



Review

Diffuse Axonal Injury: Clinical Prognostic Factors, Molecular Experimental Models and the Impact of the Trauma Related Oxidative Stress. An Extensive Review Concerning Milestones and Advances

Mauro Palmieri ^{1,*} , Alessandro Frati ^{1,2}, Antonio Santoro ¹, Paola Frati ³ , Vittorio Fineschi ³ and Alessandro Pesce ⁴

- ¹ Neurosurgery Division, A.O.U. "Policlinico Umberto I", Human Neuroscience Department, "Sapienza" University, Viale Del Policlinico 155, 00161 Rome, Italy; alessandro.frati@uniroma1.it (A.F.); antonio.santoro@uniroma1.it (A.S.)
- ² IRCCS "Neuromed", Via Atinense 18, 86077 Pozzilli, Italy
- ³ Department of Anatomical, Histological, Forensic and Orthopaedic Sciences SAIMLAL, "Sapienza" University, Viale Regina Elena 336, 00185 Rome, Italy; paola.frati@uniroma1.it (P.F.); vittorio.fineschi@uniroma1.it (V.F.)
- ⁴ Neurosurgery Division, Santa Maria Goretti Hospital, Via Lucia Scaravelli, 04100 Latina, Italy; ale_pesce83@yahoo.it
- * Correspondence: mauro.palmieri10@gmail.com; Tel.: +39-063-377-5298



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Abstract: Traumatic brain injury (TBI) is a condition burdened by an extremely high rate of morbidity and mortality and can result in an overall disability rate as high as 50% in affected individuals. Therefore, the importance of identifying clinical prognostic factors for diffuse axonal injury (DAI) in (TBI) is commonly recognized as critical. The aim of the present review paper is to evaluate the most recent contributions from the relevant literature in order to understand how each single prognostic factor determinates the severity of the clinical syndrome associated with DAI. The main clinical factors with an important impact on prognosis in case of DAI are glycemia, early GCS, the peripheral oxygen saturation, blood pressure, and time to recover consciousness. In addition, the severity of the lesion, classified on the ground of the cerebral anatomical structures involved after the trauma, has a strong correlation with survival after DAI. In conclusion, modern findings concerning the role of reactive oxygen species (ROS) and oxidative stress in DAI suggest that biomarkers such as GFAP, pNF-H, NF-L, microtubule associated protein tau, A β 42, S-100 β , NSE, AQP4, Drp-1, and NCX represent a possible critical target for future pharmaceutical treatments to prevent the damages caused by DAI.

Keywords: traumatic brain injury; diffuse axonal injury; oxidative stress; reactive oxygen species; biomarkers

1. Introduction

Traumatic brain injury (TBI) is a condition burdened by an extremely high rate of morbidity and mortality and can result in an overall disability rate as high as 50% in affected individuals [1]; in the near future, it will become the third leading cause of permanent disability and mortality worldwide [2]. TBI is a frequent cause of traumatic diffuse axonal injury, realizing a condition called diffuse axonal injury (DAI). Because of its incidence and its economic impact on national health systems [3,4], the importance of identifying clinical prognostic factors for diffuse axonal injury (DAI) in traumatic brain injury (TBI) is commonly recognized as critical [5]. More specifically, initial GCS, duration of the loss of consciousness, blood oxygenation, and blood pressure are some of the several prognostic factors that have been investigated in order to understand their impact on the clinical and neurological outcome of DAI in TBI [5–9]. Furthermore, new light has been recently shed

over the possible influence of the mechanisms managing trauma-related oxidative stress in determining the severity of the clinical phenotype of the TBI: the imbalance between the production and removal of reactive oxygen species (ROS), the release and activation of pro-inflammatory cytokines, and modifications in calcium metabolism are just a few relevant examples.

The aim of the present review paper is to encompass and analyse the most recent contributions from the relevant literature in order to understand the weight of each single prognostic factor in determining the severity of the clinical syndrome associated with diffuse axonal injury.

2. Hypoxia and Blood Pressure

The presence of hypoxia (measured by SpO₂: SpO₂ < 90%; measured by PaO₂: PaO₂ < 60 mmHg) and hypotension (measured by blood systolic pressure < 90 mmHg) is a physiological change after TBI and is associated with a major fatality rate [10]. In fact, preserving airway patency after a traumatic injury is recognised as a positive prognostic factor [11]. The relative inability to increase cerebral blood flow in response to hypoxia and hypotension after a traumatic injury exposes the injured brain to a secondary ischemic insult [12]. In the study presented by Manley et al. [12], it is reported that in patients with hypoxia alone, the mortality was 28%, while the presence of both hypoxia and hypotension increased the mortality to 57%. These findings are confirmed in other studies [13–15], and the association between hypoxia and hypotension with increased mortality has also been demonstrated in children [16]. Moreover, repeated episodes of hypotension have a negative impact on survival [12]. Regarding hypertension (blood systolic pressure ≥ 160 mmHg), Viera et al. [10] highlight the impact of elevated levels of systolic blood pressure on intracranial pressure and cerebral edema even though hypertension may be a physiological response to reduced cerebral perfusion. Therefore, the decision to treat hypertension should be made carefully since lowering the blood pressure could exacerbate cerebral ischemia itself [17]. In conclusion, according to the latest guidelines concerning the management of severe traumatic brain injury [18], the value of 90 mmHg for systolic blood pressure and of 60 mmHg of peripheral blood oxygenation are considered to be the thresholds to define hypotension and hypoxia, respectively, representing a statistically significant factor influencing the outcome.

3. Glycemia

Changes in glycemia levels in severe TBI are related to the increased blood levels of catecholamines caused by the sympathetic hyperactivity after the traumatic injury [19,20]. Hyperglycemia induces lactic acidosis, which extends secondary neuronal damage [21]. On the other hand [21], it is commonly known that hypoglycaemia induces neuronal necrosis, especially in the caudate, putamen, and hippocampus [21]. Liu-De Ryke et al. [22] analysed the association between glycemia in the first 5 days and mortality after TBI in a cohort of 380 patients. The glucose cut-off values of 7.5 mmol/L for admission and 8.89 mmol/L for the day 1 peak were demonstrated to be effective predictors of mortality [22]. More specifically, a significant difference in terms of survival was detected for the threshold of the 8.89 mmol/L day 1 peak. In fact, on day 5, 96% of the patients with glucose blood levels <8.89 mmol/L survived versus the 75% of those with glucose blood levels >8.89 mmol/L. They also highlighted the correlation between high levels of blood glucose and trauma severity. Moreover, in the same study, patients with glycemia <3.33 mmol/L showed a higher mortality rate than those with a glucose blood level >3.33 mmol/L. Regarding hyperglycemia, other studies [10,23–25] have reported that increased levels of blood glucose correlate with poor outcome. Up until now, a common threshold for hyperglycemia has not been identified; thus, modern guidelines [18] do not include pharmacological interventions for changes in glucose blood levels.

4. Pathological Anatomy and Morphologic Findings

The focal lesions caused by a severe form of DAI can be macroscopically identified postmortem [26]. In patients who do not survive, the lesions usually appear as hemorrhagic, while the identification of the lesion after weeks or months could be challenging due to the presence of gliosis [26]. In the series presented by Adams et al. [26,27] DAI has three distinctive structural features:

1. Diffused supratentorial damage to axons (grade I).
2. A focal lesion in the corpus callosum (grade II).
3. A focal lesion or multiple lesions in the rostral brain stem (grade III).

Furthermore, grade II and III lesions have typical localization. In fact, lesions in the corpus callosum typically occur in its inferior part and to one side of the midline, while lesions on the rostral brainstem typically localizes themselves in the dorsolateral quadrant or in the quadrants adjacent to the superior cerebellar peduncles [27,28], Figure 1. However, the diagnosis of DAI can only be confirmed through histological studies since axonal damage cannot be identified during macroscopical examination [28].

In order to diagnose DAI, swollen axonal varicosities and axonal bulbs [29] must be detected during microscopic examination [30]. According to modern studies [31], the two main characteristics that must be identified are the presence of diffuse/multifocal axonal damage in the white matter and this axonal damage being widespread in many brain regions, at least one of which should be located above the tentorium and one should be located below the tentorium. The morphological aspects of DAI are also influenced by the time span of interlude between the beginning and resolution of coma; in fact, the evolution of microscopical damage has been described since the 1980s [26]. In order to classify the severity of axonal injury according to microscopical findings, Gennarelli et al. [32] have divided different types of lesions into three grades of severity:

Grade 1: There are scattered axonal retraction balls in the parasagittal white matter of the cerebral hemispheres, the corpus callosum, the brain stem, and, less commonly, the cerebellum.

Grade 2: In addition to axonal damage in the white matter of the cerebral hemisphere, there is a focal lesion in the corpus callosum.

Grade 3: In addition to axonal damage in the white matter of the hemispheres, there are focal lesions present in the dorsolateral quadrant of the rostral brain stem and the corpus callosum.

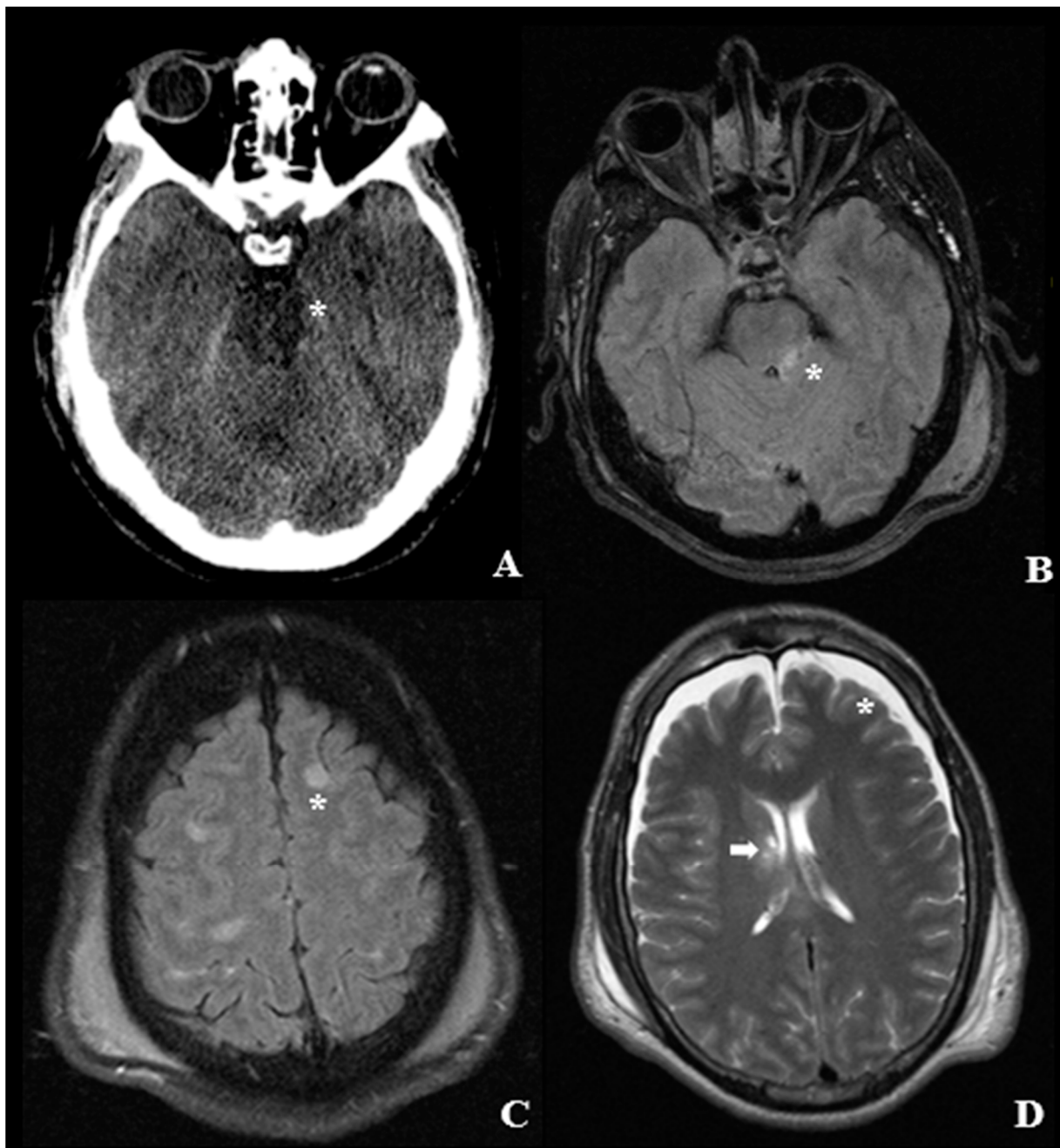


Figure 1. Radiological findings in a case of DAI—(A) First Computed tomography (CT) scan disclosing the absence of Ambiens Cistern and a slight hypodensity of the transition zone between Mesencephalon and Pons (asterisk). (B,C) Flair sequence late MRI disclosing the diffuse involvement of the subcortical with matter and of the Brachium Pontis (asterisks). (D) T2 weighted MRI scan disclosing the late atrophic changes of the Supratentorial cerebrum, with a bilateral Subdural Hygroma (asterisk). The arrow highlights a neural injury at the transition between the body of the Caudate Nucleus and the Internal Capsule.

5. Time to Recover Consciousness

The impact of the severity of DAI on the recovery of consciousness in patients with TBI is well recognized [8,9]. More specifically, a correlation has been observed between the mean time interval to recovery of consciousness in patients with DAI and the degrees of brain injuries according to the classification proposed by Gennarelli et al. [5,32]. Based on the findings of the studies [33,34] that showed the impact of magnetic resonance imaging (MRI) findings and DAI outcome, Park et al. analysed data concerning the severity of

lesions identified with MRI with the time to recover consciousness. The results outlined a faster recovery time for patients with a grade I and II injury despite grade III lesions, describing a time span of weeks for lesions involving cerebral white matter and the corpus callosum and for months for injuries located in brain stem. However, a universally accepted classification for time to recover consciousness has not been identified yet regarding patients with DAI.

6. Severity of Trauma

The Injury severity score (ISS) and the New Injury Severity Score (NISS) represent two universally accepted scales in the evaluation of the severity of trauma [35,36] and, specifically for head trauma, the Maximum Abbreviated Injury Scale (MAIS) [37] represents a more accurate tool for the evaluation of TBI. The ISS and NISS scales divide the body into different regions, and for each of them, a score from 1 to 6 is given on the basis of the severity of the lesion in that particular anatomical area [35,36]. Then, the final score is useful in order to predict mortality, morbidity, and hospitalization length after trauma. The MAIS represents a subscale of the ISS, and it represents the score of a single body region. A MAIS of 5 is related to a critical damage of the area of interest, and it relates to a poor outcome [10,37]. High scores in the ISS and NISS are related to a higher risk of poor outcome [10], and it has also been demonstrated that NISS can predict mortality better than ISS [38], even if there is scant literature on the application of NISS in DAI [10]. Furthermore, high scores in the three scales relate to dependence, which can be measured with the Glasgow Outcome Scale-Extended (GOSE) [39] 6 months after DAI [10]. The aforementioned scales take in account the clinical impact of possible concurrent lesions involving other organs or other collateral conditions, including intracranial bleeding, cranial base and vault lesions [40], and spinal cord or vertebral lesions. Moreover, it has been described that pre-existing conditions, namely other major comorbidities and aging-related changes to tissues and metabolism could worsen the injury caused by the trauma [30,40,41].

7. GCS

The Glasgow Coma Scale (GCS) [42] undoubtedly represents one of the most important neurological tools to assess the neurological performance of a patient affected by TBI. The final score of the scale ranges from 3 to 15, and it is obtained by the summation of the scores of the three subscales regarding eye response, verbal response, and motor response. A GCS score <8 represents a severe TBI, and in patient with DAI, lower GCS score levels at admission are related to higher mortality and dependence after trauma [10,25,43]. Moreover, higher levels of dependence and mortality are related also to lower GCS score levels after the withdrawal of sedation [10,44]. According to Skandsen et al. [45], GCS is correlated to outcome only if DAI happens in TBI, outlining the importance of diagnosing this kind of injury. Whereas the GCS score at admission is, according to a vast number of clinical trials, strongly associated with the prognosis of patients, it should be kept in mind that a great number of patients suffering from a TBI-related DAI (including DAI I and II) may acutely present with lower GCS scores (3–6), possibly not accurately discriminating the long-term prognostic classes of patients. The GCS score should therefore be interpreted in the context of the specific trauma of each single patient while also relating to the aforementioned biological parameters.

8. Biomarkers and the Role of Oxidative Stress

Besides clinical prognostic factors, biomarkers related to the physiopathology of DAI represent a promising solution for defining novel strategies for the assessment of the severity of the injury and for pharmacological treatment [46]. In the process of axonal swelling and neural death, the mitochondrial permeability transition pore (mPTP) is a protein that is produced after a major accumulation of calcium in the mitochondria in response to the cellular exposition to reactive oxygen species (ROS) [47–50]. Therefore, the

possible use of cyclosporin A, an inhibitor of mPTP, as a pharmacological option in the treatment of DAI is under investigation [51–56]. Buki et al. [51] conducted an experimental study on Sprague Dawley rats regarding the intrathecal administration of cyclosporin A 30 min before causing brain injury. The first result retrieved from this study shows that cyclosporin A preserves mitochondrial integrity in axons destined to be disconnected after the brain injury. According to the authors, the neuroprotection effect caused by cyclosporin A is due not only to the inhibition of mPTP but also to the reduction of calcium-induced, calpain-mediated spectrin proteolysis (CMSP) and neurofilament compaction (NFC), which are directly involved in the pathogenesis of axonal disconnection. In fact, in this experimental study [51], it was shown that the administration of cyclosporin A 30 min after brain injury resulted in a significant decrease in CMSP and NFC immunoreactivity in the corticospinal tracts and medial longitudinal fasciculi 24 h after injury. Similar findings have been described by Okonwko [52,53], who highlighted how mitochondrial damage leads to calcium-induced cytoskeletal modifications after traumatic axonal injury by specific antibodies that target spectrin proteolysis and neurofilament compaction. Immunoreactivity studies conducted in rats treated with cyclosporin A before axonal injury and in those that did not receive any premedication showed that the administration of cyclosporin A prior brain injury causes significant decreases in cytoskeletal injury (approximately 70%). Even though these promising results have been obtained in animal studies, there is still a lack of consistent evidence proving the effectiveness of cyclosporin A in humans. Aminmansour et al. [57] designed a randomized double-blind placebo-controlled study with 100 patients suffering from DAI. A total of 50 patients were treated with 5 mg/kg/24 h of cyclosporin A via a 250 mL dextrose water (DW) 5% solution (DW 5%) in the first 8 h after trauma, while the control group (n = 50) only received DW 5% in the same course. Serum values of complete blood count, blood urea nitrogen, creatinine, aspartate aminotransferase, alanine aminotransferase, and alkaline phosphatase were registered as well to assess the incidence of possible complications due to pharmacological treatment with cyclosporin A. The analysis of the GOSE score comparison between the two groups at 3 and 6 months after the trauma did not show significant differences. With the exception of a transitional elevation of blood urea nitrogen in the first 48 h and of white blood cell count at 12 h in the intervention group, no long-lasting significant differences were outlined regarding adverse effects. Similar results were obtained by other authors [58–60], suggesting that the administration of cyclosporin A still does not lead to any clinically relevant effects.

Regarding cytoskeletal injury, the role of caspase and calpain in the cytoskeletal decomposition process has been investigated extensively [61,62]. In the proteolysis process induced by these enzymes, the neurofilament protein (NF) is interesting [63], and the accumulation of its subunits after TBI is currently being studied in order to identify predictive markers for outcome. Serum levels of the phosphorylated neurofilament heavy subunit (pNF-H) at 24 and 72 h after TBI [64] and the neurofilament light chain (NF-L) [65] protein serum level has recently been advocated as a potentially promising predictor of 12-month neurological outcome. More specifically, Shibahashi et al. [65] collected blood samples from 32 human subjects after brain injury, and they included patients with a GCS score of 13 or less upon admission and assessed outcome at 6 months using the same scale. The authors showed that the pNF-H serum levels that exceeded 240 pg/mL 24 h after trauma represented a significant predictive outcome at 6 months ($p < 0.001$). The pNF-H serum levels > 80 pg/mL 72 h after TBI correlated significantly with unfavorable outcome as well at 6 months ($p < 0.01$). Regarding NF-L, Ljungqvist et al. [66] measured acute serum concentrations of this polypeptide in nine patients affected by severe DAI by means of an ultrasensitive single molecule array (Simoa) assay and analyzed their correlation with clinical outcome and MRI diffusor tensor imaging (DTI) parameters at 12 months after TBI. The blood samples were collected 6 days after TBI, and the NF-L serum levels were measured in 22 age-matched healthy controls. The statistical analysis highlighted that the patients affected by DAI had a 30-fold increase of NF-L serum concentrations than healthy controls and that the level of NF-L correlated with the severity of DAI. More specifically,

higher serum levels of NF-L were significantly associated with higher trace ($R^2 = 0.79$) and lower fractional anisotropy (FA) ($R^2 = 0.83$) DTI parameters and consequently worse clinical outcome. Even though these two biomarkers represent a promising solution for DAI assessment, they have been tested in small cohorts; therefore, further investigation must be conducted through large prospective trials to confirm their reliability.

Other biomarkers that are under investigation as potential predictors of outcome in DAI and that are related to cytoskeletal injury are the glial fibrillary acid protein (GFAP) [66,67], microtubule-associated protein tau [68], and the amyloid β peptide (A β 42) [69]. According to Bogoslovsky et al. [70], the combined measurement of the plasma levels of all three biomarkers could be useful in both acute and subacute phases of TBI. The study proposed by these authors was conducted on 34 patients affected by TBI with consequent DAI and a control group of 69 healthy volunteers. Plasma samples were collected 24 h and 30 and 90 days after the trauma. More specifically, the GFAP and microtubule-associated protein tau peaks were registered 24 h after injury, while the A β 42 levels in the plasma reached their maximal concentrations at day 30. All three biomarkers maintained high blood values up to 90 days after the trauma. Moreover, A β 42 levels after 30 days and GOSE measured at 180 after TBI showed a relationship after adjustment for age, and a weak correlation was outlined between Tau concentrations at day 30 and radiological injury severity.

The calcium binding protein S-100 β increases in the early phases of injury [71,72] although there are some limitations related to its usage. First of all, opposing evidence on the dependence of S-100 β serum concentrations from the integrity and permeability of the blood–brain barrier (BBB) have been described [73,74]. In addition, the elevation of S-100 β seems to be related to musculoskeletal injury in polytrauma patients [75], so the interpretation of high levels of S-100 β in this kind of patient should be evaluated with caution even though the consequences of polytrauma damages on the serum dosage of this protein appear to be limited to the first 12 h due to its short half-life [76]. In conclusion, in the series presented by Pelinka et al. [77], the GFAP was more reliable than S-100 β in discriminating between severe disability and a persistent vegetative state, while the data concerning mortality was demonstrated to be superimposable.

High levels of neuron-specific enolase (NSE) are measurable in cerebrospinal fluid (CSF), but despite its ability to be measured from S-100 β serum levels depends on the integrity of the BBB [78] even though conditions such as polytrauma should not influence the blood levels of NSE. Furthermore, the NSE half-life is significantly longer than S-100 β , making its sampling more manageable. Thelin et al. [76] measured NSE and S-100 β in 417 patients twice daily, and both biomarkers resulted in being independently correlated to long-term functional outcome. Moreover, S-100 β appears to have higher precision as an outcome predictor in TBI and, consequently, a more reliable biomarker than NSE [76]. Evidence regarding pediatric TBI suggests that NSE could be a good predictor for Glasgow Outcome Score (GOSE) results [79].

The expression of the nuclear factor erythroid-related factor 2 (Nrf2) has recently been identified as a protective factor towards ROS induced by DAI [80]. Nrf2 is sensitive to redox reactions [72,81], and it regulates the expression of hemeoxygenase-1 (HO-1) by binding to antioxidant the response element (ARE) [82,83]. Moreover, Nrf2 can also neutralize the effects of carcinogens or ROS directly [84]. Ishii et al. [81] first reported that oxidized low-density lipoproteins (oxLDLs) and 4-Hydroxy-2-nonenal (HNE) derived from lipid oxidation induced the nuclear translocation of Nrf2 in murine peritoneal macrophages. The expression of Nrf2 causes the consequent upregulation of CD36, which represents a scavenger receptor in the macrophages, and its role in homeostasis and anti-inflammatory responses is also regulated by PPAR- γ . The evidence retrieved from this study focused its attention on the possible role of Nrf2 in oxidative stress mediated injuries, and Gao et al. [83] demonstrated its role in DAI. In fact, after DAI-related oxidative stress, Nrf2 is released from its inhibitor Kelch sample-related protein (Keap1) and binds to ARE inside the nucleus, causing the consequent expression of NAD(P)H:quinone oxidoreductase 1 (NQO1) and HO-1 [83]. Gao et al.'s experiments confirmed that Nrf2 is increased in rats affected by DAI

while it remains normally expressed in controls through Western Blot analysis ($p < 0.01$) at 6, 24, 28, and 72 h. Moreover, regarding the expression of both HO-1 and NQO1, analogue results were retrieved after RT-PCR analyses on rat brain stems after TBI, highlighting a time-dependent increase with a peak after 24 h, high expression until 72 h after injury, and a consequent gradual decrease. The comparison with the control group showed that these changes were constantly significant ($p < 0.05$).

On the basis of these findings, Wu et al. [80] described that the expression of Nrf2 and, consequently, the HO-1 in cerebral microvessels is correlated to the progression of DAI, the severity of blood–brain barrier damage, and cell injury caused by ROS. Therefore, given its important role as an antioxidant agent, the authors highlighted how Nrf2 may represent a crucial molecular target in order to protect brain cells from ROS. In particular, they induced the expression of Nrf2 by means of an exogenous molecule administration. The role of sulphorafane (SFN) in inducing the expression of different antioxidant agents in human studies has been extensively described by Dinkova-Kostova et al. [85]. In this review, it was reported how after 2 weeks of daily consumption of 100 μmol SFN, the mRNA levels of Nrf2 and HO-1 were upregulated in a dose-dependent manner. Therefore, Wu et al. [80] proposed the use of SFN in an animal model of DAI to induce Nrf2 expression and to consequently reduce DAI-related ROS and neuronal apoptosis. The authors divided 48 Sprague Dawley rats into four equally formed groups: the control group, the DAI group, the DAI group treated with phosphate-buffered saline (PBS), and the DAI group treated with SFN. Cell samples retrieved from all of the groups were then treated for 24 h with H2O2 and/or SFN after incubation. *In vivo* and *in vitro* analysis were conducted. The results showed that the DAI-related oxidative stress damages that were significantly more represented were those where the treatment with SFN was not performed, outlining the protective effects of SNF on programmed cells death due to axonal injury. Moreover, further analysis conducted on samples with and without Nrf2 defection showed that when Nrf2 is normally expressed, treatment with SFN is more effective in suppressing neuronal apoptosis and that Nrf2/HO-1 activation due to SNF is confirmed both *in vitro* and *in vivo*.

Several authors have advocated for a potentially critical role as a biomarker and a therapeutic target of the dynamin related protein-1 (Drp-1) and the reverse-mode sodium calcium exchanger (NCX) [86]. NCX modulates the Ca^{2+} intracellular influx on the ground of an exchange mechanism based on the intracellular Na^+ concentration: under TBI-induced stress conditions, the abnormal glutamate release leads to an intracellular Na^+ increase, thus reversing the activity of this exchange protein, leading to an accumulation of Ca^{2+} in the intracellular compartment [87–89], eventually determining a diffuse mitochondrial dysfunction resulting in the creation of ROS and the activation of endoproteases, phospholipases, and endonucleases [90], resulting in axonal degeneration and neuronal death. Drp-1 is a GTP-ase involved in mitochondrial fission [86]. These potential target proteins have been found to be expressed in the homo- and contralateral cortex and the hippocampus after experimental TBI at 1 h and 24 h after the TBI, with a progressive reduction and a return to the baseline levels within 5 days [86]. In order to test Drp-1 and NCX as possible therapeutic targets, Omelchenko et al. [86] used a microfabricated culture platform to create a model of *in vitro* DAI samples. The authors observed that Drp-1 upregulation was connected with TOMM20, a mitochondrial marker, in the proximity of injured axons. Similarly, NCX was clustered along the axonal processes in the control group, while in the post-stretch injury group, NCX localized to the axonal beads. Therefore, selective inhibitors of Drp-1, dynasore, and of NCH, SN-6, were tested immediately post-injury and every hour for 6 h. After examination, both dynasore and SN-6 showed protective effects in preventing increases of axons bead side damage after 6 h. However, a minor increase of post-traumatic axonal swelling was observed but only in the Dynasore treatment group. Regarding mitochondrial fragmentation induced by axonal strain, the authors compared the morphology of the axonal mitochondria before and 1 min after injury treatment with Dynasore and SN-6. The comparison with the control group highlighted that both drugs are effective in avoiding morphological and dimensional modifications, suggesting that NCX

and Drp-1 are directly involved in the molecular cascade responsible for mitochondrial fragmentation [86].

Further analyses were conducted to assess the role of dynasore and SN-6 in preventing oxidative stress damage related to DAI, but only dynasore was able to attenuate H₂O₂-induced apoptosis and increase of ROS in all hippocampal regions. In conclusion, potential therapeutic agents such as dynasore and SN-6 have shown promising results in attenuating the stretch injury-induced swelling of axonal varicosities and mitochondrial fragmentation, thus expressing potential clinical significance as therapeutic agents in the clinical management of DAI. Moreover, Drp-1 could represent a specific molecular target for preventing oxidative stress injury related to DAI.

Recent studies regarding aquaporin-4 (AQP4) have focused on the increase of its expression and subcellular relocation of this protein after hypoxia-induced cell swelling through a calmodulin-mediated mechanism [91,92]. AQP4 is the most common isoform of aquaporin proteins in the central nervous system (CNS), and it has been identified as the major regulator of water homeostasis through the control of the exchange of CSF with interstitial fluid [92]. Therefore, modifications in AQP4 expression are consequently related to the development of brain edema. More specifically, an increase of AQP4 activity in brain edema is caused by the upregulation of calmodulin when hypoxia-mediated injury occurs. Calmodulin directly binds the AQP4 carboxyl terminus, causing a conformational change and the consequent cell-surface location of AQP4, which leads to cell swelling due to a massive increase of intracellular water [91]. It has been described by Kitchen et al. [91] that trifluoperazine (TFP) inhibits the calmodulin-mediated cell location of AQP4 by directly binding to calmodulin *in vivo* by creating a crush model injury at T8 and a cortical stab injury in rodents. The consequent development of CNS edema can be prevented by the administration of TFP. In this study [91], the authors proved that in both the crush injury and in the cortical stab injury models treated with TFP, there is a significant difference in terms of the attenuation of CNS edema and improvement in electrophysiological, sensory, and locomotor function. Sylvian et al. [93] confirmed these findings by using a photothrombotic stroke model in rats. TFP (10 mg/kg) was administered subcutaneously at either 30 min before stroke or 1 h after stroke. A significant brain edema reduction was only observed in the group treated 1 h after the photothrombotic stroke, and TFP had not shown any effects on the investigated brain neurochemical biomarkers, electrolytes, or elemental homeostasis. This promising evidence suggests a possible future perspective for AQP4 inhibitors to be tested to treat CNS edema after TBI even though there is still a lack of studies in humans.

9. Conclusions

DAI is a leading cause of mortality and permanent morbidity worldwide. Despite numerous efforts, the clinical prognosis of the high-grade DAI remains, to date, in many cases, dismal. Multiple prognostic factors have been assessed in the last decade. Among those factors, the importance of early GCS, peripheral oxygen saturation, glucose serum levels, and blood pressure have confirmed their clinical significance in high-impact trials. The role of the oxidative stress in regard to the intracellular ROS balance and Ca²⁺ cellular metabolism is increasingly being investigated, with noteworthy preliminary results. Markers such as GFAP, pNF-H, NF-L, microtubule-associated protein tau, A β 42, S-100 β , NSE, AQP4, Drp-1, and NCX have demonstrated significant effectiveness in predicting clinical outcomes in animal studies, but, at present, their assessment in human studies is still confined to small case series groups and is not included in the guidelines for the management of TBI-related DAI. Regarding possible pharmacological agents that could control and reduce morphological and ROS damages related to DAI, SFN, and TFP, Dynasore and SN-6 have shown promising results *in vitro* and in animal studies; however, further investigations are needed to verify their safety and effectiveness though it has been demonstrated that the administration of cyclosporin A does not lead to any clinically relevant effects in humans.

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Abbreviations

| | |
|--------------|---|
| TBI | Traumatic Brain Injury |
| DAI | Diffuse Axonal Injury |
| ROS | Reactive Oxygen Species |
| ISS | Injury Severity Score |
| NISS | New Injury Severity Score |
| MAIS | Maximum Abbreviated Injury Scale |
| GOSE | Glasgow Outcome Scale-Extended |
| GCS | Glasgow Coma Scale |
| mPTP | Mitochondrial Permeability Transition Pore |
| NF | Neurofilament Protein |
| pNF-H | Neurofilament Heavy Subunit |
| NF-L | Neurofilament Light Chain |
| GFAP | Glial Fibrillary Acid Protein |
| BBB | Blood–Brain Barrier |
| NSE | Neuron Specific Enolase |
| CSF | Cerebrospinal Fluid |
| A β 42 | Amyloid β Peptide |
| MRI | Magnetic Resonance Imaging |
| CT | Computed Tomography |
| NCX | Reverse-mode Sodium Calcium Exchanger |
| Drp-1 | Dynamin Related Protein-1 |
| Nrf 2 | Nuclear factor erythroid 2-related factor 2 |
| HO-1 | Hemeoxygenase-1 |
| ARE | Antioxidant Response Element |
| SNF | Sulphoraphane |
| Drp-1 | Dynamin related protein-1 |
| DTI | Diffuse Tensor Imaging |
| oxLDLs | oxidized low-density lipoproteins |
| HNE | 4-Hydroxy-2-nonenal |
| NQO1 | NAD(P)H:quinone oxidoreductase 1 |
| Keap1 | Kelch sample related protein |
| PBS | Phosphate-Buffered Saline |
| AQP4 | Aquaporin-4 |
| TFP | Trifluoperazine |
| CNS | Central Nervous System |

References

1. Gerber, L.M.; Chiu, Y.L.; Carney, N.; Härtl, R.; Ghajar, J. Marked reduction in mortality in patients with severe traumatic brain injury. *J. Neurosurg.* **2013**, *119*, 1583–1590. [[CrossRef](#)]
2. Finfer, S.R.; Cohen, J. Severe traumatic brain injury. *Resuscitation* **2001**, *48*, 77–90. [[CrossRef](#)]
3. Johnson, V.E.; Stewart, W.; Smith, D.H. Axonal pathology in traumatic brain injury. *Exp. Neurol.* **2013**, *246*, 35–43. [[CrossRef](#)]
4. Finkelstein, E.A.; Corso, P.S.; Miller, T.R. *The Incidence and Economic Burden of Injuries in the United States*; Oxford University Press: New York, NY, USA, 2006.
5. Park, S.J.; Hur, J.W.; Kwon, K.Y.; Rhee, J.J.; Lee, J.W.; Lee, H.K. Time to recover consciousness in patients with diffuse axonal injury: Assessment with reference to magnetic resonance grading. *J. Korean Neurosurg. Soc.* **2009**, *46*, 205–209. [[CrossRef](#)]
6. Gennarelli, T.A.; Spielman, G.M.; Langfitt, T.W.; Gildenberg, P.L.; Harrington, T.; Jane, J.A.; Marshall, L.F.; Miller, J.D.; Pitts, L.H. Influence of the type of intracranial lesion on outcome from severe head injury. *J. Neurosurg.* **1982**, *56*, 26–32. [[CrossRef](#)]
7. Levi, L.; Guilburd, J.N.; Lemberger, A.; Soustiel, J.F.; Feinsod, M. Diffuse axonal injury: Analysis of 100 patients with radiological signs. *Neurosurgery* **1990**, *27*, 429–432. [[CrossRef](#)]
8. Park, S.W.; Park, K.; Kim, Y.B.; Min, B.K.; Hwang, S.N.; Suk, J.S.; Choi, D.Y. Prognostic factors in diffuse axonal injuries of brain. *J. Korean Neurosurg. Soc.* **1991**, *20*, 983–990.
9. Williams, J.M.; Gomes, F.; Drudge, O.W.; Kessler, M. Predicting outcome from closed head injury by early assessment of trauma severity. *J. Neurosurg.* **1984**, *61*, 581–585. [[CrossRef](#)]
10. Vieira, R.C.; Paiva, W.S.; de Oliveira, D.V.; Teixeira, M.J.; de Andrade, A.F.; de Sousa, R.M. Diffuse axonal injury: Epidemiology, outcome and associated risk factors. *Front. Neurol.* **2016**, *20*, 178. [[CrossRef](#)]
11. Sobuwa, S.; Hartzenberg, H.B.; Geduld, H.; Uys, C. Predicting outcome in severe traumatic brain injury using a simple prognostic model. *S. Afr. Med. J.* **2014**, *104*, 492–494. [[CrossRef](#)]
12. Manley, G.; Knudson, M.M.; Morabito, D.; Damron, S.; Erickson, V.; Pitts, L. Hypotension, hypoxia, and head injury: Frequency, duration, and consequences. *Arch. Surg.* **2001**, *136*, 1118–1123. [[CrossRef](#)]
13. Newfield, P.; Pitts, L.; Kaktis, J.; Hoff, J. The influence of shock on mortality after head trauma. *Crit. Care Med.* **1980**, *8*, 254. [[CrossRef](#)]
14. Jeffreys, R.V.; Jones, J.J. Avoidable factors contributing to the death of head injury patients in general hospitals in Mersey Region. *Lancet* **1981**, *29*, 459–461. [[CrossRef](#)]
15. Hill, D.A.; Abraham, K.H.; West, R.H. Factors affecting outcome in the resuscitation of severely injured patients. *Aust. N. Z. J. Surg.* **1993**, *63*, 604–609. [[CrossRef](#)]
16. Mayer, T.; Walke, M.L.; Shasha, I.; Matlak, M.; Johnson, D.G. Effect of multiple trauma on outcome of pediatric patients with neurologic injuries. *Childs Brain* **1981**, *8*, 189–197. [[CrossRef](#)]
17. Haddad, S.H.; Arabi, Y.M. Critical care management of severe traumatic brain injury in adults. *Scand. J. Trauma Resusc. Emerg. Med.* **2012**, *3*, 12. [[CrossRef](#)]
18. Carney, N.; Totten, A.M.; O'Reilly, C.; Ullman, J.S.; Hawryluk, G.W.; Bell, M.J.; Bratton, S.L.; Chesnut, R.; Harris, O.A.; Kissoon, N.; et al. Guidelines for the management of severe traumatic brain injury, fourth edition. *Neurosurgery* **2017**, *1*, 6–15. [[CrossRef](#)]
19. Clifton, G.L.; Ziegler, M.G.; Grossman, R.G. Circulating catecholamines and sympathetic activity after head injury. *Neurosurgery* **1981**, *8*, 10–13. [[CrossRef](#)]
20. Rosner, M.J.; Newsome, H.H.; Becker, D.P. Mechanical brain injury: The sympathoadrenal response. *J. Neurosurg.* **1984**, *61*, 76–86. [[CrossRef](#)]
21. Bilotta, F.; Giovannini, F.; Caramia, R.; Rosa, G. Glycemia management in neurocritical care patients: A review. *J. Neurosurg. Anesthesiol.* **2009**, *21*, 2–9. [[CrossRef](#)]
22. Liu-DeRyke, X.; Collingridge, D.S.; Orme, J.; Roller, D.; Zurasky, J.; Rhoney, D.H. Clinical impact of early hyperglycemia during acute phase of traumatic brain injury. *Neurocrit. Care* **2009**, *11*, 151–157. [[CrossRef](#)]
23. Young, B.; Ott, L.; Dempsey, R.; Haack, D.; Tibbs, P. Relationship between admission hyperglycemia and neurologic outcome of severely brain-injured patients. *Ann. Surg.* **1989**, *210*, 466–472. [[CrossRef](#)]
24. Lam, A.M.; Winn, H.R.; Cullen, B.F.; Sundling, N. Hyperglycemia and neurological outcome in patients with head injury. *J. Neurosurg.* **1991**, *75*, 545–551. [[CrossRef](#)]
25. Chelly, H.; Chaari, A.; Daoud, E.; Dammak, H.; Medhioub, F.; Mnif, J.; Hamida, C.B.; Bahloul, M.; Bouaziz, M. Diffuse axonal injury in patients with head injuries: An epidemiologic and prognosis study of 124 cases. *J. Trauma* **2011**, *71*, 838–846. [[CrossRef](#)]
26. Adams, J.H.; Graham, D.I.; Murray, L.S.; Scott, G. Diffuse axonal injury due to nonmissile head injury in humans: An analysis of 45 cases. *Ann. Neurol.* **1982**, *12*, 557–563. [[CrossRef](#)]
27. Adams, J.H.; Doyle, D.; Ford, I.; Gennarelli, T.A.; Graham, D.I.; McLellan, D.R. Diffuse axonal injury in head injury: Definition, diagnosis and grading. *Histopathology* **1989**, *15*, 49–59. [[CrossRef](#)]
28. Adams, J.H.; Doyle, D.; Graham, D.I.; Lawrence, A.E.; McLellan, D.R. Microscopic diffuse axonal injury in cases of head injury. *Med. Sci. Law* **1985**, *25*, 265–269. [[CrossRef](#)]
29. Hill, C.S.; Coleman, M.P.; Menon, D.K. Traumatic axonal injury: Mechanisms and translational opportunities. *Trends Neurosci.* **2016**, *39*, 311–324. [[CrossRef](#)]
30. Frati, A.; Cerretani, D.; Fiaschi, A.I.; Frati, P.; Gatto, V.; La Russa, R.; Pesce, A.; Pinchi, E.; Santurro, A.; Fraschetti, F.; et al. Diffuse axonal injury and oxidative stress: A comprehensive review. *Int. J. Mol. Sci.* **2017**, *18*, 2600. [[CrossRef](#)]

31. Davceva, N.; Basheska, N.; Balazic, J. Diffuse axonal injury—A distinct clinicopathological entity in closed head injuries. *Am. J. Forensic Med. Pathol.* **2015**, *36*, 127–133. [[CrossRef](#)]
32. Gennarelli, T.A.; Thibault, L.E.; Adams, J.H.; Graham, D.I.; Thompson, C.J.; Marcincin, R.P. Diffuse axonal injury and traumatic coma in the primate. *Ann. Neurol.* **1982**, *12*, 564–574. [[CrossRef](#)]
33. Kim, C.H.; Lee, H.K.; Koh, Y.C.; Hwang, D.Y. Clinical analysis of diffuse axonal injury (DAI) diagnosed with magnetic resonance image (MRI). *J. Korean Neurosurg. Soc.* **1997**, *26*, 241–248.
34. Kim, H.J.; Park, I.S.; Kim, J.H.; Kim, K.J.; Hwang, S.H.; Kim, E.S.; Jung, J.M.; Han, J.W. Clinical analysis of the prognosis of the patients with cerebral diffuse axonal injuries, based on gradient-echo MR imaging. *J. Korean Neurosurg. Soc.* **2001**, *30*, 168–172.
35. Baker, S.P.; O'Neill, B.; Haddon, W.; Long, W.B. The injury severity score: A method for describing patients with multiple injuries and evaluating emergency care. *J. Trauma* **1974**, *14*, 187–196. [[CrossRef](#)]
36. Osler, T.; Baker, S.P.; Long, W. A modification of the injury severity score that both improves accuracy and simplifies scoring. *J. Trauma* **1997**, *43*, 922–925. [[CrossRef](#)]
37. Association for the Advancement of Automotive Medicine—AAAM. The Abbreviated Injury Scale (AIS): 2005, Update 2008. Available online: <https://www.aaam.org/abbreviated-injury-scale-ais/> (accessed on 5 June 2020).
38. Nogueira, L.S.; Domingues, C.A.; Campos, M.A.; Sousa, R.M.C. Ten years of new injury severity score (NISS): Is it a possible change? *Rev. Lat. Am. Enfermagem.* **2008**, *16*, 314–319. [[CrossRef](#)]
39. Wilson, J.T.; Pettigrew, L.E.; Teasdale, G.M. Structured interviews for the Glasgow Outcome Scale and the extended Glasgow Outcome Scale: Guidelines for their use. *J. Neurotrauma* **1998**, *15*, 573–585. [[CrossRef](#)]
40. Wierzbicki, V.; Pesce, A.; Marrocco, L.; Piccione, E.; Colonnese, C.; Caruso, R. How old is your cervical spine? Cervical spine biological age: A new evaluation scale. *Eur. Spine J.* **2015**, *24*, 2763–2770. [[CrossRef](#)]
41. Armocida, D.; Pesce, A.; Frati, A.; Miscusi, M.; Paglia, F.; Raco, A. Pneumoventricle of unknown origin: A personal experience and literature review of a clinical enigma. *World Neurosurg.* **2019**, *122*, 661–664. [[CrossRef](#)]
42. Teasdale, G.; Jennett, B. Assessment of coma and impaired consciousness. A practical scale. *Lancet* **1974**, *304*, 81–84.
43. Matsukawa, H.; Shinoda, M.; Fujii, M.; Takahashi, O.; Murakata, A.; Yamamoto, D. Acute alcohol intoxication, diffuse axonal injury and intraventricular bleeding in patients with isolated blunt traumatic brain injury. *Brain Inj.* **2013**, *27*, 1409–1414. [[CrossRef](#)]
44. Calvi, M.R.; Beretta, L.; Dell'Acqua, A.; Anzalone, N.; Licini, G.; Gemma, M. Early prognosis after severe traumatic brain injury with minor or absent computed tomography scan lesions. *J. Trauma* **2011**, *70*, 447–451.
45. Skandsen, T.; Kvistad, K.A.; Solheim, O.; Strand, I.H.; Folvik, M.; Vik, A. Prevalence and impact of diffuse axonal injury in patients with moderate and severe head injury: A cohort study of early magnetic resonance imaging findings and 1-year outcome. *J. Neurosurg.* **2010**, *113*, 556–563. [[CrossRef](#)]
46. Pinchi, E.; Frati, A.; Cipolloni, L.; Aromatario, M.; Gatto, V.; La Russa, R.; Pesce, A.; Santurro, A.; Frascchetti, F.; Frati, P.; et al. Clinical-pathological study on β -APP, IL-1 β , GFAP, NFL, Spectrin II, 8OHdG, TUNEL, miR-21, miR-16, miR-92 expressions to verify DAI-diagnosis, grade and prognosis. *Sci. Rep.* **2018**, *5*, 2387. [[CrossRef](#)]
47. Cheng, G. Mitochondria in traumatic brain injury and mitochondrial-targeted multipotential therapeutic strategies. *Br. J. Pharmacol.* **2012**, *167*, 699–719. [[CrossRef](#)]
48. Halestrap, A.P. What is the mitochondrial permeability transition pore? *J. Mol. Cell Cardiol.* **2009**, *46*, 821–831. [[CrossRef](#)]
49. Mazzeo, A.T. The role of mitochondrial transition pore, and its modulation, in traumatic brain injury and delayed neurodegeneration after TBI. *Exp. Neurol.* **2009**, *218*, 363–370. [[CrossRef](#)]
50. Barsukova, A.G. Focal increases of axoplasmic Ca²⁺, aggregation of sodium–calcium exchanger, N-type Ca²⁺ channel, and actin define the sites of spheroids in axons undergoing oxidative stress. *J. Neurosci.* **2012**, *32*, 12028–12037. [[CrossRef](#)]
51. Büki, A.; Okonkwo, D.O.; Povlishock, J.T. Postinjury cyclosporin A administration limits axonal damage and disconnection in traumatic brain injury. *J. Neurotrauma* **1999**, *16*, 511–521. [[CrossRef](#)]
52. Okonkwo, D.O.; Povlishock, J.T. An intrathecal bolus of cyclosporin A before injury preserves mitochondrial integrity and attenuates axonal disruption in traumatic brain injury. *J. Cereb. Blood Flow Metab.* **1999**, *19*, 443–451. [[CrossRef](#)]
53. Okonkwo, D.O.; Buki, A.; Siman, R.; Povlishock, J.T. Cyclosporin A limits calcium-induced axonal damage following traumatic brain injury. *Neuroreport* **1999**, *10*, 353–358. [[CrossRef](#)]
54. Barrientos, S.A. Axonal degeneration is mediated by the mitochondrial permeability transition pore. *J. Neurosci.* **2011**, *31*, 966–978. [[CrossRef](#)]
55. Staal, J.A. Initial calcium release from intracellular stores followed by calcium dysregulation is linked to secondary axotomy following transient axonal stretch injury. *J. Neurochem.* **2010**, *112*, 1147–1155. [[CrossRef](#)]
56. Forte, M.; Gold, B.G.; Marracci, G.; Chaudhary, P.; Basso, E.; Johnsen, D.; Yu, X.; Fowlkes, J.; Rahder, M.; Stem, K.; et al. Cyclophilin D inactivation protects axons in experimental autoimmune encephalomyelitis, an animal model of multiple sclerosis. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 7558–7563. [[CrossRef](#)]
57. Aminmansour, B.; Fard, S.A.; Habibabadi, M.R.; Moein, P.; Norouzi, R.; Naderan, M. The efficacy of Cyclosporine-A on diffuse axonal injury after traumatic brain injury. *Adv. Biomed. Res.* **2014**, *3*, 35.
58. Empey, P.E.; McNamara, P.J.; Young, B.; Rosbolt, M.B.; Hatton, J. Cyclosporin A disposition following acute traumatic brain injury. *J. Neurotrauma* **2006**, *23*, 109–116. [[CrossRef](#)]

59. Hatton, J.; Rosbolt, B.; Empey, P.; Kryscio, R.; Young, B. Dosing and safety of cyclosporine in patients with severe brain injury. *J. Neurosurg.* **2008**, *109*, 699–707. [[CrossRef](#)]
60. Mazzeo, A.T.; Brophy, G.M.; Gilman, C.B.; Alves, O.L.; Robles, J.R.; Hayes, R.L.; Povlishock, J.T.; Bullock, M.R. Safety and tolerability of cyclosporin a in severe traumatic brain injury patients: Results from a prospective randomized trial. *J. Neurotrauma* **2009**, *26*, 2195–2206. [[CrossRef](#)]
61. Büki, A.; Okonkwo, D.; Wang, K.K.; Povlishock, J.T. Cytochrome c release and caspase activation in traumatic axonal injury. *J. Neurosci.* **2000**, *20*, 2825–2834. [[CrossRef](#)]
62. McAllister, T.W. Neurobiological consequences of traumatic brain injury. *Dialogues Clin. Neurosci.* **2011**, *13*, 287–300.
63. Gatson, J.W.; Barillas, J.; Hynan, L.S.; Diaz-Arrastia, R.; Wolf, S.E.; Minei, J.P. Detection of neurofilament-H in serum as a diagnostic tool to predict injury severity in patients who have suffered mild traumatic brain injury. *J. Neurosurg.* **2014**, *121*, 1232–1238. [[CrossRef](#)]
64. Shibahashi, K.; Doi, T.; Tanaka, S.; Hoda, H.; Chikuda, H.; Sawada, Y.; Takasu, Y.; Chiba, K.; Nozaki, T.; Hamabe, Y.; et al. The serum phosphorylated neurofilament heavy subunit as a predictive marker for outcome in adult patients after traumatic brain injury. *J. Neurotrauma* **2016**, *33*, 1826–1833. [[CrossRef](#)]
65. Ljungqvist, J.; Zetterberg, H.; Mitsis, M.; Blennow, K.; Skoglund, T. Serum neurofilament light protein as a marker for diffuse axonal injury: Results from a case series study. *J. Neurotrauma* **2017**, *1*, 1124–1127. [[CrossRef](#)]
66. Nylén, K.; Ost, M.; Csajbok, L.Z.; Nilsson, I.; Blennow, K.; Nellgård, B.; Rosengren, L. Increased serum-GFAP in patients with severe traumatic brain injury is related to outcome. *J. Neurol. Sci.* **2006**, *240*, 85–91. [[CrossRef](#)]
67. Neri, M.; Frati, A.; Turillazzi, E.; Cantatore, S.; Cipolloni, L.; Di Paolo, M.; Frati, P.; La Russa, R.; Maiese, A.; Scopetti, M.; et al. Immunohistochemical Evaluation of Aquaporin-4 and its Correlation with CD68, IBA-1, HIF-1 α , GFAP, and CD15 Expressions in Fatal Traumatic Brain Injury. *Int. J. Mol. Sci.* **2018**, *19*, 3544. [[CrossRef](#)]
68. Zemlan, F.P.; Jauch, E.C.; Mulchahey, J.J.; Gabbita, S.P.; Rosenberg, W.S.; Speciale, S.G.; Zuccarello, M. C-tau biomarker of neuronal damage in severe brain injured patients: Association with elevated intracranial pressure and clinical outcome. *Brain Res.* **2002**, *947*, 131–139. [[CrossRef](#)]
69. Roberts, G.W.; Gentleman, S.M.; Lynch, A.; Murray, L.; Landon, M.; Graham, D.I. Beta amyloid protein deposition in the brain after severe head injury: Implications for the pathogenesis of Alzheimer's disease. *J. Neurol. Neurosurg. Psychiatry* **1994**, *57*, 419–425. [[CrossRef](#)]
70. Bogoslovsky, T.; Wilson, D.; Chen, Y.; Hanlon, D.; Gill, J.; Jeromin, A.; Song, L.; Moore, C.; Gong, Y.; Kenney, K.; et al. Increases of plasma levels of glial fibrillary acidic protein, tau, and amyloid β up to 90 days after traumatic brain injury. *J. Neurotrauma* **2017**, *34*, 66–73. [[CrossRef](#)]
71. Raabe, A.; Grolms, C.; Sorge, O.; Zimmermann, M.; Seifert, V. Serum S-100B protein in severe head injury. *Neurosurgery* **1999**, *45*, 477–483. [[CrossRef](#)]
72. Romner, B.; Ingebrigtsen, T.; Kongstad, P.; Børgesen, S.E. Traumatic brain damage: Serum S-100 protein measurements related to neuroradiological findings. *J. Neurotrauma* **2000**, *17*, 641–647. [[CrossRef](#)]
73. Ingebrigtsen, T.; Romner, B. Biochemical serum markers for brain damage: A short review with emphasis on clinical utility in mild head injury. *Restor. Neurol. Neurosci.* **2003**, *21*, 171–176.
74. Kleindienst, A.; Schmidt, C.; Parsch, H.; Emtmann, I.; Xu, Y.; Buchfelder, M. The passage of S100B from brain to blood is not specifically related to the blood-brain barrier integrity. *Cardiovasc. Psych. Neurol.* **2010**, *2010*, 801295. [[CrossRef](#)]
75. Anderson, R.E.; Hansson, L.O.; Nilsson, O.; Dijlai-Merzoug, R.; Settergren, G. High serum S100B levels for trauma patients without head injuries. *Neurosurgery* **2001**, *48*, 1255–1258.
76. Thelin, E.P.; Jeppsson, E.; Frostell, A.; Svensson, M.; Mondello, S.; Bellander, B.M.; Nelson, D.W. Utility of neuron-specific enolase in traumatic brain injury; relations to S100B levels, outcome, and extracranial injury severity. *Crit. Care* **2016**, *8*, 285. [[CrossRef](#)]
77. Pelinka, L.E.; Kroepfl, A.; Leixnering, M.; Buchinger, W.; Raabe, A.; Redl, H. GFAP versus S100B in serum after traumatic brain injury: Relationship to brain damage and outcome. *J. Neurotrauma* **2004**, *21*, 1553–1561. [[CrossRef](#)]
78. Kövesdi, E.; Lückl, J.; Bukovics, P.; Farkas, O.; Pál, J.; Czeiter, E.; Szellár, D.; Dóczy, T.; Komoly, S.; Büki, A. Update on protein biomarkers in traumatic brain injury with emphasis on clinical use in adults and paediatrics. *Acta Neurochir.* **2010**, *152*, 1–17. [[CrossRef](#)]
79. Berger, R.P.; Beers, S.R.; Richichi, R.; Wiesman, D.; Adelson, P.D. Serum biomarker concentrations and outcome after pediatric traumatic brain injury. *J. Neurotrauma* **2007**, *24*, 1793–1801. [[CrossRef](#)] [[PubMed](#)]
80. Wu, D.M.; Zheng, Z.H.; Fan, Z.H.; Zhang, Z.F.; Chen, G.Q.; Lun, J. Sulforaphane administration alleviates diffuse axonal injury (DAI) via regulation signaling pathway of NRF2 and HO-1. *J. Cell Biochem.* **2020**, *121*, 430–442. [[CrossRef](#)]
81. Ishii, T.; Itoh, K.; Ruiz, E.; Leake, D.S.; Unoky, H.; Yamamoto, M.; Mann, G.E. Role of Nrf2 in the regulation of CD36 and stress protein expression in murine macrophages: Activation by oxidatively modified LDL and 4-hydroxynonenal. *Circ. Res.* **2004**, *94*, 609–616. [[CrossRef](#)]
82. Wakabayashi, N.; Slocum, S.L.; Skoko, J.J.; Shin, S.; Kensler, T.W. When NRF2 talks, who's listening? *Antioxid. Redox Signal.* **2010**, *13*, 1649–1663. [[CrossRef](#)]
83. Gao, P.; Zhou, M.; Ouyang, Y.; Wang, H. Activation of the nuclear factor erythroid 2-related factor 2-antioxidant response element signal in rats with diffuse axonal injury. *Neuroreport* **2019**, *30*, 389–396. [[CrossRef](#)]

84. Kang, K.W.; Lee, S.J.; Kim, S.G. Molecular mechanism of nrf2 activation by oxidative stress. *Antioxid. Redox Signal.* **2005**, *7*, 1664–1673. [[CrossRef](#)]
85. Dinkova Kostova, A.T.; Kostov, R.V. Glucosinolates and isothiocyanates in health and disease. *Trends Mol. Med.* **2012**, *18*, 337–347. [[CrossRef](#)]
86. Omelchenko, A.; Shrirao, A.B.; Bhattiprolu, A.K.; Zhan, J.D.; Schloss, R.S.; Dickson, S.; Meaney, D.F.; Boustany, N.N.; Yarmush, L.L.; Firestein, B.L. Dynamin and reverse-mode sodium calcium exchanger blockade confers neuroprotection from diffuse axonal injury. *Cell Death Dis.* **2019**, *10*, 727. [[CrossRef](#)]
87. Chou, A.C.; Ju, Y.T.; Pan, C.Y. Calmodulin interacts with the sodium/calcium exchanger NCX1 to regulate activity. *PLoS ONE* **2015**, *10*, e0138856.
88. Morrison, B., III; Elkin, B.S.; Dolle, J.P.; Yarmush, M.L. In vitro models of traumatic brain injury. *Annu. Rev. Biomed. Eng.* **2011**, *13*, 91–126. [[CrossRef](#)]
89. Jeffs, G.J.; Meloni, B.P.; Bakker, A.J.; Knuckey, N.W. The role of the Na⁺/Ca²⁺ exchanger (NCX) in neurons following ischaemia. *J. Clin. Neurosci.* **2007**, *14*, 507–514. [[CrossRef](#)]
90. Weber, J.T. Altered calcium signaling following traumatic brain injury. *Front. Pharmacol.* **2012**, *3*, 60. [[CrossRef](#)]
91. Kitchen, P.; Salman, M.M.; Halsey, A.M.; Clarke-Bland, C.; MacDonald, J.A.; Ishida, H.; Vogel, H.J.; Almutiri, S.; Logan, A.; Kreida, S.; et al. Targeting aquaporin-4 subcellular localization to treat central nervous system edema. *Cell* **2020**, *181*, 784–799.e19.
92. Salman, M.M.; Kitchen, P.; Halsey, A.; Wang, M.X.; Tornroth-Horsefield, S.; Conner, A.C.; Badaut, J.; Iliff, J.J.; Bill, R.M. Emerging roles for dynamic aquaporin-4 subcellular relocalization in CNS water homeostasis. *Brain* **2021**, online ahead of print. [[CrossRef](#)]
93. Sylvain, N.J.; Salman, M.M.; Pushie, M.J.; Hou, H.; Meher, V.; Herlo, R.; Peeling, L.; Kelly, M.E. The effects of trifluoperazine on brain edema, aquaporin-4 expression and metabolic markers during the acute phase of stroke using photothrombotic mouse model. *Biochim. Biophys. Acta Biomembr.* **2021**, *1863*, 183573. [[CrossRef](#)] [[PubMed](#)]