Mutual Information for Testing Gene-Environment Interaction

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Abstract

Despite current enthusiasm for investigation of gene-gene interactions and gene-environment interactions, the essential issue of how to define and detect gene-environment interactions remains unresolved. In this report, we define gene-environment interactions as a stochastic dependence in the context of the effects of the genetic and environmental risk factors on the cause of phenotypic variation among individuals. We use mutual information that is widely used in communication and complex system analysis to measure gene-environment interactions. We investigate how gene-environment interactions generate the large difference in the information measure of gene-environment interactions between the general population and a diseased population, which motives us to develop mutual information-based statistics for testing gene-environment interactions. We validated the null distribution and calculated the type 1 error rates for the mutual information-based statistics to test gene-environment interactions using extensive simulation studies. We found that the new test statistics were more powerful than the traditional logistic regression under several disease models. Finally, in order to further evaluate the performance of our new method, we applied the mutual information-based statistics to three real examples. Our results showed that P-values for the mutual information-based statistics were much smaller than that obtained by other approaches including logistic regression models.

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Introduction

Complex diseases are the consequence of the interplay of genetic and environmental factors. Development of disease is a dynamic process of gene-gene and gene-environment interactions within a complex biological system which is organized into complicated interacting networks [1]. Modern complex theory assumes that the complexity is attributed to the interactions among the components of the system, therefore, interaction has been considered as a sensible measure of complexity of the biological systems. The more interactions between the components, the more complex system. We argue that the interactions hold a key for dissecting the genetic structure of complex diseases. Ignoring gene-environment interactions will likely mask the detection of a genetic effect and may lead to inconsistent association results across studies [2,3].

Despite current enthusiasm for investigation of gene-environment interactions, published results that document these interactions in humans are limited, and the essential issue of how to define and detect gene-environment interactions remains unresolved. The concept of gene-environment interactions is often used, but rarely specified with precision [4]. Over the last three decades, epidemiologists have debated intensely about how to define and measure interaction in epidemiologic studies [5]. Many researchers indicated the importance of distinguishing biological interaction and statistical interaction [6-10]. Biological interaction between the gene and environment is often defined as the interdependent operation of genetic and environmental factors that cause diseases. In contrast, statistical interaction between the gene and environment is defined as the interdependence between the effects of genetic and environmental risk factors in the context of a statistical model. The effects of disease risk factors are often measured by relative risks and odds ratios. The classical definition of statistical interaction has the following limitations. First, both relative risks and odds ratios are mainly defined for binary variables. Their extensions to multiple categorical risk factors (for example, three genotypes and multiple categorical environments) are cumbersome. Second, statistical interactions are essentially model dependent. Linear models and generalized linear models (logistic regressions and log-linear models) of the genetic effects of the risk factors are often used to define statistical interactions. In the classical logistic regressions and log-linear models of the geneenvironment interactions, the genetic effects of the risk factors are decomposed into main effects and interaction effects (or product term) in the model. But, such decomposition may not reflect the true nonlinear interaction between the gene and environment. In addition, in these models, the major part of the true biological interactions between the gene and environment is often partitioned into the marginal effects. The remaining part of the geneenvironment interactions which is treated as a departure (or residual) from the logistic regression and log-lineal models is small and hard to detect. Third, the classical interaction models can hardly be applied to study interactions (including pair-wise and high-order interactions) among the components of the biological systems and their complexity.

To overcome the limitations of the classical definition of the statistical interaction, we propose a new definition of interaction that is based on interdependence among the risk factors causing disease. Interactions between genes and the environment can generally be defined as a stochastic dependence between genetic and environmental risk factors in causing phenotypic variation among individuals. This definition does not require specifying the statistical models of the risks, and is similar, although not exactly identical, to the definition of biological interaction. The concept of mutual information proposed by Shannon [11] can serve as a general measure of interaction (dependence) between two random variables [12-14]. An additional asset is that mutual information measures more than linear dependence [15-16]. As we will show in the methods section, mutual information between the gene and environment has a close relationship with the classical measures of the gene-environment interactions such as odds ratio and relative risk.

As Liu [17] pointed out, "the success of investigation of $G \times E$ interactions depends greatly on the selection of the optimal study design, the most accurate and precise assessment of genetic and environmental factors, and the most efficient statistical analysis". Developing efficient analytic methods for evaluation of the gene-environment interactions is critical to the investigation of gene-environment interactions [18].

Odds ratio calculations, logistic regression analysis, data mining and multifactor dimensionality reduction (MDR) are some of the existing methods available to evaluate the gene-environment interactions [19–30]. These methods have their merits, but also they have limitations. As an alternative to these widely used methods for testing gene and environment interactions, we propose mutual information-based methods to detect gene and environment interactions.

The main purpose of this report is to use information theory as a general framework for developing statistical methods to detect geneenvironment interactions. To accomplish this, we first developed a novel definition of gene-environment interactions. Then we studied how to use mutual information to measure gene-environment interactions. We investigated how gene-environment interactions generate the large difference in aforementioned measures between the general population and disease population. This provided the motivation to develop mutual information-based statistics for testing gene-environment interactions. Using extensive simulation studies, we validated the null distribution and type 1 error rates of the mutual information-based statistics for testing gene-environment interactions. To reveal the merit and limitation of the mutual informationbased statistics to detect gene-environment interactions, we compared their power for detecting gene-environment interactions with the logistic regression. We found that the new test statistics have higher power than the traditional logistic regression under several disease models. Finally, in order to further evaluate the performance of our new method, we applied the mutual information-based statistics to real data examples. Our results showed that P-values for the mutual information-based statistics were smaller than that obtained by other approaches including logistic regression models.

Methods

Information measure of the gene-environment interaction

Consider a disease susceptibility locus G and an environment E. The locus G has three genotypes coded as 0, 1, and 2. The environmental exposure is coded as E = 1, otherwise E is coded as 0. Let D be an indicator of disease. Mutual information measures dependence between two random variables. The mutual information between the gene and environment in the general population is defined as

$$I(G; E) = \sum_{i=0}^{2} \sum_{j=0}^{1} P(G=i, E=j) \log \frac{P(G=i, E=j)}{P(G=i)P(E=j)}.$$
 (1)

Information theory [11] shows that mutual information I(G; E) is equal to zero if and only if

$$P(G=i,E=j) = P(G=i)P(E=j), \quad (i=0,1,2; j=0,1)$$

i.e., gene and environment variables are independent.

The mutual information between the gene and environment in the disease population is given by

$$I(G; E|D) = \sum_{i=0}^{2} \sum_{j=0}^{1} P(G=i, E=j|D=1)$$

$$\log \frac{P(G=i, E=j|D=1)}{P(G=i|D=1)P(E=j|D=1)}$$
(2)

while Equation (2) can be reduced to

$$I(G; E|D) = \sum_{i=0}^{2} \sum_{j=0}^{1} P(G=i, E=j|D=1) \log \frac{P(G=i, E=j)}{P(G=i)P(E=j)} + \sum_{i=0}^{2} \sum_{j=0}^{1} P(G=i, E=j|D=1)$$
(3)
$$\log \frac{P(D=1|G=i, E=j)/P_D}{\frac{P(D=1|G=i)}{P_D} \frac{P(D=1|E=j)}{P_D}}$$

where $P_D = P(D=1)$ is the prevalence of the disease.

Equation (3) shows that mutual information I(G; E|D) has two components. The first term in equation (3) is due to the dependence between the gene and environment in the general population. The second term in equation (3) is due to interaction. Thus, we define information measure of the interaction between the gene and environment as

$$I_{GE} = \sum_{i=0}^{2} \sum_{j=0}^{1} P(G=i, E=j | D=1)$$

$$\log \frac{P(D=1 | G=i, E=j) / P_D}{\frac{P(D=1 | G=i)}{P_D} \frac{P(D=1 | E=j)}{P_D}}$$
(4)

which implies that $I_{GE} = 0$ if and only if

$$\frac{P(D=1|G=i,E=j)}{P_D} =$$

$$\frac{P(D=1|G=i)}{P_D} \frac{P(D=1|E=j)}{P_D} \quad (i=0,1,2,j=0,1).$$
(5)

Information measure of interaction has two remarkable features. First, it is defined in terms of penetrance and hence related to the cause of the disease. Second, the interaction is measured by the interdependent operation of the gene and environment in causing disease. Absence of the gene and environment interaction indicates that equation (5) should hold.

If we assume that the gene and environment variables in the general population are independent, then

$$I(G; E|D) = I_{GE}.$$

In this case, the mutual information between the gene and environment in the disease population is equal to the information measure of the interaction between gene and environment. This provides an easy way to calculate the information measure of gene-environment interactions.

To gain understanding of the information measure of the gene and environment interaction, we studied several special cases.

Case 1: G is not the disease locus. If we assume that G is only a marker and will not cause disease, then we have

$$P(D=1|G=i,E=j) = P(D=1|E=j)$$
 and $P(D=1|G=i) = P_D$

which implies that

$$\frac{\frac{P(D=1|G=i,E=j)/P_D}{P_D}=1}{\frac{P(D=1|G=i)}{P_D}\frac{P(D=1|E=j)}{P_D}}$$

Thus, we obtain $I_{GE} = 0$. In other words, if the locus G is a marker, there is no interaction between the locus G and environment. The interaction measure I_{GE} between the marker and environment should be equal to zero. Hence, our information measure of the gene-environment interactions correctly characterizes the marker case.

Case 2: Environmental exposure will not cause disease. If the environmental exposure will not cause disease, there will be no interaction between the gene and environment. We expect that the information measure of gene and environment interaction should be equal to zero. Indeed, by the same argument as provided in case 1, we can show this.

Test statistics

In the previous section, we show that the information measure of the gene-environment interactions is related to the dependency of the gene and environment variables in the disease population. The interaction can be detected by testing the independence of the gene and environment. Before defining the test statistic, we introduce the following notations. Let

$$f_{ij} = P(G = i, E = j) \log \frac{P(G = i, E = j)}{P(G = i)P(E = j)} \quad (i = 0, 1, 2, j = 0, 1)$$

and $f_{D_{ij}} = P(G=i,E=j|D=1)\log \frac{P(G=i,E=j|D=1)}{P(G=i|D=1)P(E=j|D=1)}$ (i=0,1,2,j=0,1). Let $f = [f_{11},f_{12},f_{21},f_{22},f_{31}]^T$ and $f_D = [f_{D_{11}},f_{D_{12}},f_{D_{21}},f_{D_{22}},f_{D_{31}},f_{D_{32}}]^T P_{ij} = P(G=i,E=j)$ and $P_{D_{ij}} = P(G=i,E=j|D=1)$. Define

$$P = [P_{11}, P_{12}, P_{21}, P_{22}, P_{31}, P_{32}]^T \text{ and}$$
$$P_D = [P_{D_{11}}, P_{D_{12}}, P_{D_{21}}, P_{D_{22}}, P_{D_{31}}, P_{D_{32}}]^T.$$

The joint probabilities of the gene and environment variables in both the general population and disease population follow multinomial distributions with the following covariance matrices.

$$\Sigma = diag(P) - PP^T$$
 and $\Sigma_D = diag(P_D) - P_D P_D^T$

Let the Jacobean matrices of f and f_D with respect to P and P_D be $B = \left(\frac{\partial f_D}{\partial P_D^T}\right)$ and $C = \left(\frac{\partial f}{\partial P_D^T}\right)$, respectively. It is easy to see that

$$\frac{\partial f_{ij}}{\partial P_{ij}} = \log \frac{P_{ij}}{P_{i.}P_{.j}} - \frac{P_{ij}}{P_{i.}} - \frac{P_{ij}}{P_{.j}} + 1, \frac{\partial f_{ij}}{\partial P_{il}} = -\frac{P_{ij}}{P_{i.}},$$
$$\frac{\partial f_{ij}}{\partial P_{kj}} = -\frac{P_{ij}}{P_{.j}}, \frac{\partial f_{ij}}{\partial P_{kl}} = 0$$
$$_{(k \neq i)}$$

where $P_{i.} = \sum_{j=0}^{1} P_{ij}$, and $P_{.j} = \sum_{i=0}^{2} P_{ij}$. The partial derivatives of the function $f_{D_{ij}}$ with respect to $P_{D_{kl}}$ can be similarly defined. Let n_A be the number of sampled individuals in the cases and n_G be the number of sampled individuals in the controls. Define

$$\Lambda = \frac{B\Sigma_D B^T}{n_A} + \frac{C\Sigma C^T}{n_G}.$$

The statistic for testing the gene-environment interactions is then defined as

$$T_{GE} = \left(\hat{f}_D - \hat{f}\right)^T \hat{\Lambda}^- \left(\hat{f}_D - \hat{f}\right)$$
(12)

where \hat{f} , \hat{f}_D , and $\hat{\Lambda}$ are the estimators of f, f_D , and Λ . $\hat{\Lambda}^-$ is a generalized inverse of the matrix $\hat{\Lambda}$

When the sample size is sufficiently large enough to ensure application of the large sample theory, the test statistic T_{GE} is asymptotically distributed as a central $\chi^2_{(2)}$ distribution under the null hypothesis of the no gene-environment interactions, if we assume that the gene and environment variables in the general population are independent (Appendix S1).

We can also develop a statistic for testing interaction between each genotype and environment. For example, for genotype G=i, let

$$f_{i} = \begin{bmatrix} f_{i1} \\ f_{i2} \end{bmatrix} \text{ and } f_{D_{i}} = \begin{bmatrix} f_{D_{i1}} \\ f_{D_{i2}} \end{bmatrix},$$
$$C_{i} = \begin{bmatrix} \frac{\partial f_{i}}{\partial P^{T}} \end{bmatrix}, B_{i} = \begin{bmatrix} \frac{\partial f_{D_{i}}}{\partial P^{T}_{D}} \end{bmatrix}, \Lambda_{i} = \frac{B_{i} \Sigma_{D} B_{i}^{T}}{n_{A}} + \frac{C_{i} \Sigma C_{i}^{T}}{n_{G}} (i = 1, 2, 3)$$

then, the statistic for testing interaction between the genotype G = i and environment is defined as

$$T_{G_{i}E} = \left(\hat{f}_{D_{i}} - \hat{f}_{i}\right)^{T} \hat{\Lambda}_{i}^{-} \left(\hat{f}_{D_{i}} - \hat{f}_{i}\right).$$
(13)

Under the null hypothesis of no interaction between the genotype G=i and the environment the statistic T_{G_iE} is asymptotically distributed as a central $\chi^2_{(1)}$ distribution.

Results

Null distribution of test statistics

In the previous section we stated that the test statistic T_{GE} and $T_{G,E}$ under the null hypothesis are asymptotically distributed as a

central $\chi^2_{(2)}$ distribution and a central $\chi^2_{(1)}$ distribution, respectively, if we assume that the gene and environment variables are independent in the general population. To validate this statement we performed a series of simulation studies. The computer program SNaP [31] was used to generate the genotype data of the individuals and MATLAB was used to randomly generate the environment data of the individuals. Individuals (n = 100,000) with independent genotype and environmental exposure where the frequencies of two alleles at the locus were equal, and the frequency of the environmental exposure was equal to 0.2 (P(E=1)=0.2) were generated and equally divided into cases and controls. A total of 20,000 simulations were repeated. We plot Figures 1–4 showing the histograms of the test statistics T_{GE} and T_{G_iE} for testing the interaction between the gene and environment, with sample sizes $n_A = n_G = 400$, where n_A and n_G are the number of sampled individuals in the cases and controls. Figures 1-4 show that the null distributions of the test statistics $T_{GE}, T_{G_1E}, T_{G_2E}$ and T_{G_3E} are similar to the theoretical central $\chi^2_{(2)}$ and $\chi^2_{(1)}$ distributions, respectively. Type I error rates were calculated by random sampling 200–

Type I error rates were calculated by random sampling 200– 1,000 individuals from each of the cases and controls. In Tables 1 and 2 we listed type I error rates for T_{GE} and T_{GiE} , assuming $OR_g = 1$ and $OR_e = 1$. In Table 3 we listed type I error rates for T_{GE} , assuming $OR_g = 2$ and $OR_e = 2$ (For T_{GiE} , in case of $OR_g = 2$ and $OR_e = 2$ we can obtain the similar results (data not shown). Tables 1–3 demonstrated that the estimated Type I error rates for the statistics T_{GE} and T_{GiE} to test the gene and environment interactions were not appreciably different from the nominal levels $\alpha = 0.05$, $\alpha = 0.01$ and $\alpha = 0.001$, which were independent of the gene and environment odds ratios OR_g and OR_e .

Power evaluation

To evaluate the performance of the mutual information-based statistic for testing gene-environment interactions, we compared its power to that of the logistic model. The computer program SNaP [31] was used to generate the genotype data of the sampled individuals and MATLAB was used to randomly generate the environmental data of the sampled individuals. A population of 500,000 individuals with independent genotype and environmental exposure where the minor allele frequency (MAF) at the locus



Figure 1. Null Distribution of the statistic T_{GE} for 400 cases and controls respectively. doi:10.1371/journal.pone.0004578.g001



Figure 2. Null Distribution of the statistic T_{G_1E} for 400 cases and controls respectively. doi:10.1371/journal.pone.0004578.g002

were equal to 0.3 and the frequency of the environmental exposure was equal to 0.2 (P(E=1)=0.2) was generated. The model of the disease with the gene and environment interaction was defined by the penetrance. Gene-environment interactions effects were simulated with penetrance functions as given in Appendix S2. We assume the prevalence of the disease P(D=1)=0.01.

We consider two cases: (1) genetic and environmental odds ratios: $OR_g = 1$ and $OR_e = 1$, and (2) $OR_g > 1$ and $OR_e > 1$. In case (1), definition of the absence of the gene-environment interactions by information measure and gene-environment odds ratio in the logistic regression model is equivalent. In case (2), the information measure of interaction covers more situations which are interacted under the definition of information measure, but not interacted under definition of logistic models.

With this disease model, we randomly generated a disease population with 10,000 affected individuals and a general population with 10,000 unaffected individuals from the population



Figure 3. Null Distribution of the statistic T_{G_2E} for 400 cases and controls respectively. doi:10.1371/journal.pone.0004578.g003



Figure 4. Null Distribution of the statistic T_{G_3E} for 400 cases and controls respectively. doi:10.1371/journal.pone.0004578.g004

of 500,000 individuals. We then randomly sampled 500 individuals (cases) from the disease population and 500 individuals (controls) from the general population. We repeated 20,000 simulations. We presented six panels of Figures to compare the power of the proposed mutual information-based statistic and logistic regression models. Power calculation of logistic regression is based on the model $P(D=1|G,E) = \frac{e^{\alpha + \beta_g G + \beta_e E + \beta_{ge} G E}}{1 + e^{\alpha + \beta_g G + \beta_e E + \beta_{ge} G E}}$. Figures 5–7 plot the power of the test statistic T_{GE} and logistic regression to detect the gene-environment interactions in case (1) $(OR_g = 1 \text{ and } OR_e = 1)$ as a function of the gene-environment interactions odds ratios under the significance level $\alpha = 0.05$ for sample sizes 300, 400 and 500, respectively. Figures 8-10 plot the power of the test statistic T_{GE} and logistic regression to detect the gene-environment interactions in case (2) $(OR_g = 2 \text{ and } OR_e = 2)$ as a function of the gene-environment interactions odds ratios under the significance level $\alpha = 0.05$ for sample sizes 300, 400 and 500, respectively. These figures showed that the power of the mutual information-based statistic is much higher than that of the logistic regression even if in case (1) where the definition of absence of the gene-environment interactions by both the information measure and odds ratio in the logistic regression is equivalent. We also find that the difference in the power between the mutual

Table 1. Type 1 error rates for the test statistic T_{GE} to test gene-environment interaction, assuming $OR_g = 1$ and $OR_e = 1$.

Sample size	Nominal levels						
	$\alpha = 0.05$	$\alpha = 0.01$	$\alpha = 0.001$				
200	0.04641	0.00845	0.00076				
300	0.04618	0.00871	0.00089				
400	0.05033	0.00964	0.00098				
500	0.04811	0.00902	0.00082				
600	0.05012	0.01002	0.00082				
700	0.04991	0.00948	0.00096				
800	0.04804	0.00953	0.00098				
900	0.04737	0.00840	0.00088				
1000	0.04926	0.00979	0.00107				
	0.0-920	0.00979	0.00107				

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information-based statistic and the logistic regression model became larger as the significance level increases (data are not shown).

Application to real data example

To further evaluate its performance for testing gene-environment interactions, the mutual information-based statistics T_{GE} and T_{G_iE} were applied to real data examples. The first example studied the interaction between the polymorphism of the gene excision repair cross-complementing group 2 (ERCC2) and smoking exposure in lung cancer [32], where two ERCC2 polymorphisms Asp312Asn and Lys751Gln were typed in 1,092 Caucasian lung cancer patients and 1,240 spouse and friend controls collected at Massachusetts General Hospital. Both ERCC2 polymorphisms in the controls were in Hardy-Weinberg equilibrium. Smoking exposure was classified into four categories: non smoking, mild smoking, moderate smoking and heavy smoking. For simplicity of comparison, we performed only crude analysis. In other words, analysis was performed only for the raw data that were not adjusted for age and gender. Both the mutual information-based statistics and logistic regression analysis were used to test interaction between the polymorphism of ERCC2 and smoking in lung cancer. The results were summarized in Table 4. In

Table 2. Type 1 error rates for the test statistic	T_{G_iE} to test gene-e	environment interaction,	assuming $OR_g = 1$	and $OR_e = 1$.
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Sample size	$T_{G_{l}E}$			T_{G_2E}	T_{G_2E}			T_{G_3E}		
	$\alpha = 0.05$	$\alpha = 0.01$	$\alpha = 0.001$	$\alpha = 0.05$	$\alpha = 0.01$	$\alpha = 0.001$	$\alpha = 0.05$	$\alpha = 0.01$	$\alpha = 0.001$	
200	0.0495	0.0097	0.0010	0.0507	0.0104	0.0013	0.0486	0.0093	0.0009	
300	0.0482	0.0094	0.0008	0.0503	0.0100	0.0010	0.0473	0.0094	0.0009	
400	0.0494	0.0092	0.0009	0.0490	0.0103	0.0011	0.0493	0.0098	0.0010	
500	0.0491	0.0089	0.0008	0.0493	0.0105	0.0014	0.0479	0.0095	0.0010	
600	0.0480	0.0096	0.0010	0.0498	0.0098	0.0011	0.0494	0.0096	0.0010	
700	0.0500	0.0100	0.0012	0.0492	0.0095	0.0008	0.0484	0.0102	0.0009	
800	0.0494	0.0097	0.0011	0.0494	0.0096	0.0010	0.0474	0.0090	0.0008	
900	0.0489	0.0092	0.0008	0.0497	0.0103	0.0013	0.0494	0.0095	0.0009	
1000	0.0482	0.0095	0.0009	0.0506	0.0108	0.0013	0.0488	0.0090	0.0007	

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Table 3. Type 1 error rates for the test statistic T_{GE} to test gene-environment interaction, assuming $OR_g = 2$ and $OR_e = 2$.

	Nominal levels							
Sample size								
	$\alpha = 0.05$	$\alpha = 0.01$	$\alpha = 0.001$					
300	0.0513	0.0102	0.0012					
400	0.0473	0.0096	0.0007					
500	0.0470	0.0087	0.0011					
600	0.0482	0.0100	0.0008					
700	0.0513	0.0102	0.0015					
800	0.0479	0.0100	0.0011					
900	0.0493	0.0089	0.0005					
1000	0.0494	0.0102	0.0010					

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general, the logistic regression will not be used to test interaction between a single genotype and environment, thus there was no pvalue to test interaction between the single genotype and environment for logistic regression in Table 4. Two features emerge from Table 4. First, in general, the p-values of the global test statistic T_{GE} were smaller than that of the T_{GiE} for testing interaction between the particular genotype (single genotype) and environment. Second, in most cases, the p-values of the mutual information-based global test statistic T_{GE} were smaller than that of the logistic regression analysis. ERCC2 is a major DNA repair gene. DNA repair genes play a key role in protecting the genome from damage caused by smoking [32].

The second example is to study the interaction between the gene SULT1A1 and smoking/alcohol consumption for squamous cell carcinoma of the oesophagus [33]. The gene SULT1A1 catalyses sulfation that is related to the metabolism of a broad range of compounds such as phenolic xenobiotics, hydroxylated aromatic amines and drugs. The gene SULT1A1 is suspected to play a role in oesophageal cancer (OC). We applied the developed mutual information-based statistics and logistic regression to this dataset to test for gene-environment interactions in causing OC. The data in Table 5 were from Dandara's Table 4 [33] for the Mixed Ancestry South African group. The P-values in Dandara's Table 4 were obtained by the statistic based on odds ratios which tested for both the gene and environment interaction effects and the genetic effect. Thus, instead of using the P-values provided by Dandara, we used logistic regression to recalculate the P-values to test for interaction between the gene SULT1A1 and smoking or/ and alcohol consumption using data from Table 4 in Dandara [33]. The P-values of both mutual information-based statistics and logistic regressions were listed in Table 5. We can see that using the mutual information-based statistics we detected the interaction between the gene SULT1A1 and smoking, or the combination of smoking and alcohol consumption in causing OC in the Mixed Ancestry South African group, however logistic regression analysis failed to make a similar detection. The mutual information-based statistic also needs much less time than logistic regression analysis. For this example, if we use Intel Pentium(R) (D CPU 2.66 GHz×2.66 GHz, 2G memory, Windows XP) the computation time for the mutual information-based statistic and logistic



Figure 5. Power of the statistic *T_i* and logistic regression analysis for 300 cases and controls respectively. doi:10.1371/journal.pone.0004578.g005



Figure 6. Power of the statistic T_l and logistic regression analysis for 400 cases and controls respectively. doi:10.1371/journal.pone.0004578.g006



Figure 7. Power of the statistic *T_I* and logistic regression analysis for 500 cases and controls respectively. doi:10.1371/journal.pone.0004578.g007

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Figure 8. Power of the statistic T_I and logistic regression analysis for sample size 300, $OR_G = 2$ and $OR_E = 2$. doi:10.1371/journal.pone.0004578.g008



Figure 9. Power of the statistic T_1 and logistic regression analysis for sample size 400, $OR_G = 2$ and $OR_E = 2$. doi:10.1371/journal.pone.0004578.g009

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Figure 10. Power of the statistic T_I and logistic regression analysis for sample size 500, $OR_G = 2$ and $OR_E = 2$. doi:10.1371/journal.pone.0004578.g010

regression analysis was about 2.81×10^{-4} seconds and 1.59×10^{-2} seconds, respectively.

The third example is case-control study of interaction between smoking and HLA-DR SE (shared epitope) gene in the development of anticitrulline antibody-positive rheumatoid arthritis (RA) in the Swedish Epidemiological Investigation of Rheumatoid Arthritis (EIRA) study [34,35]. The major environmental risk factor and genetic risk factor are smoking and HLA-DA shared epitope (SE) gene, respectively. We analyzed data from Klareskog [34] which consisted of 827 RA patients and 1216 controls and from Kallberg [35] which consisted of 1883 RA patients and 1589 controls. Clearly, the second study [35] is the extension of the first study [34]. Both the mutual information-based statistic and logistic regression were applied to the dataset to test for interaction between the smoking and HLA-DR SE genes in the development of anticitrulline antibody-positive RA. The results were summarized in Table 6. They confirmed the recently pronounced interaction between smoking and the HLA-DR SE gene in the development of RA[34,36–37]. We also see that as the sample size increased in the second dataset, the P-values became

Table 4. Comparison of p-values for the mutual information-based statistics and logistic regression to the interaction between ERCC2 polymorphisms and smoking in lung cancer.

Genotype	Smoking									
	Mild		Moderate		Heavy	Heavy				
	P-values		P-values		P-values					
	T_{GE} or T_{G_iE}	Logistic regression	T_{GE} or T_{G_iE}	Logistic regression	T_{GE} or T_{G_iE}	Logistic regression				
Asp312Asn	0.0028	0.0151	5.70 E-4	0.0114	<10 ⁻¹⁰	2.53E-05				
Asp/Asp	0.0679		0.3447		0.00051					
Asp/Asn	0.1116		0.2462		0.4735					
Asn/Asn	0.0082		0.0094		0.0036					
Lys751Gln	0.0535	0.1611	1.24E-08	5.24E-01	<10 ⁻¹⁰	0.00197				
Lys/Lys	0.2010		0.0872		0.00095					
Lys/Gln	0.6391		0.3875		0.2417					
Gln/Gln	0.2364		0.4702		0.0399					

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Table 5. Comparison of p-values for the mutual information-
based statistics and logistic regression to the interaction
between the gene SULT1A1 and smoking (alcohol
consumption) in the Mixed Ancestry South African group.

-								
SULT1A1 genotype Patients		ts	Controls		P-value	P-values		
					T_{GE} or T_{G_iE}	Logistic regression		
Tobacco smokii	ng							
	no	yes	no	yes	0.0194	0.5196		
SULT1A1*1/*1	3	45	15	37	0.0536			
SULT1A1*1/*2	1	16	2	27	0.0096			
SULT1A1*2/*2	2	27	3	10	0.9152			
Alcohol consum	nption							
	no	yes	no	yes	0.0863	0.1847		
SULT1A1*1/*1	12	36	31	21	0.2443			
SULT1A1*1/*2	2	15	13	16	0.4105			
SULT1A1*2/*2	9	20	5	8	0.1648			
Both smoking a	and alcoh	ol consui	nption					
	no	yes	no	yes	0.0017	0.1902		
SULT1A1*1/*1	2	35	15	21	0.0082			
SULT1A1*1/*2	1	15	2	16	0.0124			
SULT1A1*2/*2	2	20	2	7	0.4310			

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much smaller (from 0.000925 to $<10^{-10}$). The results also again showed that the P-values of the mutual information-based statistics are usually smaller than that of the logistic regression and that the P-values of the global test statistic T_{GE} using all information at the locus are in general smaller than that of the test statistic $T_{G/E}$.

Discussion

Over the last three decades, epidemiologists have debated intensely about how to define and measure interaction in epidemiologic studies [5]. The distinction between biological interaction and statistical interaction becomes an important issue [6,39]. Biological interaction is often defined as interdependent operation of genetic and environmental factors that cause diseases. In other words, biological interaction means that joint presence of the genetic and environmental factors is the necessary condition for causing disease.

Due to the complexity of the development of the diseases, as Rothman [38] pointed out, there is no way to directly observe biological interactions. Biological interactions are often indirectly inferred. Our aim is to estimate the magnitude of the biological interaction as accurate as possible and develop efficient statistics to detect biological interactions. The purpose of this report is to address several issues for achieving this goal.

The first issue is how to define biological interaction mathematically. The major challenge is to come up with a definition that is mathematically explicit. In this report, we chose to use the classical concept of conditional probability to define biological interaction. A key component to biological interaction is the dependence of developing disease with the presence of both genetic and environmental factors. Therefore, the conditional dependence of the genetic factor on the environmental factor in causing disease is a natural expression for biological interaction. This mathematical definition is an alternative to the definition of biological interaction as a departure from additivity [38,39]. With this definition, interaction has a broader meaning and divergent statistical and computational tools available for further analysis.

The second issue we addressed is how to measure geneenvironment interactions. Mutual information is widely used in communication systems and complex adaptive systems analysis as a general measure of stochastic dependence between two random variables. In this report, we extended mutual information to measure gene-environment interactions. Widely used measures of interaction include relative risks or odds ratios which were originally defined for binary data. As a consequence, we often code genetic and environmental factors as binary variables for calculation of relative risks and odds ratios even if the genetic and environmental factors take multiple values or even continuous values. Mutual information can be defined for genetic and environmental factors with multiple values (or even continuous values, but not discussed here). Therefore, mutual information can cover broader cases than the relative risks and odds ratios.

The third issue addressed how to develop efficient statistics to detect gene-environment interactions. Despite current enthusiasm for investigation of interactions between the gene and environment, the essential issue of how to detect gene-environment interactions remains unresolved. Developing efficient analytical methods for evaluation of the gene-environment interactions is central to the investigation of gene-environment interactions [18]. Logistic regression is predominantly used to test for gene-environment interactions in epidemiology [38]. It depends on how to decompose the genetic effect. Most researchers use logistic regressions to model odds as the additive combination of main effects of a single-locus and the environment, and a residual term. The residual term in the model is defined as a statistical interaction between the gene and environment. As a consequence, the major part of functional (or biological) gene-environment interactions may be included in the main effects. The remaining part of the functional gene-environment interactions which is treated as a residual term in the mathematical model is small and hard to detect.

In this report, we presented mutual information-based statistics to detect gene-environment interactions. Through extensive simulation studies, we showed that the null distribution of the mutual information-based statistics was close to a central χ^2 distribution. We also calculated type 1 error rates of the mutual informationbased statistic by simulation. Our results showed that type 1 error rates were close to the nominal significance levels. We also investigated the power of the new statistic to detect the geneenvironment interactions by analytical methods. It showed that the mutual information-based test statistics have a much higher power in detecting the interaction than logistic regression methods even when $OR_g = 1$ and $OR_e = 1$ where definition of absence of interaction by both the information measure and odds ratio measure in the logistic regression are equivalent. To further evaluate their performance to detect the gene-environment interactions, the proposed mutual information-based statistics were applied to three published data sets. Our results showed that, in many cases, P-values of the mutual information-based statistics were much smaller than the results of the logistic regression analysis.

Since the computation time for the mutual information-based statistic is small, it is feasible to perform the genome-wide geneenvironment interaction analysis using PC machines. As we reported in the previous section that the computation time for the mutual information-based statistic to test one interaction between the gene and environment (94 cases and 94 controls) was only 2.81×10^{-4} sec, the total time required for testing the geneenvronment interaction for 1,000.000 SNPs and thousands of cases and controls will be about one hour. **Table 6.** Comparison of p-values for the mutual information-based statistics and logistic regression to the interaction between smoking and HLA-DR SE genes in the development of anticitrulline antibody-positive RA.

Sex, anti-CCP status and	.		Control		Declara	
HLA-DR SE genes	Case		Control		P-values	
	Never smoked	Ever smoked	Never smoked	Ever smoked	T_{GE} or T_{G_iE}	Logistic regression
The data were from Klareskog [[34]					
Male and Female						
Anti-CCP ⁺					9.25E-04	0.0198
No SE	20	58	87	184	0.01490	
Single SE	72	192	104	146	0.7090	
Double SE	36	126	31	31	0.03250	
Anti-CCP ⁻					0.2245	0.1989
No SE	65	84	87	184	0.2037	
Single SE	64	76	104	146	0.4170	
Double SE	18	18	31	31	0.4585	
Female						
Anti-CCP ⁺					0.2180	0.1378
No SE	18	41	74	115	0.4437	
Single SE	58	130	75	109	0.4989	
Double SE	30	89	25	25	0.08492	
Anti-CCP ⁻					0.3128	0.8859
No SE	50	62	74	115	0.4577	
Single SE	45	52	75	109	0.9805	
Double SE	15	11	25	25	0.8092	
Male						
Anti-CCP ⁺					6.72E-10	0.1172
No SE	2	17	13	69	2.55E-09	
Single SE	14	63	29	37	0.0574	
Double SE	6	37	6	6	0.2240	
Anti-CCP ⁻					0.0273	0.01472
No SE	15	24	13	69	0.0244	
Single SE	19	24	29	37	0.1519	
Double SE	3	7	6	6	0.1423	
Date were from Kallberg [35]						
Anti-CCP ⁺					<10 ⁻¹⁰	0.0059
No SE	35	71	137	242	0.0240	
Single SE	105	270	138	198	0.6392	
Double SE	61	179	39	39	0.0455	
Anti-CCP ⁻					0.3844	0.2979
No SE	86	115	140	242	0.2795	
Single SE	87	123	138	198	0.3946	
Double SE	25	26	39	39	0.6170	

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Although the preliminary results are appealing, the mutual information-based statistics for detection of gene-environment interactions also suffer from several limitations. First, they require an assumption that the genetic and environmental variables in the general population are independent. Deviation from independent assumption will affect the false positive rates. Second, they need to calculate the generalized inverse of the singular covariance matrix, which may lead to instability of numerical calculations. Third, in this report, we only compared the power of the mutual informationbased statistic with that of the logistic regression. A comparison with other methods including methods based on defining interaction as a departure from additive effects is in progress.

Gene-environment interactions are an important, but complex concept. There are a number of ways to define gene-environment interactions. How the definition of gene-environment interactions in population level reflects their biochemical or physiological interaction is still a mystery. We hope that this work provides further motivation to conduct theoretical research and large-scale data analysis in deciphering the genetic and physiological meaning of gene-environment interactions and to develop more statistical methods for testing gene-environment interactions. In the coming years, to integrate geneenvironment interactions into genome-wide association analysis will be a major task in genetic studies of complex diseases.

Supporting Information

Appendix S1

Found at: doi:10.1371/journal.pone.0004578.s001 (0.11 MB DOC)

References

- 1. Ay N (2002) Locality of global stochastic interaction in directed acyclic networks. Neural Comput 14: 2959-80.
- Andrieu N, Goldstein AM (1998) Epidemiologic and genetic approaches in the study of gene-environment interaction: an overview of available methods. Epidemiol Rev 20(2): 137-47.
- 3. Manolio TA, Bailey-Wilson JE, Collins FS (2006) Genes, environment and the value of prospective cohort studies. Nat Rev Genet 7(10): 812-820.
- 4. Jakulin A (2005) Machine learning based on attribute interaction. Ph.D. Dissertation. University of Ljubljana, Sezana.
- Ottman R (1996) Gene-environment interaction: definitions and study design. 5. Preventive Medicine 25: 764-770.
- 6. Rothman KJ, Greenland S, Walker AM (1980) Concepts of interaction. Am J Epidemiol 112(4): 467-70.
- Cheverud JM, Routman EJ (1995) Epistasis and its contribution to genetic 7. variance components. Genetics 139: 1455-1461.
- 8. Hansen TF, Wagner GP (2001) Modeling genetic architecture a multilinear theory of gene interaction. Theor Popul Biol 59: 61-86.
- 9. Puniyani A, Liberman U, Feldman MW (2004) On the meaning of non-epistatic selection. Theor Popul Biol 66: 317-321
- 10. Liberman U, Puniyani A, Feldman MW (2007) On the evolution of epistasis II: A generalized Wright-Kimura framework. Theor Popul Biol March 71(2): 230-238
- 11. Cover TM, Thomas JA (1991) Elements of information theory. New York: John Wiley & Sons, Inc
- 12. Jakulin A, Bratko I, Smrke D, Demsar J, Zupan B (2003) Attribute interactions in medical data analysis. Proceedings of the 9th Conference on Artificial Intelligence in Medicine in Europe (AIME 2003), Protaras, Cyprus, October 18-22, 2003. Dojat M, Keravnou E, Barahona P, eds. Lecture Notes in Artificial Intelligence 2780: 229-238
- 13. Matsuda H (2000) Physical nature of higher-order mutual information: Intrinsic correlations and frustration. Physical Review E 62(3): 3096-3102.
- 14. Nakahara H, Nishimura S, Inoue M, Hori G, Amari S (2003) Gene interaction in DNA microarray data is decomposed by information geometric measure. Bioinformatics 19: 1124-1131.
- Brillinger DR (2004) Some data analyses using mutual information. 15. Brazilian J Probability Statistics 18: 163-183.
- 16. McGill (1954) Multivariate information transmission. Psychometrika 19: 97 - 116
- 17. Liu X, Fallin MD, Kao WH (2004) Genetic dissection methods: designs used for tests of gene-environment interaction. Current Opinion Genetics Development 14: 241-245
- 18. Garcia-Closas M, Lubin JH (1999) Power and sample size calculations in casecontrol studies of gene-environment interactions: comments on different approaches. Am J Epidemiol 149: 689-92.
- 19. Winslow RL, Boguski MS (2003) Genome informatics: current status and future prospects. Circ Res 92: 953-961
- Yoon Y, Song J, Hong SH, Kim JQ (2003) Analysis of multiple single nucleotide 20. polymorphisms of candidate genes related to coronary heart disease susceptibility by using support vector machines. Clin Chem Lab Med 41: 529-534
- 21. Hahn LW, Ritchie MD, Moore JH (2003) Multifactor dimensionality reduction software for detecting gene-gene and gene-environment interactions. Bioinformatics 19: 376-382
- 22. Luan JA, Wong MY, Day NE, Wareham NJ (2001) Sample size determination for studies of gene-environment interaction. Int J Epidemiol 30(5): 1035-40.

Appendix S2

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Author Contributions

Conceived and designed the experiments: LJ MX. Analyzed the data: XW. Wrote the paper: MX.

- 23. Gauderman WJ (2002) Sample size requirements for matched case-control studies of gene-environment interaction. Stat Med 21(1): 35-50.
- 24. Goldstein AM, Dondon MG, Andrieu N (2006) Unconditional analyses can increase efficiency in assessing gene-environment interaction of the case-combined-control design. Int J Epidemiol 35(4): 1067–73.
- 25. Goodman M, Dana Flanders W (2006) Study design options in evaluating geneenvironment interactions: Practical considerations for a planned case-control study of pediatric leukemia. Pediatr Blood Cancer [Epub ahead of print].
- 26. Chatterjee N, Kalaylioglu Z, Moslehi R, Peters U, Wacholder S (2006) Powerful multilocus tests of genetic association in the presence of gene-gene and geneenvironment interactions. Am J Hum Genet 79(6): 1002-100.
- 27. Marchini J, Donnelly P, Cardon LR (2005) Genome-wide strategies for detecting multiple loci that influence complex diseases. Nat Genet 37: 413-417.
- 28. Chung Y, Lee SY, Elston RC, Park T (2007) Odds ratio based multifactordimensionality reduction method for detecting gene-gene interactions. Bioinformatics 23(1): 71-6
- Ritchie MD, Hahn LW, Roodi N, Bailey LR, Dupont WD, et al. (2001) 29. Multifactor-dimensionality reduction reveals high-order interactions among estrogen-metabolism genes in sporadic breast cancer. Am J Hum Genet 69: 138 - 147
- 30. Bush WS, Dudek SM, Ritchie MD (2006) Parallel multifactor dimensionality reduction: a tool for the large-scale analysis of gene-gene interactions. Bioinformatics 22(17): 2173-4.
- 31. Nothnagel M (2002) Simulation of LD block-structured SNP haplotype data and its use for the analysis of case-control data by supervised learning methods. Am J Hum Genet 71(Suppl): A2363.
- 32. Zhou W, Liu G, Miller DP, Thurston SW, Xu LL, et al. (2002) Geneenvironment interaction for the ERCC2 polymorphisms and cumulative cigarette smoking exposure in lung cancer. Cancer Res 62(5): 1377-81.
- 33. Dandara C, Li DP, Walther G, Parker MI (2006) Gene-environment interaction: the role of SULT1A1 and CYP3A5 polymorphisms as risk modifiers for squamous cell carcinoma of the oesophagus. Carcinogenesis 27: 791-7.
- 34. Klareskog L, Stolt P, Lundberg K, Kallberg H, Bengtsson C, et al. (2006) A new model for an etiology of rheumatoid arthritis: smoking may trigger HLA-DR (shared epitope)-restricted immune reactions to autoantigens modified by citrullination. Arthritis Rheum 54: 38-46.
- 35. Kallberg H, Padyukov L, Plenge RM, Ronnelid J, Gregersen PK, et al. (2007) Gene-gene and gene-environment interactions involving HLA-DRB1, PTPN22, and smoking in two subsets of rheumatoid arthritis. Am J Hum Genet 80: 867-75.
- 36. Lundberg K, Nijenhuis S, Vossenaar ER, Palmblad K, Van Venrooij WJ, et al. (2005) Citrullinated proteins have increased immunogenicity and arthritogenicity and their presence in arthritic joints correlates with disease severity. Arthritis Res Ther 7: R458-67.
- Linn-Rasker SP, Van der Helm-Van Mil AH, Van Gaalen FA, Kloppenburg M, 37. De Vries RR, et al. (2006) Smoking is a risk factor for anti-CCP antibodies only in rheumatoid arthritis patients who carry HLA-DRB1 shared epitope alleles. Ann Rheum Dis 65: 366-71.
- Rothman KJ (2002) Epidemiology: An introduction. New York: Oxford 38 University Press.
- 39. Ahlbom A, Alfredsson L (2005) Interaction: A word with two meanings creates confusion. Eur J Epidemiol 20: 563-4.