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ORIGINAL ARTICLE

The pro-domains of neurotrophins, including BDNF, are linked to Alzheimer's disease through a toxic synergy with Aβ

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Abstract

Brain-derived neurotrophic factor (BDNF) has a crucial role in learning and memory by promoting neuronal survival and modulating synaptic connectivity. BDNF levels are lower in the brains of individuals with Alzheimer's disease (AD), suggesting a pathogenic involvement. The *Drosophila* orthologue of BDNF is the highly conserved Neurotrophin 1 (DNT1). BDNF and DNT1 have the same overall protein structure and can be cleaved, resulting in the conversion of a full-length polypeptide into separate pro- and mature-domains. While the BDNF mature-domain is neuroprotective, the role of the pro-domain is less clear. In flies and mammalian cells, we have identified a synergistic toxic interaction between the amyloid- β peptide (A β_{1-42}) and the pro-domains of both DNT1 and BDNF. Specifically, we show that DNT1 pro-domain acquires a neurotoxic activity in the presence of A β_{1-42} . In contrast, DNT1 mature-domain is protective against A β_{1-42} toxicity. Likewise, in SH-SY5Y cell culture, BDNF pro-domain is toxic only in the presence of A β_{1-42} . Western blots indicate that this synergistic interaction likely results from the A β_{1-42} -induced upregulation of the BDNF pro-domain receptor p75^{NTR}. The clinical relevance of these findings is underlined by a greater than thirty fold increase in the ratio of BDNF pro- to mature-domains in the brains of individuals with AD. This unbalanced BDNF pro:mature-domain ratio in patients represents a possible biomarker of AD and may offer a target for therapeutic intervention.

Introduction

Alzheimer disease (AD) is a devastating neurodegenerative disorder that manifests as a progressive decline in cognitive function (1). Pathologically, AD is characterized by the accumulation of plaques containing amyloid- β (A β), tau-laden neurofibrillary tangles and progressive synaptic and neuronal loss. Emerging evidence suggests that brain-derived neurotrophic factor (BDNF) may be involved in the pathogenesis of AD. BDNF is the most functionally diverse member of the neurotrophin (NT) family. As well as promoting the differentiation and migration of peripheral and central neurons and glia during development (2,3), BDNF also supports neuronal survival and synaptic plasticity in adulthood and also having effects on memory and mood (4,5).

All NTs are synthesized as precursors (pro-NTs) that then form homodimers (6). Pro-NTs may be secreted from cells (7) or

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cleaved intracellularly by furin or proconvertases to yield the mature C-terminal NT. The processing of pro-BDNF may also be completed extracellularly by plasmin or metalloproteinases, including MMP3, MMP7 and MMP9 (7,8). BDNF exerts many of its neuroprotective effects by binding to the TrkB receptor, a receptor tyrosine kinase (9). However, the consequences of its binding to its alternative receptor p75^{NTR} are thought to include promotion of myelination (3) and neuronal migration (10) but also neuronal process retraction (11) and neuronal apoptosis (12). While the application of mature BDNF reduces sensory neuronal loss in dorsal root ganglia after peripheral nerve injury, in contrast the precursor form of BDNF increases the death of axotomized neurons (13). This toxic effect requires the expression levels of the TrkB and p75^{NTR} high-affinity receptors. Likewise, the proand mature-domains of nerve growth factor (NGF) have different effects on neurones, specifically the precursor form of NGF preferentially activates p75^{NTR} resulting in apoptosis, while the NGF mature-domain signals through the TrkA receptor resulting in neurotrophic activity (7).

Reduced levels of hippocampal and cortical pro-BDNF or BDNF are a consistent feature of AD (14,15); however, the mechanistic basis for this correlation is unclear. There is some evidence that that $A\beta$ may reduce TrkB-mediated neurotrophic signaling (16); conversely, a reduction in BDNF may favor the amyloidogenic processing of the amyloid precursor protein, favouring $A\beta$ generation (17). However, little is known regarding the particular role of BDNF pro-domain despite it harbouring a common sequence polymorphism (Val66Met) linked to accelerated cognitive decline in AD (18,19). DNT1 is the Drosophila orthologue of BDNF and as a member of the NT superfamily it has structural and sequence orthologues throughout the phylogenetic spectrum (20).

In the present study, we investigate a potential role for the DNT1/BDNF pro-domain in mediating $A\beta$ toxicity and, in clinical samples, show that AD is accompanied by profound changes in the pro:mature-domain ratio of BDNF.

Results

Drosophila DNT1 mRNA levels are high during development but suppressed in adulthood

Low levels of BDNF have been correlated with increased agerelated risk of AD (16,21) and so we used quantitative RT-PCR to quantify the levels of DNT1 mRNA in flies during development and for the first 10 days of adult life. We found that levels of endogenous DNT1 mRNA were high during development but fell rapidly in adult life whether or not A β was also co-expressed (Fig. 1). Consequently knock-down of DNT1, by RNAi or the hemizygous introduction of a null allele (Supplementary Material, Fig. S1A), had no effect on longevity and locomotor phenotypes in the adult (Supplementary Material, Fig. S1B–E).

The subdomains of DNT1 have differential effects on neurotoxic phenotypes in the fly

Exogenous application of BDNF improves cognitive and behavioural deficits and ameliorates A β pathology in murine and primate models of AD (22,23). To test whether overexpression of DNT1 rescues neurotoxic phenotypes induced by Arc A β_{1-42} peptide in flies, we expressed three forms of the protein (20): (1) full-length DNT1, (2) pro-domain of DNT1, (3) mature-domain of DNT1 in all neurons and measured the interaction with Arc A β_{1-42} expression. The overexpression of both mature- and



Figure 1. Drosophila DNT1 mRNA levels are high during development but suppressed in adulthood. (A) DNT1 and RP49 mRNA were measured at Days 0 (D0) and 10 (D10) by quantitative real-time PCR in control flies and those expressing the $A\beta_{1-40}$ and $A\beta_{1-42}$ peptides. DNT1 mRNA levels were highest during larval development (black bar) and fall rapidly before eclosion (Day 0, grey bars). An essentially identical pattern of age-related decline in DNT1 was seen in 'Control' flies (elav-gal4; +) and in 'A β_{1-40} ' (elav-gal4; UAS-Abeta40) and 'A β_{1-42} ' (elav-gal4; UAS-Abeta42) flies. Data are presented as the ratio of DNT1 to RP49 mRNA levels (±SD). Statistical comparisons were made by one-way analysis variance test. **P < 0.01 and ***P < 0.001.

full-length DNT1 prolonged the lifespan of control flies at 29°C, increasing their median survival by 30%. Notably, overexpression of DNT1 pro-domain had no effect on the longevity of control flies (Fig. 2A). As expected Arc $A\beta_{1-42}$ -expression resulted in a 50% reduction in longevity and this effect was significantly suppressed by co-expression of DNT1 mature-domain (Fig. 2B). Remarkably, the co-expression of DNT1 pro-domain with Arc $A\beta_{1-42}$ enhanced the reduction in longevity, significantly reducing the median survival by a further 30%. Interestingly when full-length DNT1, containing both pro- and mature-domains, was co-expressed with Arc $A\beta_{1-42}$, the effects of the component domains cancelled each other out, resulting in no overall effect on the longevity (Fig. 2B). These results were confirmed at 25°C when transgene expression levels were expected to be lower (Supplementary Material, Fig. S2).

To further investigate the effect of DNT1 on A β -linked toxicity, we assessed the average climbing velocity of flies overexpressing three forms of DNT1 in the absence, or presence, of the Arc A β_{1-42} . Wild-type control flies showed the expected age-related reduction in climbing velocity that is not modified by the overexpression of any DNT1 construct (Fig. 2C). Arc A β_{1-42} flies exhibited a much earlier and more rapid decline in locomotor activity but this was not affected by the overexpression of DNT1 pro-domain (Fig. 2D). However, full-length DNT1 and the mature-domain of DNT1 did rescue locomotor deficits as witnessed by higher velocities from Days 1–5 when compared with flies expressing Arc A β_{1-42} flies beyond Day 9 due to their short life span.

Co-expression of various domains of DNT1 does not change the abundance or conformation of $A\beta$

To understand whether the modulation of A β toxicity results from changes in the handling of the peptide, we western blotted SDS–PAGE gels of brain homogenates using the 6E10 monoclonal antibody (Fig. 3A, representative of n = 3). While multiple



Figure 2. The subdomains of DNT1 have differential effects on neurotoxic phenotypes in the fly. (A) Expression of full length (elav-gal4; UAS-DNT1 full length; +) and mature-domain (elav-gal4; UAS-DNT1 mature-domain; +) of DNT1 prolonged the lifespan of control flies (elav-gal4; +; +), whereas DNT1 pro-domain (elav-gal4; UAS-DNT1 pro-domain; +) had no effect. The flies were cultured at 29°C. Each graph presents the mean of 10 estimates of median survival ±SEM. Statistical comparisons were made by one-way analysis variance test. ***P < 0.001. (B) The overexpression of DNT1 mature-domain (elav-gal4; UAS-DNT1 mature-domain; UAS-ArcAbeta42) significantly increased median survival of Arc A β_{1-42} flies (elav-gal4; +; UAS-ArcAbeta42) while DNT1 pro-domain (elav-gal4; UAS-DNT1 pro-domain; UAS-ArcAbeta42) further reduced median survival by 30%. Full-length DNT1 (elav-gal4; UAS-DNT1 full length; UAS-ArcAbeta42) had no overall effect. The flies were cultured at 29°C. Each graph presents the mean of 10 estimates of median survival by 30%. Full-length DNT1 (elav-gal4; UAS-DNT1 full length; UAS-ArcAbeta42) had no overall effect. The flies were cultured at 29°C. Each graph presents the mean of 10 estimates of median survival ±SEM. Statistical comparisons were made by one-way analysis variance test. ***P < 0.001. (C) The mean climbing velocity of control flies was not affected by the overexpression of any DNT1 constructs. The genotypes were the same as (B). Mean velocity ±SEM. (D) Some rescue of the locomotor deficits associated with Arc A β_{1-42} expression (red squares) was seen upon co-expression of full-length DNT1 (purple diamonds, ***P < 0.001 and ****P < 0.001 on Day 1 only) and DNT1 mature-domain (blue inverted triangles, **P < 0.01 and ****P < 0.001 on days 1, 3 and 5), but not DNT1 pro-domain (black triangles). The genotypes were the same as (B). Mean velocity ±SEM. Statistical comparisons were made with controls at all times points. Mean velocity ±SEM. Statistical comparisons were the same as (B). Mean veloci



Figure 3. Co-expression of various domains of DNT1 does not change the abundance or conformation of $A\beta$. (**A**) Western blots of SDS–PAGE gels of *Drosophila* brain extracts containing the SDS-soluble $A\beta$ fraction. Immunodetection using the monoclonal antibody 6E10 revealed a number of bands in all Arc $A\beta_{1-42}$ lines. There were no consistent changes in the individual $A\beta$ bands between genotypes across three independent experiments. β -Actin expression was used as a loading control. (**B**) The multiple $A\beta$ -specific band intensities were normalized to β -actin and the data were plotted as a bar chart (band intensities ±SEM, n = 3). There was no consistent change in the total intensity of $A\beta$ bands upon co-expression of various domains of DNT1. Genotypes: '–': elav-gal4; +; UAS-Abeta42, 'pro': elav-gal4; UAS-DNT1 pro-domain; UAS-Abeta42, 'mature': elav-gal4; UAS-DNT1 full length; UAS-DNT1 full length; UAS-Abeta42.

A β -specific bands representing monomeric, dimeric and oligomeric forms were visible in all Arc A β_{1-42} lines, there was no consistent change in their intensities, relative to each other or to the β -actin control, upon expression of the various domains of DNT1 (Fig. 3B). Thus, it appeared unlikely that DNT1 constructs were regulating the clearance or aggregation of A β in the fly brain and so alternative mechanisms were investigated in mammalian cell culture.

The pro-domain of BDNF and $A\beta_{1-42}$ exhibit synergistic toxicity in SH-SY5Y human neuroblastoma cell cultures

We now sought to determine whether the synergy between the pro-domain of DNT1 and $A\beta$, seen in the longevity experiments in Drosophila, is also observed in mammalian cell culture. We

used oligomeric preparations of $A\beta_{1-42}$ (Supplementary Material, Fig. S3, lanes 2–3) to induce cell death in cultures of SH-SY5Y human neuroblastoma cells and showed that there was a nonlinear dose-dependent reduction in cell viability as measured by the MTT assay. Significant cell death (20–30%) was seen 48– 60 h after exposure to $A\beta_{1-42}$ oligomer preparation (Fig. 4A). To test whether the BDNF pro-domain could enhance $A\beta_{1-42}$ -induced cytotoxicity, we treated cells with both wild type (Val66) and variant (Met66) recombinant pro-domain of BDNF with, or without, 25 nM oligomeric $A\beta_{1-42}$. The MTT results indicated that the variant Met66 BDNF pro-domain alone did not affect cell viability (Fig. 4B); however, the variant BDNF pro-domain (200 ng/ml) combined with $A\beta_{1-42}$ oligomers (25 nM) resulted in greater cell death when compared with $A\beta_{1-42}$ alone (Fig. 4C, P < 0.01). When we consider all 26 paired experiments (for n = 9



Figure 4. The pro-domain of BDNF and $A\beta_{1-42}$ exhibits synergistic toxicity in SH-SY5Y human neuroblastoma cell cultures. (A) Addition of 25, 100 and 1000 nM oligomeric $A\beta_{1-42}$ to the culture medium of SH-SY5Y human neuroblastoma cells for 48–60 h resulted in significant toxicity when compared with vehicle. Each bar represents mean ± SEM (*n* = 3). Statistical comparisons were made by one-way analysis variance test. * *P* < 0.05. (B) Cells were treated with 200 ng/ml recombinant pro-domain of BDNF for 48–60 h. Met66 BDNF pro-domain alone did not have an effect on cell viability in control cells. (C) Cells were treated with Met66 pro-domain of BDNF (200 ng/ml) in combination with 25 nM $A\beta_{1-42}$ for 48–60 h. In the presence of $A\beta_{1-42}$ cell death was increased upon addition of Met66 BDNF pro-domain (*n* = 9) performed. Each bar represents mean ± SEM (*n* = 9). Statistical comparisons were made by paired t test. **P < 0.01. (D) Twenty-six paired experiments indicated that cell death was higher when a particular preparation of $A\beta_{1-42}$ (25 nM, *n* = 9 preparations of $A\beta_{1-42}$) was supplemented with Met66 BDNF pro-domain (*y*-axis) than for that preparation of $A\beta_{1-42}$ alone (*x*-axis, dashed line indicates the null hypothesis of no effect of the Met66 BDNF pro-domain). (E) Similar results were seen when non-differentiated and (F) differentiated cells were treated with the Met66 BDNF pro-domain (200 ng/ml) in combination with 25 nM $A\beta_{1-42}$ oligomers for 48–60 h before SDS–PAGE and western blotting with antibodies against p75^{NTR} and sortilin. β -Actin expression was used as a loading control. Western blots indicated that levels of p75^{NTR} but not sortilin were higher following $A\beta_{1-42}$ oligomer treatment in cultures of human neuroblastoma cells. Representative blots are shown (*n* = 3). (H) The p75^{NTR} and (I) sortilin intensities from the western blots were normalized against the level of β -actin. Each bar represents mean ± SEM (*n* = 3). Statistical comparisons were made by one-

preparations of $A\beta$), we saw more cell death when a particular preparation of $A\beta$ was supplemented with the variant Met66 BDNF pro-domain when compared with treatment with the same preparation of $A\beta_{1-42}$ alone (Fig. 4D, paired MTT assays of Aβ toxicity ±Met66 BDNF pro-domain). No such interaction was seen between $A\beta$ and the wild-type Val66 pro-domain of BDNF (Supplementary Material, Fig. S4A and B, nine paired experiments, n = 3 preparations of A β). As expected the mature domain of BDNF alone did protect the cells from A_β toxicity (Supplementary Material, Fig. S4C and D). These data indicate that, similar to the findings in the fly model, the NT pro-domain acts synergistically with $A\beta_{1-42}$ to exert a toxic effect. Using higher concentrations (1 μ M) of A β_{1-42} , similar results were also seen for non-differentiated (Fig. 4E, nine paired experiments, n = 3 preparations of $A\beta$), differentiated (Fig. 4F, nine paired experiments, n = 3 preparations of A β) and SH-SY5Y cells. A β has previously been reported to increase sortilin and $p75^{NTR}$ levels (24,25) and we sought to confirm this observation in our cultures of SH-SY5Y cells treated with 25 nm oligomeric $A\beta_{1-42}$. Western blots indicated that levels of p75^{NTR}, but not sortilin, were higher following $A\beta_{1-42}$ treatment (Fig. 4G–I). The pro-domain of BDNF did not modulate this $A\beta_{1\text{-}42}\text{-}mediated$ upregulation of $p75^{\text{NTR}}$

An elevated ratio of pro- to mature-domain of BDNF correlates with disease status in elderly individuals

(Fig. 4G and H).

We next tested whether the levels of BDNF pro- and maturedomains were different in extracts of age-matched post-mortem hippocampal samples from 10 patients with advanced AD (Braak Stages 5–6) and 10 healthy individuals (Table 1). Western blots of SDS-PAGE gels (Fig. 5A) showed that, as expected, BDNF mature-domain levels were, on average, 6-fold lower in

 Table 1. Demographics of Alzheimer's disease cases and control individuals (as provided by the source)

Diagnosis	Braak Stage	Age (years)	Gender
1. Controls		89	F
2. Controls		85	М
3. Controls		85	F
4. Controls		89	F
5. Controls		82	F
6. Controls		88	М
7. Controls		67	М
8. Controls		87	F
9. Controls		91	М
10. Controls		63	F
Mean		82.6	
SEM		3.056	
11. AD	5	68	М
12. AD	5	90	М
13. AD	5	84	F
14. AD	5/6	65	F
15. AD	5	84	F
16. AD	5	95	F
17. AD	5	87	М
18. AD	5	92	F
19. AD	5	82	М
20. AD	5	83	F
Mean		83	
SEM		3.059	

Hippocampal tissue was taken from age and sex matched AD cases and controls.

cases when compared with controls (Supplementary Material, Table S1). In contrast, the levels of BDNF pro-domain were, on average, 16-fold higher in cases. Combining these findings as a pro:mature-domain ratio revealed a 30-fold relative difference for cases and controls (Fig. 5B; Supplementary Material, Table S1). Furthermore, we observed a robust correlation between the BDNF pro:mature domain ratio and the accumulation of urea-solubilizable A β in the brain extracts (Fig. 5C). Of particular note is the control individual (age 63, Fig. 5B and C marked with #) who, despite being asymptomatic and free of obvious AD pathology, is nevertheless the only control with significant A β accumulation, elevated pro-domain levels and a pro:mature domain ratio that overlaps with AD patients.

Discussion

NTs are a major class of molecules promoting neuronal survival in vertebrates. They are synthesized as larger precursor forms that are proteolytically processed to yield a mature, biologically active ligand (6,7). Among the NTs, BDNF has emerged as a major regulator of synaptic plasticity, neuronal survival and differentiation, and also as a potential molecular target for the treatment of neurological disease (26). Several studies indicate that the cortex and hippocampus, areas of the brain associated with learning and memory, not only exhibit extensive amyloid pathology but also show decreased levels of BDNF in AD (14,15,27). Interestingly, precursor and mature forms of BDNF are significantly decreased in preclinical and early stages of AD, and this reduction correlates with clinical neuropsychological scores (15). Low levels of BDNF may favor AD pathogenesis by failing to adequately support neurones and allowing them to succumb to other toxic insults. Furthermore, several studies have linked polymorphisms, specifically Val66Met and Cys270Thr, in the pro-domain of BDNF to an increased risk for AD (28,29).

BDNF exerts many of its neuroprotective effects by binding to the TrkB receptor, a member of the tumour necrosis factor receptor family (9). Indeed, some reports indicate that $A\beta$ may have a negative effect on neuronal survival by down-regulating the TrkB receptor (16). For example, levels of TrkB are reduced in the temporal and frontal cortex of AD brain (30). In addition, BDNF-induced TrkB autophosphorylation and the activation of the downstream enzymes AKT and ERK are all suppressed in the hippocampus of APP/PS1 mice (31).

The consequences of BDNF binding to its alternative receptor p75^{NTR} and sortilin are less well understood, although they are thought to include promotion of myelination (3) and neuronal migration (10) but also neuronal process retraction (11) and neuronal apoptosis (12). It is proposed that the balance between cell death and survival is determined by the relative activity of the precursor versus mature forms of BDNF; indeed, in dorsal root ganglion lesion models in neonatal rats, the signaling appears to be affected by the relative levels of the relevant receptors, namely TrkB, p75^{NTR} and sortilin. Notably, BDNF promotes the death of cultured neurons in vitro when p75^{NTR} is upregulated and TrkB downregulated (13). Precursor BDNF can also cause cell death in both in vitro and in vivo model systems (32) and the prevailing view is that the apoptotic signal is generated by the pro-domain. Similar domain-specific activities have also been observed for NGF; specifically, the precursor form of this related NT preferentially activates p75^{NTR} resulting in apoptosis, while mature NGF preferentially activates TrkA receptor with neurotrophic effects (7).

Mammalian NTs are similar in many ways to their insect orthologues. Sequence analysis has identified Drosophila



Figure 5. An elevated ratio of pro- to mature-domain BDNF correlates with disease status in elderly individuals. (A) Frozen post-mortem hippocampal tissues from 10 patients with advanced AD were homogenized and proteins were separated by SDS–PAGE and then subjected to western blot analysis with antibodies against BDNF pro-domain, mature-domain and $A\beta$. β -Actin expression was used as a loading control. Western blots show that the levels of BDNF pro-domain were higher, and mature-domain levels lower, in the hippocampal formation of AD cases than in control brains. (B) The pro- and mature-domains of BDNF were quantified by densitometry and normalized against the corresponding β -actin band. As expected, mature-domain BDNF levels were, on average, 6-fold lower in cases when compared with controls. In contrast, the levels of pro-domain BDNF were 16-fold higher in cases. Combining these findings as a pro:mature domain ratio yielded a relative difference for cases and controls of 30-fold. (C) There was a robust correlation between the BDNF pro:mature domain ratio and the accumulation of ureas solubilizable $A\beta$ in the brain extracts. Of particular note is the control individual marked with #.

neurotrophin (DNT1), also called Spatzle 2 (Spz 2), as the closest fly orthologue of human BDNF. Like the NTs, the Spz polypeptides are synthesized with a signal peptide, followed by a pro-domain and then the cysteine knot-containing mature domain (33,34). The characteristic NT cysteine knot, formed by antiparallel β -sheets held together by three disulfide bonds, is conserved in the crystal structures of both Spz and NGF (35,36). There is also functional conservation between DNT1, Spz and the mammalian NTs in the nervous system (20,37,38). Indeed, during Drosophila embryogenesis DNT1 is expressed in neurones and muscle cells where it promotes neuronal survival and suppresses apoptosis (20). We have shown that the high levels of endogenous DNT1 mRNA that are present during larval development are rapidly suppressed at the beginning of adult life in both control flies and similarly in those expressing various isoforms of the AB peptide (Fig. 1).

Because of the functional similarities between mammalian and Drosophila NTs, we undertook a study of the interaction between A β and DNT1 in the fruit fly. We found that transgenic expression of the DNT1 mature-domain protected flies against A β toxicity (Fig. 2) and that this benefit was not due to spurious reductions in A β levels (Fig. 3). Similarly, in human SH-SY5Y culture, the mature-domain of BDNF protected cells from A β toxicity (Supplementary Material, Fig. S3B). In both the fly and the mammalian models, the pro-domains of both DNT1 and BDNF, while being harmless alone, were nevertheless toxic when added in combination with A β (Figs 2 and 4). In SH-SY5Y cells, this synergistic toxic interaction between A β and the BDNF pro-domain was only apparent for the Met66 variant and was absent for the wild-type Val66 isoform (Fig. 4; Supplementary Material, Fig. S4A). This finding provides mechanistic underpinning for the clinical observation that the Met66 polymorphism is linked to poor prognosis, particularly in individuals with high A β burden on PiB photon emission tomography brain scans (18,39).

The expression of the receptor $p75^{NTR}$ is upregulated in both SH-SY5Y cells following A β treatment and also in transgenic mice expressing the human APPswe transgene (Fig. 4G and H) (40,41). A β treatment also enhances sortilin expression via $p75^{NTR}$, which is thought to activate the downstream effectors JNK and Rho (24,25), resulting in apoptotic cell death. Furthermore, the accumulation of A β in humans is also accompanied by an increased hippocampal membrane-associated $p75^{NTR}$

(42). However, it seems unlikely that A β binds directly to $p75^{NTR}$ (43), rather we suggest that A β amplifies the pro-apoptotic signaling of pro-domain BDNF by upregulating $p75^{NTR}$ in the human SH-SY5Y cells (Fig. 4G and H).

Our subsequent work with post-mortem hippocampal tissue from age-matched healthy elderly and patients with AD has underlined the clinical significance of these experimental findings. As expected, the mature-domain of BDNF is reduced in AD when compared with control brain tissue (15). In our series of 10 cases and controls, we found that control subjects fell into two groups-those with levels of BDNF mature-domain that are up to 20 times higher than AD cases (Fig. 5); however, an equal number of controls have mature-domain levels that are equivalent to AD patients. The separation between cases and controls is better when we measure BDNF pro-domain levels: cases have elevated levels of pro-domain, spread over almost 2 orders of magnitude while all but one of the controls (outlier labelled with #) are closely grouped. The average pro-domain level is ~16-fold higher in AD cases when compared with controls. These reciprocal changes in the levels of pro- and mature-BDNF are surprising considering that the peptides are generated stoichiometrically. Receptormediated clearance is an unlikely explanation because the levels of TrkB are low and p75 high in AD cases when compared with controls (30,31,42). Conceivably, in AD the BDNF pro-domain is being stabilized by binding to another protein (44); this intriguing possibility requires further investigation.

When these relative changes in pro- and mature-domains are combined as a ratio, we see that cases and controls have an average 30-fold difference and only one of the controls overlaps with the AD range. This particular control individual is interesting because, despite being symptom-free and relatively young (aged 63), she was the only control to have accumulated significant A β . In fact, she had the highest level of A β amongst all cases and controls. That this control individual's pro:mature domain ratio was in the AD range raises the tantalizing possibility that we are able to predict disease changes before the onset of symptoms.

In conclusion, we have shown for the first time that the proand mature-domains of NTs have opposing effects on A β neurotoxicity *in vitro* and *in vivo*. The mature-domains of NTs protected against A β_{1-42} toxicity, whereas the pro-domains acquired a toxic role in the presence of A β_{1-42} . In our study of post-mortem clinical brain samples, the ratio of pro- to mature-domain of BDNF was significantly higher in patients with AD when compared with controls. Taken together, the finding that patients with AD have elevated levels of BDNF pro-domain underlines the importance of its synergistic toxic interaction with A β . Conceivably, individuals with an unfavourable pro:mature domain ratio could be targeted with therapy aimed at restoring a more neurotrophic environment in the brain.

Materials and Methods

Fly stocks and maintenance

All fly stocks were maintained at 25 or 29°C on a 12:12-h light:dark cycle at constant 60% humidity on a standard cornmeal-based Drosophila medium. Adult-onset neuronal-specific expression of DNT1, human $A\beta_{1-40}$, $A\beta_{1-42}$ and $A\beta_{1-42}$ with Arctic mutation (Arc $A\beta_{1-42}$) was achieved by using the gal4-UAS system. The elav^{C155}-Gal4 driver was used for all experiments. The transgenic fly lines carrying human $A\beta$ peptides have been described previously (45). The transgenic fly lines carrying three forms of DNT1: (1) full-length DNT1 (UAS-DNT1 full length); (2) pro-domain of

DNT1 (lacking the Cysknot) (UAS-DNT1 pro-domain); (3) Cysknot DNT1 (UAS-DNT1 mature-domain, lacking the C-terminal extension) as described by Zhu *et al.* (20) were kind gifts from Dr Alicia Hidalgo (University of Birmingham, UK). Flies transgenic for anti-DNT1 hairpin loop RNAi constructs were obtained from the Vienna Drosophila Resource Center (Transformant ID: 26115, Construct ID: 10831).

Quantitative RT-PCR

Total RNA was extracted from 50 fly heads using the RNA isolation kit (Quiagen) according to the manufacturer's instructions. The concentration of total RNA purified for each sample was measured using a spectrometry on the basis of optical density measurements. One microgram of total RNA was then subjected to DNA digestion using DNAse I (Invitrogen), immediately followed by reverse transcription using the Superscript II system (Promega) with oligo(dT) primers. Quantitative PCR was performed using an SYBR Green PCR mixture (Bio-Rad) according to the manufacturer's instructions. Each sample was analysed in triplicate with both target gene (DNT1) and control gene (RP49) primers in parallel. The DNT1 primers amplified the cDNA coding for the Cysknot: forward 5'-TAGACGACTGCAATT AAAAACAA-3'; reverse 5'-GCCTGTTCAAACTTTAATTCGT-3'. The RP49 primers were as follows: forward 5'-ATGACCATCCGCC CAGCATCAGG-3'; reverse 5'-ATCTCGCCGCAGTAAACG-3'.

Longevity assay

Male flies carrying UAS-Arc $A\beta_{1-42}$ ($A\beta_{1-42}$ with E22G substitution), UAS-DNT1 (various forms of the DNT1 protein), UAS-DNT1RNAi (RNAi against DNT1) or DNT1^{null} (null mutant) were crossed with virgin elav^{C155}-Gal4 driver lines. From the progeny, mated female flies were collected 24 h post-eclosion and divided between 10 tubes, each containing 10 flies. The flies were then cultured with the surviving flies counted on Days 1, 3 and 5 of a 7 days cycle. The median survival of the flies in each of the 10 tubes was plotted for each treatment condition. The mean was indicated ±SEM. The statistical significance of differences in longevity was determined by considering each tube to be an independent estimate of the population median survival and using one-way analysis variance test (*n* = 10).

Locomotor activity assay

Flies carrying UAS-Arc $A\beta_{1-42}$, UAS-DNT1, UAS-DNT1RNAi or DNT1^{null} were crossed with elav^{C155}-Gal4 driver lines. From the progeny, mated female flies were collected 24 h post-eclosion and for each condition five flies were placed in each of three different tubes of 2 cm diameter and 10 cm height. Three 30 s videos were taken on Days 1, 3, 7, 9, 12, 14 and 16 using the iFly apparatus and the walking velocity of the flies' negative geotactic response was determined as described previously (46). Experiments were repeated more than twice, and representative results are shown.

Generation of Aß oligomers

 β -Amyloid (1–42) prepared in HFIP was purchased from Eurogentec. Oligomeric amyloid was prepared as described by Dahlgren et al. (47). Briefly, A β that was lyophilized as a hexafluoroisopropanol film was dissolved in dry dimethyl sulfoxide (DMSO, Sigma) to a concentration of 1 mM and then sonicated for 180 s. Dry DMSO was added to a concentration of 100 or 25 μ M and Ham's F-12 (Invitrogen) was then added to bring the peptide to a final

concentration of 10 or 2.5 $\mu M,$ and the samples were rotated on a rotary shaker at 4°C for 5 days and then RT for 2 days.

Cell culture and treatment

The human neuroblastoma SH-SY5Y cell line from European collection (ECACC) were cultured in medium containing equal volumes of minimum essential medium (MEM, Gibco) and Nutrient Mixture Ham's F-12 (Gibco), supplemented with 1% (v/v) penicillin/streptomycin (antibiotics, Invitrogen), 1% (v/v) non-essential amino acids (Gibco) and 10% (v/v) foetal bovine serum (FBS, Gibco) in a humidified atmosphere containing 5% (v/v) CO_2 at 37°C. Cells were plated 3×10^4 cells/cm² into culture dishes and maintained in growing medium. Twenty four hours after plating, cells were washed and maintained in 0.5% (v/v) FBS-containing medium for 18 h. After that cells were replaced with 0.2% (v/v) FBS-containing medium and then incubated further 48-72 h in the presence of $A\beta_{1-42}$ oligomers with or without recombinant human wild type (Val66) or variant (Met66) BDNF pro-domain (Alomone labs). For differentiation of SH-SY5Y cells, cell were plated 3.5×10^4 cells/cm² into culture dishes and maintained in growing medium. Twenty four hours after plating, cells were washed and replaced with MEM, 1% (v/v) N-2 supplements (Invitrogen), 2 mM L-glutamine (Invitrogen), 1% (v/v) antibiotics and 10 µM all-trans RA (Sigma). The medium was replaced every 2-3 days. After 5-6 days of RA differentiation, fresh medium was provided and incubation continued for a further 48 h in the presence of $A\beta_{1-42}$ oligomers with or without recombinant human variant Met66 BDNF pro-domain.

Measurement of in vitro cell viability

Cell viability was measured using the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT)-based cytotoxicity assay (Promega). Cells were seeded in 24-well plates and then treated with increasing dose of $A\beta_{1-42}$ oligomers or recombinant BDNF, or both reagents at adequate concentrations and then maintained for 60–72 h in a 5% (v/v) CO₂ incubator. MTT was added to all wells and allowed to incubate at 37°C for 4 h, followed by cell lysis and spectrophotometric measurement at 570 nm.

Cell and tissue extraction

The cells were lysed in cell lysis buffer (Abcam) containing 0.216% (w/v) β -glycerophosphate, 1 mM sodium orthovanadate, 1 μ g/ml leupeptin, 1 mM EGTA, 10% (v/v) Triton-X-100, 50 mM Tris-HCl, pH 7.5, 150 mм NaCl, 1 mм sodium EDTA, 1.12% (w/v) sodium pyrophosphate decahydrate and supplemented with phenylmethylsulfonyl fluoride (Sigma), homogenized, sonicated and centrifuged at 20 000g for 30 min at 4°C. The supernatant was then retained. Frozen hippocampal tissue (2-3 mg) was homogenized in ice-cold RIPA buffer containing Tris-HCl buffer, pH 7.5, 105 mм NaCl, 1% (v/v) NP-40, 1% (v/v) Triton X-100, 1% (w/v) sodium deoxycholate, 0.1% (w/v) SDS, 2 mм EDTA supplemented with protease inhibitor cocktail (Roche), sonicated for 480 s. The tissue homogenates were centrifuged at 18 000g for 20 min at 4°C and the supernatants were centrifuged again at 18000g for 10 min at 4°C. The protein concentration was determined using the DC Protein Assay Kit (Bio-Rad).

SDS-PAGE and western blots

Samples for western blotting were diluted into the $4 \times$ NuPage LDS sample buffer (Invitrogen). For western blots of A β from human brain samples the buffer was supplemented with 8 M urea. The

samples were boiled and equal amounts of protein were loaded on to each well of NuPAGE 4-12% (w/v) Bis-Tris Gel and transferred to a 0.45 µm nitrocellulose membrane. The membranes were fixed with 2.5% (v/v) glutaraldehyde (Sigma) in PBS, pH 7.4, for detecting the BDNF pro-domain. The membranes were blocked with 5% (w/v) milk, incubated with primary antibodies against Aβ (6E10; Signet Laboratories), p75^{NTR} (Millipore), Sortilin (Abcam), BDNF pro-domain (GeneCopoeia), BDNF maturedomain (Santa Cruz Biotechnology) or β-actin (Abcam). The membranes were then washed and then incubated with appropriate secondary antibody and developed using ECL detection reagents (Thermo Scientific) before exposing to film (Sigma-Aldrich). Film was digitized and bands were quantified in Image J by measuring the mean grey value. For each western blot, one representative of at least three independent experiments was shown.

Statistical analysis

All data are expressed as means \pm SD or means \pm SEM from at least three independent experiments. For comparison of multiple groups, the one- or two-way analysis of variance (ANOVA) with a Tukey's post hoc test for was used to define statistically significant differences among the groups. The differences between two samples were detected with the two-tailed Mann–Whitney test or Student t-test. Probability values <0.05 were considered as significant.

Supplementary Material

Supplementary Material is available at HMG online.

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Conflict of Interest statement. None declared.

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References

- 1. Querfurth, H.W. and LaFerla, F.M. (2010) Alzheimer's disease. N. Engl. J. Med., **362**, 329–344.
- McAllister, A.K., Katz, L.C. and Lo, D.C. (1997) Opposing roles for endogenous BDNF and NT-3 in regulating cortical dendritic growth. Neuron, 18, 767–778.

- 3. Cosgaya, J.M., Chan, J.R. and Shooter, E.M. (2002) The neurotrophin receptor p75NTR as a positive modulator of myelination. *Science*, **298**, 1184–1186.
- Lyons, W.E., Mamounas, L.A., Ricaurte, G.A., Coppola, V., Reid, S.W., Bora, S.H., Wihler, C., Koliatsos, V. and Tessarollo, L. (1999) Brain-derived neurotrophic factor-deficient mice develop aggressiveness and hyperphagia in conjunction with brain serotonergic abnormalities. Proc. Natl. Acad. Sci. USA, 96, 15239–15244.
- Hariri, A.R., Goldberg, T.E., Mattay, V.S., Kolachana, B.S., Callicott, J.H., Egan, M.F. and Weinberger, D.R. (2003) Brain-derived neurotrophic factor val66met polymorphism affects human memory-related hippocampal activity and predicts memory performance. J. Neurosci., 23, 6690–6694.
- Kolbeck, R., Jungbluth, S. and Barde, Y.A. (1994) Characterisation of neurotrophin dimers and monomers. *Eur. J. Biochem.*, 225, 995–1003.
- Lee, R., Kermani, P., Teng, K.K. and Hempstead, B.L. (2001) Regulation of cell survival by secreted proneurotrophins. Science, 294, 1945–1948.
- Mizoguchi, H., Nakade, J., Tachibana, M., Ibi, D., Someya, E., Koike, H., Kamei, H., Nabeshima, T., Itohara, S., Takuma, K. et al. (2011) Matrix metalloproteinase-9 contributes to kindled seizure development in pentylenetetrazole-treated mice by converting pro-BDNF to mature BDNF in the hippocampus. J. Neurosci., **31**, 12963–12971.
- Huang, E.J. and Reichardt, L.F. (2001) Neurotrophins: roles in neuronal development and function. Annu. Rev. Neurosci., 24, 677–736.
- Carter, A.R., Berry, E.M. and Segal, R.A. (2003) Regional expression of p75NTR contributes to neurotrophin regulation of cerebellar patterning. Mol. Cell Neurosci., 22, 1–13.
- Gehler, S., Gallo, G., Veien, E. and Letourneau, P.C. (2004) p75 neurotrophin receptor signaling regulates growth cone filopodial dynamics through modulating RhoA activity. J. Neurosci., 24, 4363–4372.
- Bamji, S.X., Majdan, M., Pozniak, C.D., Belliveau, D.J., Aloyz, R., Kohn, J., Causing, C.G. and Miller, F.D. (1998) The p75 neurotrophin receptor mediates neuronal apoptosis and is essential for naturally occurring sympathetic neuron death. J. Cell Biol., 140, 911–923.
- Zhou, X.F., Li, W.P., Zhou, F.H., Zhong, J.H., Mi, J.X., Wu, L.L. and Xian, C.J. (2005) Differential effects of endogenous brain-derived neurotrophic factor on the survival of axotomized sensory neurons in dorsal root ganglia: a possible role for the p75 neurotrophin receptor. *Neuroscience*, 132, 591–603.
- Connor, B., Young, D., Yan, Q., Faull, R.L., Synek, B. and Dragunow, M. (1997) Brain-derived neurotrophic factor is reduced in Alzheimer's disease. Brain Res. Mol. Brain Res., 49, 71–81.
- Peng, S., Wuu, J., Mufson, E.J. and Fahnestock, M. (2005) Precursor form of brain derived neurotrophic factor and mature brain-derived neurotrophic factor are decreased in the preclinical stages of Alzheimer's disease. J. Neurochem., 93, 1412–1421.
- Tong, L., Balazs, R., Thornton, P.L. and Cotman, C.W. (2004) Beta-amyloid peptide at sublethal concentrations downregulates brain-derived neurotrophic factor functions in cultured cortical neurons. J. Neurosci., 24, 6799–6809.
- Matrone, C., Ciotti, M.T., Mercanti, D., Marolda, R. and Calissano, P. (2008) NGF and BDNF signaling control amyloidogenic route and Abeta production in hippocampal neurons. Proc. Natl. Acad. Sci. USA, 105, 13139–13144.

- Lim, Y.Y., Villemagne, V.L., Laws, S.M., Ames, D., Pietrzak, R. H., Ellis, K.A., Harrington, K.D., Bourgeat, P., Bush, A.I., Martins, R.N. et al. (2014) Effect of BDNF Val66Met on memory decline and hippocampal atrophy in prodromal Alzheimer's Disease: a preliminary study. PLoS ONE, 9, e86498.
- Lim, Y.Y., Villemagne, V.L., Laws, S.M., Pietrzak, R.H., Snyder, P.J., Ames, D., Ellis, K.A., Harrington, K.D. and Rembach, A. (2014) APOE and BDNF polymorphisms moderate amyloid β-related cognitive decline in preclinical Alzheimer's disease. Mol. Psychiatry, doi: 10.1038/mp.2014.123.
- Zhu, B., Pennack, J.A., McQuilton, P., Forero, M.G., Mizuguchi, K., Sutcliffe, B., Gu, C.J., Fenton, J.C. and Hidalgo, A. (2008) Drosophila neurotrophins reveal a common mechanism for nervous system formation. PLoS Biol., 6, 2476–2495.
- Garzon, D.J. and Fahnestock, M. (2007) Oligomeric amyloid decreases basal levels of brain-derived neurotrophic factor (BDNF) mRNA via specific downregulation of BDNF transcripts IV and V in differentiated human neuroblastoma cells. J. Neurosci., 27, 2628–2635.
- Mucke, L., Masliah, E., Yu, G.Q., Mallory, M., Rockenstein, E.M., Tatsuno, G., Hu, K., Kholodenko, D., Johnson-Wood, K. and McConlogue, L. (2000) High level neurona lexpression of Aβ 1-42 in wild-type human amyloid protein precursor transgenic mice: synaptotoxicity without plaque formation. *J. Neurosci.*, **20**, 4050–4058.
- 23. Nagahara, A.H., Merrill, D.A., Coppola, G., Tsukada, S., Schroeder, B.E., Shaked, G.M., Wang, L., Blesch, A., Kim, A., Conner, J.M. et al. (2009) Neuroprotective effects of brainderived neurotrophic factor in rodent and primate models of Alzheimer's disease. Nat. Med., 15, 331–337.
- Yao, M., Nguyen, T.V. and Pike, C.J. (2005) Beta-amyloidinduced neuronal apoptosis involves c-Jun N-terminal kinase-dependent downregulation of Bcl-w. J. Neurosci., 25, 1149–1158.
- 25. Saadipour, K., Yang, M., Lim, Y., Georgiou, K., Sun, Y., Keating, D., Liu, J., Wang, Y.R., Gai, W., Zhong, J. et al. (2013) Amyloid beta₁₋₄₂ ($A\beta_{42}$) up-regulates the expression of sortilin via the p75^{NTR}/RhoA signaling pathway. J. Neurochem., **127**, 152–162.
- Binder, D.K. and Scharfman, H.E. (2004) Brain-derived neurotrophic factor. Growth Factors, 22, 123–131.
- Phillips, H.S., Hains, J.M., Armanini, M., Laramee, G.R., Johnson, S.A. and Winslow, J.W. (1991) BDNF mRNA is decreased in the hippocampus of individuals with Alzheimer's disease. *Neuron*, 7, 695–702.
- Kunugi, H., Ueki, A., Otsuka, M., Isse, K., Hirasawa, H., Kato, N., Nabika, T., Kobayashi, S. and Nanko, S. (2001) A novel polymorphism of the brain-derived neurotrophic factor (BDNF) gene associated with late-onset Alzheimer's disease. Mol. Psychiatry, 6, 83–86.
- Bodner, S.M., Berrettini, W., van Deerlin, V., Bennett, D.A., Wilson, R.S., Trojanowski, J.Q. and Arnold, S.E. (2005) Genetic variation in the brain derived neurotrophic factor gene in Alzheimer's disease. Am. J. Med. Genet. B Neuropsychiatr. Genet., 134B, 1–5.
- Allen, S.J., Wilcock, G.K. and Dawbarn, D. (1999) Profound and selective loss of catalytic TrkB immunoreactivity in Alzheimer's disease. Biochem. Biophys. Res. Commun., 264, 648–651.
- Li, N. and Liu, G.T. (2010) The novel squamosamide derivative FLZ enhances BDNF/TrkB/CREB signaling and inhibits neuronal apoptosis in APP/PS1 mice. Acta Phamacol. Sin., 31, 265–272.

- Fan, Y.J., Wu, L.L.Y., Li, H.Y., Wang, Y.J. and Zhou, X.F. (2008) Differential effects of pro-BDNF on sensory neurons after sciatic nerve transection in neonatal rats. *Eur. J. Neurosci.*, 27, 2380–2390.
- DeLotto, Y. and DeLotto, R. (1998) Proteolytic processing of the Drosophila Spatzle protein by easter generates a dimeric NGF-like molecule with ventralising activity. Mech. Dev., 72, 141–148.
- Gay, N.J. and Gangloff, M. (2007) Structure and function of Toll receptors and their ligands. Annu. Rev. Biochem., 76, 141–165.
- Hoffmann, A., Funkner, A., Neumann, P., Juhnkes, S., Walter, M., Schierhorn, A., Weininger, M., Balbach, J., Penter, G. and Stubbs, M.T. (2008) Biophysical characterization of refolded Drosophila Spatzle, a cystine knot protein, reveals distinct properties of three isoforms. J. Biol. Chem., 283, 32598–32609.
- Arnot, C.J., Gay, N. and Gangloff, M. (2010) Molecular mechanism that induces activation of Spatzle, the ligand for the Drosophila Toll receptor. J. Biol. Chem., 285, 19502–19509.
- Sutcliffe, B., Forero, M.G., Zhu, B., Robinson, I.M. and Hidalgo, A. (2013) Neuron-type specific functions of DNT1, DNT2 and Spz at the Drosophila neuromuscular junction. PLoS ONE, 8, e75902.
- Ballard, S.L., Miller, D.L. and Ganetzky, B. (2014) Retrograde neurotrophin signaling through Tollo regulates synaptic growth in Drosophila. J. Cell Biol., 204, 1157–1172.
- Lim, Y.Y., Villemagne, V.L., Laws, S.M., Ames, D., Pietrzak, R.H., Ellis, K.A., Harrington, K.D., Bourgeat, P., Salvado, O., Darby, D. et al. (2013) BDNF Val66Met, Aβ amyloid, and cognitive decline in preclinical Alzheimer's disease. Neurobiol. Aging, 34, 2457–2464.
- Chakravarthy, B., Gaudet, C., Menard, M., Atkinson, T., Brown, L., Laferla, F.M., Armato, U. and Whiteld, J. (2010) Amyloidbeta peptides stimulate the expression of the p75 (NTR)

neurotrophin receptor in SHSY5Y human neuroblastoma cells and AD transgenic mice. J. Alzheimers Dis., **19**, 915–925.

- 41. Ito, S., Menard, M., Atkinson, T., Gaudet, C., Brown, L., Whiteld, J. and Chakravarthy, B. (2012) Involvement of insulin-like growth factor 1 receptor signaling in the amyloidbeta peptide oligomers-induced p75 neurotrophin receptor protein expression in mouse hippocampus. J. Alzheimers Dis., **31**, 493–506.
- 42. Chakravarthy, B., Menard, M., Ito, S., Gaudet, C., Dal Pra, I., Armato, U. and Whiteld, J. (2012) Hippocampal membraneassociated p75NTR levels are increased in Alzheimer's disease. J. Alzheimers Dis., **30**, 675–684.
- 43. Yu, H., Yang, M., Wang, Y., Xiao, R. and Zhou, X.F. (2012) p75NTR is mainly responsible for Aβ toxicity but not for its internalization: a primary study. *Neurol. Sci.*, 33, 1043–1050.
- 44. Wu, L.L., Fan, Y., Li, S., Li, X.J. and Zhou, X.F. (2010) Huntingtinassociated protein-1 interacts with pro-brain-derived neurotrophic factor and mediates its transport and release. J. Biol. Chem., 285, 5614–5623.
- Crowther, D.C., Kinghorn, K.J., Miranda, E., Page, R., Curry, J.A., Duthie, F.A.I., Gub, D.C. and Lomas, D.A. (2005) Intraneuronal Abeta, non-amyloid aggregates and neurodegeneration in a Drosophila model of Alzheimer's disease. *Neuroscience*, 132, 123–135.
- Jahn, T.R., Kohlhoff, K.J., Scotta, M., Tartaglia, G.G., Lomase, D.A., Dobson, C.M., Vendruscolo, M. and Crowther, D.C. (2011) Detection of early locomotor abnormalities in a Drosophila model of Alzheimer's disease. J. Neurosci. Methods, 197, 186–189.
- Dahlgren, K.N., Manelli, A.M., Stine, W.B. Jr, Baker, L.K., Krafft, G.A. and LaDU, M.J. (2002) Oligomeric and fibrillar species of amyloid-beta peptides differentially affect neuronal viability. J. Biol. Chem., 277, 32046–32053.