

Train-the-Trainer as an Effective Approach to Building Global Networks of Experts in Genomic Surveillance of Antimicrobial Resistance (AMR)

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Advanced genomics and sequencing technologies are increasingly becoming critical for global health applications such as pathogen and antimicrobial resistance (AMR) surveillance. Limited resources challenge capacity development in low- and middle-income countries (LMICs), with few countries having genomics facilities and adequately trained staff. Training research and public health experts who are directly involved in the establishment of such facilities offers an effective, but limited, solution to a growing need. Instead, training them to impart their knowledge and skills to others provides a sustainable model for scaling up the much needed capacity and capability for genomic sequencing and analysis locally with global impact. We designed and developed a Train-the-Trainer course integrating pedagogical aspects with genomic and bioinformatics activities. The course was delivered to 18 participants from 12 countries in Africa, Asia, and Latin America. A combination of teaching strategies culminating in a group project created a foundation for continued development at home institutions. Upon follow-up after 6 months, at least 40% of trainees had initiated training programs and collaborations to build capacity at local, national, and regional level. This work provides a framework for implementing a training and capacity building program for the application of genomics tools and resources in AMR surveillance.

Keywords. train-the-trainer; antimicrobial resistance; AMR surveillance; genomics; bioinformatics.

Capacity Building for Genomic Surveillance of AMR

Antimicrobial resistance (AMR) is a major global challenge that is increasing the burden on health systems and impacting on food sustainability, environmental wellbeing, and socioeconomic development globally [1]. To strengthen knowledge and evidence base and inform decision making, the World Health Organization (WHO) has recommended the application of advanced technologies such as whole genome sequencing (WGS) for surveillance of pathogens and AMR. However, in low- and middle-income countries (LMICs), implementation of genomic surveillance of AMR is challenged by the high costs of infrastructure and equipment, and the lack of specialized workforce [2].

To build capacity for pathogen surveillance and genomics, a critical mass of expertise in sequencing techniques and

bioinformatics is essential. The establishment of local and regional networks and trainers will raise the impact and ensure the sustainability of the long-term implementation of the genomic surveillance of AMR.

In 2018, the Centre of Genomic Pathogen Surveillance (CGPS) was funded as a partner within the NIHR Global Health Research Unit (GHRU) on Genomic Surveillance of Antimicrobial Resistance, with the overall aim to establish intelligent genomic surveillance of bacterial pathogens through appropriate sampling and analysis with partners in Colombia, India, Nigeria, and the Philippines.

To address the need for expertise, in 2019, the GHRU collaborated with Wellcome Genome Campus Advanced Courses and Scientific Conferences (ACSC) to design and deliver a Train-the-Trainer (TtT) course, which focused on the application of genomics in AMR surveillance [3]. The goal was to establish a global network of trainers, capable of delivering direct training and presentation of concepts to varied audiences and contributing to building capacity for the genomic surveillance of AMR laboratories in LMICs.

An Effective Method for Building Sustainable Global Networks of Experts

Train-the-Trainer refers to a program or course where subject domain specialists receive training in a given subject and pedagogical skills needed to train and share their expertise with others. Scaling up training programs in various domains has

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improved coverage and access, reduced costs, utilized local trainers who are knowledgeable about local issues, and encouraged collaboration among researchers and staff from neighboring research facilities [4] (see Figure 1).

Several organizations have developed networks of training expertise aimed at addressing the knowledge gaps in fast-changing fields, such as genomics and bioinformatics. TtT courses teach researchers the skills to train others in topics that are broad in scope, such as data analysis (eg, carpentries), bioinformatics (eg, H3ABioNet in Africa, EBI-CABANA in Latin America, Next Generation Sequencing Bioinformatics program in Australia), and digital skills (ARDC, a national initiative to support Australian research) [5–9].

The ACSC Program also developed tailored models of training trainers, which were delivered in addition to the usual training in biomedicine and genomics for researchers and healthcare professionals worldwide. Participants were equipped with tools for training others following good educational practice. A key outcome of the ACSC TtT program was the establishment of regional instructor teams, who collaborate with international experts to deliver training in LMIC regions where the need for trainer expertise is most critical.

Course Development

The goal of the TtT course was to equip participants with skills for the application of effective methods for training adult learners in laboratory techniques and bioinformatics required for the implementation of genomic surveillance of AMR toward establishing global networks of expertise. To effectively train others, participants' learning outcomes included demonstration of knowledge in (a) setting up a genomic surveillance laboratory and bioinformatics pipelines; (b) implementation of laboratory and bioinformatics procedures for generation, analysis, and interpretation of genome data for AMR surveillance; and (c) teaching genomic surveillance of AMR to varied audiences.

The course instructor team included experts in education, bioinformatics, epidemiology, genomics, and AMR laboratory methods. Prior to developing the course, the instructor team undertook a TtT course, which provided guidance for the development of an integrated program encompassing pedagogical aspects, and laboratory and computational techniques.

The course was attended by 18 scientists, working in academic research centers or national reference laboratories in 12 countries (Argentina, Brazil, Colombia, Cuba, India, Malaysia, Nigeria, South Africa, Sudan, Thailand, the Philippines, and Vietnam).

The Structure of the Course

The course consisted of 12 modules, which were run either in parallel or as joint sessions (Figure 2). The course ran over 6 days and included didactic lectures, interactive sessions, and seminars, totaling 47.5 contact hours.

The pedagogical modules and refresher sessions were run with the whole cohort of attendees. Later, the participants were split into 2 workstreams specific to their interests and expertise. The Laboratory workstream covered processes that enable WGS of bacterial pathogens, from sample collection to sequencing quality assurance. The Bioinformatics workstream covered various aspects, from building the infrastructure and pipelines required for WGS data analysis, to interpretation applied to global contexts. The course concluded with a session in which participants learned aspects needed to be considered when communicating with policymakers and other decision makers.

Four guest lectures given by scientists and public health experts in AMR surveillance offered complementary views on the challenges faced in the implementation of genomic AMR surveillance in the United Kingdom versus LMICs.

Participants were introduced to a Training and Learning Toolkit, an online book of worksheets, where they collected their own work during the course, as well as their notes on course/module design, networks, resources, and reflections.

A Strong Emphasis on Pedagogy

The TtT course was designed to support social learning and teaching, where both the instructors that facilitated the training and course participants took active roles and responsibilities for their own teaching and learning. Learning and teaching took several formats: direct teaching/training through presentation or demonstration followed by Q&A session (eg, direct instruction on use of scripts/programs, on setting up labs, or on different lab procedures); group work (for peer-learning on any topic covered in the course); individual reflection, and consolidation. The course considered training on many levels, and the course instructors and facilitators modeled behavior to be adapted and adopted by the participants in the last module of the course but also in their future teaching and learning.

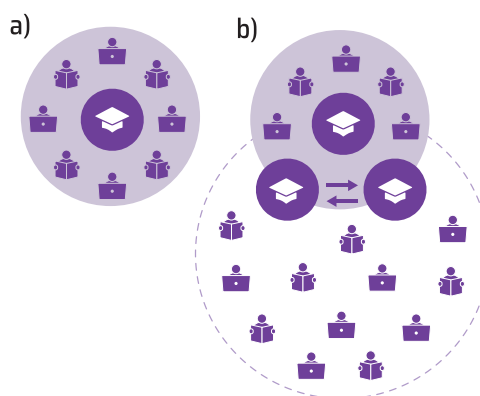


Figure 1. Difference in sizes of key beneficiaries of training, between (a) conventional training courses, where a trainer (mortarboard hat) teaches participants that gain individual skills, which they then apply to their own data (audience is shown as people with books and laptops); and (b) Train-the-Trainer courses, where the trainer (mortarboard hat) teaches other trainers, who go on to teach more people (audience), and the final audience is larger.

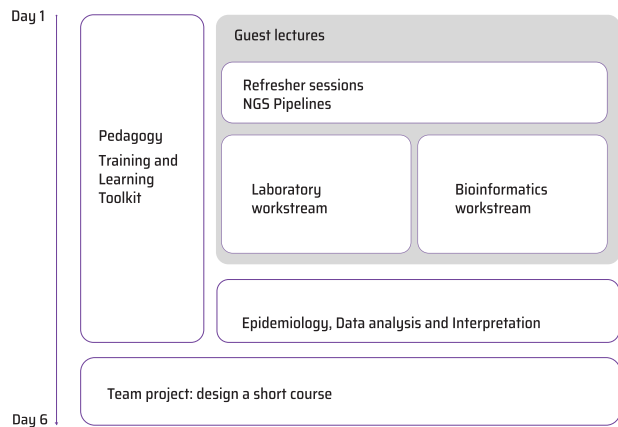


Figure 2. Structure of the course in time, with some modules run in parallel. Abbreviation: NGS, next-generation sequencing.

The expanded learning outcomes (LOs) and module content purposefully integrated the various theoretical and practical aspects of both the pedagogy and the scientific training (Table 1). Learning outcomes 1–4 were covered in the pedagogy sessions introducing educational theory and active learning pedagogy. Topics included elements of course design, writing LOs, and

assessments. Scientific content focused on LOs 5 and 6, providing refresher content and demonstration of key techniques and pipelines required for effective AMR genomic surveillance. This further developed to higher-level outcomes 7 and 8, where participants collaboratively designed and presented their modules combined with feedback from the instructors.

Teaching Approaches

The teaching on the course was integrated and made uniform through the use of templates for instructional design and through the application of the adult and active learning principles throughout [12]. In addition, the instructors used customized teaching approaches to bioinformatics and laboratory topics (Table 2). The main characteristics of bioinformatics and laboratory-specific teaching on this course are Table 2.

Assessment for Learning

Formative assessment was implemented throughout the course: quizzes, discussions, Q&A sessions, group work and reporting, peer critiquing, module/course presentation with peer and instructors' feedback, pitching of a genomics training plan with instructors' feedback.

Table 1. Pedagogical Learning Outcomes Mapped Onto Content

Pedagogical Learning Outcomes	Modules																
1. Discuss learning and teaching (or educational) theory in the context of active learning pedagogy	<ul style="list-style-type: none"> Introduction to pedagogical theory, training and learning techniques Part 1 																
2. Apply good practice learning/teaching skills of communication, presentation, and group work rules																	
3. List and describe the basic elements of course design	<ul style="list-style-type: none"> Introduction to pedagogical theory, training and learning techniques Part 2 																
4. Illustrate how underpinning pedagogy guides course design	<ul style="list-style-type: none"> Bioinformatics and WGS Lab refresher training (joint session) Integration and interpretation of lab and genomic data (joint session) 																
5. Discuss, and share learning design and course design practices in various environments	<table border="0"> <tr> <td>Laboratory stream</td> <td>Bioinformatics stream</td> </tr> <tr> <td>Teaching of the following topics:</td> <td></td> </tr> <tr> <td>o Genome Sequencing and its Applications</td> <td>o Bioinformatics infrastructure and pipelines</td> </tr> <tr> <td>o Assembling the WGS technology suite</td> <td>o Databases</td> </tr> <tr> <td>o Phenotypic characterization—IS/AST</td> <td>o De novo assembly and Reference-based approaches, pros and cons</td> </tr> <tr> <td>o Laboratory Quality Assurance</td> <td>o Phylogeny</td> </tr> <tr> <td>o Sequencing Quality Assurance</td> <td>o Databases</td> </tr> <tr> <td></td> <td>o Genotypic AMR prediction</td> </tr> </table>	Laboratory stream	Bioinformatics stream	Teaching of the following topics:		o Genome Sequencing and its Applications	o Bioinformatics infrastructure and pipelines	o Assembling the WGS technology suite	o Databases	o Phenotypic characterization—IS/AST	o De novo assembly and Reference-based approaches, pros and cons	o Laboratory Quality Assurance	o Phylogeny	o Sequencing Quality Assurance	o Databases		o Genotypic AMR prediction
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6. Identify opportunities for creating learning experiences from subject content																	
7. Design, develop, and present short course or module(s) encompassing aspects of genomic surveillance	<ul style="list-style-type: none"> Module design practical Module project presentation by participants 																
8. Reflect on created (or existing) modules and identify areas for improvement																	

Abbreviations: AMR, antimicrobial resistance; AST, antimicrobial susceptibility testing; WGS, whole genome sequencing.

Table 2. Teaching Approaches

Strategy	Description	Implementation Examples
Challenge and problem-based tasks	Problem-solving and decision-making exercises	<ul style="list-style-type: none"> • Use card-based blueprint to order processes and procedures in a typical WGS for AMR surveillance pipeline • Use of fictive case scenarios involving training others in genomics aspects
Case studies	Use of real cases, as published in the literature	<ul style="list-style-type: none"> • Identify MDR <i>Salmonella enterica</i> subsp. <i>enterica</i> serotype Kentucky lineage expansion [13] • Investigation of a Brazilian epidemic of <i>Mycobacterium abscessus</i> species complex [14] • <i>Klebsiella pneumoniae</i> outbreak investigation [15]
Historical and chronological presentation of processes	Demonstration of lab procedures, visual tours, lab visits, exhibition tour	<ul style="list-style-type: none"> • Visit to the sequencing facilities at the Sanger Institute • Visit to the exhibition marking the start of the Human Genome Project • Demonstration of library preparation for WGS • Demonstration of best practices for antimicrobial susceptibility testing
Comparative analysis of tools and platforms	Presenting alternative ways of achieving the same/similar result	<ul style="list-style-type: none"> • Comparison of data merging and consolidation techniques—interactively with spreadsheets versus programmatically with R and Python • Comparison of different databases and software tools used to determine AMR phenotype based on genomic data • Comparison of different ways of interpreting bacterial population structures—MLST trees versus phylogenetic trees obtained from WGS data
Project planning and management techniques	Considering standards and quality control, constraints and limitations, time management; project planning, and constraints (financial, scaling and other)	<ul style="list-style-type: none"> • Exercise on how to build a computer server that needs to accomplish certain tasks, given a list of hardware components and a fixed budget • Exercise on how to set up a WGS laboratory • QC assessment of libraries and sequencing
Provision of resources	Provision of links to resources and portals, quality control standards, virtual and cloud tools, toolkits	<ul style="list-style-type: none"> • Software tools: • AMR prediction tools: ARIBA [16], Pathogenwatch [17], AMRFinder [18] • AMR databases: CARD [19], ResFinder [20] • Nucleotide archives: Genbank [21], European Nucleotide Archive [22] • Data integration tools: dataflo (https://dataflo.io/) • Data visualization: Microreact [11] • Pipelines: Nextflow [23] • Phylogeny: RAxML [24] • Genome Assembly: SPAdes [25] • QC tools: FastQC [26], Quast [27], Bactinspector [28], Qualifyr [29] • Typing tools: PubMed MLST (www.pubmlst.org) • The Training and Learning Toolkit
Analysis and evaluation	Consideration and evaluation of participants' specific contexts	<ul style="list-style-type: none"> • Big-picture exercise where participants evaluate their own circumstances

Abbreviations: AMR, antimicrobial resistance; MDR, multidrug resistance; MLST, multilocus sequence typing; QC, quality control; WGS, whole genome sequencing.

To implement all the learning activities and formative assessment on this course, a number of specific tools, services, and learning material were used, including Post-it Notes, printed cards, bioinformatics online tools and platforms, online quiz tools, presentation tools (PowerPoint, flipcharts), live documents, and lab settings.

Project-Based Learning

Integrated teaching and learning on this course resulted in the hands-on design of different genomics modules by participants who worked in small groups, so they could collaborate and bring in personal expertise. To guide them with the design, participants used the Toolkit, an online tool shared with the instructors, which provided them with formative feedback during the project and gave them electronic access to course content and outputs throughout the course. The project required the application of the course design principles taught in the pedagogy modules. A well-established methodology was presented, which considers aspects of course design, such as the course aim, target audience, and the level of the course,

the formulation of the learning outcomes using revised Bloom's taxonomy of learning, and outlining of the course content and assessment. How to create specific learning activities taking into account the underpinning pedagogy, and how to plan for the course/module delivery and evaluation, considering specific circumstances, were also discussed. The same process applied in the design, development, delivery, and evaluation of the TtT course was presented to the course participants and used in the final project. This provided the course organizers and instructors with formative feedback. The projects also encouraged creative thinking, collaboration, communication, and presentation skills, and created opportunities for networking. Instructors reviewed the projects and provided constructive critique to the training plans designed by the participants. Project presentations with feedback from instructors and participants concluded the major training activities of the course.

FEEDBACK AND EVALUATION

A post-course survey asked participants about course organization, course content, what participants most enjoyed in the

course, and any suggestions for changes or improvements. All participants were satisfied with the course materials and indicated that the course had been very relevant to their research or work. Although the course was intensive, participants reported that the modules were well designed and met expectations, and that the combination of active learning strategies was beneficial to their learning and interactions with each other. However, participants also felt that networking and time for discussions could be improved. Since participants were separated by workstream, a suggestion was made to provide a more overlapping structure, whereby bioinformaticians could have an overview of project management approaches for laboratory pipelines. Feedback on the course from organizers, trainers, and trainees was also filmed, and the recording is available online [30].

A follow-up survey was conducted 6 months and 1 year post-course to understand whether and how participants had applied their knowledge in their research or professional practice. Respondents indicated that they had implemented, or were planning, specific training in microbiology, AMR, sequencing, and bioinformatics techniques to a range of audiences, including university students, epidemiologists, scientists involved in AMR surveillance. Four of the trainees had facilitated regional courses on genomic epidemiology and bioinformatics analysis workflows for AMR, and more than half have continued to network with other course participants informally. One of the respondents was awarded an international grant to start a pilot training program in their region [31, 32]. Respondents also reported plans or implementation of remodeling their AMR laboratories for WGS and designed SOPs, and tailored WGS data analysis workflows. Participants expressed challenges in implementation due limited infrastructure set-up and delays due to the coronavirus disease 2019 (COVID-19) pandemic. However, some participants reported on how the TtT course had impacted on their personal skills, including improved teaching skills and use of active learning and learner-oriented approaches. Others reported improved collaborative and scientific skills including analysis of WGS data and improved understanding of the principles for all aspects of sequencing, including troubleshooting and workflow procedures.

DISCUSSION

This article describes the first delivery to a regionally diverse audience of a TtT course for building capacity for the application of genomics tools and resources in AMR surveillance. Advances in genomics and emerging infectious disease and environmental threats create a need to train more scientists in pathogen surveillance and bioinformatics skills [33]. This TtT course provided a timely intervention coinciding with global AMR strategies to build capacity in genomic surveillance and the establishment of global centers [34–37].

In LMICs, limited expertise in genomics applications in the areas of epidemiology, surveillance, and public health results in short specialized courses being offered through collaborations with regional and international experts. However, this is severely challenged by the time limitations of trainers and experts, and funding and travel constraints. In addition, public health scientists may not be able to take time away from their core duties or secure the necessary resources to travel for training. This work established a framework to develop skilled local trainers who are knowledgeable about local issues, encouraging local and regional collaboration.

The integrated domain and application-specific approach enabled participants to refresh and reflect on their knowledge and expertise, while learning how to effectively train others. The course applied an innovative combination of teaching approaches, emphasizing the elements of how to teach various audiences in bioinformatics and laboratory techniques used in the genomic surveillance of AMR. Although some domain-specific teaching approaches were similar to those more commonly used in general bioinformatics and laboratory education, some of them were uniquely applied to this course (Table 2) [38].

The integration of pedagogy with scientific aspects emphasized that “what to teach” is as important as “how to teach.” Ideally, this should be accompanied by an understanding of how students best learn specific topics, and by reviewing well established methodologies for the training of laboratory techniques or aligning bioinformatics educational approaches with those in computer sciences (eg, [38]).

The successful implementation of the course was highlighted in the feedback and follow-up evaluation, with participants reporting that they had applied what they learned to establish experimental and analysis pipelines in their institutions and train people in their countries and regions. This highlights the role of the TtT course in building capacity and applying genomics technologies locally with global impact.

Global health partnerships integrate collaborative TtT courses as a key part of effective and sustainable capacity building programs [39]. This TtT course established a scalable model for up-skilling domain expert trainers capable of training others and locally cascading skills development in a context-specific and sustainable way. This provided a foundation to build a global network of trainers and experts, thereby empowering scientists to work more collaboratively on the common goals of tackling AMR.

There is a clear requirement within such courses to establish frameworks for measuring impact and success. Continued follow-up of trainees is possible through the network platform and will be complimented by a more rigorous impact evaluation approach, which is within the scope of future projects. To strengthen efforts initiated by participants and other professionals in their local settings, we are working toward building a network of mentors to provide guidance for running courses independently in

the future. This will be guided by a competency framework for training in genomic epidemiology and AMR surveillance.

CONCLUSION AND FUTURE WORK

Global health consortia and collaborative projects across continents play an important role in establishing centers of excellence, providing infrastructure, facilities, and shared resources for genomic surveillance of AMR [40]. To complement these efforts, training trainers is a sustainable way to build local and regional capacity and networks for research and public health. Our TtT framework presents a potentially effective model for building capacity and developing sustainable trainer networks locally for global impact. Materials for this course are available on demand for anyone interested in developing training materials for other related TtT courses. This course was also used as a case study for demonstrating effective strategies for capacity building through training for genomic surveillance of AMR (<https://www.futurelearn.com/courses/train-the-trainer-design-genomics-training>). To complement these efforts, future work will involve the development of standardized trainer resources and best practice guidelines. In addition, digitalization of such a course is important in widening reach across regions where resources and expertise are a major challenge. Finally, various initiatives both at regional and global levels are underway aimed at developing networks and training resources (eg, SAGESA <https://sagesa.africa/>, PHA4GE <https://pha4ge.org/>) strengthening capacity and making training accessible globally.

Notes

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