

Inhibitory effect of MyoD on the proliferation of breast cancer cells

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Abstract. Skeletal muscle is rich in lymphatic vessels, with an abundant blood supply, and it is an infrequent site of cancer metastasis. Previous studies have demonstrated that enhanced secretion of MyoD may occur when skeletal muscle is injured or becomes cancerous. It was hypothesized that MyoD may act as an endogenous cytokine to inhibit the proliferation of cancer cells. To verify the possible effect of this protein on tumor cell proliferation, C2C12 mouse skeletal muscle cells and 4T1 mouse breast cancer cells were co-cultured using embedded Transwell plates. Following co-culture, cell cycle analysis revealed that C2C12 muscle cells were able to inhibit the proliferation of the breast cancer cells. Subsequently, MyoD was silenced in C2C12 cells to assess its effect on 4T1 cell proliferation. Following co-culture with MyoD-silenced cells, a 5-ethynyl-20-deoxyuridine assay indicated that MyoD silencing prevented the reduction in proliferation of 4T1 cells induced by untransfected C2C12 cells. In summary, the results indicated that MyoD inhibits the proliferation of breast cancer cells and may be a tumor suppressor factor.

Introduction

MyoD was first cloned in 1987 and termed MyoD1 (1). The protein has a basic helical three-dimensional crystal structure containing a basic helix-loop-helix domain that is able to bind to other proteins that also possess this domain, including myocyte-enhancing factor 2, myogenin and creatine kinase (CK). Its adjacent basic region is required for it to bind to the promoters or enhancers of numerous muscle-specific genes, including CK and myogenin (1,2). The N-terminus of MyoD contains a histidine-cysteine domain and a transcription activation domain, which are associated with the transcriptional activation of MyoD target genes, and the C-terminus contains

a facultative helical (helix III) domain that may be associated with chromatin remodeling (3-5).

As a member of the muscle transcription factor family, MyoD has decisive roles in muscle differentiation, including muscle conversion and the maintenance of muscle differentiation (1,2). Recent studies have demonstrated that a synthetic MyoD polypeptide has a high affinity for the inhibitor of DNA-binding proteins (ID), and thus may inhibit the binding of ID with DNA, thereby inhibiting the proliferation of cancer cells (6). In addition, Dey *et al* (7) identified that MyoD is an important cytokine in cerebellar development and a tumor suppressor gene in medulloblastoma. These previous studies strongly indicate the existence of a close association between MyoD and cancer cells.

As a major organ, skeletal muscle is rich in lymphatic vessels with an abundant blood supply. However, few studies have demonstrated cancer metastasis to skeletal muscle tissue (8-12). MyoD expression may be increased following skeletal muscle injury or its invasion by cancer cells (13,14). The present study aimed to test the hypothesis that MyoD may act as an endogenous cytokine to inhibit the growth of metastatic cancer. Its expression was assessed in breast cancer tissue and cell lines and in C2C12 skeletal muscle cells, and the proliferation of breast cancer cells was evaluated following co-culture with control or MyoD-silenced skeletal muscle cells.

Materials and methods

Cell culture and co-culture. The immortalized mouse myoblast cell line C2C12 and the mouse breast tumor cell line 4T1 (each gifted by the Xiangya Central Experiment Laboratory, Changsha, China) were maintained at 37°C in an atmosphere of 5% CO₂ in Dulbecco's modified Eagle's medium (DMEM) supplemented with 100 U/ml penicillin, 100 U/ml streptomycin and 10% heat-inactivated fetal bovine serum (all purchased from Sigma-Aldrich China, Inc., Shanghai, China).

Transwell chambers (0.4- μ m pore size; Corning Incorporated, Corning, NY, USA) were placed into 6-well plates. The interior of the Transwell plate was designated the upper chamber, while the space between the plates formed the lower chamber, and the chambers were separated by a polycarbonate membrane. Due to the permeability of the polycarbonate membranes, components in the lower-layer medium are able to affect the growth and movement of cells

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placed in the upper chamber. In order to study the impact of cytokines secreted by skeletal muscle cells on cancer cells, Transwell chambers were used to form a co-culture, with skeletal muscle cells in the lower chamber and cancer cells in the upper chamber (15). C1C12 and 4T1 cells were firstly cultured in a culture flask to a cell concentration of 5×10^5 /ml for ~48 h until they reached 70% confluence. The C2C12 cells were subsequently transplanted onto a 6-well plate (Corning Incorporated) for 24 h, and the 4T1 cells were cultured in Transwell (Corning Incorporated). The cells were co-cultured for 48 h with the 4T1 cells in the upper chambers and the C1C12 cells in the lower chambers.

Immunohistochemical analysis. Breast cancer tissues and adjacent non-cancer tissues were obtained from 7 randomly selected patients diagnosed with breast cancer at the Xiangya Hospital of Central South University (Changsha, China). Breast cancer tissue was dissected away from normal tissue, fixed with 4% paraformaldehyde, embedded in paraffin and cut into 5- μ m sections. A primary mouse monoclonal anti-MyoD antibody (#sc-32758; Santa Cruz Biotechnology, Inc., Santa Cruz, CA, USA) was used to detect MyoD expression. Briefly, endogenous peroxidase was inhibited by soaking tissue sections in 3% H_2O_2 . After rinsing in phosphate-buffered saline (PBS), sections were incubated with goat serum (Sigma-Aldrich China, Inc.) to block the non-specific binding of antibodies, and sections were then incubated overnight at 4-8°C with the anti-MyoD primary antibody (dilution, 1:50). After washing in PBS, the sections were incubated with biotinylated goat anti-rabbit IgG polyclonal antibody (dilution, 1:1,000; #A6667; Sigma-Aldrich China, Inc.) for 1 h at room temperature and washed again. A streptavidin-biotin-peroxidase complex (#RPN1051-2ML; GE Healthcare Life Sciences, Shanghai, China) was then incubated with the sections for 60 min at room temperature. After washing in PBS, the signal was detected with 3,3-diaminobenzidine. A negative control in which the primary antibody was omitted was included for each biopsy. Written informed consent was obtained from all patients and ethical approval was provided by the Medical Ethics Committee of the Basic Medical College of Central South University (Changsha, China).

Immunofluorescence. Sections were freed from the paraffin, rehydrated, subjected to antigen retrieval in 10 mM sodium citrate, and treated with hydrogen peroxide. Sections were then blocked with 5% goat serum containing 3% Triton X-100 and incubated with the mouse monoclonal anti-MyoD antibody (dilution, 1:200) at 10 μ g/ml for 1 h at room temperature. Next, slides were incubated with ABC reagent (from the VECTASTAIN® Elite ABC kit; Vector Laboratories, Inc., Burlingame, CA, USA) and Alexa Fluor 568-conjugated goat anti-mouse IgG (dilution, 1:1,000; #A-11004; Thermo Fisher Scientific, Inc., Carlsbad, CA, USA), washed, and incubated with Tyramide Signal Amplification reagent (Thermo Fisher Scientific, Inc.) according to the manufacturer's instructions. Finally, the slides were washed and mounted using fluorescence mounting medium (Dako Omnis; Agilent Technologies, Santa Clara, CA, USA) containing 500 μ g/l 4',6-diamidino-2-phenylindole for nuclei staining. Nuclei were then counterstained with 2% purified methyl green for 2 min.

Table I. siRNA sequences and properties.

siRNA	Sequences	GC%
01	5'-GCCUGAGCAAAGUGAAUGA-dTdT-3'	39
	3'-dTdT-CGGACUCGUUUCACUUACU-5'	39
02	5'-CAGCAGACGACUUCUAUGA-dTdT-3'	39
	3'-dTdT-GUCGUCUGCUGAAGAUACU-5'	39
03	5'-CCAACUGCUCUGAUGGCAU-dTdT-3'	43
	3'-dTdT-GGUUGACGAGACUACCGUA-5'	43

The slides incubated in the dark for 24 h prior to examination with a Zeiss LSM 510 laser-scanning confocal microscope (Zeiss GmbH, Jena, Germany).

MyoD staining intensity was determined using a color video camera (Sony DXC-950P; Sony Corporation, Tokyo, Japan) connected to a Leica Q500 IW Imaging Workstation with MoticFluo software v1.0 (Leica Microsystems, Cambridge, UK). A semi-quantitative scoring method was employed by three independent observers who were blinded to the conditions in order to record MyoD staining expression; scores were assigned on a scale of 0-4 (0, no staining; 4, maximum staining) according to the staining intensities.

Small interfering RNA (siRNA) synthesis and transfection. Candidate siRNAs directed against MyoD mRNA were designed by Guangzhou RiboBio Co., Ltd. (Guangzhou, China). Three potential siRNAs were selected corresponding with the prediction of single-strand domains within the mRNA secondary structure (Table I). BLAST analyses (<http://blast.ncbi.nlm.nih.gov/Blast.cgi>) were performed to ensure that no additional significantly matching mouse transcripts would be targeted by these siRNAs. MyoD and nonsense siRNAs were synthesized by Guangzhou RiboBio Co., Ltd., and transfections were conducted with Invitrogen Lipofectamine 2000 (Thermo Fisher Scientific, Inc.), according to the manufacturer's instructions. Reverse transcription (RT)-polymerase chain reaction (PCR) and western blotting were used to detect MyoD expression following siRNA transfection.

Semi-quantitative RT-PCR. The sequences of the primers for mouse MyoD were as follows: MyoD forward, 5'-CTC CTTGAGACAGCAGACGACTT-3', and reverse, 5'-AAA TCGCATTGGGGTTTGAGCCTG-3'; and β -actin forward, 5'-GAAACTACCTTCAACTCCATC-3', and reverse, 5'-CGA GGCCAGGATGGAGCCCGCC-3'. Primers were designed by Primer-BLAST (www.ncbi.nlm.nih.gov/tools/primer-blast/). β -actin was used to ascertain the presence of an equal amount of cDNA in each reaction. A TRIzol® kit (Thermo Fisher Scientific, Inc.) was used to extract genomic RNA. Total RNA (1 μ g) purified from siRNA-transfected cells was reverse transcribed into cDNA using AMV reverse transcriptase (Qiagen, Venlo, Netherlands) with an RNase inhibitor (Thermo Fisher Scientific, Inc.) and oligo(dT) primer (Thermo Fisher Scientific, Inc.) at 40°C for 50 min, followed by heating at 90°C for 5 min. Next, 1 μ l reverse-transcriptase was added to a 30 μ l PCR mixture [Bio-Rad Laboratories (Singapore) Pte. Ltd., Singapore] for 30 cycles. Taq polymerase was added

from the SuperScript® III One-Step RT-PCR system (Thermo Fisher Scientific, Inc.), and each PCR cycle consisted of 93°C for 30 sec and 54°C for 60 sec. Negative controls consisted of an equal volume of water substituted for the volume of RNA in the RT reaction. mRNA expression data for sample-to-sample variability in RNA input, RNA quality and reverse transcription efficiency was normalized to β -actin.

Western blotting. MyoD protein expression was detected by western blotting using the aforementioned MyoD monoclonal antibody. For the preparation of cell extracts, cells from different groups were washed three times with ice-cold PBS and then lysed in lysis buffer [50 mM Tris-HCl (pH 8.0), 150 mM NaCl, 1 mM ethylenediaminetetraacetic acid, 1% Triton X-100 and 100 μ g/ml phenylmethylsulfonyl fluoride] on ice for 20 min. Following centrifugation at 16,000 \times g for 2 min at 4°C, supernatants were separated using 10% sodium dodecyl sulfate-polyacrylamide gel electrophoresis, and then electrophoretically transferred to nitrocellulose membranes (Invitrogen; Thermo Fisher Scientific, Inc.). After blocking for 2 h with 5% fat-free milk at room temperature, the membranes were incubated with the primary mouse monoclonal anti-MyoD antibody or control mouse monoclonal anti- β -actin secondary antibody (dilution, 1:1,000; #A1978; Sigma-Aldrich China, Inc.) for 24 h at 4°C. The membranes were then incubated with a secondary biotinylated goat anti rabbit IgG polyclonal antibody (dilution, 1:1,000; #A6667; Sigma-Aldrich China, Inc.) for 1 h at room temperature. Protein bands were visualized using Pierce enhanced chemiluminescence reagent (Thermo Fisher Scientific, Inc.) and Odyssey v1.2 software (LI-COR Biosciences, Lincoln, NE, USA). The intensity of expression was measured by comparing the target and control bands.

Cell cycle analysis using propidium iodide (PI) and flow cytometry. Cell cycle analysis was conducted at 72 h after transfection. 4T1 cells (5×10^5) from the tested groups were harvested by brief trypsinization, washed twice with PBS, fixed in 70% ethanol overnight and stained with PI (final concentration, 20 mg/ml)/Triton X-100 solution containing 10 mg/ml RNase (DNase-free (Thermo Fisher Scientific, Inc.). Following incubation at 37°C for 30 min, the samples were analyzed using a FACSCalibur flow cytometer (BD Biosciences, Franklin Lakes, NJ, USA), and the populations of cells in G1, S and G2 were quantified. The BD FACSDiva software (BD Biosciences) can export data files in FCS 2.0 or 3.0 default formats. The ModFit LT v3.0 software package (Verity Software House, Topsham, ME, USA) was used. The Click-iT® EdU Alexa Fluor® 488 Imaging Kit (Thermo Fisher Scientific, Inc.) is a superior alternative to traditional proliferation assays that is optimized for fluorescence microscopy applications.

5-ethynyl-20-deoxyuridine (EdU) assay. EdU is a nucleoside analog of thymidine that is incorporated into DNA during active DNA synthesis by proliferating cells, and may be visualized by the addition of a fluorescent molecule. Thus, proliferating 4T1 cells were detected using a Cell-Light™ EdU Apollo®567 *in vitro* Imaging Kit (Guangzhou RiboBio Co., Ltd.) according to the manufacturer's protocol. Briefly, cells were incubated with 500 μ l of 50 μ M EdU for 3 h before

fixation, permeabilization and visualization of EdU staining. Cell nuclei were stained with 10 mg/ml Hoechst 33342 (Guangzhou RiboBio Co., Ltd) for 30 min. Quantification of the staining intensity was determined using a color video camera (DXC-950P; Sony, Tokyo, Japan). The camera was connected to a Leica Imaging Workstation with MoticFluo 1.0 imaging software (Leica Q500IW; Leica, Cambridge, UK).

Statistical analysis. All experiments were performed at least five times. All data are presented as the mean \pm standard error of the mean. For all determinations, the differences were considered significant when $P < 0.05$. The unpaired *t*-test was used for comparing two groups. All statistical analyses were performed using SPSS software version 18 (SPSS, Inc., Chicago, IL, USA).

Results

Proliferation of 4T1 cells may be suppressed by C2C12 cells. To identify whether skeletal muscle cells can inhibit the proliferation of cancer cells, mouse breast tumor cells (4T1 cells) and mouse myoblast cells (C2C12 cells) were co-cultured on Transwell plates. Mouse breast tumor cells and mouse breast tumors were co-cultured as controls. PI staining and flow cytometry were used to detect the proliferation of the 4T1 cells. As shown in Fig. 1, at 48 h after co-culture, 65% of the cells were in G1 phase and 33% were in S phase in the experimental group, compared to 31% in G1 phase and 56% that were in S phase in the control group (G1: $P = 0.0376$ vs. control group; S: $P = 0.0396$ vs. control group; G2/M: $P = 0.0479$ vs. control group; $n = 6$), demonstrating that the proliferation of the 4T1 cells was inhibited following co-culture with the C2C12 cells (Fig. 1A and B).

Silencing efficiency of MyoD siRNAs. RNA interference was used to generate a C2C12 cell line with targeted silencing of MyoD. To verify the silencing efficiency of MyoD siRNA, three candidate siRNAs were designed and numbered 01, 02 and 03, respectively. The expression of MyoD mRNA in C2C12 cells was detected using RT-PCR following siRNA transfection at the recommended concentration of 100 nM. As shown in Fig. 2A and B, all three siRNAs effectively downregulated MyoD expression in the C2C12 cells ($P = 0.00134$ vs. control siRNA; $n = 3$), and the silencing efficiency of siRNA 01 reached ~70%. Therefore siRNA 01 was selected for use in subsequent experiments.

siRNA 01 was transfected into C2C12 cells using Lipofectamine 2000 and western blotting was performed to investigate MyoD protein expression at 72 h after transfection (Fig. 2C and D). The results demonstrated that the expression of MyoD was markedly reduced compared with the control group, which consisted of untransfected cells. Gray value analysis indicated that MyoD expression in the experimental group was decreased by ~70% compared with the control group ($P = 0.00149$ vs. control siRNA; $n = 3$), which was consistent with the RT-PCR results, and the nonsense siRNA had no effect on MyoD expression.

Proliferation of 4T1 cells is inhibited by MyoD. To explore the effects of MyoD and other cytokines from skeletal muscle

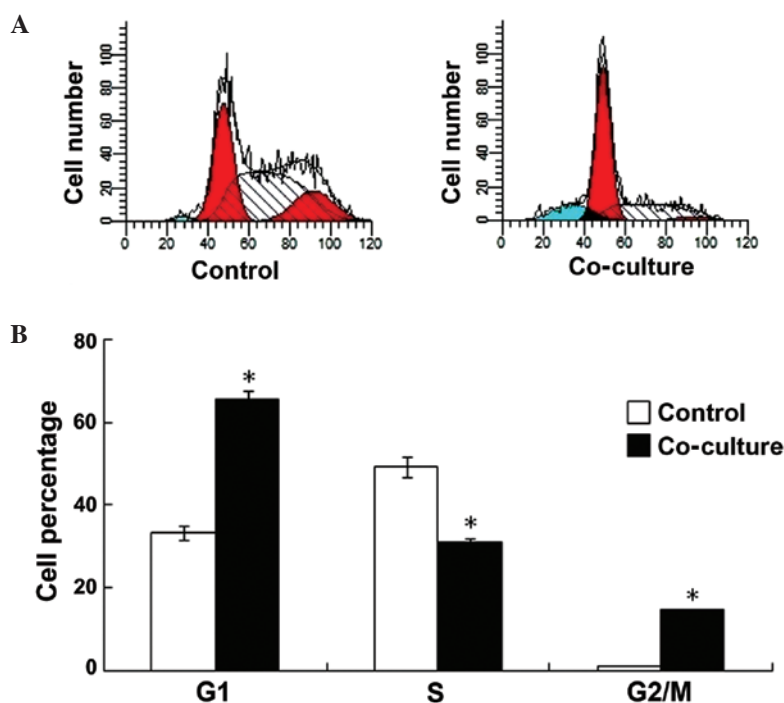


Figure 1. Proliferation of 4T1 cells was suppressed by co-culture with C2C12 skeletal muscle cells. (A) After 48 h of co-culture, the populations of cells in G1, S and G2/M phases from the different groups were separated and counted. (B) Cell populations in the G1, S and G2/M phases are shown in histograms produced using ModFit LT v3.0 software package. * $P < 0.05$ vs. control (4T1 cells cultured in the absence of C2C12 cells).

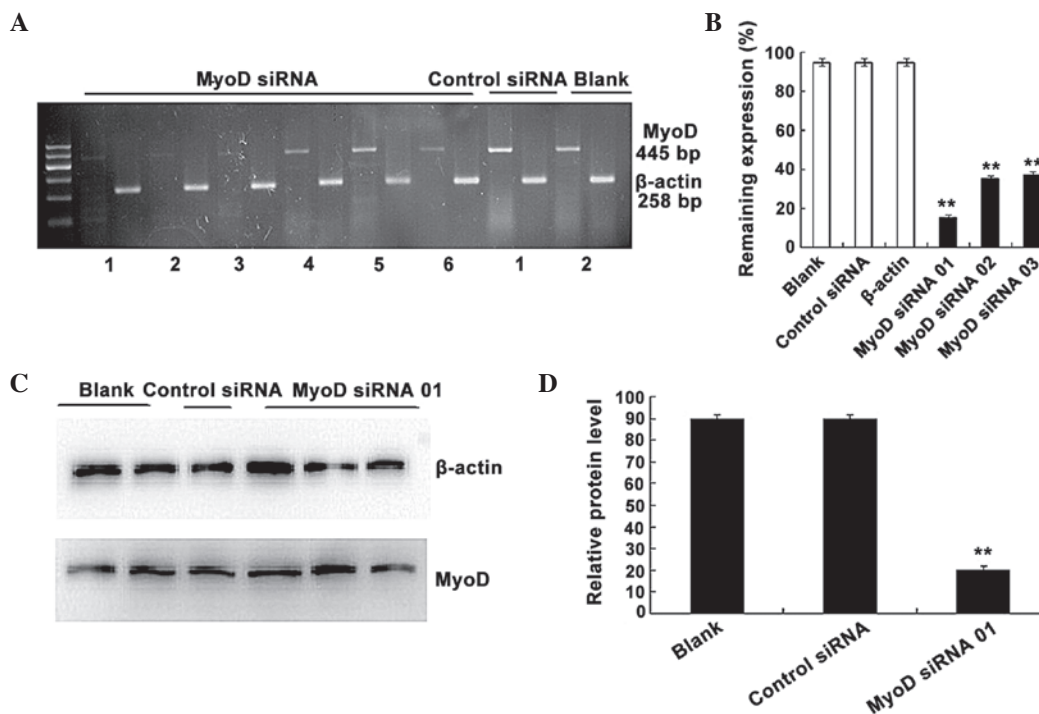


Figure 2. Silencing efficiency of MyoD siRNAs. (A) Silencing of MyoD mRNA in C2C12 cells. Three candidate MyoD siRNAs [siRNA 01 (lanes 1-3), siRNA 02 (lanes 4-5) and siRNA 03 (lane 6)], and 100 nmol of non-silencing control siRNA was transfected into C2C12 cells in triplicate. Semi-quantitative RT-PCR was conducted at 48 h after siRNA transfection. (B) Semi-quantification of RT-PCR normalized to β -actin levels (** $P < 0.01$ vs. control). (C) The most effective siRNA, siRNA 01, and the control siRNA were prepared and transfected into C2C12 cells. Three days after transfection, MyoD protein expression was detected by western blotting. (D) Quantification of western blot results normalized to the β -actin expression level. ** $P < 0.01$ vs. control.

cells on cancer cells, MyoD-silenced C2C12 cells were co-cultured with 4T1 cells in Transwell plates and EdU assays were performed to detect cancer cell proliferation following 48 h of co-culture (Fig. 3A and B). Fluorescence microscopy

clearly showed that the 4T1 cells alone actively proliferated and exhibited extremely strong EdU fluorescent labeling (~50% of cell population; $n=3$). The EdU fluorescence of the 4T1 cells co-cultured with the untransfected C2C12 cells

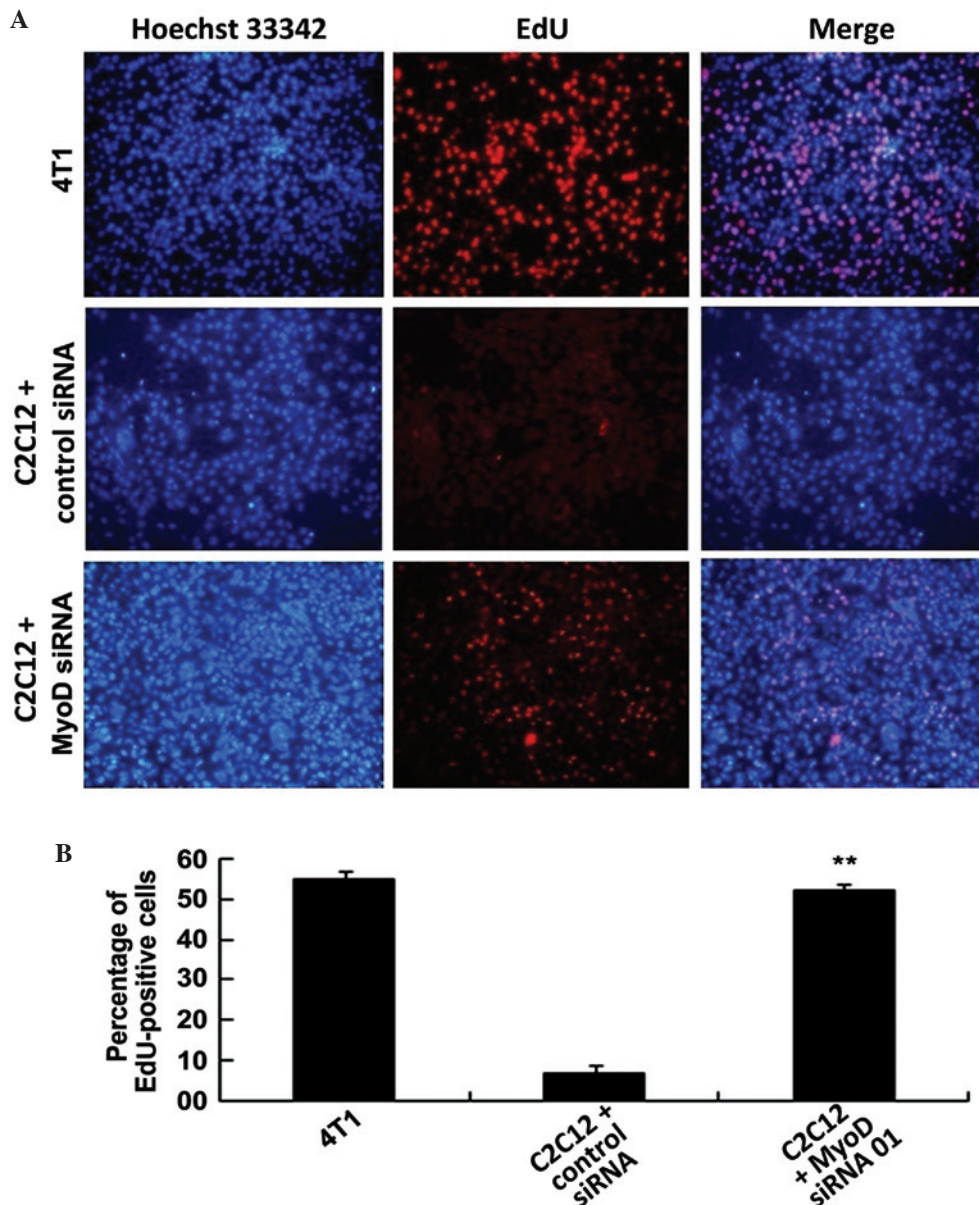


Figure 3. Proliferation of 4T1 cells can be inhibited by MyoD. (A) 4T1 cells were labeled with EdU following co-culture with MyoD siRNA-transfected or control siRNA-transfected C2C12 cells, or with only 4T1 cells, in Transwell chambers. The Click-iT reaction revealed EdU staining (red), and cell nuclei were stained with Hoechst 33342 (blue). The images are representative of the results obtained. (B) The percentage of EdU-positive 4T1 cells was quantified. The data are presented as the mean \pm standard error of the mean. ** $P < 0.01$ vs. 4T1.

was extremely sparse and was detected in $\sim 10\%$ of all cells (untransfected C2C12 vs. 4T1, $P = 0.00648$; $n = 3$). However, the EdU fluorescence of the 4T1 cells co-cultured with the C2C12 cells that had been transfected with MyoD siRNA was moderately bright, accounting for $\sim 40\%$ of all cells with gray signals (C2C12+MyoD siRNA vs. 4T1, $P = 0.00130$; $n = 3$). The proliferation of the 4T1 cells co-cultured with the C2C12 cells transfected with nonsense siRNA was similar to that of the 4T1 cells co-cultured with the untransfected C2C12 cells (C2C12+control siRNA vs. 4T1, $P = 0.00539$; $n = 3$). These results indicate that MyoD is able to markedly inhibit the proliferation of the 4T1 cells.

MyoD is weakly expressed in 4T1 and human breast cancer cells. To detect whether cancer cells express MyoD, TRIzol was used to extract genomic RNA and total protein from 4T1

cells after 48 h of culturing in 6-well plates and then performed RT-PCR and western blotting. Only faint bands were observed by RT-PCR ($P = 0.00278$ vs. β -actin; $n = 3$) and western blotting ($P = 0.00324$ vs. β -actin; $n = 3$), indicating weak MyoD expression in the 4T1 cells (Fig. 4A-D).

To further characterize MyoD expression in breast cancer tissue, breast cancer and normal tissue samples were obtained from 7 randomly selected breast cancer patients and assessed its expression using immunofluorescence and immunohistochemistry. The two assays revealed weak MyoD expression in the breast cancer and control human breast tissues (immunofluorescence: 0.97 ± 0.01 , $P = 0.00448$ vs. control, $n = 7$, unpaired t -test; immunohistochemistry: 0.95 ± 0.01 , $P = 0.00377$ vs. control, $n = 7$, unpaired t -test) (Fig. 4E-H). In addition, Fig. 4 shows evident differences between the cancer and control tissues. The present study aimed to test

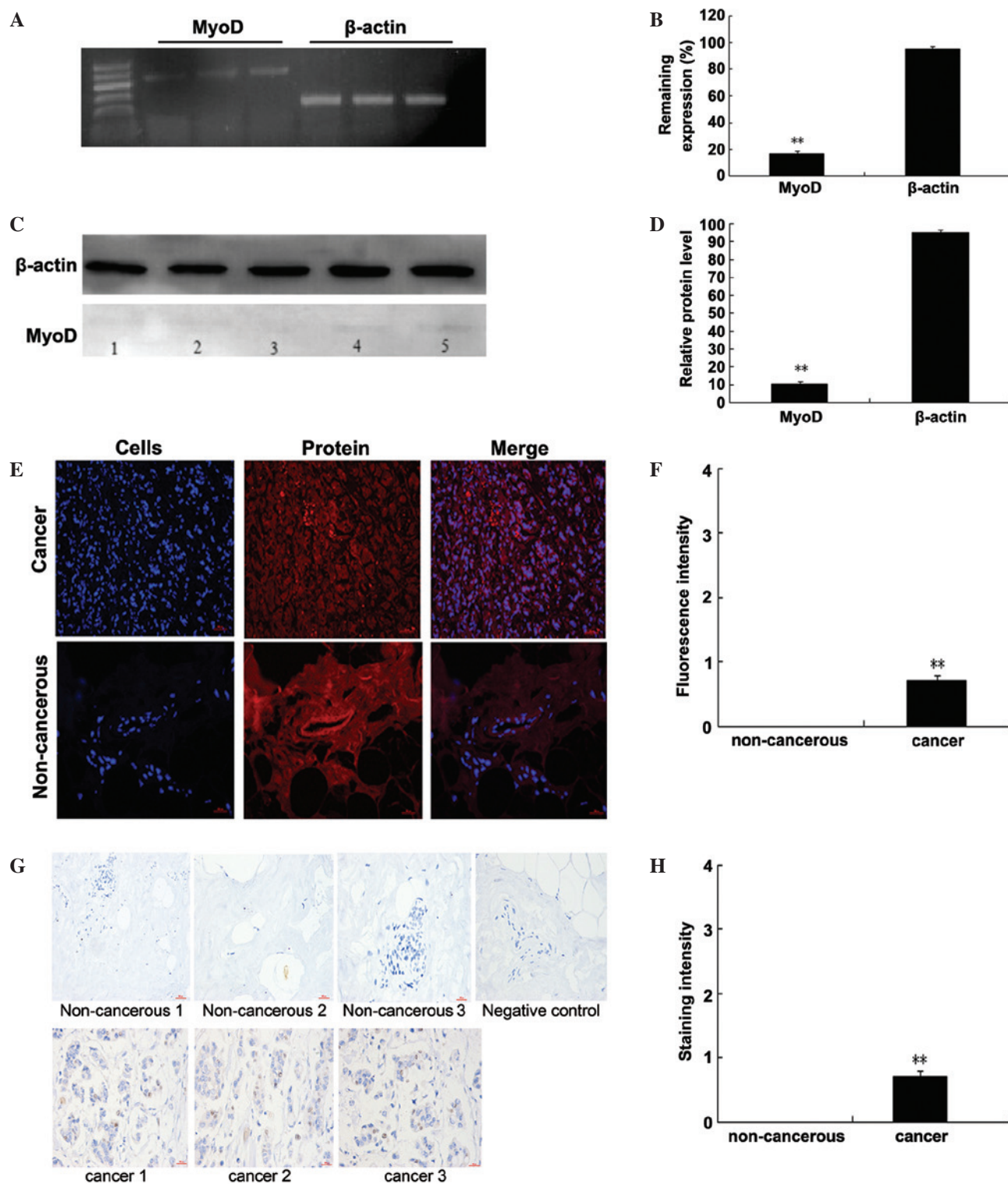


Figure 4. MyoD was weakly expressed in breast cancer tissues. (A) Results of reverse transcription-polymerase chain reaction analysis of MyoD mRNA expression and (B) semi-quantification relative to β -actin (** $P<0.01$ vs. β -actin). (C) Results of western blot analysis of MyoD protein expression and (D) quantification relative to β -actin (** $P<0.01$ vs. β -actin). (E) Representative MyoD expression in human breast cancer and control tissues as revealed by immunofluorescence and (F) semi-quantitative scores (** $P<0.01$ vs. non-cancerous). (G) Representative MyoD expression in human breast cancer and control tissues as revealed by immunohistochemistry and (H) semi-quantitative scores (** $P<0.01$ vs. non-cancerous).

the expression of MyoD in the tissues. The other differences between the control tissues and the cancer may be due to the atypia of cancer cells. To the best of our knowledge, this is the first report of low levels of MyoD expression in breast cancer tissue.

Discussion

Skeletal muscle is widely distributed and is an infrequent site of cancer metastasis (9). The current study was conducted to test the hypothesis that an endogenous tumor suppressor factor

may be associated with the low incidence of cancer metastasis in skeletal muscle. MyoD secretion increases when skeletal muscle is injured or invaded by cancer cells, suggesting that there is an association between MyoD and tissue wound repair (1). We hypothesized that MyoD may be an endogenous tumor suppressor factor that is also associated with the low occurrence of cancer in skeletal muscle (16).

MyoD is a DNA-binding protein that also has a significant role in skeletal muscle differentiation due to its importance in muscle conversion (6). Recently, Dey *et al* (7) demonstrated that MyoD is an important cytokine during cerebellar development and is a tumor suppressor gene in medulloblastoma. In fact, MyoD may regulate gene expression as a DNA-binding protein. Chen *et al* (6) used synthetic peptide fragments of MyoD to block the binding of DNA with ID, which is an important regulator of cell proliferation. After ID binding is blocked, cancer cell proliferation decreases (17), indicating a possible pathway by which MyoD may inhibit the proliferation of these cells. However, MyoD is a large protein, and its ability to enter the cell and affect DNA duplication require further verification (18,19).

As it is difficult to monitor the biological activity of MyoD *in vitro*, a MyoD-silenced model of mouse myoblast C2C12 cells was constructed in the present study. The C2C12 cells were co-cultured with 4T1 mouse breast cancer cells in Transwell chambers to explore the effects of MyoD on the proliferation of cancer cells. PI and EdU assays were used to assess cancer cell proliferation and the results revealed that the proliferation of the 4T1 cells was markedly inhibited by the C2C12 cells. PI staining results revealed that the population of cancer cells in S phase was 20% lower following co-culture with skeletal muscle cells, and the population in G1 phase was 35% higher than that of the control group. These results indicate that skeletal muscle cells may inhibit cancer cell proliferation by regulating the cell cycle.

To further assess the effects of MyoD on cancer cell proliferation, mouse breast tumor cells were co-cultured with MyoD-silenced mouse myoblast cells. Proliferation of the 4T1 cancer cells was significantly inhibited in the group that was co-cultured with the control (untransfected) C2C12 cells. However, in the group that was co-cultured with the MyoD-silenced C2C12 cells, the 4T1 cells exhibited no change in proliferative activity compared with the 4T1 cells cultured in the absence of C2C12 cells. These results suggest that MyoD was responsible for inhibiting the proliferation of the 4T1 cells, indicating that this protein may act as an endogenous factor to inhibit cancer cell proliferation (20).

The current study also evaluated MyoD expression in 4T1 cells by RT-PCR and western blotting. MyoD protein was found to be expressed at low levels in the 4T1 cells. In addition, its expression was assessed in the control human breast tissue and breast cancer tissue samples obtained from randomly selected breast cancer patients; no obvious MyoD expression was observed in the non-cancerous tissues, whereas this protein was expressed in the breast cancer tissue. Similarly to other tumor suppressor genes that are expressed only in cancer cells, such as α -fetoprotein (21), MyoD was expressed in the cancer cells at levels insufficient to inhibit their proliferation. A recent study performed by Dey *et al* (7) has shown that MyoD is a tumor suppressor gene in medulloblastoma. The current results

indicated that MyoD may act as a tumor suppressor gene in 4T1 cells; however, this conclusion requires further verification. In addition, the low incidence of skeletal muscle metastasis is not limited to one type of cancer cell, suggesting that MyoD may be a suppressor of multiple types of cancer (8,9).

MyoD also serves important roles in muscle transformation in the skeletal muscle microenvironment. Several previous studies have reported the successful transformation of fat cells into skeletal muscle cells by MyoD transfection *in vitro* (22), and this technique has been widely applied in chicken, pork and beef production (23-25). Skeletal muscle spends a long period of time undergoing tissue differentiation, maturation and repair (26,27). Cancer cells are a class of cells with high proliferative abilities (28-30). We suspect that it may also be possible to transform cancer cells into normal muscle tissue by taking advantage of the muscle conversion function of MyoD, the accomplishment of which would represent progress in cancer research.

In conclusion, the current study demonstrates for the first time that MyoD plays a critical role in cancer development by inhibiting the proliferation of cancer cells. Furthermore, it may act as a tumor suppressor gene in multiple types of cancer cells. These results will aid in the elucidation of the mechanisms underlying the low incidence of cancer metastasis in skeletal muscle.

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