



Closed Genome Sequence of Octadecabacter temperatus SB1, the First Mesophilic Species of the Genus Octadecabacter

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The Gram-negative alphaproteobacterium *Octadecabacter temperatus* SB1 (DSM 26878) belongs to the marine *Roseobacter* clade. The genome of this strain is the smallest closed genome of the *Roseobacter* clade. *O. temperatus* SB1 is the first described nonpolar mesophilic isolate of the genus *Octadecabacter* and the type strain of the species.

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"he Roseobacter clade (family Rhodobacteraceae) is a group of exclusively marine bacteria within the alphaproteobacteria. In sea ice microbial communities (SIMCO), roseobacters are represented by the genus Octadecabacter. O. antarcticus constitutes up to 1% of the total bacterial community in southern ocean SIMCOs, while O. arcticus comprises up to 23% of the total bacterial community in Arctic SIMCOs (1). The nonpolar clusters are represented only by 16S rRNA gene sequences from uncultured organisms (2). A recently described mesophilic species has been reclassified as Pseudooctadecabacter jejudonensis (3, 4). Octadecabacter temperatus SB1 (DSM 26878) was isolated from a surface water sample collected in the harbor of the island of Helgoland in the North Sea (4). It is the first described nonpolar mesophilic isolate of the genus Octadecabacter and the type strain of the species. The strain was grown in marine broth (MB) medium at 20°C (4). Chromosomal DNA was isolated with the MasterPure complete DNA purification kit as recommended by the manufacturer (Epicentre, Madison, WI, USA). Whole-genome sequencing was performed with a combined approach using the 454 GS-FLX + pyrosequencer (Roche Life Science, Mannheim, Germany) and the Genome Analyzer IIx (Illumina, San Diego, CA, USA). Shotgun libraries for 454 sequencing as well as Nextera XT paired-end libraries for Illumina sequencing were prepared according to the manufacturer's protocols. Sequencing resulted in 54,153 pyrosequencing reads and 5,626,412 paired-end Illumina reads. Hybrid assembly of Illumina and 454 data was performed de novo with Mira v3.4.0.1 (5). Illumina data was also assembled with SPAdes v2.5.0 (6). Both assemblies were combined resulting in 8 contigs and a 158-fold coverage. Remaining gaps were closed by PCRbased techniques and Sanger sequencing of the recovered products using BigDye 3.0 chemistry on an ABI3730XL capillary sequencer (Applied Biosystems, Life Technologies GmbH, Darmstadt, Germany), and by employing Gap4 v4.11 of the Staden Package (7). The closed genome sequence of O. temperatus SB1 consists of one chromosome (3.23 Mb) and one plasmid (31.6 kb). To date this genome represents the smallest closed genome within the Roseobacter clade. Protein-encoding genes were identified and

annotated with the Prokka annotation pipeline using Prodigal v2.6 (8). The closed genome harbored 1 rRNA operon, 39 tRNA genes, and 3,284 potential protein-encoding genes, of which 2,711 have a predicted function. The plasmid encodes a replication system of the RepABC-type with highest similarity to the RepC9-subtype, which is also present in the plasmids of the polar *Octadecabacter* species (9). Moreover, genes for the biosynthesis and export of lipopolysaccharides are encoded by the plasmid. Like the polar *Octadecabacter* species, *O. temperatus* SB1 lacks a gene cluster for aerobic anoxygenic photosynthesis. In addition, genes encoding xanthorhodopsins are absent in the genome of *O. temperatus*. This is in contrast to the polar *Octadecabacter* species, which harbor such genes as putative adaption for living in icy environments (2).

Nucleotide sequence accession numbers. This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession numbers CP012160 (chromosome) and CP012161 (plasmid).

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REFERENCES

- 1. Brinkmeyer R, Knittel K, Jürgens J, Weyland H, Amann R, Helmke E. 2003. Diversity and structure of bacterial communities in Arctic versus Antarctic pack ice. Appl Environ Microbiol 69:6610–6619. http://dx.doi.org/10.1128/AEM.69.11.6610-6619.2003.
- Vollmers J, Voget S, Dietrich S, Gollnow K, Smits M, Meyer K, Brinkhoff T, Simon M, Daniel R. 2013. Poles apart: Arctic and Antarctic Octadecabacter strains share high genome plasticity and a new type of xanthorhodopsin. PLoS One 8:e63422. http://dx.doi.org/10.1371/journal-.pone.0063422.
- Park S, Yoon J-H. 2014. Octadecabacter jejudonensis sp. nov., isolated from the junction between the ocean and a freshwater spring and emended description of the genus Octadecabacter. Int J Syst Evol Microbiol 64:719–724. http://dx.doi.org/10.1099/ijs.0.057513-0.
- Billerbeck S, Orchard J, Tindall BJ, Giebel H-A, Brinkhoff T, Simon M. 2015. Description of Octadecabacter temperatus sp. nov., isolated from the

- southern North Sea, emended description of the genus *Octadecabacter* and its species and reclassification of *Octadecabacter jejudonensis* Park and Yoon 2014 as *Pseudooctadecabacter jejudonensis* gen. nov., comb. Int J Syst Evol Microbiol 65:1967–1974. http://dx.doi.org/10.1099/ijs.0.000205.
- 5. Chevreux B, Pfisterer T, Drescher B, Driesel AJ, Müller WE, Wetter T, Suhai S. 2004. Using the miraEST assembler for reliable and automated mRNA transcript assembly and SNP detection in sequenced ESTs. Genome Res 14:1147–1159. http://dx.doi.org/10.1101/gr.1917404.
- 6. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin
- AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. http://dx.doi.org/10.1089/cmb.2012.0021.
- Staden R. 1996. The Staden sequence analysis package. Mol Biotechnol 5:233–241. http://dx.doi.org/10.1007/BF02900361.
- Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. Bioinformatics 30:2068–2069. http://dx.doi.org/10.1093/bioinformatics/btu153.
- Petersen J, Brinkmann H, Pradella S. 2009. Diversity and evolution of repABC type plasmids in *Rhodobacterales*. Environ Microbiol 11: 2627–2638. http://dx.doi.org/10.1111/j.1462-2920.2009.01987.x.