



Draft Genome Sequence of *Clostridium botulinum* Subtype *bont/A5(B2')*

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ABSTRACT Here, we report the draft genome sequence of *Clostridium botulinum* strain CDC76130, which harbors a rare botulinum toxin gene (*bont*) complex arrangement of *bont/A5* and truncated *bont/B2* within the same *ha* toxin gene cluster.

Clostridium botulinum is a Gram-positive, spore-forming, anaerobic bacterium that is capable of producing neurotoxins known as botulinum neurotoxins (BoNTs), some of the most potent and threatening toxins to human and animal health (1). According to their ability to neutralize serotype-specific antisera, BoNTs are classified into seven distinct serotypes (A to G) and subdivided in more than 40 subtypes, which are determined by at least 2.6% differences in the amino acid sequences of the BoNT gene (*bont*) (2).

The *bont* genes are associated with protein accessory genes, arranged in two possible clusters, *ha* or *orfX* (3). Depending on the BoNT serotype or subtype, the gene cluster may vary in sequence and organization, for example, *bont/B* and *bont/A5* are always found within a *ha* cluster, and *bont/A2* to *bont/A4*, *bont/A6* to *bont/A8*, and *bont/F* are located in an *orfX* cluster. Additionally, *bont/A1* and bivalent strains are uniquely capable of harboring either of the two *bont* cluster types (4).

During a routine outbreak investigation of an infant botulism case from the Midwest region of the United States, CDC76130 was isolated from a fecal sample (5). A single colony was inoculated into Trypticase-peptone-glucose-yeast (TPGY) broth and grown anaerobically for 16 h prior to genomic DNA extraction. DNA extraction was performed using a modified MasterPure complete DNA and RNA purification kit (Lucigen, Middleton, WI) (6). The protocol utilized in this study was approved by the Centers for Disease Control and Prevention (CDC) Institutional Review Board (protocol approval number 6911).

Libraries were generated with a DNA preparation kit and sequenced using a MiniSeq system (2 × 150-bp chemistry) (Illumina, San Diego, CA), according to the manufacturer's instructions. Quality assurance and control and assembly of a total of 4,539,446 paired-end reads were performed using the SneakerNet pipeline v0.20.1 (7). The quality of the assembly was estimated with QUAST v5.1.0rc1 (8), and the draft genome assembly yielded a total read length of 3,931,884 bp (N_{50} 435,765 bp), assembled into 30 contigs, with an average coverage depth of 134× and a GC content of 28.02%. Default settings were used for all software unless otherwise noted.

Traditional seven-gene multilocus sequence typing (MLST) (9, 10) revealed that isolate CDC76130 harbored a new *mdh* allele. The new allele and new sequence type (ST), ST-153, were submitted to the *C. botulinum* MLST database (<http://pubmlst.org/cbotulinum>). Using the CLC Genomics Workbench v20.0.2 (Qiagen, Denmark) Map Reads to Reference tool with default parameters, subtype *bont/A5* and a truncated *bont/B2* were identified. The genome was annotated with the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) v5.3 (11), which resulted in the identification of 3,672 genes, including 3,609 coding sequences (CDSs) and 63 total RNA genes.

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Sequence analysis of the *bont* gene cluster revealed a *ha* gene cluster arrangement very similar to the ones identified for *C. botulinum* strains IBCA94-0216 (12) and H04402 065 (13), with the same deletion between the *ha-33* gene and the *botR* gene and two copies of IS3 transposases preceding the truncated, nonfunctional *bont*/B2.

Data availability. The draft genome assembly is available in NCBI GenBank under the accession number [JAKETJ000000000](https://doi.org/10.1093/bioinformatics/btt086), and the raw reads have been deposited in the NCBI Sequence Read Archive (SRA) under the accession number [SRR17486303](https://doi.org/10.1093/bioinformatics/btt086).

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REFERENCES

1. Sobel J. 2005. Botulism. *Clin Infect Dis* 41:1167–1173. <https://doi.org/10.1086/444507>.
2. Peck MW, Smith TJ, Anniballi F, Austin JW, Bano L, Bradshaw M, Cuervo P, Cheng LW, Derman Y, Dörner BG, Fisher A, Hill KK, Kalb SR, Korkeala H, Lindstrom M, Lista F, Luquez C, Mazuet C, Pirazzini M, Popoff MR, Rossetto O, Rummel A, Sesardic D, Singh BR, Stringer SC. 2017. Historical perspectives and guidelines for botulinum neurotoxin subtype nomenclature. *Toxins (Basel)* 9:38. <https://doi.org/10.3390/toxins9010038>.
3. Hill KK, Xie G, Foley BT, Smith TJ, Munk AC, Bruce D, Smith LA, Brettin TS, Detter JC. 2009. Recombination and insertion events involving the botulinum neurotoxin complex genes in *Clostridium botulinum* types A, B, E and F and *Clostridium butyricum* type E strains. *BMC Biol* 7:66. <https://doi.org/10.1186/1741-7007-7-66>.
4. Rossetto O, Pirazzini M, Montecucco C. 2014. Botulinum neurotoxins: genetic, structural and mechanistic insights. *Nat Rev Microbiol* 12:535–549. <https://doi.org/10.1038/nrmicro3295>.
5. Centers for Disease Control and Prevention. 1998. Botulism in the United States (1899–1996): handbook for epidemiologists, clinicians, and laboratory workers. Centers for Disease Control and Prevention, Atlanta, GA.
6. Halpin JL, Foltz V, Dykes JK, Chatham-Stephens K, Lúquez C. 2021. *Clostridium botulinum* type B isolated from a wound botulism case due to injection drug use resembles other local strains originating from Hawaii. *Front Microbiol* <https://doi.org/10.3389/fmicb.2021.678473>.
7. Griswold T, Kapsak C, Chen J, Bakker H, Williams G, Kelley A, Vidyaprakash E, Katz L. 2021. SneakerNet: a modular quality assurance and quality check workflow for primary genomic and metagenomic read data. *J Open Source Softw* 6:2334. <https://doi.org/10.21105/joss.02334>.
8. Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. *Bioinformatics* 29:1072–1075. <https://doi.org/10.1093/bioinformatics/btt086>.
9. Jacobson MJ, Lin G, Whittam TS, Johnson EA. 2008. Phylogenetic analysis of *Clostridium botulinum* type A by multi-locus sequence typing. *Microbiology (Reading)* 154:2408–2415. <https://doi.org/10.1099/mic.0.2008/016915-0>.
10. Jolley KA, Maiden MC. 2010. BIGSdb: scalable analysis of bacterial genome variation at the population level. *BMC Bioinformatics* 11:595. <https://doi.org/10.1186/1471-2105-11-595>.
11. Tatusova T, DiCuccio M, Badretdin A, Chetvermin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>.
12. Dover N, Barash JR, Arnon SS. 2009. Novel *Clostridium botulinum* toxin gene arrangement with subtype A5 and partial subtype B3 botulinum neurotoxin genes. *J Clin Microbiol* 47:2349–2350. <https://doi.org/10.1128/JCM.00799-09>.
13. Carter AT, Mason DR, Grant KA, Franciosa G, Aureli P, Peck MW. 2010. Further characterization of proteolytic *Clostridium botulinum* type A5 reveals that neurotoxin formation is unaffected by loss of the *cntR* (*botR*) promoter sigma factor binding site. *J Clin Microbiol* 48:1012–1013. <https://doi.org/10.1128/JCM.01774-09>.