022.11.011.
- 64. Pan Z, Yang G, Cui J, Li W, Li Y, Gao P, Jiang T, Sun Y, Dong L, Song Y, Zhao G. A Pilot Phase 1 Study of Intrathecal Pemetrexed for Refractory Leptomeningeal Metastases From Non-small-cell Lung Cancer. Front Oncol. 2019;9:838. https://doi.org/10.3389/ fonc.2019.00838.
- Mrugala MM, Kim B, Sharma A, Johnson N, Graham C, Kurland BF, Gralow J. Phase II Study of Systemic High-dose Methotrexate and Intrathecal Liposomal Cytarabine for Treatment of Leptomeningeal Carcinomatosis From Breast Cancer. Clin Breast Cancer. 2019;19(5):311–6. https://doi.org/10.1016/j.clbc.2019. 04.004.
- Beauchesne P. Intrathecal chemotherapy for treatment of leptomeningeal dissemination of metastatic tumours. Lancet Oncol. 2010;11(9):871–9. https://doi.org/10.1016/S1470-2045(10) 70034-6.
- Byrnes DM, Vargas F, Dermarkarian C, Kahn R, Kwon D, Hurley J, Schatz JH. Complications of Intrathecal Chemotherapy in Adults: Single-Institution Experience in 109 Consecutive Patients. J Oncol. 2019;2019:4047617. https://doi.org/10.1155/2019/4047617.



- 68. Kim DY, Lee KW, Yun T, Park SR, Jung JY, Kim DW, Kim TY, Heo DS, Bang YJ, Kim NK. Comparison of intrathecal chemotherapy for leptomeningeal carcinomatosis of a solid tumor: methotrexate alone versus methotrexate in combination with cytosine arabinoside and hydrocortisone. Jpn J Clin Oncol. 2003;33(12):608–12. https://doi.org/10.1093/jjco/hyg118.
- 69. Glitza IC, Rohlfs M, Guha-Thakurta N, Bassett RL Jr, Bernatchez C, Diab A, Woodman SE, Yee C, Amaria RN, Patel SP, Tawbi H, Wong M, Hwu WJ, Hwu P, Heimberger A, McCutcheon IE, Papadopoulos N, Davies MA. Retrospective review of metastatic melanoma patients with leptomeningeal disease treated with intrathecal interleukin-2. ESMO Open. 2018;3(1):e000283. https://doi.org/10.1136/esmoopen-2017-000283.
- 70. Dan X, Huang M, Sun Z, Chu X, Shi X, Chen Y. Case report: Concurrent intrathecal and intravenous pembrolizumab for metastatic melanoma with leptomeningeal disease. Front Oncol. 2024;14:1344829. https://doi.org/10.3389/fonc.2024.1344829.
- 71. Glitza Oliva IC, Ferguson SD, Bassett R Jr, Foster AP, John I, Hennegan TD, Rohlfs M, Richard J, Iqbal M, Dett T, Lacey C, Jackson N, Rodgers T, Phillips S, Duncan S, Haydu L, Lin R, Amaria RN, Wong MK, Diab A, Yee C, Patel SP, McQuade JL, Fischer GM, McCutcheon IE, O'Brien BJ, Tummala S, Debnam M, Guha-Thakurta N, Wargo JA, Carapeto FCL, Hudgens CW, Huse JT, Tetzlaff MT, Burton EM, Tawbi HA, Davies MA. Concurrent intrathecal and intravenous nivolumab in leptomeningeal disease: phase 1 trial interim results. Nat Med. 2023;29(4):898-905. https://doi.org/10.1038/s41591-022-02170-x.
- 72. Kumthekar PU, Avram MJ, Lassman AB, Lin NU, Lee E, Grimm SA, Schwartz M, Bell Burdett KL, Lukas RV, Dixit K, Perron I, Zhang H, Gradishar WJ, Pentsova EI, Jeyapalan S, Groves MD, Melisko M, Raizer JJ. A phase I/II study of intrathecal trastuzumab in human epidermal growth factor receptor 2-positive (HER2-positive) cancer with leptomeningeal metastases: Safety, efficacy, and cerebrospinal fluid pharmacokinetics. Neuro Oncol. 2023;25(3):557–65. https://doi.org/10.1093/neuonc/noac195.
- Trifănescu OG, Mitrea D, Galeș LN, Ciornei A, Păun MA, Butnariu I, Trifănescu RA, Motaș N, Toma RV, Bîlteanu L, Gherghe M. Therapies beyond Physiological Barriers and Drug Resistance: A Pilot Study and Review of the Literature Investigating If Intrathecal Trastuzumab and New Treatment Options Can Improve Oncologic Outcomes in Leptomeningeal Metastases from HER2-Positive Breast Cancer. Cancers. 2023 Apr 27;15(9):2508. https://doi.org/10.3390/cancers15092508
- 74. Wu SA, Jia DT, Schwartz M, Mulcahy M, Guo K, Tate MC, Sachdev S, Kostelecky N, Escobar DJ, Brat DJ, Heimberger AB, Lukas RV. HER2+ esophageal carcinoma leptomeningeal metastases treated with intrathecal trastuzumab regimen. CNS Oncol. 2023;12(3):CNS99. https://doi.org/10.2217/cns-2022-0018.
- 75. Lazaratos AM, Maritan SM, Quaiattini A, Darlix A, Ratosa I, Ferraro E, Griguolo G, Guarneri V, Pellerino A, Hofer S, Jacot W, Stemmler HJ, van den Broek MPH, Dobnikar N, Panet F, Lahijanian Z, Morikawa A, Seidman AD, Soffietti R, Panasci L, Petrecca K, Rose AAN, Bouganim N, Dankner M. Intrathecal trastuzumab versus alternate routes of delivery for HER2-targeted therapies in patients with HER2+ breast cancer leptomeningeal metastases. Breast. 2023;69:451–68. https://doi.org/10. 1016/j.breast.2023.04.008.
- 76. Sener U, Ruff MW, Campian JL. Immunotherapy in Glioblastoma: Current Approaches and Future Perspectives. Int J Molecul Sci. 2022;23(13):7046. https://doi.org/10.3390/ijms23137046.
- Robert C. A decade of immune-checkpoint inhibitors in cancer therapy. Nat Commun. 2020;11(1):3801. https://doi.org/10.1038/ s41467-020-17670-y.
- Naidoo J, Schreck KC, Fu W, Hu C, Carvajal-Gonzalez A, Connolly RM, Santa-Maria CA, Lipson EJ, Holdhoff M, Forde PM, Douville C. Pembrolizumab for patients with leptomeningeal

- metastasis from solid tumors: efficacy, safety, and cerebrospinal fluid biomarkers. J Immunother Cancer. 2021;9(8):e002473. https://doi.org/10.1136/jitc-2021-002473.
- 79. Brastianos PK, Lee EQ, Cohen JV, Tolaney SM, Lin NU, Wang N, Chukwueke U, White MD, Nayyar N, Kim A, Alvarez-Breckenridge C, Krop I, Mahar MK, Bertalan MS, Shaw B, Mora JL, Goss N, Subramanian M, Nayak L, Dietrich J, Forst DA, Nahed BV, Batchelor TT, Shih HA, Gerstner ER, Moy B, Lawrence D, Giobbie-Hurder A, Carter SL, Oh K, Cahill DP, Sullivan RJ. Single-arm, open-label phase 2 trial of pembrolizumab in patients with leptomeningeal carcinomatosis. Nat Med. 2020;26(8):1280-4. https://doi.org/10.1038/s41591-020-0918-0.
- 80. Long GV, Atkinson V, Lo S, Sandhu S, Guminski AD, Brown MP, Wilmott JS, Edwards J, Gonzalez M, Scolyer RA, Menzies AM, McArthur GA. Combination nivolumab and ipilimumab or nivolumab alone in melanoma brain metastases: a multicentre randomised phase 2 study. Lancet Oncol. 2018;19(5):672–81. https://doi.org/10.1016/S1470-2045(18)30139-6.
- 81. Brastianos PK, Strickland MR, Lee EQ, Wang N, Cohen JV, Chukwueke U, Forst DA, Eichler A, Overmoyer B, Lin NU, Chen WY, Bardia A, Juric D, Dagogo-Jack I, White MD, Dietrich J, Nayyar N, Kim AE, Alvarez-Breckenridge C, Mahar M, Mora JL, Nahed BV, Jones PS, Shih HA, Gerstner ER, Giobbie-Hurder A, Carter SL, Oh K, Cahill DP, Sullivan RJ. Phase II study of ipilimumab and nivolumab in leptomeningeal carcinomatosis. Nat Commun. 2021;12(1):5954. https://doi.org/10.1038/ s41467-021-25859-y.
- 82. Hendriks LEL, Bootsma G, Mourlanette J, Henon C, Mezquita L. Ferrara R. Audigier-Valette C. Mazieres J. Lefebyre C. Duchemann B, Cousin S, le Pechoux C, Botticella A, De Ruysscher D, Dingemans AC, Besse B. Survival of patients with non-small cell lung cancer having leptomeningeal metastases treated with immune checkpoint inhibitors. Eur J Cancer. 2019;116:182-9. https://doi.org/10.1016/j.ejca.2019.05.019.
- 83. Palmisciano P, Haider AS, Nwagwu CD, Wahood W, Yu K, Ene CI, O'Brien BJ, Aoun SG, Cohen-Gadol AA, El Ahmadieh TY. The Role of Immune Checkpoint Inhibitors in Leptomeningeal Disease: A Systematic Review. Anticancer Res. 2021;41(11):5333-42. https://doi.org/10.21873/anticanres. 15346.
- 84. Yang JCH, Kim SW, Kim DW, Lee JS, Cho BC, Ahn JS, Lee DH, Kim TM, Goldman JW, Natale RB, Brown AP, Collins B, Chmielecki J, Vishwanathan K, Mendoza-Naranjo A, Ahn MJ. Osimertinib in Patients With Epidermal Growth Factor Receptor Mutation-Positive Non-Small-Cell Lung Cancer and Leptomeningeal Metastases: The BLOOM Study. J Clin Oncol. 2020;38(6):538-47. https://doi.org/10.1200/JCO.19.00457.
- 85. Ahn MJ, Chiu CH, Cheng Y, Han JY, Goldberg SB, Greystoke A, Crawford J, Zhao Y, Huang X, Johnson M, Vishwanathan K, Yates JWT, Brown AP, Mendoza-Naranjo A, Mok T. Osimertinib for Patients With Leptomeningeal Metastases Associated With EGFR T790M-Positive Advanced NSCLC: The AURA Leptomeningeal Metastases Analysis. J Thorac Oncol. 2020;15(4):637-48. https://doi.org/10.1016/j.jtho.2019.12.113.
- 86. Bian DJH, Lazaratos AM, Maritan SM, Quaiattini A, Zeng Z, Zhu Z, Sener U, Malani R, Kim YJ, Ichihara E, Cohen V, Rose AAN, Bouganim N, Dankner M. Osimertinib is associated with improved outcomes in pre-treated non-small cell lung cancer leptomeningeal metastases: A systematic review and meta-analysiss. Heliyon. 2024;10(9):e29668. https://doi.org/10.1016/j.heliyon. 2024.e29668.
- 87. Wilcox JA, Boire AA. Leveraging Molecular and Immune-Based Therapies in Leptomeningeal Metastases. CNS Drugs. 2023;37(1):45-67. https://doi.org/10.1007/s40263-022-00975-5.
- Zhu VW, Lin YT, Kim DW, Loong HH, Nagasaka M, To H, Ang YL, Ock CY, Tchekmedyian N, Ou SI, Syn NL,



- Reungwetwattana T, Lin CC, Soo RA. An International Real-World Analysis of the Efficacy and Safety of Lorlatinib Through Early or Expanded Access Programs in Patients With Tyrosine Kinase Inhibitor-Refractory ALK-Positive or ROS1-Positive NSCLC. J Thorac Oncol. 2020;15(9):1484–96. https://doi.org/10.1016/j.jtho.2020.04.019.
- 89. Ou SH, Sommers KR, Azada MC, Garon EB. Alectinib induces a durable (>15 months) complete response in an ALK-positive non-small cell lung cancer patient who progressed on crizotinib with diffuse leptomeningeal carcinomatosis. Oncologist. 2015;20(2):224–6. https://doi.org/10.1634/theoncologist. 2014-0309.
- Gainor JF, Sherman CA, Willoughby K, Logan J, Kennedy E, Brastianos PK, Chi AS, Shaw AT. Alectinib salvages CNS relapses in ALK-positive lung cancer patients previously treated with crizotinib and ceritinib. J Thorac Oncol. 2015;10(2):232–6. https://doi.org/10.1097/JTO.0000000000000455.
- Kim DW, Barcena E, Mehta UN, Rohlfs ML, Kumar AJ, Penas-Prado M, Kim KB. Prolonged survival of a patient with metastatic leptomeningeal melanoma treated with BRAF inhibitionbased therapy: a case report. BMC Cancer. 2015;15:400. https:// doi.org/10.1186/s12885-015-1391-x.
- Burger MC, Ronellenfitsch MW, Lorenz NI, Wagner M, Voss M, Capper D, Tzaridis T, Herrlinger U, Steinbach JP, Stoffels G, Langen KJ, Brandts C, Senft C, Harter PN, Bahr O. Dabrafenib in patients with recurrent, BRAF V600E mutated malignant glioma and leptomeningeal disease. Oncol Rep. 2017;38(6):3291–6. https://doi.org/10.3892/or.2017.6013.
- 93. McLoughlin EM, Fadul CE, Patel SH, Hall RD, Gentzler RD. Clinical and Radiographic Response of Leptomeningeal and Brain Metastases to Encorafenib and Binimetinib in a Patient With BRAF V600E-Mutated Lung Adenocarcinoma. J Thorac Oncol. 2019;14(12):e269–71. https://doi.org/10.1016/j.jtho. 2019.07.019.
- 94. Niikura N, Yamanaka T, Nomura H, Shiraishi K, Kusama H, Yamamoto M, Matsuura K, Inoue K, Takahara S, Kita S, Yamaguchi M, Aruga T, Shibata N, Shimomura A, Ozaki Y, Sakai S, Kiga Y, Izutani T, Shiosakai K, Tsurutani J. Treatment with trastuzumab deruxtecan in patients with HER2-positive breast cancer and brain metastases and/or leptomeningeal disease (ROSET-BM). NPJ Breast Cancer. 2023;9(1):82. https://doi.org/10.1038/s41523-023-00584-5.
- Alder L, Trapani D, Bradbury C, Van Swearingen AED, Tolaney SM, Khasraw M, Anders CK, Lascola CD, Hsu L, Lin NU, Sammons S. Durable responses in patients with HER2+ breast cancer and leptomeningeal metastases treated with trastuzumab deruxtecan. NPJ Breast Cancer. 2023;9(1):19. https://doi.org/10.1038/ s41523-023-00519-0.
- Yan F, Rinn KJ, Kullnat JA, Wu AY, Ennett MD, Scott EL, Kaplan HG. Response of Leptomeningeal Metastasis of Breast Cancer With a HER2/neu Activating Variant to Tucatinib: A

- Case Report. J Natl Compr Canc Netw. 2022;20(7):745–52. https://doi.org/10.6004/jnccn.2022.7006.
- O'Brien BJ, Murthy RK, Berry DA, Raghavendra AS, Gule-Monroe M, Johnson JM, Schwartz-Gomez J, Topletz-Erickson A, Lobbous M, Melisko ME, Morikawa A, Ferguson SD, Groot JFd, Krop IE, Valero V, Rimawi MF, Wolff AC, Tripathy D, Lin NU, Stringer-Reasor EM. Tucatinib-trastuzumab-capecitabine for treatment of leptomeningeal metastasis in HER2+ breast cancer: TBCRC049 phase 2 study results. J Clinic Oncol. 2024;42(16_suppl):2018–2018. https://doi.org/10.1200/JCO.2024.42.16_suppl.2018.
- 98. Stringer-Reasor EM, O'Brien BJ, Topletz-Erickson A, White JB, Lobbous M, Riley K, Childress J, LaMaster K, Melisko ME, Morikawa A, Groot JFD, Krop IE, Valero V, Rimawi MF, Wolff AC, Tripathy D, Lin NU, Murthy RK (2021) Pharmacokinetic (PK) analyses in CSF and plasma from TBCRC049, an ongoing trial to assess the safety and efficacy of the combination of tucatinib, trastuzumab and capecitabine for the treatment of leptomeningeal metastasis (LM) in HER2 positive breast cancer. J Clinic Oncol 39 (15_suppl):1044-1044. https://doi.org/10.1200/JCO.2021.39.15_suppl.1044
- Sener U, Islam M, Webb M, Kizilbash SH (2024) Antiangiogenic exclusion rules in glioma trials: Historical perspectives and guidance for future trial design. Neurooncol Adv 6 (1):vdae039. https://doi.org/10.1093/noajnl/vdae039
- Reijneveld JC, Brandsma D, Boogerd W, Bonfrer JG, Kalmijn S, Voest EE, Geurts-Moespot A, Visser MC, Taphoorn MJ. CSF levels of angiogenesis-related proteins in patients with leptomeningeal metastases. Neurology. 2005;65(7):1120–2. https://doi. org/10.1212/01.wnl.0000178981.39984.c2.
- 101. Wu PF, Lin CH, Kuo CH, Chen WW, Yeh DC, Liao HW, Huang SM, Cheng AL, Lu YS. A pilot study of bevacizumab combined with etoposide and cisplatin in breast cancer patients with leptomeningeal carcinomatosis. BMC Cancer. 2015;15:299. https://doi.org/10.1186/s12885-015-1290-1.
- Sasich LD, Sukkari SR. The US FDAs withdrawal of the breast cancer indication for Avastin (bevacizumab). Saudi Pharm J. 2012;20(4):381–5. https://doi.org/10.1016/j.jsps.2011.12.001.
- 103. Ariyasu R, Horiike A, Koyama J, Saiki M, Sonoda T, Kawashima Y, Takano N, Oguri T, Nishikawa S, Kitazono S, Yanagitani N, Ohyanagi F, Nishio M. Efficacy of bevacizumab and erlotinib combination for leptomeningeal carcinomatosis after failure of erlotinib. Anticancer Drugs. 2017;28(5):565–7. https://doi.org/10.1097/CAD.00000000000000489.

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

