Research Article

Transcriptional Expressions of CXCL9/10/12/13 as Prognosis Factors in Breast Cancer

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CXCLs play critical roles in antitumor immunity by activating tumor-specific immune responses and stimulating tumor proliferation, thus affecting patient outcomes. However, the expression and prognostic values of CXCLs in breast cancer have not been well clarified. The aim of this study was to investigate the impact of CXCLs transcriptional expression on breast cancer patients. Oncomine database, GEPIA (Gene Expression Profiling Interactive Analysis), UALCAN, Kaplan–Meier Plotter, TIMER (Tumor Immune Estimation Resource), and DAVID were used in our study. The transcriptional levels of CXCL9/10/11/13 in breast cancer tissues were significantly elevated while the transcriptional levels of CXCL1/2/3/12 were decreased based on intersections of Oncomine database and GEPIA. Among them, breast cancer patients with high transcriptional levels of CXCL2/9/ 10/12/13 and low transcriptional level of CXCL3 were associated with a better prognosis. We also found that most of CXCLs expressions are significantly correlated with known prognostic factors, such as patient's age, major subclasses, individual cancer stages, and nodal metastasis status. In addition, the expression of CXCL9/10/12/13 was also indicated to be correlated with the infiltration of six types of immune cells (B cells, CD8+ T cells, CD4+ T cells, macrophages, neutrophils, and dendritic cells). The functions of differentially expressed CXCLs are primarily related to the immune response and cytokine-cytokine receptor interactions. Our results may provide novel evidence of new prognostic or predictive biomarkers for breast cancer patients.

1. Introduction

CXCLs, in which C stands for cysteine and X represents any amino acid, are a family of structurally similar inflammatory chemokines. By binding to cognate CXCRs, these CXCLs play indispensable roles in various biological processes, including angiogenic, angiostatic, tumor initiation, promotion, and progression [1–3]. CXCLs chemokine family were separated into two structurally distinct groups, ELR⁺ and ELR⁻, based on the presence or absence of a Glu-Leu-Arg motif at the N-terminus, which appears to be important in ligand/receptor interactions on evoking immune cells to eliminate tumor cells. The ELR⁺ members cover CXCL1 to CXCL3, CXCL5 to CXCL8, and CXCL17 while the ELR⁻ members include CXCL4, CXCL9 to CXCL13, and CXCL16. The latest study has revealed that CXCL family was vital to regulate tumor progression by interaction between tumor and the tumor microenvironment in pancreatic cancer [4]. As to renal cell carcinoma, low transcriptional levels of CXCL1/2/3/5/13 were associated with a significantly better prognosis [5]. Conversely, CXCL1 and CXCL2 were found upregulated in colorectal cancer, which play an important role in treatment resistance and metastasis [6]. Therefore, we surmised that CXCLs could be utilized as potential therapeutic targets and prognostic biomarkers for neoplasm.

Breast cancer is the most common malignant tumor and the leading cause of cancer deaths among women worldwide, accounting for 24.2% of newly diagnosed cancers and 15.2% of mortality [7]. Approximately 5–10% of female breast cancer patients might have metastatic disease at presentation and 13% of women with primary stage IV breast cancer survive 10 years after diagnosis [8]. Breast cancer is a highly heterogeneous disease and may be divided into three major subtypes according to different molecular features as follows: luminal group, basal-like group, and HER-2 overexpressing group [9]. Although some progress has been made in identifying the therapeutic targets and prognostic biomarkers, some biomarkers involved in tumor microenvironment may uncover new possible avenues to individualized treatment.

To date, several studies have demonstrated the expression profile and function of some CXCLs in breast cancer. For example, silencing CXCL7 could significantly reduce invasive and metastatic properties of breast cancer stem cells, thus leading to an improved outcome [10]. CXCL10 plays indispensable role in breast cancer patients for immune checkpoint-based therapies [11], while CXCL1 had been shown to be involved in resistance to cancer chemotherapies [12]. However, the role of distinct CXCLs family members remained unknown in the development and progression of breast cancer. We performed this study by analyzing the expression patterns and potential functions of different CXCLs family members in patients with breast cancer based on some public databases, thus mining some therapeutic targets and prognostic biomarkers.

2. Materials and Methods

2.1. Oncomine Database. Oncomine database (http://www. oncomine.org) is an online cancer microarray database and synthetic gene-wide data-mining platform [13]. We compared the transcriptional levels of 15 CXCLs members in different cancer tissues with their corresponding adjacent normal controls from Oncomine database, using Student's ttest to generate a p value. The cutoffs of p value and fold change were defined as 0.05 and 2, respectively.

2.2. Gene Expression Profiling Interactive Analysis. The Gene Expression Profiling Interactive Analysis (GEPIA) (http:// gepia.cancer-pku.cn/) integrates tremendous amount of tumor and non-tumor samples from The Cancer Genome Atlas (TCGA) and the Genotype-Tissue Expression (GTEx) database, providing differential expression analysis, correlation analysis, and patient survival analysis online [14]. In our study, GEPIA was used to analyze the expression of CXCLs in breast tumors with corresponding breast tissues. Cutoff of p value was 0.05 and Log 2 FC was 1 (fold change was 2). We also obtained the top 100 similar expression protein-coding genes with certain CXCLs family members in the "TCGA Tumor"—BRCA Tumor dataset by similar genes module for Gene Ontology (GO), Kyoto Encyclopedia of Genes and Genomes (KEGG) analysis.

2.3. UALCAN. UALCAN (http://ualcan.path.uab.edu) is a comprehensive web resource based on TCGA database level

3 RNA-seq and clinical data from 31 cancer types [15]. It can be used to estimate relative transcriptional expression of query gene between tumor and normal samples as well as relative clinicopathologic parameters on patient survival. In this study, UALCAN was used to analyze the association between mRNA expressions of 15 CXCLs family members in breast invasive carcinoma and known prognostic factors (patient's age, individual cancer stages, major subclasses, and nodal metastasis status).

2.4. Kaplan-Meier Plotter Analysis. The association of CXCLs expression with overall survival (OS) of patients with breast cancers was analyzed by Kaplan-Meier Plotter (http:// www.kmplot.com), which is able to assess the effect of 54k genes on survival in 21 cancer types [16]. Patients with breast cancer were divided into high and low expression group by best cutoff. The number-at-risk cases, HRs, and log rank p values can be found at the Kaplan-Meier Plotter web page. Relapse-free survival (RFS) of ER- (estrogen receptor-) positive group and ER-negative group was furthermore assessed. Statistically significant difference was considered when a log rank p value <0.05.

2.5. Tumor Immune Estimation Resource Database Analysis. Tumor Immune Estimation Resource (TIMER) (https:// cistrome.shinyapps.io/timer/) acts as a web server for systematical analysis of tumor-infiltrating immune cells across 32 tumor types from TCGA [17]. Gene module was used to explore correlation between CXCLs expression and abundance of immune infiltrates, including tumor purity, B cells, CD8+ T cells, CD4+ T cells, macrophages, neutrophils, and dendritic cells (DCs) by Spearman's correlation. Cancer types are listed as BRCA (breast invasive carcinoma), BRCA-Basal, BRCA-Her2, and BRCA-Luminal. The expression levels of CXCLs are log 2 RSEM.

2.6. Gene Ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) Analysis. DAVID (https://david.ncifcrf. gov/home.jsp) is an integrated database that helps investigators better understand the biological meaning behind a large list of genes. GO and KEGG analysis were conducted by DAVID and visualized with R project using a "ggplot2" package [18, 19]. Biological processes, cellular components, and molecular functions were included in the GO enrichment analysis, while KEGG analysis defined the pathways related to the CXCLs.

3. Results

3.1. Transcription Levels of CXCLs in Patients with Breast Cancer. Fifteen different CXCLs family members have been identified in *Homo sapiens*. In order to explore the potential prognostic and therapeutic value of different CXCLs members in breast cancer patients, we compared the transcriptional levels of CXCLs by using Oncomine database and GEPIA. As shown in Figure 1, Oncomine database revealed that mRNA expressions of CXCL9/10/11 were significantly higher in human breast cancer than in normal tissues in

	Cancer														
	vs.														
Analysis type by cancer	normal														
	CXCL1	CXCL2	CXCL3	CXCL4	CXCL5	CXCL6	CXCL7	CXCL8	CXCL9	CXCL10	CXCL11	CXCL12	CXCL13	CXCL16	CXCL17
Bladder cancer		1 2	1			1 2		1	2	2		5	1		
Brain and cns cancer	5		1	1	1	1 1		4 1	1	2	1 1	4	1	8	
Breast cancer	15	27	19	4	7	1 5	4		15	16	12	2 16	2	2 2	1
Cervical cancer	3		1		1	1		4	2	2	2	1	2	1	
Colorectal cancer	21	14 1	23	4	10	7	2	19	1 1	4	12	8	5	5	3
Esophageal cancer	7	4	4	2		3	1	5	1	1	1	1	_	1	2
Gastric cancer	4				1 1			4	1		1	4		1	8
Head and neck cancer	10 1	3	3		1	2		5	6	7	6	3	4		2
Kidney cancer	1 3		2	2	2 2	1	7		4	4	3	7	1 2	3	
Leukemia	9	1 3	1 3	1 10	1		2 8	1 3	1		1	1 3	1	1	
Liver cancer	1 1	6			1	4	1	2	4 1	5	3	1 4			
Lung cancer	2	12	6	10	2		11		2			3	7	4	1
Lymphoma		3 2		1		1		5	21	16	7	9	13 1	3	
Melanoma	1				1 1			1	1	1		1			
Myeloma	1	1	1	1	1			1				2			
Other cancer	3	2 2	1 1		3	1	2 1	6	6 1	11	1	2	3	3 2	1
Ovarian cancer	1		1					1		1	1	1	2		
Pancreatic cancer		2	2		5	1	1	3	1	1		3	1	2	1
Prostate cancer	1	1				1	1	1	1		1	2 1	2		
Sarcoma	1	7			1 3	1	2		2	4	2	2 2	2		1
Significant unique analyses	58 33	30 64	37 24	7 29	28 16	24 10	8 34	57 10	72 3	76	53 2	18 70	38 12	29 9	6 14
Total unique analysis	447	440	407	433	392	409	440	464	452	435	392	458	413	296	253
1 5 10 10 5 1															

FIGURE 1: Transcriptional expression of CXCLs in 20 different types of cancers (Oncomine database).

multiple datasets and transcription levels of CXCL1/2/4/5/7 were prominently low. Then, GEPIA was applied to measure expression levels based on TCGA and the GTEx database. Expressions on box plots are presented in Figure 2. The results indicated that the expression levels of CXCL9, CXCL10, CXCL11, and CXCL13 were marked higher in breast cancer tissues than in normal tissues, while the expression levels of CXCL1, CXCL2, CXCL3, and CXCL12 were lower inversely.

3.2. Prognostic Value of mRNA Expression of CXCLs in Breast Cancer Patients. Furthermore, we explored the prognostic values of mRNA expression of CXCLs in breast cancer patients by using Kaplan-Meier survival function. The Kaplan-Meier curve and log rank test analyses revealed that the high CXCL2, CXCL6, CXCL9, CXCL10, CXCL12, and CXCL13 mRNA levels were significantly associated with higher overall survival (OS) (p < 0.05) (Figure 3) of breast cancer patients, while low mRNA levels of CXCL3, CXCL8, and CXCL17 were predicted to have higher OS. Presented in Supplementary Figures S1 and S2, we also found that high CXCL1/2/3/12/16 expressions were significantly correlated with better RFS (relapse-free survival) in ER-positive breast cancer, and high CXCL8/9/13 expressions were significantly correlated with better RFS in ER-negative breast cancer. It is interesting to note that high CXCL10/11 expressions were significantly correlated with poor RFS in ER-positive breast cancer while better RFS in ER-negative breast cancer.

3.3. Association of mRNA Expression of CXCLs with Known Prognostic Factors of Breast Cancer Patients. Correlations between the mRNA levels of CXCLs and known prognostic factors of breast cancer patients were further analyzed in UALCAN. As presented in Figure 4, CXCLs are differently expressed in different age groups. And we can draw conclusions from Figure 5 that CXCL1/2/3/5/6/7 are found significantly expressed in Luminal, CXCL8/11/17 in HER2 positive, and CXCL9/10/12/13/16 in triple negative breast cancer. Also, the transcriptional expressions of CXCLs family members were correlated with patients' individual cancer stages and nodal metastasis status, especially for CXCL2, CXCL3, CXCL4, CXCL7, CXCL9, CXCL10, CXCL11, and CXCL12 (shown in Figures 6 and 7).

3.4. Correlation between CXCLs mRNA Expression and Immune Cells by TIMER. Drawn from the above results, CXCL2, CXCL3, CXCL9, CXCL10, and CXCL12 together with CXCL13 were all significantly expressed and associated with the overall survival in breast cancer patients. The expressions were also connected to tumor stages. We then performed correlation analysis between the six CXCLs family members and tumor immune cells by TIMER in patients with breast cancer (BRCA) and different subtypes (BRCA-Basal, BRCA-Her2, and BRCA-Luminal) to assess the effectiveness of immunotherapy. CXCL9 expression level had obviously positive correlations with infiltrating levels of B cells (r = 0.508, p = 4.21e - 65), CD8+ T cells (r = 0.499, p = 1.84e - 62, CD4+ T cells (r = 0.483, p = 2.47e - 57), neutrophils (r = 0.496, p = 2.98e - 60), and dendritic cells (r = 0.586, p = 1.35e - 88) in BRCA as presented in Figure 8, identified to be a significantly favorable factor in BRCA-Basal, BRCA-Her2, and BRCA-Luminal. CXCL10 showed similar correlations of B cells (r = 0.472, p = 2.51e - 55), CD8+ T cells (r = 0.389, p = 1.45e - 36), CD4+ T cells p = 7.66e - 37),(r = 0.393,neutrophils (r = 0.585,p = 1.70e - 88),and dendritic cells (r = 0.574,p = 1.36e - 84). In addition, CXCL13 expression had



FIGURE 2: The expression of CXCLs in breast cancer (GEPIA). *p < 0.05 (red for tumor, black for normal).

significant correlations with immune cells irrespective of subgroups. It is interesting to note that the mRNA expression of CXCL12 was meaningful associated with macrophages (r=0.399, p = 7.69e – 39) in BRCA, especially in BRCA-Her2 (r=0.445, p = 4.71e – 04) and BRCA-Luminal (r=0.417, p = 2.93e – 24).

3.5. Function Enrichment Analysis of Favorable CXCLs. We first obtained the top 100 similar expression proteincoding genes with CXCL9, CXCL10, CXCL12, and CXCL13, respectively, by similar genes module in GEPIA. We then performed GO and KEGG analysis by DAVID online (https://david.ncifcrf.gov/) and R project. Presented in Figure 9, biological processes such as immune response, positive regulation of immune system process, and cell activation were remarkably regulated by the four CXCLs in breast cancer patients. Cellular components were mainly focused on plasma membrane, extracellular region, and plasma membrane part. Protein dimerization activity, protein homodimerization activity, and carbohydrate binding function correlated closely with the mRNA expression of CXCL9, CXCL10, CXCL12, and CXCL13. Among the top 10 KEGG analysis, cytokine-cytokine



FIGURE 3: The prognostic value of mRNA level of CXCLs family members in breast cancer patients in the overall survival (OS) curve (Kaplan–Meier Plotter).

receptor interaction, cell adhesion molecules (CAMs), T cell receptor signaling pathway, and natural killer cell mediated cytotoxicity held the majority.

4. Discussion

CXCLs were recently discovered to play important roles in various cancer types, including colorectal cancer, pancreatic cancer, lung cancer, and hepatocellular carcinoma [4, 6, 20, 21]. However, the prognostic value and biological function of CXCLs in breast cancer have not been well illustrated. CXCL9/10/12/13 are all angiostatic ELR-members. CXCL9 and CXCL10 both bind to CXCR3, which is associated with T cell function [22]. CXCL9 has been identified as a candidate biomarker in breast cancer. Denkert's study showed that high CXCL9 expression conferred a significantly increased pathologic complete response rate (pCR) in breast cancer patients who received neoadjuvant anthracycline/taxane-based chemotherapy [23]. However, CXCL9 contributes to not only tumor inhibition but also tumor promotion. Ejaeidi et al. showed that the levels of CXCL9 and CXCL10 were markedly high in 40 HR-(hormone receptor-) positive metastatic breast cancer patients when compared to HR-negative patients and healthy

controls in plasma. CXCL10, also known as IFN-g-inducible Protein 10, plays an important role in promoting the homing of immune cells that mediates the subsequent death of cancer cells in breast cancer. Our study showed high CXCL9/10 expression significantly correlated with favorable survival outcome in breast cancer. Combined with the results in ovarian cancer [25], we surmise that CXCL9 and CXCL10 exert tumor-suppressive function by TIL recruitment through the JAK/STAT and NF-*k*B pathways. What is more, studies also revealed that PARP inhibitor-Olaparib induced T cell recruitment is mediated through activation of the cGAS/STING pathway, whose major effector, IRF3, has been reported to regulate expression of CXCL10 [26]. CXCL13, originally named B cell attracting chemokine 1, is also identified to be correlated with CXCL9 (rho = 0.52, p < 0.001) in early breast cancer [27]. Interestingly, the combined expression was also associated with TIL. CXCL13 has been reported to contribute to cancer progression in breast cancer by recruiting B lymphocytes into tumor microenvironment [4, 28]. Zeng et al. also found the correlation of CXCL9/10/13 with immune infiltration in renal cell carcinoma, which was similar to the results in our study [5]. CXCL12 binds to CXCR4 and CXCR7 to promote cancer progression and promotes chemoresistance invasion and migration by activating a number of intracellular signaling



FIGURE 4: Correlation between mRNA expression of distinct CXCLs family members and age of breast invasive carcinoma patients (UALCAN). *p < 0.05, **p < 0.01, ***p < 0.001.

Journal of Oncology



FIGURE 5: Correlation between mRNA expression of distinct CXCLs family members and major subclasses of breast invasive carcinoma patients (UALCAN). *p < 0.05, **p < 0.01, ***p < 0.001.



FIGURE 6: Correlation between mRNA expression of distinct CXCLs family members and individual cancer stages of breast invasive carcinoma patients (UALCAN). *p < 0.05, **p < 0.01, ***p < 0.001.



FIGURE 7: Correlation between mRNA expression of distinct CXCLs family members and nodal metastasis status of breast invasive carcinoma patients (UALCAN). *p < 0.05, **p < 0.01, ***p < 0.001.



1.00 0.0 0.1 0.2 0.3 0.40.0 0.2 0.6 0.2 0.3 0.40.5 0.1 0.2 0.3 0.40.0 0.40.0 0.1

Infiltration level

0.2 0.3 0.0 0.0

0.1

0.5

1.0

(b) FIGURE 8: Continued.

> 0.25 0.50 0.75



FIGURE 8: Continued.



(f)

FIGURE 8: Association of CXCLs (CXCL2, CXCL3, CXCL9, CXCL10, CXCL12, and CXCL3) expression with immune infiltration level in breast cancer patients (TIMER).



FIGURE 9: GO functional enrichment analysis on CXCL9, CXCL10, CXCL12, and CXCL13 as well as their 100 frequently altered neighbor genes in breast cancer patients. (a) Biological process. (b) Cellular components. (c) Molecular functions. (d) KEGG pathway analysis on CXCL9, CXCL10, CXCL10, CXCL12, and CXCL13 as well as their 100 frequently altered neighbor genes in breast cancer patients.

molecules, including Akt, EGFR, mTOR, NF- κ B, and Src [29, 30]. A recent study indicates enhanced infiltration of T lymphocytes and natural killer cells in the tumor microenvironment by blocking CXCL12 [31].

Several limitations should also be mentioned in our study. Firstly, multicenter, large-scale prospective studies with long-term follow-up are still urgently needed, for conclusions drawn from different databases may be inconsistent and not wholly reliable Secondly, it is noted that chemokines play a dual role in tumor progression. However, variant mechanisms have not been well clarified in different cancers. For example, it is worth a deeper look at the complicated relation among CXCLs and the NF- κ B signaling pathway. In conclusion, our study found additional evidence that CXCL9/10/12/13 could be used as potential prognosis biomarkers in breast cancer.

5. Conclusions

In this study, we analyzed the expression of 15 different CXCLs family members in breast cancer and adjacent tissues or normal tissues and assessed the relationship

between CXCLs expression and overall survival, as well as tumor staging. Our findings indicated that high CXCL2/9/ 10/12/13 expression and low CXCL3 expression were associated with favorable OS in breast cancer patients. All of them are significantly linked with patients' known prognostic factors, such as patient's age, major subclasses, individual cancer stages, and nodal metastasis status. Tumorinfiltrating lymphocytes (TILs) are recently found predicting neoadjuvant chemotherapy response in different breast cancer subtypes [32]. It is considered that immune cells have a large impact on the tumor immune network and tumor development. We then utilized TIMER to perform the relationships between CXCL2/3/9/10/12/13 expression and abundance of immune infiltrates, as well as conducting the GO and KEGG analysis. The results suggested that CXCL9/10/13 expressions were significantly associated with infiltrating levels of B cells, CD8+ T cells, CD4+ T cells, neutrophils, and DCs in breast cancer, while CXCL12 was conversely correlated with macrophages. In recent years, with the breakthroughs in the research of immunological checkpoint, patients have received unprecedented long-lasting anti-tumor response, where immune microenvironment, tumor-infiltrating cells, and immune biomarkers play important roles. Our study indicated that the mRNA expressions of CXCL9/10/12/13 are correlated with immune cells. Therefore, we speculated that they could be used to predict the effectiveness of immune blockade therapy in breast cancer patients.

Data Availability

The datasets generated and analyzed during the current study are available in the Oncomine database (http://www.oncomine.org), GEPIA (http://gepia.cancer-pku.cn/), UALCAN (http://ualcan.path.uab.edu), Kaplan-Meier Plotter (http://www.kmplot.com), TIMER (https://cistrome.shinyapps.io/timer/), and DAVID (https://david.ncifcrf.gov/home.jsp).

Conflicts of Interest

The authors declare that there are no conflicts of interest regarding the publication of this article.

Authors' Contributions

Yan Li, Mingqiang Liang, and Yuxiang Lin contributed equally to this work.

Supplementary Materials

Figure S1: the prognostic value of mRNA level of CXCLs family members in ER-positive breast cancer patients in the relapse-free survival (RFS) curve (Kaplan–Meier Plotter). Figure S2: the prognostic value of mRNA level of CXCLs family members in ER-negative breast cancer patients in the relapse-free survival (RFS) curve (Kaplan–Meier Plotter). (*Supplementary Materials*)

References

- C. Bizzarri, A. R. Beccari, R. Bertini, M. R. Cavicchia, S. Giorgini, and M. Allegretti, "ELR⁺ CXC chemokines and their receptors (CXC chemokine receptor 1 and CXC chemokine receptor 2) as new therapeutic targets," *Pharmacology* & *Therapeutics*, vol. 112, no. 1, pp. 139–149, 2006.
- [2] K. Koizumi, S. Hojo, T. Akashi, K. Yasumoto, and I. Saiki, "Chemokine receptors in cancer metastasis and cancer cellderived chemokines in host immune response," *Cancer Science*, vol. 98, no. 11, pp. 1652–1658, 2007.
- [3] J. Vandercappellen, J. Van Damme, and S. Struyf, "The role of CXC chemokines and their receptors in cancer," *Cancer Letters*, vol. 267, no. 2, pp. 226–244, 2008.
- [4] N.-H. Lee, M. Nikfarjam, and H. He, "Functions of the CXC ligand family in the pancreatic tumor microenvironment," *Pancreatology*, vol. 18, no. 7, pp. 705–716, 2018.
- [5] Q. Zeng, S. Sun, Y. Li, X. Li, Z. Li, and H. Liang, "Identification of therapeutic targets and prognostic biomarkers among CXC chemokines in the renal cell carcinoma microenvironment," *Frontiers in Oncology*, vol. 9, p. 1555, 2019.
- [6] S. C.-d. l. Heras and E. Martínez-Balibrea, "CXC family of chemokines as prognostic or predictive biomarkers and

possible drug targets in colorectal cancer," *World Journal of Gastroenterology*, vol. 24, no. 42, pp. 4738–4749, 2018.

- [7] F. Bray, J. Ferlay, I. Soerjomataram, R. L. Siegel, L. A. Torre, and A. Jemal, "Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries," *CA: A Cancer Journal for Clinicians*, vol. 68, no. 6, pp. 394–424, 2018.
- [8] L. G. Eng, S. Dawood, V. Sopik et al., "Ten-year survival in women with primary stage IV breast cancer," *Breast Cancer Research and Treatment*, vol. 160, no. 1, pp. 145–152, 2016.
- [9] C. M. Perou, T. Sørlie, M. B. Eisen et al., "Molecular portraits of human breast tumours," *Nature*, vol. 406, no. 6797, pp. 747–752, 2000.
- [10] S. Liu, C. Ginestier, S. J. Ou et al., "Breast cancer stem cells are regulated by mesenchymal stem cells through cytokine networks," *Cancer Research*, vol. 71, no. 2, pp. 614–624, 2011.
- [11] E. E. Parkes, S. M. Walker, L. E. Taggart et al., "Activation of STING-dependent innate immune signaling by S-Phase-Specific DNA damage in breast cancer," *Journal of the National Cancer Institute*, vol. 109, no. 1, Article ID djw199, 2017.
- [12] S. Acharyya, T. Oskarsson, S. Vanharanta et al., "A CXCL1 paracrine network links cancer chemoresistance and metastasis," *Cell*, vol. 150, no. 1, pp. 165–178, 2012.
- [13] D. R. Rhodes, J. Yu, K. Shanker et al., "ONCOMINE: a cancer microarray database and integrated data-mining platform," *Neoplasia*, vol. 6, no. 1, pp. 1–6, 2004.
- [14] Z. Tang, C. Li, B. Kang, G. Gao, C. Li, and Z. Zhang, "GEPIA: a web server for cancer and normal gene expression profiling and interactive analyses," *Nucleic Acids Research*, vol. 45, no. W1, pp. W98–W102, 2017.
- [15] D. S. Chandrashekar, B. Bashel, S. A. H. Balasubramanya et al., "UALCAN: a portal for facilitating tumor subgroup gene expression and survival analyses," *Neoplasia*, vol. 19, no. 8, pp. 649–658, 2017.
- [16] B. Györffy, A. Lanczky, A. C. Eklund et al., "An online survival analysis tool to rapidly assess the effect of 22,277 genes on breast cancer prognosis using microarray data of 1,809 patients," *Breast Cancer Research and Treatment*, vol. 123, no. 3, pp. 725–731, 2010.
- [17] T. Li, J. Fan, B. Wang et al., "TIMER: a web server for comprehensive analysis of tumor-infiltrating immune cells," *Cancer Research*, vol. 77, no. 21, pp. e108–e110, 2017.
- [18] D. W. Huang, B. T. Sherman, and R. A. Lempicki, "Systematic and integrative analysis of large gene lists using DAVID bioinformatics resources," *Nature Protocols*, vol. 4, no. 1, pp. 44–57, 2009.
- [19] G. Dennis Jr., B. T. Sherman, D. A. Hosack et al., "DAVID: database for annotation, visualization, and integrated discovery," *Genome Biology*, vol. 4, no. 9, 2003.
- [20] H. Imai, N. Sunaga, Y. Shimizu et al., "Clinicopathological and therapeutic significance of CXCL12 expression in lung cancer," *International Journal of Immunopathology and Pharmacology*, vol. 23, no. 1, pp. 153–164, 2010.
- [21] Q. Ding, Y. Xia, S. Ding et al., "Potential role of CXCL9 induced by endothelial cells/CD133+ liver cancer cells coculture system in tumor transendothelial migration," *Genes Cancer*, vol. 7, no. 7-8, pp. 254–259, 2016.
- [22] J. R. Groom and A. D. Luster, "CXCR3 in T cell function," *Experimental Cell Research*, vol. 317, no. 5, pp. 620–631, 2011.
- [23] Q. Ding, P. Lu, Y. Xia et al., "CXCL9: evidence and contradictions for its role in tumor progression," *Cancer Medicine*, vol. 5, no. 11, pp. 3246–3259, 2016.
- [24] A. A. Ejaeidi, B. S. Craft, and L. V. Puneky, "Hormone receptor-independent CXCL10 production is associated with

the regulation of cellular factors linked to breast cancer progression and metastasis," *Experimental and Molecular Pathology*, vol. 99, no. 1, pp. 1636–172, 2015.

- [25] H. Bronger, J. Singer, C. Windmüller et al., "CXCL9 and CXCL10 predict survival and are regulated by cyclooxygenase inhibition in advanced serous ovarian cancer," *British Journal* of Cancer, vol. 115, no. 5, pp. 553–563, 2016.
- [26] C. Pantelidou, O. Sonzogni, M. De Oliveria Taveira et al., "PARP inhibitor efficacy depends on CD8+ T-cell recruitment via intratumoral STING pathway activation in BRCA-deficient models of triple-negative breast cancer," *Cancer Discovery*, vol. 9, no. 6, pp. 722–737, 2019.
- [27] E. Razis, K. T. Kalogeras, I. Kotsantis et al., "The role of CXCL13 and CXCL9 in early breast cancer," *Clinical Breast Cancer*, vol. 20, no. 1, pp. e36–e53, 2020.
- [28] M. Ammirante, S. Shalapour, Y. Kang, C. A. M. Jamieson, and M. Karin, "Tissue injury and hypoxia promote malignant progression of prostate cancer by inducing CXCL13 expression in tumor myofibroblasts," *Proceedings of the National Academy of Sciences*, vol. 111, no. 41, pp. 14776–14781, 2014.
- [29] M. K. Conley-LaComb, L. Semaan, R. Singareddy et al., "Pharmacological targeting of CXCL12/CXCR4 signaling in prostate cancer bone metastasis," *Molecular Cancer*, vol. 15, no. 1, p. 68, 2016.
- [30] X. Zhu, Q. Bai, Y. Lu et al., "Expression and function of CXCL12/CXCR4/CXCR7 in thyroid cancer," *International Journal of Oncology*, vol. 48, no. 6, pp. 2321–2329, 2016.
- [31] D. Zboralski, K. Hoehlig, D. Eulberg, A. Frömming, and A. Vater, "Increasing tumor-infiltrating T cells through inhibition of CXCL12 with NOX-A12 synergizes with PD-1 blockade," *Cancer Immunology Research*, vol. 5, no. 11, pp. 950–956, 2017.
- [32] M. Kochi, T. Iwamoto, N. Niikura et al., "Tumourinfiltrating lymphocytes (TILs)-related genomic signature predicts chemotherapy response in breast cancer," *Breast Cancer Research and Treatment*, vol. 167, no. 1, pp. 39–47, 2018.