

resistant organisms may be inhibited by a carbapenem antibiotic until sufficient carbapenemase production has been achieved and traditional AST platforms must wait to make MIC calls. More accurate carbapenem MICs can be determined by implementing a carbapenemase test alongside rapid AST.

Methods. We demonstrate a novel, proprietary test to detect carbapenemase production that enables rapid MIC testing for carbapenem antibiotics. The test is processed in parallel with the Selux next-generation phenotyping (NGP) AST method, enabling rapid, <6-hour, accurate MIC determinations. The carbapenemase assay utilizes high concentrations of intact bacteria. After 3 hours of incubation, a fluorescent pH indicator is read spectroscopically. The solution pH is lowered by carbapenemase-mediated imipenem degradation and is indicative of enzyme activity.

Results. This assay accurately identifies carbapenemases across multiple enzyme classes and bacterial species. Figure 1 shows the accuracy and speed of NGP AST at determining MICs for representative isolates from the FDA-CDC antimicrobial resistance bank compared with results from overnight broth microdilution (BMD). To date, over 100 challenge strains of *Enterobacteriaceae*, *Pseudomonas aeruginosa*, and *Acinetobacter baumannii* have been tested with no very major errors and an average time-to-result of 5.3 hours.

Conclusion. By incorporating a rapid, on-board carbapenemase activity assay, the NGP AST platform rapidly delivers accurate carbapenem results. Combined with NGP's comprehensive antibiotic menus, this platform will therefore ensure prompt delivery of personalized antibiotic therapies for all patients, including those infected with MDROs, and enable streamlined antibiotic stewardship coordination.

Isolate	Carbapenemase	Overnight BMD MIC	NGP IMP MIC	NGP Time to Result (hrs)
<i>E cloacae</i> AR 154	VIM	≥ 8	8	5
<i>E cloacae</i> AR 161	IMP	4	8	5
<i>E cloacae</i> AR 32	KPC	2	8	5
<i>E cloacae</i> AR 38	NDM	≥ 8	≥ 8	5
<i>E coli</i> AR 61	KPC	4	4	5
<i>E coli</i> AR 69	NDM	8	8	5
<i>K aerogenes</i> AR 7	None	0.5	0.5	5
<i>K pneumoniae</i> AR 153	NDM	≥ 8	≥ 8	5
<i>K pneumoniae</i> AR 361	KPC	8	8	5
<i>K pneumoniae</i> AR 504	OXA	4	4	5
<i>A baumannii</i> AR 52	OXA	8	≥ 8	5
<i>K pneumoniae</i> ATCC 700603	None	< 0.12	< 0.12	5
<i>P aeruginosa</i> AR 54	VIM	≥ 8	≥ 8	6

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2160. Performance of the Cepheid Rapid PCR Test for Patient Screening and Association with Efficacy of Suvratoxumab, A Novel Anti-*Staphylococcus aureus* Monoclonal Antibody, During the Phase 2 SAATELLITE study

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Background. Patients with lower airway *Staphylococcus aureus* (SA) colonization are at great risk (> 20%) of early-onset ventilator-associated pneumonia (VAP). Thus, a rapid test is required to identify patients at risk. Suvratoxumab (formerly MEDI4893) is a human monoclonal antibody that neutralizes SA alpha toxin. SAATELLITE, a phase 2 study of safety and efficacy of suvratoxumab for reducing the incidence of SA pneumonia (NCT02296320) was conducted and recently completed within the consortium for Combating Bacterial Resistance in Europe. We investigated the performance of a

rapid PCR test (Xpert MRSA/SA SST1[®], Cepheid) as a screening tool during the study and the association between SA load and suvratoxumab efficacy.

Methods. The PCR assay was used to detect SA and methicillin-resistant SA (MRSA) in lower respiratory tract (LRT) samples. Culture was performed on PCR SA+ LRT samples according to local procedures. PCR SA+ subjects were randomized 1:1 to either a single intravenous infusion of 5000 mg suvratoxumab (*n* = 96) or placebo (*n* = 100) and followed for 190 days post dose. Efficacy of suvratoxumab was defined as relative risk reduction (RRR) in incidence of SA pneumonia within 30 days post-dose compared with placebo.

Results. 299 (41.5%) out of 720 screened subjects were SA+ by PCR. Of 209 subjects with culture data, there were 162 (77.5%) SA+, 47 (22.5%) SA- and 9 (5.6%) MRSA by culture. Culture results could have been affected by antibiotic use and site variability in limits of detection ranging from 3.3 to 100,000 colony-forming units per mL (CFU/mL). No discordance was noted between PCR and culture for MRSA detection. An inverse linear correlation was observed between the PCR cycle threshold (Ct) values for SA protein A gene (*spa*) and SA CFU/mL counts from quantitative culture. In subjects with low SA load (Ct ≥ 29; *n* = 72), suvratoxumab provided 66.7% RRR [90% confidence interval (CI): 21.3%, 86.2%] compared with 31.9% RRR [90% CI: -7.5%, 56.8%] in total study population.

Conclusion. Cepheid Xpert PCR assay was easy to perform, sensitive and standardized, and provided better sensitivity than conventional culture for detection of SA. Additionally, quantitative PCR Ct output was associated with the efficacy of suvratoxumab in reducing SA pneumonia incidence.

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2161. Organism-Specific Turn Around Time Improvement in Urinary Specimens as a Result of Microbiological Laboratory Automation

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Background. University Medical Center in Lubbock, TX is one of few medical centers using Becton Dickinson (BD) Kiestra Total Laboratory Automation (TLA) system since May 2015. The impact on organism-specific turn around time (TAT) in urinary specimens after implementation of TLA was evaluated.

Methods. After approval from the Quality Improvement Review Board, a retrospective analysis of microbiological data from urinary specimens in BD research database was performed. Before vs. after implementation (2013 vs. 2016) TAT was compared. Ten clinically relevant organisms were analyzed. Statistical analysis was performed with SAS software version 9.2. Data were analyzed using *Chi-squared test*. A *P*-value of < 0.05 was considered statistically significant.

Results. Overall, 2282 specimens from 2013 and 2306 specimens from 2016 were analyzed. Compared with before vs. after implementation of TLA, an overall improvement in TAT was observed (expressed as mean hours for each organism): *Enterococcus faecalis* (55.2 vs. 38.8), *Enterococcus faecium* (68.4 vs. 43.8), *Escherichia coli* (44.2 vs. 41.0), *Klebsiella pneumoniae* (45.0 vs. 44.0), *Proteus mirabilis* (44.8 vs. 38.6), *Pseudomonas aeruginosa* (58.9 vs. 37.7), *Staphylococcus aureus* (49.2 vs. 36.0), *Streptococcus agalactiae* (49.2 vs. 31.4), *Streptococcus pneumoniae* (51.7 vs. 61.8), *Streptococcus pyogenes* (62.6 vs. 26.6). It was also observed that improvement in TAT was more pronounced for Gram-positive organisms than Gram-negative organisms. *P*-value was < 0.01 for all organisms except *Streptococcus pneumoniae* (0.7985) and *Streptococcus pyogenes* (0.2562). The number of specimens with these two organisms was too small to be considered significant.

Conclusion. Automation of microbiology laboratory leads to significant TAT improvement in urinary specimens, making early data availability to clinicians. This improves efficiency as well as supporting earlier antibiotic switch, antimicrobial stewardship and optimal patient care in treating urinary tract infections.

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2162. Comparison of Plazomicin Disk Diffusion vs. Gradient Diffusion Susceptibility Testing Results Against Drug-Resistant Clinical *Enterobacteriaceae* Isolates

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Background. Plazomicin, a novel aminoglycoside, is active against carbapenem-resistant *Enterobacteriaceae* (EB) and is not inhibited by most aminoglycoside modifying enzymes that affect gentamicin and tobramycin. We investigated the activity of plazomicin against resistant EB clinical isolates and compared disk diffusion (DD) vs. gradient diffusion (GD) results.

Methods. EB isolates that were carbapenem resistant and/or resistant to both gentamicin and tobramycin were retrieved from the UW Health clinical isolate repository. Each isolate was tested against plazomicin using both DD (MAST Group Ltd. Plazomicin disk 30 µg) and GD (Liofilchem Plazomicin MIC Test Strip 0.16–256 µg/