

MM with bortezomib, a proteasome inhibitor known for its remarkable efficacy in treating extramedullary MM.

Although the pathogenesis of MPE is unknown, it is theorized that it may be a direct extension of thoracic myelomatous involvement. A review of 57 cases [9] demonstrated that half of the patients with MPE had concomitant thoracic skeletal, lung parenchymal, or chest wall plasmacytomas, which would provide a source for MPEs. Similarly, both of our patients had a pulmonary nodule, which likely represented metastatic disease.

Genetic analysis showed that the patient in our first case had a trisomy at chromosome 3 and monosomy at chromosome 13. In addition to the t(4;14) translocation, this complex karyotype is associated with unfavorable prognosis [5]. Given that the median survival time for high-risk patients without malignant pleural effusions is 3 years, it is likely that the progression of the myeloma and development of the pleural effusions contributed significantly to the eventual death of the first patient. In our second case, the patient had no trisomy, but she did have monosomy of chromosome 13 in addition to the t(4;14) translocation. This chromosome 13 abnormality was also seen in 77.8% of patients in the Cho *et al.* [10] case series.

Although rare, more cases of MPE are being described in the literature, with evidence indicating its poor prognosis and lack of efficacious treatment [5, 12]. Because of the severity of MPE, we recommend that patients with pleural effusions and suspicion of myeloma undergo protein electrophoresis, flow cytometry, cytologic examination of the pleural fluid, or pleural biopsy examination to identify MPE and begin treatment promptly [12, 13].

Akshay Amaraneni¹, Usman Saeed², Devin Malik¹, Megan Brown³, Sreenivasa R. Chandana⁴

¹Department of Internal Medicine, ²Department of Internal Medicine-Pediatrics, Western Michigan University, Homer Stryker M.D. School of Medicine, Kalamazoo, ³Michigan State University, College of Human Medicine, East Lansing, ⁴Division of Hematology and Oncology, West Michigan Cancer Center, Kalamazoo, MI, USA

Correspondence to: Sreenivasa R. Chandana
Division of Hematology and Oncology,
West Michigan Cancer Center,
200 N Park Street, Kalamazoo, MI 49007, USA
E-mail: schandana@wmcc.org

Received on May 9, 2015; Revised on May 27, 2015; Accepted on Jun. 15, 2015

<http://dx.doi.org/10.5045/br.2016.51.2.142>

Authors' Disclosures of Potential Conflicts of Interest

No potential conflicts of interest relevant to this article were reported.

REFERENCES

1. Siegel R, Naishadham D, Jemal A. Cancer statistics, 2013. *CA*

Cancer J Clin 2013;63:11-30.

2. Kumar SK, Rajkumar SV, Dispenzieri A, et al. Improved survival in multiple myeloma and the impact of novel therapies. *Blood* 2008;111:2516-20.
3. Bladé J, Rosiñol L. Complications of multiple myeloma. *Hematol Oncol Clin North Am* 2007;21:1231-46.
4. Yosunkaya S, Maden E, Toy H, Yazici R, Ozer F, Reisl I. A multiple myeloma case presenting with bilateral pleural involvement. *Tuberk Toraks* 2007;55:285-9.
5. Mikhael JR, Dingli D, Roy V, et al. Management of newly diagnosed symptomatic multiple myeloma: updated Mayo stratification of myeloma and risk-adapted therapy (mSMART) consensus guidelines 2013. *Mayo Clin Proc* 2013;88:360-76.
6. Kintzer JS Jr, Rosenow EC 3rd, Kyle RA. Thoracic and pulmonary abnormalities in multiple myeloma. A review of 958 cases. *Arch Intern Med* 1978;138:727-30.
7. Meoli A, Willsie S, Fiorella R. Myelomatous pleural effusion. *South Med J* 1997;90:65-8.
8. Kamble R, Wilson CS, Fassas A, et al. Malignant pleural effusion of multiple myeloma: prognostic factors and outcome. *Leuk Lymphoma* 2005;46:1137-42.
9. Kim YJ, Kim SJ, Min K, et al. Multiple myeloma with myelomatous pleural effusion: a case report and review of the literature. *Acta Haematol* 2008;120:108-11.
10. Cho YU, Chi HS, Park CJ, Jang S, Seo EJ, Suh C. Myelomatous pleural effusion: a case series in a single institution and literature review. *Korean J Lab Med* 2011;31:225-30.
11. Mangiacavalli S, Varettoni M, Zappasodi P, Pica G, Lazzarino M, Corso A. A striking response to bortezomib in a patient with pleural localization of multiple myeloma. *Leuk Res* 2009;33:577-8.
12. Keklik M, Sivgin S, Pala C, et al. Flow cytometry method as a diagnostic tool for pleural fluid involvement in a patient with multiple myeloma. *Mediterr J Hematol Infect Dis* 2012;4:e2012063.
13. Oudart JB, Maquart FX, Semouma O, Lauer M, Arthuis-Demoulin P, Ramont L. Pleural effusion in a patient with multiple myeloma. *Clin Chem* 2012;58:672-4.

A rare case of diffuse large B cell lymphoma-associated hemophagocytic syndrome initially present in the bone marrow with a favorable clinical course

TO THE EDITOR: Lymphoma-associated hemophagocytic syndrome (LAHS) is a hematological disorder associated with malignant lymphoma. It is characterized by clinical features and laboratory findings associated with hemophagocytic lymphohistiocytosis (HLH), such as fever, cytopenia, hyperferritinemia, hypofibrinogenemia, and hemophagocytosis in the bone marrow (BM) [1]. The development of LAHS can be accounted for by various types of lymphoma,

but most of them are associated with T cell or natural killer (NK) cell lymphomas such as aggressive NK/T cell lymphoma, peripheral T cell lymphoma, anaplastic large cell lymphoma, or extranodal NK/T cell lymphoma [2-7]. Development of LAHS from B cell lymphoma is rarely reported, and occurs in old age with a low frequency of BM involvement at diagnosis [6, 7]. To date, seven cases of LAHS associated with large B cell lymphoma initially manifesting in the BM have been reported and the majority of these cases were of the non-germinal center (GC) type, with a complex karyotype and very poor prognosis [8]. We report here a rare case of diffuse large B cell lymphoma (DLBCL)-associated hemophagocytic syndrome initially manifesting in the BM and exhibiting a favorable clinical course.

In February 2015, a 73-year-old female was admitted with fever (body temperature, 38.1°C), poor oral intake,

and general weakness, which had developed 2 weeks prior to admission. She was diagnosed as diabetes mellitus (DM) 8 years ago and had been on treatment with a hypoglycemic agent, but had no medical history related to hematological malignancy, including hepatosplenomegaly. The patient's hemogram results at admission showed pancytopenia (white blood cell [WBC] count $3.57 \times 10^9/L$ [absolute neutrophil count $2.82 \times 10^9/L$], hemoglobin 7.6 g/dL, platelet count $32.0 \times 10^9/L$). Her peripheral blood smear showed the presence of atypical lymphocytes (3% of total nucleated cells). She underwent BM aspiration and biopsy due to pancytopenia with presence of atypical lymphocytes.

The patient's BM aspirates (Fig. 1A, B) and touch print (Fig. 1C, D) showed normocellular marrow with increased infiltration of large-sized neoplastic lymphoid cells with a frequency of 20.0% (indicated by the orange arrows).

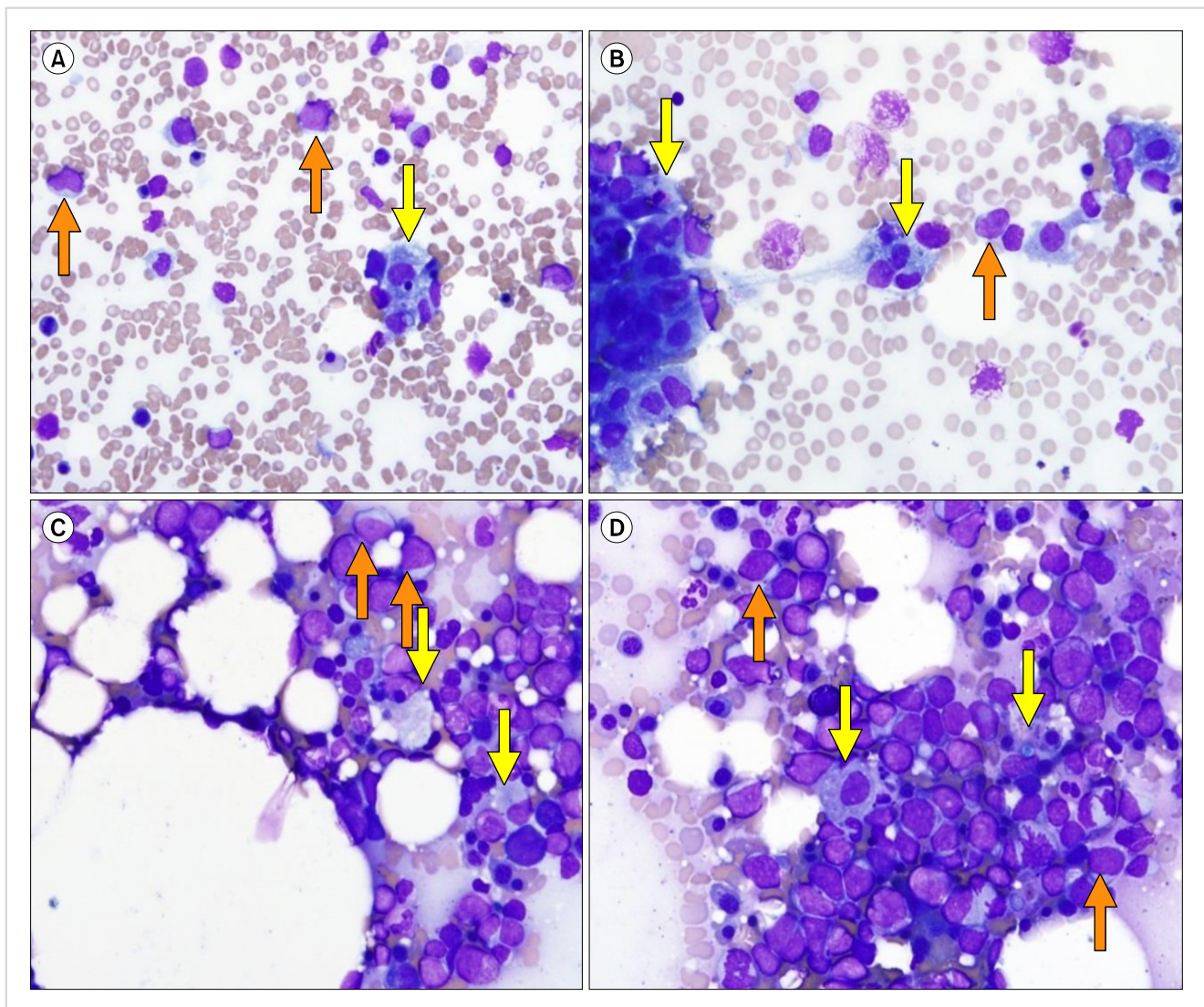


Fig. 1. Bone marrow aspirates (A, B, Wright stain, $\times 400$) and touch print (C, D, Wright stain, $\times 400$). The bone marrow aspirates and touch print show normocellular marrow with infiltration of neoplastic lymphoid cells (20.0% of total nucleated cells, orange arrows). In addition, hemophagocytic histiocytes (11.0% of total nucleated cells, yellow arrows) were identified.

In addition, histiocytes with active hemophagocytosis were occasionally identified with a frequency of 11.0% (indicated by the yellow arrows). Flow cytometric analysis results of BM aspirates demonstrated the presence of a clonal B lymphocyte population with kappa light chain restriction: 19.0% positivity for CD19, 22.6% positivity for CD20, 35.0% positivity for kappa light chain, and 5.2% positivity for lambda light chain. However, T-cell markers showed no evidence of clonality, with variable positivity for CD3, CD2, CD5, CD7, CD4 and CD8, (range, 10.9–48.1%).

The BM biopsy (Fig. 2A) showed that 65% of the marrow was cellular. Diffuse infiltration of CD20-positive neoplastic lymphoid cells (indicated by the orange arrows) and increased infiltration of CD68-positive hemophagocytic histiocytes (indicated by the yellow arrows) was demonstrated by immunohistochemical stains for CD3, CD20 and CD68

(DAKO, Glostrup, Denmark) (Fig. 2B–D, respectively). Serum ferritin, fibrinogen, triglyceride, and soluble interleukin-2 (IL-2) receptor levels were 1,198.0 ng/mL (reference range, 33–298 ng/mL), 300.5 mg/dL (reference range, 170–380 mg/dL), 149 mg/dL (reference range, 58–250 mg/dL) and 6,530.0 U/mL (reference range, 124–466 U/mL), respectively. The patient's karyotype analysis results showed 46–48,XX,add(1)(q25),add(1)(q42),add(2)(q33),del(4)(q21),del(5)(p13p15.3),add(6)(q21),del(8)(q22),+16,add(17)(q25),add(19)(p13.3),+20[cp11]/46,XX[9], indicating the presence of a complex karyotype in 55% of all analyzed cells. Based on these results, and because her clinical and laboratory findings fulfilled the 2008 diagnostic criteria of HLH (fever, cytopenia, hyperferritinemia, hemophagocytosis in BM, high soluble IL-2 receptor levels) [9], the patient was initially diagnosed with B cell LAHS which was finally revised as

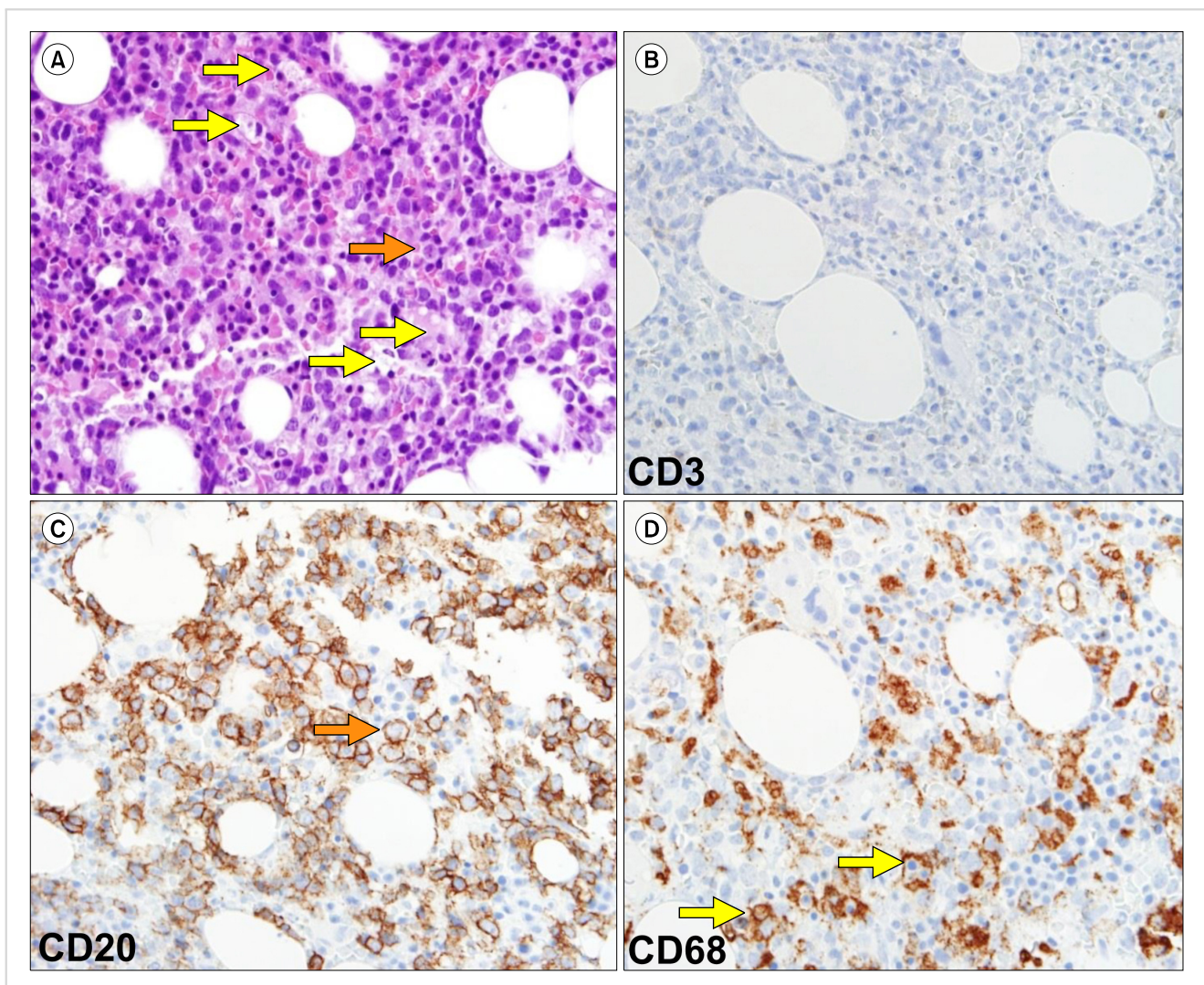


Fig. 2. Bone marrow biopsy (A, hematoxylin and eosin stain, $\times 400$) and immunohistochemical staining results for CD3, CD20 and CD68 (B–D, $\times 400$, respectively). The bone marrow biopsy shows normocellular marrow with diffuse infiltration of CD20-positive neoplastic lymphoid cells (orange arrows) accompanied by an increase of CD68-positive histiocytes with occasional hemophagocytosis (yellow arrows), indicating the presence of B-cell lymphoma associated with hemophagocytic histiocytosis.

DLBCL (non-GC type)-associated hemophagocytic syndrome. She was treated with 5 cycles of R-CHOP (cyclophosphamide, doxorubicin, vincristine, prednisone plus rituximab) chemotherapy. At follow-up after 3 months of chemotherapy, the patient's hemogram results had improved significantly (WBC $4.45 \times 10^9/L$, hemoglobin 10.7 g/dL, platelet count $148 \times 10^9/L$). In the BM study, residual neoplastic lymphoid cells were not identified and active hemophagocytosis was not observed, despite persistent histiocytic hyperplasia. The patient has been discharged in an improved condition and is waiting to undergo additional chemotherapy.

According to the literature, B cell LAHS cases are rather uncommon compared with T- or NK-cell lymphoma cases and the most common type is DLBCL, which occurs in older patients [6, 7]. To date, there have been seven reports of large B cell lymphoma initially manifesting in the BM, with consistent clinical features such as occurrence in old age (range, 54–80 yr), non-GC type (six cases), a complex karyotype, aggressive clinical course (four cases died before treatment), and no association with Epstein-Barr virus (EB) infection (all seven cases) [8]. The clinical features of our patient were generally similar to those of the previous cases: a complex karyotype, non-GC type of DLBCL, old age at diagnosis. However, our patient also demonstrated an uncommon clinical feature of B cell LAHS initially manifesting in the BM: a favorable clinical outcome after chemotherapy. Since our patient did not undergo serologic or molecular EBV studies, we could not confirm that EBV infection was not associated with B cell LAHS initially manifesting in the BM, which was suggested by the previous study [8]. Further studies will be required to confirm the clinical features of B cell LAHS initially manifesting in the BM.

In conclusion, we report a rare case of DLBCL-associated hemophagocytic syndrome initially manifesting in the BM and exhibiting a favorable clinical outcome. More extensive studies will be needed to investigate the clinical features of these patients.

*This work was supported by the year 2015 clinical research grant from Pusan National University Hospital.

Sang Hyuk Park¹, Eun Yup Lee¹, Joo Seop Chung²

¹Department of Laboratory Medicine and Biomedical Research Institute, ²Division of Hematology-Oncology, Department of Internal Medicine, Pusan National University Hospital, Pusan National University School of Medicine, Busan, Korea

Correspondence to: Sang Hyuk Park

Department of Laboratory Medicine and Biomedical Research Institute, Pusan National University Hospital, 179 Gudeok-ro, Seo-gu, Busan 49241, Korea
E-mail: korailman-1@hanmail.net

Received on May 26, 2015; Revised on Jun. 3, 2015; Accepted on Jun. 18, 2015

<http://dx.doi.org/10.5045/br.2016.51.2.144>

Authors' Disclosures of Potential Conflicts of Interest

No potential conflicts of interest relevant to this article were reported.

REFERENCES

1. Usmani GN, Woda BA, Newburger PE. Advances in understanding the pathogenesis of HLH. *Br J Haematol* 2013;161:609–22.
2. Cheung MM, Chan JK, Lau WH, et al. Primary non-Hodgkin's lymphoma of the nose and nasopharynx: clinical features, tumor immunophenotype, and treatment outcome in 113 patients. *J Clin Oncol* 1998;16:70–7.
3. Falini B, Pileri S, De Solas I, et al. Peripheral T-cell lymphoma associated with hemophagocytic syndrome. *Blood* 1990;75:434–44.
4. Florena AM, Iannitto E, Quintini G, Franco V. Bone marrow biopsy in hemophagocytic syndrome. *Virchows Arch* 2002;441:335–44.
5. Shimazaki C, Inaba T, Shimura K, et al. B-cell lymphoma associated with haemophagocytic syndrome: a clinical, immunological and cytogenetic study. *Br J Haematol* 1999;104:672–9.
6. Han AR, Lee HR, Park BB, et al. Lymphoma-associated hemophagocytic syndrome: clinical features and treatment outcome. *Ann Hematol* 2007;86:493–8.
7. Sano H, Kobayashi R, Tanaka J, et al. Risk factor analysis of non-Hodgkin lymphoma-associated haemophagocytic syndromes: a multicentre study. *Br J Haematol* 2014;165:786–92.
8. Yeh YM, Chang KC, Chen YP, et al. Large B cell lymphoma presenting initially in bone marrow, liver and spleen: an aggressive entity associated frequently with haemophagocytic syndrome. *Histopathology* 2010;57:785–95.
9. Jordan MB, Filipovich AH. Hematopoietic cell transplantation for hemophagocytic lymphohistiocytosis: a journey of a thousand miles begins with a single (big) step. *Bone Marrow Transplant* 2008;42:433–7.