



Proteoglycans and Immunobiology of Cancer—Therapeutic Implications

George Tzanakakis¹, Monica Neagu^{2,3}, Aristidis Tsatsakis⁴ and Dragana Nikitovic^{1*}

¹ Laboratory of Anatomy-Histology-Embryology, School of Medicine, University of Crete, Heraklion, Greece, ² Immunology Department, "Victor Babes" National Institute of Pathology, Bucharest, Romania, ³ Pathology Department, Colentina Clinical Hospital, Bucharest, Romania, ⁴ Laboratory of Toxicology, University of Crete, Heraklion, Greece

Disparity during the resolution of inflammation is closely related with the initiation and progression of the tumorigenesis. The transformed cells, through continuously evolving interactions, participate in various exchanges with the surrounding microenvironment consisting of extracellular matrix (ECM) components, cytokines embedded in the ECM, as well as the stromal cells. Proteoglycans (PGs), complex molecules consisting of a protein core into which one or more glycosaminoglycan (GAG) chains are covalently tethered, are important regulators of the cell/matrix interface and, consecutively, biological functions. The discrete expression of PGs and their interacting partners has been distinguished as specific for disease development in diverse cancer types. In this mini-review, we will critically discuss the roles of PGs in the complex processes of cancer-associated modulation of the immune response and analyze their mechanisms of action. A deeper understanding of mechanisms which are capable of regulating the immune response could be harnessed to treat malignant disease.

OPEN ACCESS

Edited by:

Toshiyuki Murai, Osaka University, Japan

Reviewed by:

James McCarthy, University of Minnesota Twin Cities, United States Alberto Passi, University of Insubria, Italy

*Correspondence:

Dragana Nikitovic nikitovic@uoc.gr

Specialty section:

This article was submitted to Cancer Immunity and Immunotherapy, a section of the journal Frontiers in Immunology

> Received: 31 January 2019 Accepted: 05 April 2019 Published: 24 April 2019

Citation:

Tzanakakis G, Neagu M, Tsatsakis A and Nikitovic D (2019) Proteoglycans and Immunobiology of Cancer—Therapeutic Implications. Front. Immunol. 10:875. doi: 10.3389/fimmu.2019.00875 Keywords: proteoglycans, immunobiology, cancer, extracellular matrix, remodeling

INTRODUCTION

Cancer initiation is a multi-faceted process with a contribution of genetic, metabolic and environmental factors. Tumorigenesis is closely associated with chronic inflammation, with approximately 20% of cancer incidences being directly correlated to chronic infections (1). Indeed, all tumors independently of etiology are distinguished by an early inflammatory milieu and characterized by discrete interactions with the immune system at all stages of disease progression (2, 3). During malignant transformation, cells obtain complex biological characteristics correlated with more efficient survival, invasion, metastasis and the ability to evade the immune response. The transformed cells, through continuously evolving interactions, communicate with and alter the surrounding microenvironment consisting of extracellular matrix (ECM) components, cytokines embedded in the ECM, and the stromal cells (e.g., fibroblasts, endothelial cells, adipocytes, and immune cells) (4, 5). The resulting ECM remodeling crucially contributes to the abnormal tumor inflammatory pattern (5, 6).

Proteoglycans (PGs) are complex molecules consisting of a protein core into which one or more glycosaminoglycan (GAG) chains are covalently tethered. The bound GAGs can be the heparan sulfate (HS), the chondroitin sulfate/dermatan sulfate (CS/DS), or the keratan sulfate (KS) type. In mammalian cells PGs are associated to the plasma membranes, released into the ECM or intracellularly localized. Presently, 45 PGs have been identified with each member characterized by immense alterability attributed to the modifications of the protein core and by the type and different stoichiometry of the GAG chain substitutions (7, 8). Thus, PGs have highly specific

1

and multifaceted biological roles, including: (i) contributing to ECM superstructure (9); (ii) defining ECM biochemical and physicochemical properties (9); (iii) acting as receptors of diverse responsiveness as well as a pool of various biologic effectors such as growth factors (10-12).

It is well-established that malignant tumors have discrete PG expression profiles, which are immediately correlated with their behavior and differentiation status. Thus, epithelial tumors exhibit a different PG characterization as compared to mesenchymal tumors (13, 14). Proteolytic cleavage of PGs, due to the action of matrix metalloproteinases (MMPs), cathepsins, and bone-morphogenetic protein-1, can release bioactive fragments or matrikines with roles in the propagation of tumorigenesis separate from that of parent molecules (15). Importantly, PGs are highly implicated in the processes of cancer-associated inflammation (16, 17). Indeed, PGs are suggested to modulate key events respective to both innate and adaptive immunity (18, 19).

In this review, we provide a critical overview of PGs' roles in the inflammatory cancer milieu and consecutively extrapolate to potential options for the development of targeted cancer therapies.

IMMUNOBIOLOGY OF CANCER

The input of the immune system, introduced as cancer immunoediting, consists of three phases: elimination (i.e., cancer immunosurveillance), equilibrium, and escape (20). Importantly, the tumor ECM contributes to the development of an immunosuppressive network where stromal cells intertwine with inflammatory immune cells and with cells appending to the vascular system. Within this complex, a newly formed network, secreted cytokines and chemokines can sustain the tumor immune escape (21, 22). The chain of tumorigeneis can be initiated by injury of normal tissue, independently of the causative agent, and concomitant triggering of acute inflammation (23, 24). The maintenance of inflammatory conditions due to various effectors leads to chronic inflammation which may evolve to precancerous lesion (25-29). The evolvement of the pre-cancerous lesion can be attenuated by an active immune defense or driven to primary tumor development (30), as schematically presented in Figure 1. Thus, the immune system has the ability to perceive and destroy many tumors early on in their development, whereas during the stage of equilibrium, a restraint of the tumor is attained. Some tumors will succeed in escaping from the growth restriction maintained by the immune system, and become clinically apparent (20, 31).

The resolution of disease and response to therapy is likewise affected by the tumor microenvironment as two major subsets of tumors with distinct mechanisms of resistance to immunemediated destruction have been recognized. Tumors exhibiting the inflamed immunophenotype present with the recruitment of $CD8^+$ cytotoxic T cells, B cells and macrophages. Immune resistance in this case is due to the action of microenvironmentoriginating negative immune regulators. In non-inflamed tumors an absence of T-cells and innate immunity regulators is evident and leads to immediate immune failure (32, 33). Indeed, the ability of the tumor cells to escape impairment by the immune system has been suggested as a novel "hallmark of cancer" (6).

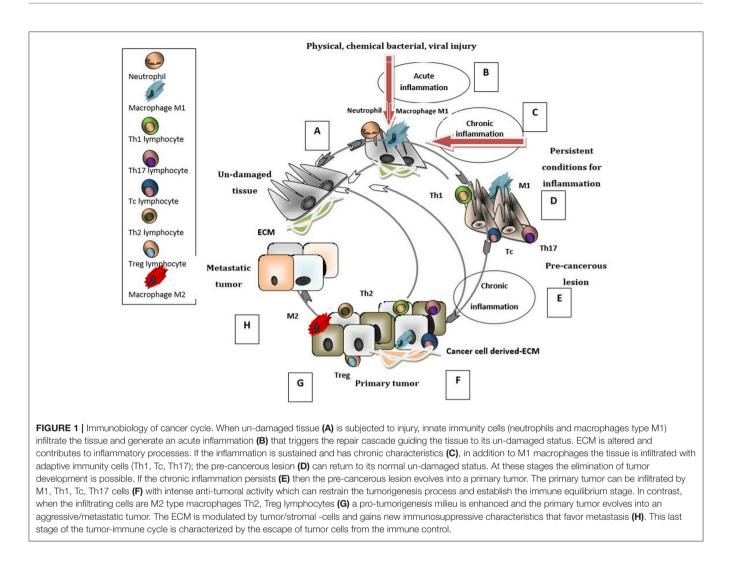
REMODELING OF THE CANCER ENVIRONMENT

During tumor progression, an extensive remodeling of the ECM with correlated release of pro-tumorigenic factors and orchestration of surrounding "stroma" cells is initiated. This remodeling of the tumor microenvironment is closely associated to the modulation of the immune response (34, 35). Thus, in cancer pathogenesis the resulting "desmoplastic reaction" among resident fibroblasts, endothelial cells, pericytes, leukocytes and surrounding ECM is directly correlated with invasion and poor patient prognosis (36, 37). The restructuring of the ECM is due to: (i) modulation in the synthesis and release of ECM components; (ii) degradation of the ECM owing to enzyme action or chemical degradation due to radical oxygen species (ROS) (4, 38). Specific ECM remodeling will ultimately result in: (i) the detachment of tumor cells from each other, from adjacent stromal cells or from matrix; (ii) enhanced growth and mobility of tumor cells; (iii) modulation of the immune system and finally in sustaining of the tumorigenic microenvironment (39, 40).

PGs ARE ACTIVE MEDIATORS OF CANCER-ASSOCIATED INFLAMMATORY MILIEU

Small Leucine Rich Proteoglycans Have a Dual Role in the Regulation of Cancer Inflammatory Setting

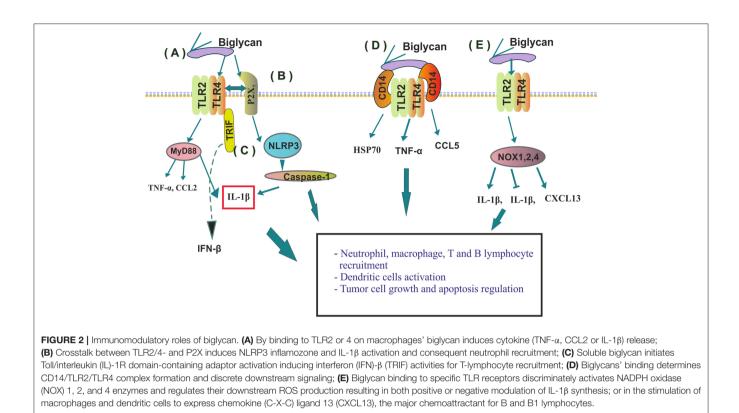
The family of small leucine rich PGs (SLRPGs) was initially correlated with the regulation of innate immunological responses, noteworthy due to the fact that the triggering of these very responses can lead to the initiation of tumorigenesis (41). The role of a pro-inflammatory molecule has been designated to a class I SLRP member, biglycan (17). Importantly, this SLRP is overexpressed and secreted by various cancers including gastric (42), pancreatic (43), ovarian (44), and colon cancer (45). This SLRP can be either freed from the ECM through proteolytic degradation initiated upon tissue injury or de novo produced by activated macrophages and resident cells with various immunological roles (Figure 2) (46). Soluble biglycan can undertake the role of signaling mediator by binding to the Toll-like receptors (TLR)-2 and -4 on the surface of macrophages. The formation of the ligandreceptor complex initiates sterile inflammation and can facilitate pathogen-mediated inflammation through the production of pro-inflammatory cytokines and chemokines, including the tumor necrosis factor (TNF)-α, interleukin (IL)-1β or chemokine (C-C motif) ligand (CCL)2 (46, 47). It was shown that soluble biglycan utilizes TLR2/4 signaling pathways to activate adaptor molecule myeloid differentiation primary response 88 (MyD88), for the recruitment of neutrophils and macrophages or to initiate Toll/interleukin (IL)-1R domain-containing adaptor inducing interferon (IFN)-β (TRIF) activities for T-lymphocyte



recruitment (48). Moreover, the role of a danger signal (DAMP) that triggers the NLRP3 inflammasome through upstream TLR2/4 and P2X receptors signaling has been attributed to biglycan (49), by specifically regulating the crosstalk between TLR2/4- and P2X7-NLRP3-caspase-1 (50, 51). Increasing data proposes that the initiation of TLR2/TLR4 signaling enhances tumor cell growth, downregulates apoptosis, and upregulates the synthesis of growth factors and inflammation-associated cytokines by tumor and stromal cells (50, 52, 53). Recently, biglycan was shown to bind with high affinity to macrophage CD14, an established GPI-anchored TLRs co-receptor. CD14 is mandatory for biglycan-dependent TLRs activation, where biglycan seems to have the role of a re-router as complexing with specific TLR members induces a discrete response. Thus, in macrophages, the biglycan/CD14/TLR2,4 complex induces the TNF-α expression, thebiglycan/CD14/TLR2 co-localization results in HSP70 release, whereas the biglycan/CD14/TLR4 complex initiates CCL5 secretion (54). Moreover, in a mouse model of renal injury, a deficiency of CD14 prevented biglycan-mediated cytokine expression, recruitment of macrophages, and M1 macrophage polarization (54).

Moreover, biglycan has been implicated in the regulation of important to tumorigenesis (4), ROS generation (55). Indeed, biglycan through activating discrete TLR receptors discriminately stabilizes NADPH oxidase (NOX) 1, 2, and 4 enzymes and regulates their downstream ROS production, resulting in either a positive or negative modulation of IL-1β synthesis (18). Furthermore, TLR2/4 induced ROS generation by biglycan (56) stimulates macrophages and dendritic cells to express chemokine (C-X-C) ligand 13 (CXCL13), the major chemoattractant for B and B1 lymphocytes (56). In addition to chemoattracting B cells, biglycan through TLR2/4 stimulates RANTES, MCP-1, and MIP-1a secretion, inducing macrophages and T cells recruitment into the kidney. Thus, these developments indicate that biglycan signaling can bridge innate and adaptive immunity (56), with the ability to act at different check points of the tumor immune cycle.

On the other hand, by enhancing NOX-generated ROS, biglycan induces genomic volatility coupled with chromosomal DNA modifications, which results in increased tumor cell growth, viability, and a metastatic ability of inflammation-associated tumors (57). Even though the majority of reports



suggest that the activities of biglycan are pro-oncogenic, recent studies provide evidence that biglycan can promote an antiinflammatory response that is key for the resolution of acute inflammation and hence the switch to acquired immunity response (18). Indeed, it is now proposed that biglycan, even though exhibiting affinity to both TLR2 and 4, will discriminately bind to only one TLR, which will, in combination with specific TLR adaptor molecules, lead to a discrete biological response (17, 18). The summarized regulation of immunity responses by biglycan can conceivably affect the development and resolution of early pre-cancerous lesions as well as the progression of inflamed tumor subtypes. Decorin, likewise an SLRP class I member, has established anti-tumorigenic properties (58). This SLRP member has the ability to initiate the TLR2/4 downstream signaling and cytokine release with an outcome different to that of biglycan. Specifically, utilizing a transforming growth factor -β (TGF-β)/oncogenic microRNA (miR)-21/tumor suppressor programmed cell death protein 4 (PDCD4) axis decorin downregulates the release of IL-10, which is an anti-inflammatory mediator and thus inhibits tumor growth (59). Moreover, decorin mobilizes mononuclear cells to the region of damaged tissue by enhancing CCL2 release and thereby, effectively upholding the inflammatory state (60). In a breast cancer xenographic model it was demonstrated that the decorin protein core inhibits genes obligatory for the regulation of the immunological response. Therefore, Buraschi et al. conclude that the "systemic administration of decorin protein core reveals a fundamental basis of action for decorin to modulate the tumor stroma as a biological mechanism for the ascribed anti-tumorigenic

properties." Indeed, the authors' gene ontology data indicated an inhibitory role in the regulation of proteins implicated in immunomodulatory responses when assigned to decorin protein core (61). The SLRP lumican has been postulated to modulate tumor-associated inflammation by affecting peripheral monocyte extravasation, and Fas–FasL signaling (14). Interestingly, lumican enhances LPS-dependent activation of TLR4 (62).

The Hyalectan Versican Is Crucial to Both Innate and Adaptive Immunity

Versican, a member of the hyalectan family of large chondroitin sulfate PGs (CSPGs) has also been implicated in inflammatory processes (19, 63). These pericellular PGs are localized, among other, to the sub-endothelial compartment where they encounter the infiltrating leukocytes and modulate their biological activities by binding to specific receptors including TLR2, and P- or Lselectins (19). Once bound to the versican-containing ECM, leukocytes degrade the ECM to generate pro-inflammatory fragments that further drive the inflammatory response (64). Versican has been established to be overexpressed in a number of cancers, including prostate, breast, malignant myeloma, glioblastoma, laryngeal, pancreatic ovarian, gastric, testicular germ-cell, and cervical cancer, as recently discussed (65).

Importantly, in cancer-associated inflammation, versican can affect the production of cytokines by both lymphoid and myeloid cells. Thus, Lewis lung carcinoma (LLC) cells overexpress versican in that, by binding to TLR2 and its co-receptors on macrophage cell membranes, it activates the latter and facilitates their TNF- α secretion (66). The initiation of this cascade by

versican and the induction of TNF- α by myeloid cells enhances LLC cell metastatic growth (66). Furthermore, an increased expression of versican V1 and V3 isoforms was positively correlated to lung metastasis in a bladder cancer murine model due to the increased release of CCL chemokine by macrophages (67). Specifically, versican was shown to facilitate bladder tumor cell migration, resulting in lung metastasis due to a mechanism involving CCL2/CCR2 secretion by macrophages. Said & Theodorescu propose that the aforementioned cytokine release generates a permissive lung inflammatory environment (68). Likewise, in mesotheliomas, versican downregulates macrophage M1 phenotype and decreases their ability to phagocytose tumor cells (69) contributing to the process of immune "escape."

Interestingly, monocytes cultured in versican-containing supernatants of colon cancer cells secrete pro-inflammatory cytokines, including IL-12, TNF α and ROS, whereas monocytes cultured in versican-free supernatants gathered from breast cancer tumors' culture exhibited a different profile of secreted cytokines. Thus, versican seems to have the ability to specifically direct the inflammatory monocyte response (19), conceivably correlated to the differential recruitment of lymphocytes, and differentiation into tumor-associated macrophages and dendritic cells (DC) (70).

DCs are proposed to engage tumor antigens and relocate to draining lymph nodes, where they trigger tumor-specific T cells (71). Importantly, versican derived from tumor ECM activates DCs' TLR2, resulting in the production of immunosuppressive IL-10 and DC dysfunction. This was associated with a TLR2induced increase of IL-6 and IL-10 cell-surface receptors and a strongly decreased cytokine concentration threshold is needed to trigger STAT3 (72, 73). Intact versican, thus, dampens DC activation (72, 73) and conceivably downstream Th and cytotoxic lymphocyte (CTL) differentiation. Likewise, intact versican secreted by macrophages was shown to have antiinflammatory properties in a mouse model of acute pulmonary inflammation (74). On the other hand, in myelomas, versican is proteolyticaly processed to the DAMP versikine, which induces the secretion of IL-1ß and IL-6 by human myeloma marrowderived macrophages (MAMs). Importantly, MAMs chiefly synthesized V1, the precursor to versikine, whereas stromal cells secreted the versican-degrading protease. This interplay is suggested to enhance the "T-cell inflammation," response and downregulate the "tolerogenic consequences of intact versican accumulation" contributing to tumor restraint (75).

Syndecans, Modulators of the Inflammatory IL-6/STAT3 Pathway

Syndecans (SDCs), cell membrane HS containing PGs (76), have also been implicated in tumor immunomodulation. SDC1defficient mouse model of colitis-related colon carcinoma exhibited higher susceptibleness to malignant transformation due to the enhanced topical release of IL-6, downstream activation of STAT3 and target genes with important roles in colonic tumorigenesis (77). Likewise, in inflammatory breast cancer, SDC1 was shown to upregulate the inflammatory IL-6/STAT3 pathway (78), a crucial part of tumor-stimulating signaling in epithelial-origin tumors (79). The well-described role of the enzyme heparanase, responsible for the cleavage of the HS chains may partly elucidate the role of the HS-bearing SDCs (80). Thus, due to the fact that HS regulates inflammatory processes at various checkpoints, e.g., including the segregation of inflammatory cytokines to the ECM, the configuration of leukocyte binding to the endothelial cells and ECM components, as well as the generation of responses respective to innate immunity by interacting with the TLRs, the reconfiguration of HS due to enzymatic heparanase affects the propagation of inflammatory events (80, 81).

Endocan, A Unique Plasma PG Blocks Leukocyte Trafficking

Endocan is an endothelial-derived soluble PG, initially identified in human umbilical vein endothelial cells (82). Interestingly, endothelium activated by inflammatory processes and/or tumor pathogenesis exhibits a strong augmentation in endocan expression correlated to poor patient prognosis (81, 83–85). Glycanated human endocan has been shown to strongly promote tumor growth (86) through its ability to bind to the integrin CD11a/CD18 (LFA-1) and thus to block leukocyte binding to endothelium and subsequent infiltration to tumor tissues (87); or through its promotion of growth factor actions (83, 88). On the other hand, the non-glycanated mouse or human endocan polypeptide was shown, in a murine model, to delay tumor expansion through the induction of pan-leukocytic infiltration of CD122+ expressing cells within tumor and stroma tissues (89).

The Intracellular PG, Serglycin, Is Implicated in Heamotological Malignancies

Serglycin, the only known intracellular PG is shown to be overexpressed in various tumor tissues and cell lines (90, 91). Importantly, serglycin is a key compound of secretory particles produced by CTL/NK cells, suggested to facilitate the safe storage of particle toxins, granzymes and perforin and affect CTL/NK cells' cytotoxic ability (92). Even though no evidence is available as such, these data implicate a plausible contribution of serglycin to processes of cancer-associated inflammation.

THERAPEUTIC IMPLICATIONS

The modulation of PGs' activities, as endogenous mediators of cancer-associated inflammation, is a novel approach in the field of cancer immunotherapy. Research efforts up to date have demonstrated the plausibility of this strategy as discussed below. Thus, the administration of the versican fragment characterized as DAMP, versikine, is suggested to "facilitate immune sensing of myeloma tumors and modulate the tolerogenic consequences of intact versican accumulation" which may facilitate tumor restraint and enhance T-cell-activating immunotherapies (75). Blocking the versican/CCL2/CCR2 signaling axis is indicated to purvey novel adjuvant strategies for detaining the emergence of clinical metastasis in bladder cancer patients (68). The non-glycanated mouse or human endocan polypeptide

restrains tumor growth by increasing leukocyte infiltration in vivo, enhancing de facto innate immunity response (89). Suppressing the shedding of SDC1 from intestinal epithelial cells decreases intestinal inflammation and putative malignant transformation by augmenting NF-kB, respective downstream signaling, and neutrophil transmigration in ulcerative colitis (93). Indeed, the modulation of SDC1 expression could facilitate immunesurveillance and the positive resolution of precancerous lesions. Biglycan is suggested to bridge innate and adaptive immunity through TLR2/4 downstream signaling due to its' ability to regulate neutrophil, macrophage, T and B lymphocyte, and dendritic cells activities (17, 18, 46-56). Thus, selective inhibition of biglycan-TLR2/TLR4 axis could be a novel therapeutic approach targeting at various checkpoints of the cancer immunobiology cycle. Modulations of PGs posttranscriptional modifications, including their respective sulfation pattern, seems to be a therapy option as inhibition of sulfatase-2 activity had cytotoxic and partial hepatoprotective activity in both in vivo and in vitro hepatocellular carcinoma models (94, 95). Indeed, a clinically relevant pattern of PG and GAG expression and structural modifications was recently determined

REFERENCES

- Aggarwal BB, Vijayalekshmi RV, Sung B. Targeting inflammatory pathways for prevention and therapy of cancer: short-term friend, long term foe. *Clin Cancer Res.* (2009) 15:425–30. doi: 10.1158/1078-0432.CCR-08-0149
- Grivennikov SI, Greten FR, Karin M. Immunity, inflammation, and cancer. Cell. (2010) 140:883–99. doi: 10.1016/j.cell.2010.01.025
- Konsoulova A. Chapter 5. Principles of cancer immunobiology and immunotherapy of solid tumors. In: Metodiev E, editor. *Immunopathology and Immunomodulation*, INTECH (2015).
- Nikitovic D, Corsini E, Kouretas D, Tsatsakis A, Tzanakaki G. ROS-major mediators of extracellular matrix remodeling during tumor progression. *Food* and Chemical Toxicology. (2013) 61:178–86. doi: 10.1016/j.fct.2013.06.013
- Nikitovic D, Berdiaki A, Spyridaki I, Krasanakis T, Tsatsakis A, Tzanakakis GN. Proteoglycans-biomarkers and targets in cancer therapy. *Front Endocrinol.* (2018) 9:69. doi: 10.3389/fendo.2018.00069
- Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. Cell. (2011) 144:646–7410. doi: 10.1016/j.cell.2011.02.013
- Iozzo RV, Schaefer L. Proteoglycan form and function: a comprehensive nomenclature of proteoglycans. *Matrix Biol.* (2015) 42:11–55. doi: 10.1016/j.matbio.2015.02.003
- Lindahl U, Couchman J, Kimata K, Esko JD. Chapter 17. Proteoglycans and sulfated glycosaminoglycans. In: Varki A, Cummings RD, Esko JD, Stanley P, Hart GW, Aebi M, et al. editors. *Essentials of Glycobiology [Internet]*. 3rd ed. Cold Spring Harbor, NY: Cold Spring Harbor Laboratory Press (2015–2017).
- Marchand M, Monnot C, Muller L, Germain S. Extracellular matrix scaffolding in angiogenesis and capillary homeostasis. *Semin. Cell Dev. Biol.* (2018) S1084–9521:30578–5. doi: 10.1016/j.semcdb.2018.08.007
- 10. Aggelidakis J, Berdiaki A, Nikitovic D, Papoutsidakis A, Papachristou DJ, Tsatsakis AM, et al. Biglycan regulates MG63 osteosarcoma cell growth through a LPR6/ β -catenin/IGFR-IR signaling Axis. *Front Oncol.* (2018) 8:470. doi: 10.3389/fonc.2018.00470
- Mytilinaiou M, Nikitovic D, Berdiaki A, Papoutsidakis A, Papachristou DJ, Tsatsakis A, et al. IGF-I regulates HT1080 fibrosarcoma cell migration through a syndecan-2/Erk/ezrin signaling axis. *Exp Cell Res.* (2017) 1:9–18. doi: 10.1016/j.yexcr.2017.09.035
- Nikitovic D, Aggelidakis J, Young MF, Iozzo RV, Karamanos NK, Tzanakakis GN. The biology of small leucine-rich proteoglycans in bone pathophysiology. *J Biol Chem.* (2012) 287:33926–33. doi: 10.1074/jbc.R112.379602

for PGs and GAG synthesizing enzymes in glioma and breast cancer, which might help in the development of personalized therapy (96).

Despite these advances, however, many issues need yet to be addressed for implementation to clinical practice (97). Furthermore, an understanding of tumor microenvironment biology and its PG component is necessary for therapy development.

AUTHOR CONTRIBUTIONS

DN designed the concept and completed the final editing of the manuscript. All authors contributed to writing of the manuscript. DN and MN prepared the figures. All authors read and approved the final version of the manuscript.

FUNDING

DN was supported by ELKE KA10038. MN was supported by PN 19.29.01.01, PN-III-P1-1.2-PCCDI-2017-0341/2018 and 7PFE/16.10.2018.

- Mytilinaiou M, Nikitovic D, Berdiaki A, Kostouras A, Papoutsidakis A, Tsatsakis AM, et al. Emerging roles of syndecan 2 in epithelial and mesenchymal cancer progression. *IUBMB Life.* (2017) 69:824–833. doi: 10.1002/iub.1678
- Nikitovic D, Papoutsidakis A, Karamanos NK, Tzanakakis GN. Lumican affects tumor cell functions, tumor-ECM interactions, angiogenesis and inflammatory response. *Matrix Biol.* (2014) 35:206–14. doi: 10.1016/j.matbio.2013.09.003
- Ricard-Blum S, Vallet SD. Matricryptins network with matricellular receptors at the surface of endothelial and tumor cells. *Front Pharmacol.* (2016) 7:11. doi: 10.3389/fphar.2016.00011
- Theocharis AD, Karamanos NK. Proteoglycans remodeling in cancer: Underlying molecular mechanisms. *Matrix Biol.* (2017) 75–6:220–59. doi: 10.1016/j.matbio.2017.10.008
- Schaefer L, Tredup C, Gubbiotti MA, Iozzo RV. Proteoglycan neofunctions: regulation of inflammation and autophagy in cancer biology. *FEBS J.* (2016) 284:10–26. doi: 10.1111/febs.13963
- Hsieh LT, Frey H, Nastase MV, Tredup C, Hoffmann A, Poluzzi C, et al. Bimodal role of NADPH oxidases in the regulation of biglycan-triggered IL-1β synthesis. *Matrix Biol.* (2016) 49:61–81. doi: 10.1016/j.matbio.2015. 12.005
- Wight TN, Kang I, Merrilees MJ. Versican and the control of inflammation. Matrix Biol. (2016) 35:152–61. doi: 10.1016/j.matbio.2014.01.015
- Dunn GP, Old LJ, Schreiber RD. The immunobiology of cancer immunosurveillance and immunoediting. *Immunity*. (2004) 21:137–48. doi: 10.1016/j.immuni.2004.07.017
- Karlou M, Tzelepi V, Efstathiou E. Therapeutic targeting of the prostate cancer microenvironment. Nat Rev Urol. (2010) 7:494-509. doi: 10.1038/nrurol.2010.134
- Vinay DS, Ryan EP, Pawelec G, Talib WH, Stagg J, Elkord E, et al. Immune evasion in cancer: mechanistic basis and therapeutic strategies. *Semin Cancer Biol.* (2015) 35(Suppl.) S185–98. doi: 10.1016/j.semcancer.2015.03.004
- Neagu M, Constantin C, Caruntu C, Dumitru C, Surcel M, Zurac S. Inflammation: A key process in skin tumorigenesis (review). Oncol Lett. (2019) 17:4068–84. doi: 10.3892/ol.2018.9735
- Mantovani A, Allavena P, Sica A, Balkwill F. Cancer-related inflammation. Nature. (2008) 454:436–44. doi: 10.1038/nature07205
- 25. Castle PE, Hillier SL, Rabe LK, Hildesheim A, Herrero R, Bratti MC, et al. An association of cervical inflammation with high-grade cervical neoplasia

in women infected with oncogenic human papillomavirus (HPV). Cancer Epidemiol Biomarkers Prev. (2001) 10:1021–7.

- Rothenberg SM, Ellisen LW. The molecular pathogenesis of head and neck squamous cell carcinoma. J Clin Investig. (2012) 122:1951–7. doi: 10.1172/JCI59889
- Arzumanyan A, Reis HM, Feitelson MA. Pathogenic mechanisms in HBVand HCV-associated hepatocellular carcinoma. *Nat Rev Cancer.* (2013) 13:123–35. doi: 10.1038/nrc3449
- Salama NR, Hartung ML, Muller A. Life in the human stomach: persistence strategies of the bacterial pathogen Helicobacter pylori. *Nat Rev Microbiol.* (2013) 11:385–99. doi: 10.1038/nrmicro3016
- Houghton AM. Mechanistic links between COPD and lung cancer. Nat Rev Cancer. (2013) 13:233–45. doi: 10.1038/nrc3477
- Meiliana A, Dewi NM, Wijaya A. The immunobiology of cancer: an update review. *Indones Biomed J.* (2017) 9:53–72. doi: 10.18585/inabj.v9i2.342
- Raval RR, Sharabi AB, Walker AJ, Drake CG, Sharma P. Tumor immunology and cancer immunotherapy: summary of the 2013 SITC primer. *J Immunother Cancer.* (2014) 2:14. doi: 10.1186/2051-1426-2-14
- Gajewski TF, Corrales L, Williams J, Horton B, Sivan A, Spranger S. Cancer immunotherapy targets based on understanding the T cell-inflamed versus non-T cell-inflamed tumor microenvironment. *Adv Exp Med Biol.* (2017) 1036:19–31. doi: 10.1007/978-3-319-67577-0_2
- Spranger S. Mechanisms of tumor escape in the context of the T-cell-inflamed and the non-T-cell-inflamed tumor microenvironment. *Int Immunol.* (2011) 28:383–91. doi: 10.1093/intimm/dxw014
- Pietras K, Ostman A. Hallmarks of cancer: interactions with the tumor stroma. Experim Cell Res. (2010) 316:1324–31. doi: 10.1016/j.yexcr.2010.02.045
- Schäfer M, Werner S. Cancer as an overhealing wound: an old hypothesis revisited. Nat Rev Mol Cell Biol. (2008) 9:628–38. doi: 10.1038/nrm2455
- Cardone A, Tolino A, Zarcone R, Borruto Caracciolo G, Tartaglia E. Prognostic value of desmoplastic reaction and lymphocytic infiltration in the management of breast cancer. *Panminerva Med.* (1997) 39:174–7.
- Avgustinova A, Iravani M, Robertson D, Fearns A, Gao Q, Klingbeil P, et al. Tumour cell-derived Wnt7a recruits and activates fibroblasts to promote tumour aggressiveness. *Nat Commun.* (2016) 7:10305. doi: 10.1038/ncomms10305
- Sung SY, Hsieh CL, Law A, Zhau HE, Pathak S, Multani AS, et al. Coevolution of prostate cancer and bone stroma in three-dimensional coculture: implications for cancer growth and metastasis. *Cancer Res.* (2008) 68:9996–10003. doi: 10.1158/0008-5472.CAN-08-2492
- Lu P, Weaver VM, Werb Z. The extracellular matrix: a dynamic niche in cancer progression. J Cell Biol. (2012) 196:395–406. doi: 10.1083/jcb.2011 02147
- Nikitovic D, Tzardi M, Berdiaki A, Tsatsakis A, Tzanakakis GN. Cancer Microenvironment and inflammation: role of hyaluronan. *Front Immun.* (2015) 6:169. doi: 10.3389/fimmu.2015.00169
- Neill T, Schaefer L, Iozzo RV. Decoding the matrix: Instructive roles of proteoglycan receptors. *Biochemistry*. (2015) 54:4583– 98. doi: 10.1021/acs.biochem.5b00653
- Hu L, Duan YT, Li JF, Su LP, Yan M, Zhu ZG, et al. Biglycan enhances gastric cancer invasion by activating FAK signaling pathway. *Oncotarget.* (2014) 5:1885–96. doi: 10.18632/oncotarget.1871
- Avellini C, Reni M, Mazzer M, Foltran L, Rossi D, Cereda S, et al. Biglycan expression and clinical outcome in patients with pancreatic adenocarcinoma. *Tumour Biol.* (2013) 34:131–7. doi: 10.1007/s13277-012-0520-2
- 44. Pan S, Cheng L, White JT, Lu W, Utleg AG, Yan X, et al. Quantitative proteomics analysis integrated with microarray data reveals that extracellular matrix proteins, catenins, and p53 binding protein 1 are important for chemotherapy response in ovarian cancers. *OMICS J Integr Biol.* (2009) 13:345–54. doi: 10.1089/omi.2009.0008
- Mikula M, Rubel T, Karczmarski J, Goryca K, Dadlez M, Ostrowski J. Integrating proteomic and transcriptomic high-throughput surveys for search of new biomarkers of colon tumors. *Funct Integr Genomics*. (2011) 11:215– 24. doi: 10.1007/s10142-010-0200-5
- 46. Schaefer L, Babelova A, Kiss E, Hausser HJ, Baliova M, Krzyzankova M, et al. The matrix component biglycan is proinflammatory which signals through the innate immunity Toll-like receptors 4 and 2 (TLR4, 2). *J Clin Invest*. (2005) 115:2223–33. doi: 10.1172/JCI23755

- Moreth K, Frey H, Hubo M, Zeng-Brouwers J, Nastase MV, Hsieh LT, et al. Biglycan-triggered TLR-2- and TLR-4-signaling exacerbates the pathophysiology of ischemic acute kidney injury. *Matrix Biol.* (2014) 35:143– 51. doi: 10.1016/j.matbio.2014.01.010
- Zeng-Brouwers J, Beckmann J, Nastase MV, Iozzo RV, Schaefer L. De novo expression of circulating biglycan evokes an innate inflammatory tissue response via MyD88/TRIF pathways. Matrix Biol. (2014) 35:132– 42. doi: 10.1016/j.matbio.2013.12.003
- Babelova A, Moreth K, Tsalastra-Greul W, Zeng-Brouwers J, Eickelberg O, Young MF, et al. Biglycan, a danger signal that activates the NLRP3 inflammasome via toll-like and P2X receptors. *J Biol Chem*. (2009) 284:24035– 42408. doi: 10.1074/jbc.M109.014266
- Schaefer L, Iozzo RV. Small leucine-rich proteoglycans, at the crossroad of cancer growth and inflammation. *Curr Opin Genet Dev.* (2012) 22:56– 7. doi: 10.1016/j.gde.2011.12.002
- Moreth K, Iozzo RV, Schaefer L. Small leucine-rich proteoglycans orchestrate receptor crosstalk during inflammation. *Cell Cycle.* (2012) 11:2084– 91. doi: 10.4161/cc.20316
- Dajon M, Iribarren K, Cremer I. Toll-like receptor stimulation in cancer: a pro- and anti-tumor double-edged sword. *Immunobiology*. (2016) 222:89– 100. doi: 10.1016/j.imbio.2016.06.009
- Farnebo L, Shahangian A, Lee Y, Shin JH, Scheeren FA, Sunwoo JB. Targeting Toll-like receptor 2 inhibits growth of head and neck squamous cell carcinoma. *Oncotarget*. (2015) 6:9897–907. doi: 10.18632/oncotarget.3393
- Roedig H, Nastase MV, Frey H, Moreth K, Zeng-Brouwers J, Poluzzi C, et al. Biglycan is a new high-affinity ligand for CD14 in macrophages. *Matrix Biol.* (2018) 17:4–22. doi: 10.1016/j.matbio.2018.05.006
- Nastase MV, Janicova A, Wygrecka M, Schaefer L. Signaling at the crossroads: matrix-derived proteoglycan and reactive oxygen species signaling. *Antioxid Redox Signal.* (2017) 27:855–73. doi: 10.1089/ars.2017.7165
- Moreth K, Brodbeck R, Babelova A, Gretz N, Spieker T, Zeng-Brouwers J, et al. The proteoglycan biglycan regulates expression of the B cell chemoattractant CXCL13 and aggravates murine lupus nephritis. *J Clin Invest.* (2010) 120:4251–72. doi: 10.1172/JCI42213
- Wu Y, Antony S, Meitzler JL, Doroshow JH. Molecular mechanisms underlying chronic inflammation-associated cancers. *Cancer Lett.* (2014) 34:164–73. doi: 10.1016/j.canlet.2013.08.014
- Gubbiotti MA, Vallet SD, Ricard-Blum S, Iozzo RV. Decorin interacting network: a comprehensive analysis of decorinbinding partners and their versatile functions. *Matrix Biol.* (2016) 55:7–21. doi: 10.1016/j.matbio.2016.09.009
- Merline R, Moreth K, Beckmann J, Nastase MV, Zeng-Brouwers J, Tralhão JG, et al. Signaling by the matrix proteoglycan decorin controls inflammation and cancer through PDCD4 and microRNA-21. *Sci Signal.* (2011) 4:75. doi: 10.1126/scisignal.2001868
- Koninger J, Giese NA, Bartel M, di Mola FF, Berberat PO, di Sebastiano P, et al. The ECM proteoglycan decorin links desmoplasia and inflammation in chronic pancreatitis. *J Clin Pathol.* (2006) 59:21–7. doi: 10.1136/jcp.2004.023135
- Buraschi S, Neill T, Owens RT, Iniguez LA, Purkins G, Vadigepalli R, et al. Decorin protein core affects the global gene expression profile of the tumor microenvironment in a triple-negative orthotopic breast carcinoma xenograft model. *PLoS ONE.* (2012) 7:e45559. doi: 10.1371/journal.pone.0045559
- Wu F, Vij N, Roberts L, Lopez-Briones S, Joyce S, Chakravarti S. A novel role of the lumican core protein in bacterial lipopolysaccharide-induced innate immune response. J Biol Chem. (2007) 282:6409–26417. doi: 10.1074/jbc.M702402200
- Frevert CW, Felgenhauer J, Wygrecka M, Nastase MVIII, Schaefer L. Dangerassociated molecular patterns derived from the extracellular matrix provide temporal control of innate immunity. *J Histochem Cytochem*. (2018) 66:213– 27. doi: 10.1369/0022155417740880
- Adair-Kirk TL, Senior RM. Fragments of extracellular matrix as mediators of inflammation. *Int J Biochem Cell Biol.* (2008) 40:1101–10. doi: 10.1016/j.biocel.2007.12.005
- Binder MJ, McCoombe S, Williams ED, McCulloch DR, Ward AC. The extracellular matrix in cancer progression: role of hyalectan proteoglycans and ADAMTS enzymes. *Cancer Lett.* (2017) 385:55–6. doi: 10.1016/j.canlet.2016.11.001

- Kim S, Takahashi H, Lin WW, Descargues P, Grivennikov S, Kim Y, et al. Carcinoma-produced factors activate myeloid cells through TLR2 to stimulate metastasis. *Nature*. (2009) 457:102–6. doi: 10.1038/nature 07623
- Said N, Sanchez-Carbayo M, Smith SC, Theodorescu D. RhoGDI2 suppresses lung metastasis in mice by reducing tumor versican expression and macrophage infiltration. *J Clin Investig.* (2012) 122:1503–18. doi: 10.1172/JCI61392
- Said N, Theodorescu D. RhoGDI2 suppresses bladder cancer metastasis via reduction of inflammation in the tumor microenvironment. *OncoImmunology*. (2012) 1:1175–17. doi: 10.4161/onci.20594
- Pappas AG, Magkouta S, Pateras IS, Skianis I, Moschos C, Vazakidou ME, et al. Versican modulates tumor-associated macrophage properties to stimulate mesothelioma growth. *Oncoimmunology*. (2018) 8:e1537427. doi: 10.1080/2162402X.2018.1537427
- Olingy CE, Dinh HQ, Hedrick CC. Monocyte heterogeneity and functions in cancer. *J Leukoc Biol.* (2019) doi: 10.1002/JLB.4RI0818-311R. [Epub ahead of print].
- 71. Finn OJ. Cancer immunology. N Engl J Med. (2008) 358:2704– 15. doi: 10.1056/NEJMra072739
- Tang M, Diao J, Cattral MS. Molecular mechanisms involved in dendritic cell dysfunction in cancer. *Cell Mol Life Sci.* (2017) 74:761–76. doi: 10.1007/s00018-016-2317-8
- Tang M, Diao J, Gu H, Khatri I, Zhao J, Cattral MS. Toll-like receptor 2 activation promotes tumor dendritic cell dysfunction by regulating IL-6 and IL-10 receptor signaling. *Cell Rep.* (2015) 13:2851– 64. doi: 10.1016/j.celrep.2015.11.053
- 74. Chang MY, Kang I, Gale MJr, Manicone AM, Kinsella MG, Braun KR, et al. Versican is produced by Trif- and type I interferon-dependent signaling in macrophages and contributes to fine control of innate immunity in lungs. *Am J Physiol Lung Cell Mol Physiol.* (2017) 313:L1069– 86. doi: 10.1152/ajplung.00353.2017
- 75. Hope C, Foulcer S, Jagodinsky J, Chen SX, Jensen JL, Patel S, et al. Immunoregulatory roles of versican proteolysis in the myeloma microenvironment. *Blood.* (2016) 128:680–5. doi: 10.1182/blood-2016-03-705780
- Afratis NA, Nikitovic D, Multhaupt HA, Theocharis AD, Couchman JR, Karamanos NK. Syndecans – key regulators of cell signaling and biological functions. *FEBS J.* (2017) 284:27–41. doi: 10.1111/febs.13940
- 77. Binder Gallimidi A, Nussbaum G, Hermano E, Weizman B, Meirovitz A, Vlodavsky I, et al. Elkin M. Syndecan-1 deficiency promotes tumor growth in a murine model of colitis-induced colon carcinoma. *PLoS ONE.* (2017) 12:e0174343. doi: 10.1371/journal.pone.0174343
- 78. Ibrahim SA, Gadalla R, El-Ghonaimy EA, Samir O, Mohamed HT, Hassan H, et al. Syndecan-1 is a novel molecular marker for triple negative inflammatory breast cancer and modulates the cancer stem cell phenotype via the IL-6/STAT3, Notch and EGFR signaling pathways. *Mol Cancer*. (2017) 16:57. doi: 10.1186/s12943-017-0621-z
- 79. Grivennikov S, Karin E, Terzic J, Mucida D, Yu GY, Vallabhapurapu S, et al. IL-6 and Stat3 are required for survival of intestinal epithelial cells and development of colitis-associated cancer. *Cancer Cell.* (2009) 15:103–13. doi: 10.1016/j.ccr.2009.01.001
- Goodall KJ, Poon IK, Phipps S, Hulett MD. Soluble heparan sulfate fragments generated by heparanase trigger the release of pro-inflammatory cytokines through TLR-4. *PLoS ONE.* (2014) 9:e109596. doi: 10.1371/journal.pone.0109596
- Vlodavsky I, Singh P, Boyango I, Gutter-Kapon L, Elkin M, Sanderson RD, et al. Heparanase: From basic research to therapeutic applications in cancer and inflammation. *Drug Resist Updates*. (2016) 29:54–75. doi: 10.1016/j.drup.2016. 10.001
- Lassalle P, Molet S, Janin A, Heyden JV, Tavernier J, Fiers W, et al. ESM-1 is a novel human endothelial cell-specific molecule expressed in lung and regulated by cytokines. J Biol Chem. (1996) 271:20458–64. doi: 10.1074/jbc.271.34.20458

- Sarrazin S, Maurage CA, Delmas D, Lassalle P, Delehedde M. Endocan as a biomarker of endothelial dysfunction in cancer. J Cancer Sci Ther. (2010) 2:47–52. doi: 10.4172/1948-5956.1000022
- Roudnicky F, Poyet C, Wild P, Krampitz S, Negrini F, Huggenberger R, et al. Endocan is upregulated on tumor vessels in invasive bladder cancer where it mediates VEGF-A-induced angiogenesis. *Cancer Res.* (2013) 73:1097– 106. doi: 10.1158/0008-5472.CAN-12-1855
- 85. Grigoriu BD, Depontieu F, Scherpereel A, Gourcerol D, Devos P, Ouatas T, et al. Endocan expression and relationship with survival in human non-small cell lung cancer. *Clin Cancer Res.* (2006) 12:4575– 82. doi: 10.1158/1078-0432.CCR-06-0185
- Kali A, Shetty KS. Endocan: a novel circulating proteoglycan. Indian J Pharmacol. (2014) 46:579–83. doi: 10.4103/0253-7613.144891
- Béchard D, Scherpereel A, Hammad H, Gentina T, Tsicopoulos A, Aumercier M, et al. Human endothelial-cell specific molecule-1 binds directly to the integrin CD11a/CD18 (LFA-1) and blocks binding to intercellular adhesion molecule-1. J Immunol. (2001) 167:3099–106. doi: 10.4049/jimmunol.167.6.3099
- Scherpereel A, Gentina T, Grigoriu B, Sénéchal S, Janin A, Tsicopoulos A, et al. Overexpression of endocan induces tumor formation. *Cancer Res.* (2003) 63:6084–9.
- Yassine H, De Freitasm Cairesm NM, Depontieum F, Scherpereel A, Awad A, Tsicopoulos A, et al. The non glycanated endocan polypeptide slows tumor growth by inducing stromal inflammatory reaction. *Oncotarget*. (2015) 6:2725–35. doi: 10.18632/oncotarget.2614
- Korpetinou A, Papachristou DJ, Lampropoulou A, Bouris P, Labropoulou VT, Noulas A, et al. Increased expression of serglycin in specific carcinomas and aggressive cancer cell lines. *Biomed Res Int.* (2015) 2015:690721. doi: 10.1155/2015/690721
- Fadnes B, Husebekk A, Svineng G, Rekdal Ø, Yanagishita M, Kolset SO, et al. The proteoglycan repertoire of lymphoid cells. *Glycoconj J.* (2012) 29:513–23. doi: 10.1007/s10719-012-9427-9
- Sutton VR, Brennan AJ, Ellis S, Danne J, Thia K, Jenkins MR, et al. Serglycin determines secretory granule repertoire and regulates natural killer cell and cytotoxic T lymphocyte cytotoxicity. *FEBS J.* (2016) 283:947– 61. doi: 10.1111/febs.13649
- Zhang Y, Wang Z, Liu J, Zhang S, Fei J, Li J, et al. Cell surfaceanchored syndecan-1 ameliorates intestinal inflammation and neutrophil transmigration in ulcerative colitis. J Cell Molec Med. (2017) 21:13– 25. doi: 10.1111/jcmm.12934
- Yang JD, Sun Z, Hu C, Lai J, Dove R, Nakamura I. Sulfatase 1 and sulfatase 2 in hepatocellular carcinoma: associated signaling pathways, tumor phenotypes, and survival. *Genes Chromo Cancer*. (2011) 50:122–35. doi: 10.1002/gcc.20838
- Alyoussef A, Al-Gayyar MMH. Cytotoxic and partial hepatoprotective activity of sodium ascorbate against hepatocellular carcinoma through inhibition of sulfatase-2 *in vivo* and *in vitro*. *Biomed Pharmacother*. (2018) 103:362– 72. doi: 10.1016/j.biopha.2018.04.060
- 96. Subbarayan K, Seliger B. Tumor-dependent effects of proteoglycans and various glycosaminoglycan synthesizing enzymes and sulfotransferases on patients' outcome. *Curr Cancer Drug Targets*. (2019) 19:210–21. doi: 10.2174/1568009618666180706165845
- 97. Karin M, Clevers H. Reparative inflammation takes charge of tissue regeneration. *Nature*. (2016) 529:307–15. doi: 10.1038/nature17039

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2019 Tzanakakis, Neagu, Tsatsakis and Nikitovic. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.