

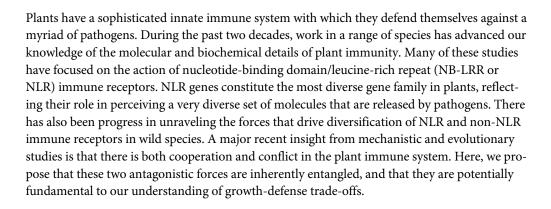
PEARLS

Cooperation and Conflict in the Plant Immune System

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Cooperation in the Immune System

Genetic studies of plant immunity have been tremendously influenced by Flor's gene-for-gene hypothesis, which posits that a single host resistance gene is matched by a single effector gene from a specific pathogen strain. It was thus unexpected when two NLR genes that are located immediately adjacent to each other in the Arabidopsis thaliana genome were found to cooperatively confer resistance to the *Hyaloperonospora arabidopsidis* ex *parasitica* isolate Cala2 [1]. Since then, several more such pairs have been discovered, with one member often encoding other protein domains in addition to the canonical "NLR" moieties [2]. Two good examples are the RPS4/RRS1 protein pair, which endows A. thaliana with resistance to both bacterial and fungal pathogens, and the RGA4/RGA5 pair in rice, which confers resistance to specific races of the fungus Magnaporthe oryzae. The dual genetic requirement translates into direct physical interaction, with RPS4 and RRS1 (as well as RGA4 and RGA5) forming heterodimers [2-5]. In both cases, one protein, RRS1 or RGA5, contains an additional specific recognition domain at the C-terminus [2,6]. Binding of a pathogen effector protein to this domain alters the interaction with its respective partner, RPS4 or RGA4, and thereby leads to host immune signaling. The addition of an effector-binding domain to one of the NLR proteins may have facilitated expansion of recognition specificity, since the evolution of the recognition domain is not necessarily coupled to constraints acting on the NLR moiety.

An attractive hypothesis is that the involvement of two physically interacting NLR proteins not only increases the specificity of the response compared to effector recognition by a single protein but also provides safeguards against inappropriate activation. In support of such a scenario, ectopic expression of RPS4 or RGA4, but not of the effector-detecting RRS1 or RGA5, causes autoimmunity [2,3,7]. The tight physical linkage in the genome would support



functional coevolution, as seen in other clustered immune-gene systems (e.g., [8]). This may also explain why functionally dependent but unlinked NLR pairs are less common [9].

Conflict in the Immune System

Another common type of protein–protein interaction in the plant immune system is the one between immune receptors (often NLR proteins) and host clients that are direct targets of pathogen effectors. From the host perspective, pathogen-targeting of the NLR clients may be undesirable when it leads to enhanced pathogen virulence; in this case, clients are considered NLR-guardees. In other cases, however, targeting may be desirable, because the NLR-clients are merely decoy versions of true targets [10]. The interaction between the NLR-guard and its guardee/decoy mediates indirect recognition of pathogen effectors [9]. Similar to the NLR-NLR interaction discussed above, the guardee/decoy partner modulates signaling of the NLR protein; if the guardee/decoy is compromised, either by effector-dependent modification or mutation, immune responses are activated. This can also occur when the NLR (or a non-NLR immune receptor) and its host client have not coevolved and are mismatched [11,12]. Such spontaneous, pathogen-independent autoimmune reactions can be observed in many genetic intra- and interspecific crosses, a phenomenon that breeders have known as hybrid necrosis for decades [13].

A. thaliana, an excellent species for the study of naturally occurring genetic variation, has proven to be valuable for the investigation of hybrid necrosis. The first report of a hybrid necrosis locus in A. thaliana, DANGEROUS MIX1 (DM1), identified an NLR protein that triggers autoimmunity when combined with a specific allele at the unlinked DM2 locus [14]. A recent systematic study using thousands of intraspecific A. thaliana crosses has not only corroborated NLR loci as major contributors to hybrid necrosis but also confirmed that the genetic architecture of hybrid necrosis is often simple, with typically one or two causal loci. Notably, several cases involve NLR–NLR interactions, including the DM1/DM2 case [15]. NLR genes show extreme sequence and copy number variation both within and between species [16], reflecting the multitude of pathogen molecules that are directly or indirectly detected by NLR proteins. It thus hardly seems surprising that such extreme diversity can lead to collateral damage in the form of hybrid necrosis—highlighting a conflict between diversification of the immune system and avoidance of unwarranted autoimmunity.

Another outcome of this conflict appears to manifest itself as the suppression of host resistance, which has been observed during attempts to pyramid valuable resistance genes in crops. At least in one such case, in wheat, resistance suppression results from mismatched interactions between NLR proteins that are encoded by different alleles of the same locus [17,18]. Given that other NLR protein pairs positively cooperate for proper function [2,3], an obvious hypothesis is that resistance suppression is caused by inappropriate physical interactions between the different NLRs. Such opposite outcomes of interference among plant immune system components—either leading to hybrid failure due to hyperimmunity or to weakened immune response—had previously been predicted from modelling the effects of pathogen diversity on the evolution of interactions within the plant immune system [19].

Towards Dissecting Trade-Offs between Growth and Immunity

Individual NLR genes can incur substantial fitness costs in the absence of the pathogen they recognize, not only in the greenhouse but also in the field (e.g., [20]). Ideally, immune receptors would, of course, be perfectly "off" in the absence of a trigger. The fact that there are apparent NLR effects, even without pathogen effectors, points to a biochemical trade-off between the robustness of the on/off switch and sensitive, highly specific activation of immune receptors.

Still, knocking out common components of NLR downstream signaling does not lead to enormous increases in growth. One can think of several explanations for why this is the case; for example, it could be that NLR genes with substantial fitness costs are the exception, with aberrant NLR activity being often suppressed by other NLR genes, or that there is an inherent limit to the resources that will normally be diverted to defense upon NLR activation. No matter what the right answer is, we certainly need a better understanding of the trade-offs both within the immune system and between immunity and growth. One approach is to investigate biochemical details of immune receptor activation, and of the connection between immune receptors and hormone signaling [21]. Similarly, phenotypic differences among different autoactive NLR mutants (e.g., [22]) and between different hybrid necrosis cases [15] may be illuminating in this regard.

A complementary approach begins with a population perspective. We hypothesize that the strong necrosis phenotypes seen in some hybrids constitute the proverbial tip of the iceberg of hybrid immune reactions. A corollary of this proposal is the ubiquity of more subtle epistatic interactions that affect both the state of the plant immune system and growth or fertility. Systematic analyses of intraspecific crosses in *A. thaliana* have already shown that subtle F_1 hybrid necrosis cases are much more common than drastic, lethal interactions [15], and there are hybrid necrosis cases that are only revealed in the F_2 generation [23,24]. We like to think about the trade-offs in the framework of phenotype space (Fig 1). In addition to the lethally immune and hypo-immune hybrid genotypes discussed in the previous section, there are natural genotypes that suffer from autoimmunity and reduced growth but, at the same time, benefit from pleiotropic broad-spectrum resistance against a wide range of pathogens [25].

Of most interest in this abstract phenotypic space is the "Pareto front" (grey line in $\underline{\text{Fig 1}}$), which describes a classical optimization problem [26]. It is clear that the preferred genotypes are those that are closest to the Pareto front (colored green in $\underline{\text{Fig 1}}$), but that the particular

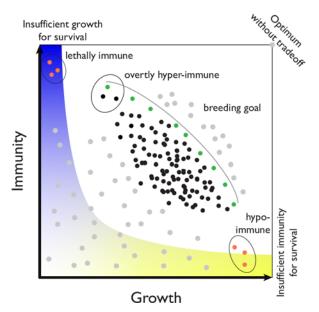


Fig 1. Hypothetical phenotypic space determined by trade-offs between growth and immunity. Common, reasonably fit genotypes are in black; genotypes that could potentially be created by crosses or mutations are in grey and orange. Green are genotypes closest to the Pareto front (grey line), which defines optimality among existing genotypes. The blue/yellow area represents forbidden phenotypic space, i.e., genotypes that die, always succumb to pathogens, or are infertile. The Pareto front can shift in response to a certain pathogen pressure or to abiotic challenge.

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pathogen pressure will determine which of these preferred genotypes is superior in a specific situation. Given that, in reality, the phenotypic space (and therefore the position of the Pareto front itself) will vary with shifts in pathogen populations and abiotic conditions, it will be important to determine the Pareto front in numerous controlled laboratory settings as well as in natural habitats of *A. thaliana*. We also note that mild necrosis in either inbred *A. thaliana* accessions or progeny of crosses is not that rare [15,23,25], suggesting that mild necrosis risk alleles are maintained at an appreciable frequency. Important goals for the future are, therefore, to better understand how the inherent trade-off between growth or fertility and immunity is managed in natural populations, to find out whether one can extrapolate from findings in *A. thaliana* to other species, and to test whether lessons from wild plants such as *A. thaliana* can be useful in crop breeding.

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References

- Sinapidou E, Williams K, Nott L, Bahkt S, Tor M, et al. (2004) Two TIR:NB:LRR genes are required to specify resistance to *Peronospora parasitica* isolate Cala2 in Arabidopsis. Plant J 38: 898–909. PMID: 15165183
- Cesari S, Kanzaki H, Fujiwara T, Bernoux M, Chalvon V, et al. (2014) The NB-LRR proteins RGA4 and RGA5 interact functionally and physically to confer disease resistance. EMBO J 33: 1941–1959.
 PMID: 25024433 doi: 10.15252/embj.201487923
- Williams SJ, Sohn KH, Wan L, Bernoux M, Sarris PF, et al. (2014) Structural basis for assembly and function of a heterodimeric plant immune receptor. Science 344: 299–303. PMID: <u>24744375</u> doi: <u>10.</u> 1126/science.1247357
- Sarris PF, Duxbury Z, Huh SU, Ma Y, Segonzac C, et al. (2015) A plant immune receptor detects pathogen effectors that target WRKY transcription factors. Cell 161: 1089–1100. PMID: 26000484 doi: 1016/j.cell.2015.04.024
- Le Roux C, Huet G, Jauneau A, Camborde L, Tremousaygue D, et al. (2015) A receptor pair with an integrated decoy converts pathogen disabling of transcription factors to immunity. Cell 161: 1074– 1088. PMID: 26000483 doi: 10.1016/j.cell.2015.04.025
- Wu CH, Krasileva KV, Banfield MJ, Terauchi R, Kamoun S (2015) The "sensor domains" of plant NLR proteins: more than decoys? Front Plant Sci 6: 134. PMID: 25798142 doi: 10.3389/fpls.2015.00134
- Wirthmueller L, Zhang Y, Jones JD, Parker JE (2007) Nuclear accumulation of the *Arabidopsis* immune receptor RPS4 is necessary for triggering EDS1-dependent defense. Curr Biol 17: 2023–2029. PMID: 17997306
- Walker BA, Hunt LG, Sowa AK, Skjodt K, Gobel TW, et al. (2011) The dominantly expressed class I molecule of the chicken MHC is explained by coevolution with the polymorphic peptide transporter (TAP) genes. Proc Natl Acad Sci USA 108: 8396–8401. PMID: <u>21536896</u> doi: <u>10.1073/pnas.</u> 1019496108
- Eitas TK, Dangl JL (2010) NB-LRR proteins: pairs, pieces, perception, partners, and pathways. Curr Opin Plant Biol 13: 472–477. PMID: 20483655 doi: 10.1016/j.pbi.2010.04.007
- van der Hoorn RA, Kamoun S (2008) From guard to decoy: a new model for perception of plant pathogen effectors. Plant Cell 20: 2009–2017. PMID: 18723576 doi: 10.1105/tpc.108.060194
- Krüger J, Thomas CM, Golstein C, Dixon MS, Smoker M, et al. (2002) A tomato cysteine protease required for Cf-2-dependent disease resistance and suppression of autonecrosis. Science 296: 744– 747. PMID: 11976458
- Alcázar R, García AV, Kronholm I, de Meaux J, Koornneef M, et al. (2010) Natural variation at Strubbelig Receptor Kinase 3 drives immune-triggered incompatibilities between *Arabidopsis thaliana* accessions. Nat Genet 42: 1135–1139. PMID: 21037570 doi: 10.1038/ng.704
- Bomblies K, Weigel D (2007) Hybrid necrosis: autoimmunity as a potential gene-flow barrier in plant species. Nat Rev Genet 8: 382–393. PMID: 17404584



- Bomblies K, Lempe J, Epple P, Warthmann N, Lanz C, et al. (2007) Autoimmune response as a mechanism for a Dobzhansky-Muller-type incompatibility syndrome in plants. PLoS Biol 5: e236. PMID: 17803357
- 15. Chae E, Bomblies K, Kim ST, Karelina D, Zaidem M, et al. (2014) Species-wide genetic incompatibility analysis identifies immune genes as hot spots of deleterious epistasis. Cell 159: 1341–1351. PMID: 25467443 doi: 10.1016/j.cell.2014.10.049
- Jacob F, Vernaldi S, Maekawa T (2013) Evolution and conservation of plant NLR functions. Front Immunol 4: 297. PMID: 24093022 doi: 10.3389/fimmu.2013.00297
- 17. McIntosh RA, Zhang P, Cowger C, Parks R, Lagudah ES, et al. (2011) Rye-derived powdery mildew resistance gene Pm8 in wheat is suppressed by the Pm3 locus. Theor Appl Genet 123: 359–367. PMID: 21509697 doi: 10.1007/s00122-011-1589-5
- 18. Stirnweis D, Milani SD, Brunner S, Herren G, Buchmann G, et al. (2014) Suppression among alleles encoding nucleotide-binding-leucine-rich repeat resistance proteins interferes with resistance in F₁ hybrid and allele-pyramided wheat plants. Plant J 79: 893–903. PMID: <u>24942051</u> doi: <u>10.1111/tpj.</u> 12592
- Ispolatov I, Doebeli M (2009) Speciation due to hybrid necrosis in plant-pathogen models. Evolution 63: 3076–3084. PMID: 19674099 doi: 10.1111/j.1558-5646.2009.00800.x
- Karasov TL, Kniskern JM, Gao L, DeYoung BJ, Ding J, et al. (2014) The long-term maintenance of a resistance polymorphism through diffuse interactions. Nature 512: 436–440. PMID: 25043057 doi: 10.38/nature13439
- 21. Huot B, Yao J, Montgomery BL, He SY (2014) Growth-defense tradeoffs in plants: a balancing act to optimize fitness. Mol Plant 7: 1267–1287. PMID: 24777989 doi: 10.1093/mp/ssu049
- 22. Igari K, Endo S, Hibara K, Aida M, Sakakibara H, et al. (2008) Constitutive activation of a CC-NB-LRR protein alters morphogenesis through the cytokinin pathway in *Arabidopsis*. Plant J 55: 14–27. PMID: 18315541 doi: 10.1111/j.1365-313X.2008.03466.x
- Alcázar R, Garcia AV, Parker JE, Reymond M (2009) Incremental steps toward incompatibility revealed by *Arabidopsis* epistatic interactions modulating salicylic acid pathway activation. Proc Natl Acad Sci USA 106: 334–339. PMID: doi: 10.1073/pnas.0811734106 PMID: 19106299
- 24. Alcázar R, von Reth M, Bautor J, Chae E, Weigel D, et al. (2014) Analysis of a plant complex resistance gene locus underlying immune-related hybrid incompatibility and its occurrence in nature. PLoS Genet 10: e1004848. PMID: 25503786 doi: 10.1371/journal.pgen.1004848
- Todesco M, Balasubramanian S, Hu TT, Traw MB, Horton M, et al. (2010) Natural allelic variation underlying a major fitness trade-off in *Arabidopsis thaliana*. Nature 465: 632–636. PMID: <u>20520716</u> doi: 10.1038/nature09083
- Shoval O, Sheftel H, Shinar G, Hart Y, Ramote O, et al. (2012) Evolutionary trade-offs, Pareto optimality, and the geometry of phenotype space. Science 336: 1157–1160. PMID: 22539553 doi: 10.1126/ science.1217405