

Local Drug Delivery Strategies for Glioblastoma Treatment

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Glioblastoma multiforme (GBM) is a brain tumor notorious for its malignancy. The key reason for the limited efficacy of standard treatment is the high recurrence rate of GBM, even after surgical resection. Hence, intensive postsurgical chemical therapies, such as the systemic delivery of various drugs and/or drug combinations, are typically followed after surgery. However, overcoming the blood-brain barrier by systemic administration to efficiently deliver drugs to the brain tumor remains a daunting goal. Therefore, various local drug delivery methods showing potential for improved therapeutic efficacy have been proposed. In particular, the recent application of electronic devices for the controlled delivery of chemotherapy drugs to GBM tissue has attracted attention. We herein review the recent progress of local drug delivery strategies, including electronics-assisted strategies, at the research and commercial level. We also present a brief discussion of the unsolved challenges and future research direction of localized chemotherapy methods for GBM.

Keywords

Drug administration routes; Chemotherapy; Blood-brain barrier; Brain tumor; Absorbable implants; Drug delivery systems.

INTRODUCTION

Glioblastoma multiforme (GBM) is notorious for its poor prognosis due to the high recurrence rate, even after surgical resection [1,2]. The highly infiltrative nature of GBM cells makes them difficult to completely remove by incision surgery without damaging normal brain tissue [3,4]. Consequently, adjuvant therapies including radiotherapy and chemotherapy are also administered as standard postoperative treatment; however, their therapeutic efficacy is limited by the inherent features of GBM [5,6]. Radiotherapy is hindered by radiation tolerance induced by genetic heterogeneity and frequent mutation of GBM cells [7,8]. Additionally, chemothera-

py (e.g., oral administration of temozolomide) is impeded by the blood-brain barrier (BBB), which lowers the efficiency of systemic drug delivery to GBM cells [9].

However, the systemic administration of drugs is still preferred among treatment options for GBM, as this method is easily accessible and less burdensome to patients [10]. Therefore, systemic drug delivery strategies to penetrate the BBB and increase delivery efficiency have been developed [11]. For example, nanoparticles with modified ligands capable of crossing the BBB have been suggested [12-14]. In addition, temporal opening of the BBB using external stimuli such as ultrasound has been studied to promote systemic drug delivery efficiency [15,16]. Although many studies have successfully improved systemic drug delivery efficiency, they are still insufficient to completely cure GBM. Systemic drug delivery strategies exhibit clear limitations as their long delivery routes induce a high probability of drug absorption by other organs, or drug clearance during blood circulation. Additionally, the

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risk of systemic toxicity remains a concern [17,18].

Thus, local drug delivery methods that directly administer drugs to the brain have been proposed as promising solutions. The local delivery approach can dramatically enhance delivery efficiency by bypassing the BBB. Various types of local drug delivery strategies, such as intracranial delivery [19,20], convection-enhanced delivery (CED) [21,22], and intracranial implant-based delivery [23,24], have been developed [10,25]. These macroscopic approaches have been further strengthened by the integration of microscopic approaches including the molecular design of drugs and/or incorporation of therapeutic nanomaterials. Recently, the use of electronic devices (e.g., sensors and actuators), which were already approached for the theranostic applications of brain [26-29], has also been proposed. The integration of electronics with soft and nanomaterials applied for other biomedical applications [30-33] shows the great potential of such approaches to GBM treatment. These advances in local drug delivery methods have allowed for the introduction of new perspectives on GBM treatment.

In this article, we review the recent progress in local drug delivery technologies for GBM treatment with a focus on electronics-assisted drug delivery methods. First, we introduce various conventional local drug delivery strategies, and the characteristics of commercialized drug delivery methods are described. Subsequently, representative research-level local drug delivery methods, classified into macroscopic and microscopic approaches, are presented. We then focused on studies using electronic devices for GBM treatment, a recent innovation in local drug delivery technology. Finally, we conclude this review by presenting the issues to be addressed in the future, and the developmental direction of local delivery approaches to achieve complete recovery from GBM.

LOCAL DRUG DELIVERY STRATEGIES FOR GBM TREATMENT

The local drug delivery strategy for GBM treatment has many advantages over systemic drug delivery, including superior drug delivery efficiency and minimal systemic side effects of the toxic chemotherapeutic drugs. However, it should be accompanied by surgery of the skull and dura to obtain direct access to the target site of the brain, resulting in a large physical and economic burden to the patient. To minimize this disadvantage, local drug delivery devices can be implanted during surgical resection of brain tumors, or applied through minimally invasive procedures [34,35]. This chapter comprises three subchapters on clinically available, research-level, and electronics-assisted local drug delivery strategies, and describes various local delivery strategies with explanations of

both their advantages and disadvantages.

Clinically-available local drug delivery strategies

Among the various local drug delivery strategies, the therapeutic efficacy and biocompatibility of several methods could be proven; these methods could thus be approved for clinical implementation in GBM patients. This subchapter describes these clinically available local drug delivery methods (e.g., intracranial injection, CED, and solid-state implants).

Intracranial injection of therapeutic agents can deploy therapeutic moieties directly into the target brain region. Since the drug is directly injected into the target site, this method exhibits the clear advantage of excellent drug delivery efficiency. Furthermore, the location of the injection can be controlled at a high spatial resolution using a catheter or syringe, which minimizes unwanted exposure of chemotherapeutic drugs to normal brain tissue. However, this bolus injection method still shows therapeutic efficacy below expectations owing to the rapid dissipation and backflow of the injected drugs. To overcome these shortcomings, macroscopic drug injection tools (e.g., infusion convection pump) (Fig. 1A) [36-38] or microscopic drug delivery vehicles (e.g., liposomal doxorubicin) (Fig. 1B) [39,40] have been used together. In particular, CED using a pressure gradient generated by the pump has shown a noteworthy potential to promote drug delivery to the deep brain tissue as well as improve the drug injection procedure.

Implantable solid-state intracranial drug reservoirs have been developed for drug release over a longer period of time; they can significantly increase the drug loading capacity, and the nonfluidic property of the drug reservoir prevents rapid drug dissipation. However, this method requires a surgical opening of the skull for implantation of the solid-state device; thus, they are usually implanted during the surgical resection of tumors.

One representative example of solid-state implants is the Gliadel wafer (Arbor Pharmaceuticals, LLC, Atlanta, GA, USA) (Fig. 1C) [24,41,42]. It includes a mixture of biodegradable polymers and alkylating agents (i.e., carmustine; 1,3-bis(2-chloroethyl)-1-nitrosourea) and is implanted in the cavity after resection of the brain tumor. After implantation, the drug is released into the brain tissue from the Gliadel wafer by natural diffusion, preventing GBM recurrence. However, its rigid cylindrical structure has a disadvantage in its application, as the drug released from the wafer is often washed away by the cerebrospinal fluid (CSF) that permeates the gap between the wafer and curved brain surface. This can diminish drug delivery efficiency and therapeutic efficacy. In addition, potential inflammatory issues exist due to the mechanical mismatch between rigid wafers and soft brain tissue.

The Ommaya reservoir (Fig. 1D), another type of solid-state

implant, provides a conduit for drug administration from outside the brain [43,44]. The device, made of soft plastic, consists of a reservoir dome located on the brain surface and a

catheter that penetrates brain tissue. The drug can be periodically administered to the target region via the Ommaya reservoir. Consequently, the type and dosage of drugs can be

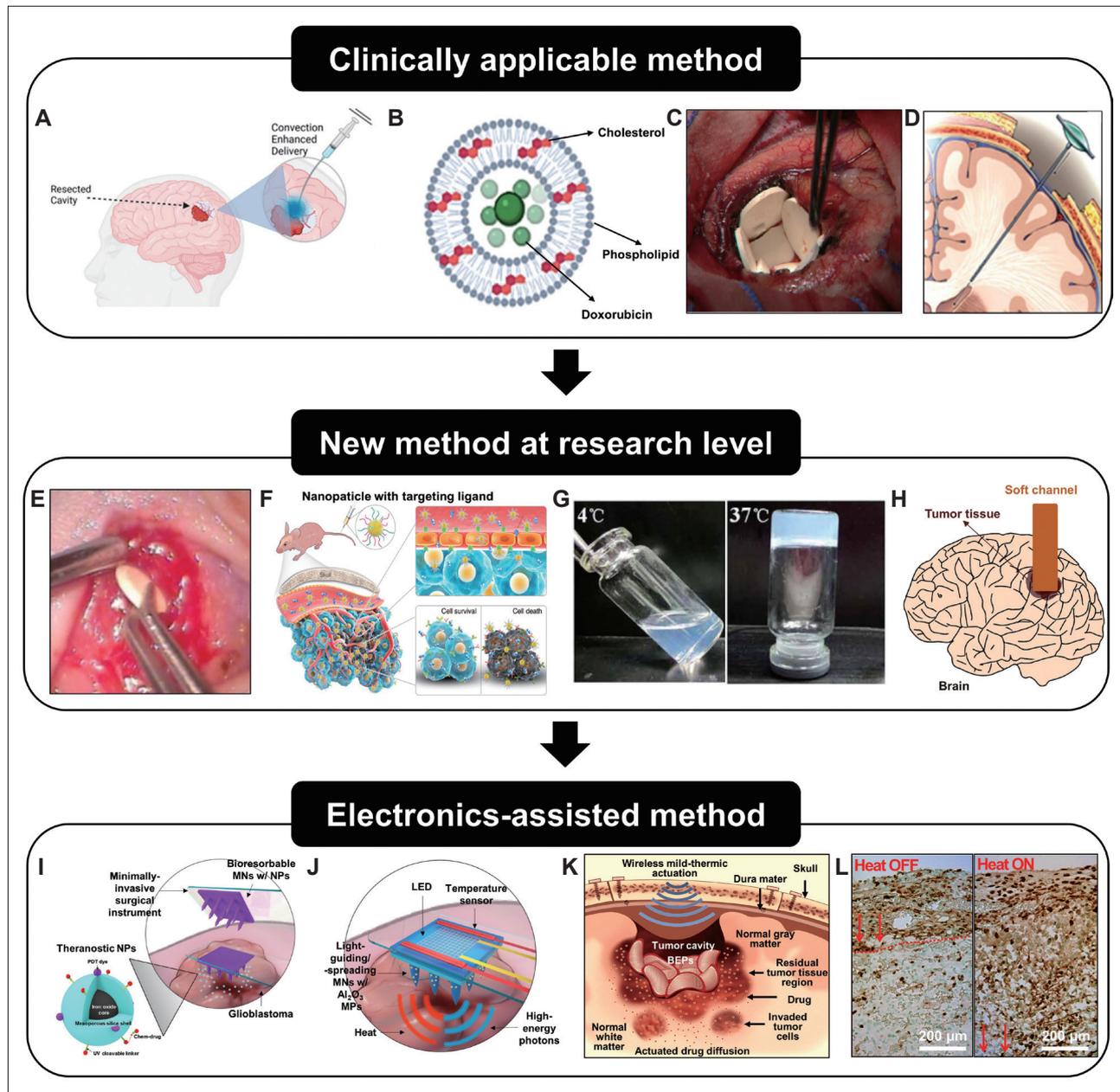


Fig. 1. Local drug delivery strategies. A: Schematic illustration of convection enhanced drug delivery compared to the normal diffusion method. B: Schematic illustration of the structure of the liposomal doxorubicin. C: Optical image of the Gliadel wafer implanted on the cavity after resection. D: Schematic illustration of the Ommaya reservoir implanted into the brain. E: Optical image of a nanofiber wafer implanted on the brain. F: Schematic illustration of the administration of the therapeutic nanoparticles conjugated with targeting moieties for the glioblastoma multi-forme treatment. G: Optical image of the GemC₁₂-lipid nanocapsules hydrogel implanted on the cavity after resection. H: Schematic illustration and optical image (inset) of the tumor-guiding conduit implanted on the tumor tissue in the brain. I: Schematic illustration of the delivery of theranostic nanoparticles (NPs) using bioresorbable microneedles (MNs). J: Schematic illustration of the delivery of the high-energy photons using light-guiding microneedles with microparticles (MPs), light-emitting diode (LED), and bioelectronics. K: Schematic illustration of drug delivery by the bioresorbable electronic patch (BEP) with magnetic actuation. L: Optical image of tumor tissues without (left) and with (right) the magnetic actuation. A: Adapted from Pena et al. *Int J Mol Sci* 2021;22:13160 [38]; B: Adapted from Ibrahim et al. *Pharmaceutics* 2022;14:254 [40]; C: Adapted from Kleinberg. *Patient Prefer Adherence* 2016;10:2397-406 [42]; D: Adapted from Lau et al. *Cureus* 2012;4:e66 [43]; E: Adapted from Ramachandran et al. *Sci Rep* 2017;7:43271 [48]; F: Adapted from Meng et al. *Nat Commun* 2020;11:594 [50]; G: Adapted from Shi et al. *Sci Rep* 2016;6:19077 [52]; K and L: Adapted from Lee et al. *Nat Commun* 2019;15:10:5205 [58]; under the Creative Commons license. I and J: Adapted from Lee et al. *Adv Mater* 2021;33:2100425, with permission from John Wiley and Sons [57].

changed to suit the patient's condition during treatment. These advantages, achieved by adopting the Ommaya reservoir, are suitable for the long-term treatment of GBM. Furthermore, it can perform additional functions—such as CSF perfusion—which may help to monitor unexpected side effects by observing the status of the CSF. However, several side effects (including inflammation and infection) may occur due to prolonged exposure of the brain to the external environment via the Ommaya reservoir [45]. Although the small size of the device (e.g., the size of a quarter) minimizes the inconvenience of patients equipped with the Ommaya reservoir, they may face uncomfortable situations in their daily lives due to the permanent installation of the external device.

Research-level local drug delivery strategies

Clinically available local drug delivery strategies have shown significant improvements in treatment efficacy. However, further improvements are still required, as the median survival can only be extended by several months. Hence, various studies on novel and/or improved local drug delivery strategies have been conducted to enhance the therapeutic effect.

One research direction involves changing the materials used in conventional drug delivery devices. For example, solid-state drug-releasing implant (e.g., Gliadel wafer) can be replaced by soft drug-releasing materials, such as hydrogels [46,47] and fibers [48,49] (Fig. 1E). As soft and deformable material properties are compatible with soft brain tissue and curved surfaces, they can be advantageous for enhancing delivery efficiency and decreasing potential side effects. For example, the softness of implants prevents neurological side effects (e.g., seizure) induced by the mechanical mismatch between soft brain tissue and rigid implants. Furthermore, deformable features enable conformal integration of the drug delivery implant with curvilinear brain surface.

Another direction is to introduce nanotechnology into GBM treatment. Nanoparticles have been suggested as drug carriers for enhancing drug delivery efficiency by encapsulating the drugs and decorating the nanoparticle surface with functional moieties (Fig. 1F) [50]. To enhance the specificity of drug delivery, a tumor-targeting or stimuli-responsive moiety (e.g., pH-responsive group) is conjugated to the surface of the nanoparticles (Fig. 1F). Additionally, other functional moieties that could control drug release behavior (e.g., control of the degradation rate) or facilitate drug penetration (e.g., PEGylation) were studied. These approaches can be integrated with other macroscopic approaches to achieve synergistic therapeutic efficacy.

A new therapeutic platform with optimized administration methods has been reported. For example, a thermo-responsive injectable hydrogel, which can be administered to the

brain via a minimally-invasive route, has been employed for GBM treatment (Fig. 1G) [51,52]. When implanted, it gels in response to body temperature without any external energy and can be fixed to the target site. These injectable-type implants can offset the shortcomings of solid-state devices that require incision surgery for implantation. The other advantages of hydrogels (e.g., softness, deformability, and high drug dose) [53,54] for GBM treatment are also valid for soft drug-releasing implants based on injectable hydrogels.

Another approach is to exploit soft materials as a migration route that induces the movement of tumor cells to the drug reservoir [55]. This soft device comprises aligned nanofibers and polymers as channels, and a hydrogel as the drug reservoir. For treatment, the channel is inserted directly into the tumor, serving as a conduit between the hydrogel drug reservoir and tumor tissue (Fig. 1H). Owing to the aligned nanostructure, tumor cells preferentially move along the implanted conduit. Finally, these cells reach the drug reservoir and undergo apoptosis. Since all material components of the channel feature softness and biocompatibility, the suggested platform is relatively free from long-term safety issues when compared to platforms using a similar administration method, such as the Ommaya reservoir.

Electronics-assisted local drug delivery strategies

Several strategies have been suggested to improve the efficacy of GBM treatment. However, these efforts have not yet achieved the desired efficacy. New functions and capabilities, such as real-time sensing and meticulous drug delivery control, may further improve therapeutic efficiency. Several recent studies have presented novel drug delivery platforms that integrate flexible and/or biodegradable electronics with drug delivery nanoparticles or drug-releasing reservoirs [56].

For example, Lee et al. [57] presented a minimally-invasive local drug delivery and therapy platform comprising two types of microneedles integrated with bioelectronics. One was a bioresorbable microneedle containing theranostic nanoparticles for drug delivery and the generation of therapeutic reactive oxygen species (ROS) (Fig. 1I); the other was a light-guiding microneedle containing light-scattering alumina microparticles integrated with flexible electronics, containing an ultraviolet light-emitting diode (LED) and temperature sensor. When the bioresorbable microneedles dissolved, the nanoparticles were delivered to the target tumor tissue. Ultraviolet light generated by the LED then activates the nanoparticles to release the loaded chemotherapeutic drugs and generate ROS for photodynamic therapy (Fig. 1J). In addition, the LED generates heat, which accelerates the diffusion of the delivered/generated therapeutic moieties. The temperature increase is monitored by a temperature sensor; thus, thermal

overshoot can be prevented.

Despite the potential of electronics-assisted local drug delivery as a minimally invasive treatment tool, its therapeutic efficacy remains insufficient. The major cause is the noncontinuous, rather than continuous long-term treatment protocol. Continuous treatment can be performed using implantable patches with onboard biodegradable electronics. For example, a wirelessly controllable bioresorbable electronic patch (BEP) was developed and implanted inside the surgical brain cavity for long-term drug delivery and tumor treatment [58]. The heater on the BEP was wirelessly actuated by an external magnetic field, promoting drug diffusion to deep brain regions (Fig. 1K). As a result, deeply infiltrated tumor cells were treated (Fig. 1L). Such actuation can be periodically conducted until the biodegradation of the heater, and subsequent drug delivery from the BEP is carried out via natural diffusion of the remaining drugs. Furthermore, a second surgery to remove the implanted device is not required, as the BEP is biodegradable. Owing to these advantages, BEP may show improved therapeutic efficacy in mouse/canine tumor models.

REMAINING CHALLENGES AND FUTURE OUTLOOK

Although local drug delivery strategies have shown progress in GBM treatment, complete recovery from GBM remains a daunting goal. The major cause of GBM recurrence is the infiltration of tumor cells, and although several electronics-assisted strategies have attempted to counter this challenge, the penetration/diffusion depth of the drug into brain tissues is still limited. Other methods, such as CED, exhibit better delivery efficiency; however, liquid-state drugs dissipate rapidly and thus often result in low treatment efficiency. While the electronics-assisted drug delivery method can potentially deliver drugs to the deep brain region over a much longer period, both the drug penetration depth and delivery period should be further improved. Additionally, the biocompatibility and biodegradability of implantable devices should be considered.

Many practical issues must be resolved for clinical translation. For example, since each GBM case exhibits various characteristics owing to the highly pathological and genetic heterogeneity of GBM, the applicability of the platform device for a broad range of personalized therapeutic moieties and drugs should be guaranteed. Furthermore, the treatment efficacy should be verified in GBM models of large animals. At present, small animal models are mostly used, and a large animal model test is critical before human applications. Thus, the development of reliable GBM models in large animals is required. The biocompatibility of implantable platforms should

also be systematically examined in a larger number of subjects. For clinical applications, additional protocols must be developed to increase friendliness of the patients and reduce the economic burden on patients. Utilizing new digital technologies, such as wearable and/or implantable bioelectronics connected to wireless networks, may provide new opportunities. Despite the many remaining challenges, there is potential for the development of novel treatment strategies to make a breakthrough in GBM treatment.

Ethics Statement

Not applicable

Availability of Data and Material

Data sharing not applicable to this article as no datasets were generated or analyzed during the study.

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Conflicts of Interest

The authors have no potential conflicts of interest to disclose.

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REFERENCES

1. Das M. Iniparib for newly diagnosed glioblastoma. *Lancet Oncol* 2018; 19:e514.
2. Tran B, Rosenthal MA. Survival comparison between glioblastoma multiforme and other incurable cancers. *J Clin Neurosci* 2010;17:417-21.
3. Holland EC. Glioblastoma multiforme: the terminator. *Proc Natl Acad Sci U S A* 2000;97:6242-4.
4. Das S, Marsden PA. Angiogenesis in glioblastoma. *N Engl J Med* 2013; 369:1561-3.
5. Komotar RJ, Otten ML, Moise G, Connolly ES Jr. Radiotherapy plus concomitant and adjuvant temozolomide for glioblastoma—a critical review. *Clin Med Oncol* 2008;2:421-2.
6. Ganipineni LP, Danhier F, Pr at V. Drug delivery challenges and future of chemotherapeutic nanomedicine for glioblastoma treatment. *J Control Release* 2018;281:42-57.
7. Jung E, Osswald M, Ratliff M, Dogan H, Xie R, Weil S, et al. Tumor cell plasticity, heterogeneity, and resistance in crucial microenvironmental niches in glioma. *Nat Commun* 2021;12:1014.
8. Dirkse A, Golebiewska A, Buder T, Nazarov PV, Muller A, Poovathingal S, et al. Stem cell-associated heterogeneity in glioblastoma results from intrinsic tumor plasticity shaped by the microenvironment. *Nat Commun* 2019;10:1787.

9. Abbott NJ, Rönnbäck L, Hansson E. Astrocyte-endothelial interactions at the blood-brain barrier. *Nat Rev Neurosci* 2006;7:41-53.
10. Cha GD, Kang T, Baik S, Kim D, Choi SH, Hyeon T, et al. Advances in drug delivery technology for the treatment of glioblastoma multiforme. *J Control Release* 2020;328:350-67.
11. Xue J, Zhao Z, Zhang L, Xue L, Shen S, Wen Y, et al. Neutrophil-mediated anticancer drug delivery for suppression of postoperative malignant glioma recurrence. *Nat Nanotechnol* 2017;12:692-700.
12. Wilhelm S, Tavares A, Dai Q, Ohta S, Audet J, Dvorak HF, et al. Analysis of nanoparticle delivery to tumours. *Nat Rev Mater* 2016;1:16014.
13. Ruan S, Hu C, Tang X, Cun X, Xiao W, Shi K, et al. Increased gold nanoparticle retention in brain tumors by in situ enzyme-induced aggregation. *ACS Nano* 2016;10:10086-98.
14. Shen Z, Liu T, Li Y, Lau J, Yang Z, Fan W, et al. Fenton-reaction-accelerated magnetic nanoparticles for ferroptosis therapy of orthotopic brain tumors. *ACS Nano* 2018;12:11355-65.
15. Wen L, Wang K, Zhang F, Tan Y, Shang X, Zhu Y, et al. AKT activation by SC79 to transiently re-open pathological blood brain barrier for improved functionalized nanoparticles therapy of glioblastoma. *Biomaterials* 2020;237:119793.
16. Alli S, Figueiredo CA, Golbourn B, Sabha N, Wu MY, Bondoc A, et al. Brainstem blood brain barrier disruption using focused ultrasound: a demonstration of feasibility and enhanced doxorubicin delivery. *J Control Release* 2018;281:29-41.
17. Nam L, Coll C, Erthal LCS, de la Torre C, Serrano D, Martínez-Máñez R, et al. Drug delivery nanosystems for the localized treatment of glioblastoma multiforme. *Materials (Basel)* 2018;11:779.
18. Jain KK. A critical overview of targeted therapies for glioblastoma. *Front Oncol* 2018;8:419.
19. Nance E, Zhang C, Shih TY, Xu Q, Schuster BS, Hanes J. Brain-penetrating nanoparticles improve paclitaxel efficacy in malignant glioma following local administration. *ACS Nano* 2014;8:10655-64.
20. Kim DG, Kim KH, Seo YJ, Yang H, Marcusson EG, Son E, et al. Anti-miR delivery strategies to bypass the blood-brain barrier in glioblastoma therapy. *Oncotarget* 2016;7:29400-11.
21. Zhang C, Nance EA, Mastorakos P, Chisholm J, Berry S, Eberhart C, et al. Convection enhanced delivery of cisplatin-loaded brain penetrating nanoparticles cures malignant glioma in rats. *J Control Release* 2017;263:112-9.
22. Stephen ZR, Kievit FM, Veisheh O, Chiarelli PA, Fang C, Wang K, et al. Redox-responsive magnetic nanoparticle for targeted convection-enhanced delivery of O6-benzylguanidine to brain tumors. *ACS Nano* 2014;8:10383-95.
23. Han D, Serra R, Gorelick N, Fatima U, Eberhart CG, Brem H, et al. Multi-layered core-sheath fiber membranes for controlled drug release in the local treatment of brain tumor. *Sci Rep* 2019;9:17936.
24. Colen RR, Zinn PO, Hazany S, Do-Dai D, Wu JK, Yao K, et al. Magnetic resonance imaging appearance and changes on intracavitary Gliadel wafer placement: a pilot study. *World J Radiol* 2011;3:266-72.
25. van Solinge TS, Nieland L, Chiocca EA, Broekman MLD. Advances in local therapy for glioblastoma - taking the fight to the tumour. *Nat Rev Neurol* 2022;18:221-36.
26. Lee WH, Cha GD, Kim DH. Flexible and biodegradable electronic implants for diagnosis and treatment of brain diseases. *Curr Opin Biotechnol* 2021;72:13-21.
27. Sunwoo SH, Han SI, Joo H, Cha GD, Kim D, Choi SH, et al. Advances in soft bioelectronics for brain research and clinical neuroengineering. *Matter* 2020;3:1923-47.
28. Joo H, Lee Y, Kim J, Yoo JS, Yoo S, Kim S, et al. Soft implantable drug delivery device integrated wirelessly with wearable devices to treat fatal seizures. *Sci Adv* 2021;7:eabd4639.
29. Koo JH, Song JK, Kim DH, Son D. Soft implantable bioelectronics. *ACS Mater Lett* 2021;3:1528-40.
30. Cho KW, Sunwoo SH, Hong YJ, Koo JH, Kim JH, Baik S, et al. Soft bioelectronics based on nanomaterials. *Chem Rev* 2022;122:5068-143.
31. Koo JH, Song JK, Yoo S, Sunwoo SH, Son D, Kim DH. Unconventional device and material approaches for monolithic biointegration of implantable sensors and wearable electronics. *Adv Mater Technol* 2020;5:202000407.
32. Yoo S, Lee J, Joo H, Sunwoo SH, Kim S, Kim DH. Wireless power transfer and telemetry for implantable bioelectronics. *Adv Healthc Mater* 2021;10:2100614.
33. Sunwoo SH, Ha KH, Lee S, Lu N, Kim DH. Wearable and implantable soft bioelectronics: device designs and material strategies. *Annu Rev Chem Biomol Eng* 2021;12:359-91.
34. Lesniak MS, Brem H. Targeted therapy for brain tumours. *Nat Rev Drug Discov* 2004;3:499-508.
35. Mathios D, Kim JE, Mangraviti A, Phallen J, Park CK, Jackson CM, et al. Anti-PD-1 antitumor immunity is enhanced by local and abrogated by systemic chemotherapy in GBM. *Sci Transl Med* 2016;8:370ra180.
36. Allard E, Passirani C, Benoit JP. Convection-enhanced delivery of nanocarriers for the treatment of brain tumors. *Biomaterials* 2009;30:2302-18.
37. Wang Y, Jiang Y, Wei D, Singh P, Yu Y, Lee T, et al. Nanoparticle-mediated convection-enhanced delivery of a DNA intercalator to gliomas circumvents temozolomide resistance. *Nat Biomed Eng* 2021;5:1048-58.
38. Pena ES, Graham-Gurysh EG, Bachelder EM, Ainslie KM. Design of biopolymer-based interstitial therapies for the treatment of glioblastoma. *Int J Mol Sci* 2021;22:13160.
39. Engelberth SA, Hempel N, Bergkvist M. Development of nanoscale approaches for ovarian cancer therapeutics and diagnostics. *Crit Rev Oncog* 2014;19:281-315.
40. Ibrahim M, Abuwatfa WH, Awad NS, Sabouni R, Hussein GA. Encapsulation, release, and cytotoxicity of doxorubicin loaded in liposomes, micelles, and metal-organic frameworks: a review. *Pharmaceutics* 2022;14:254.
41. Westphal M, Hilt DC, Bortey E, Delavault P, Olivares R, Warnke PC, et al. A phase 3 trial of local chemotherapy with biodegradable carmustine (BCNU) wafers (Gliadel wafers) in patients with primary malignant glioma. *Neuro Oncol* 2003;5:79-88.
42. Kleinberg L. Polifeprosan 20, 3.85% carmustine slow release wafer in malignant glioma: patient selection and perspectives on a low-burden therapy. *Patient Prefer Adherence* 2016;10:2397-406.
43. Lau D, Rowland N, Devasagaya S, McDermott MW. Recession of Ommaya Reservoir improves cosmesis in patients undergoing intrathecal chemotherapy for leptomeningeal disease: a technical note. *Cureus* 2012;4:e66.
44. Magill ST, Choy W, Nguyen MP, McDermott MW. Ommaya reservoir insertion: a technical note. *Cureus* 2020;12:e7731.
45. Szvalb AD, Raad II, Weinberg JS, Suki D, Mayer R, Viola GM. Ommaya reservoir-related infections: clinical manifestations and treatment outcomes. *J Infect* 2014;68:216-24.
46. Bastiancich C, Bozzato E, Luyten U, Danhier F, Bastiat G, Pr at V. Drug combination using an injectable nanomedicine hydrogel for glioblastoma treatment. *Int J Pharm* 2019;559:220-7.
47. Zhang J, Chen C, Li A, Jing W, Sun P, Huang X, et al. Immunostimulant hydrogel for the inhibition of malignant glioma relapse post-resection. *Nat Nanotechnol* 2021;16:538-48.
48. Ramachandran R, Junnuthula VR, Gowd GS, Ashokan A, Thomas J, Peethambaran R, et al. Theranostic 3-dimensional nano brain-implant for prolonged and localized treatment of recurrent glioma. *Sci Rep* 2017;7:43271.
49. Di Mascolo D, Palange AL, Primavera R, Macchi F, Catelani T, Piccardi F, et al. Conformable hierarchically engineered polymeric micromeshes enabling combinatorial therapies in brain tumours. *Nat Nanotechnol* 2021;16:820-9.
50. Meng X, Zhao Y, Han B, Zha C, Zhang Y, Li Z, et al. Dual functionalized brain-targeting nanoinhibitors restrain temozolomide-resistant glioma via attenuating EGFR and MET signaling pathways. *Nat Commun* 2020;11:594.

51. Bastiancich C, Bianco J, Vanvarenberg K, Ucakar B, Joudiou N, Gallez B, et al. Injectable nanomedicine hydrogel for local chemotherapy of glioblastoma after surgical resection. *J Control Release* 2017;264:45-54.
52. Shi K, Wang YL, Qu Y, Liao JF, Chu BY, Zhang HP, et al. Synthesis, characterization, and application of reversible PDLLA-PEG-PDLLA copolymer thermogels in vitro and in vivo. *Sci Rep* 2016;6:19077.
53. Cha GD, Lee WH, Sunwoo SH, Kang D, Kang T, Cho KW, et al. Multi-functional injectable hydrogel for in vivo diagnostic and therapeutic applications. *ACS Nano* 2022;16:554-67.
54. Cha GD, Lee WH, Lim C, Choi MK, Kim DH. Materials engineering, processing, and device application of hydrogel nanocomposites. *Nanoscale* 2020;12:10456-73.
55. Jain A, Betancur M, Patel GD, Valmikinathan CM, Mukhatyar VJ, Vakharia A, et al. Guiding intracortical brain tumour cells to an extra-cortical cytotoxic hydrogel using aligned polymeric nanofibres. *Nat Mater* 2014;13:308-16.
56. Cha GD, Kang D, Lee J, Kim DH. Bioresorbable electronic implants: history, materials, fabrication, devices, and clinical applications. *Adv Healthc Mater* 2019;8:1801660.
57. Lee Y, Kang T, Cho HR, Lee GJ, Park OK, Kim S, et al. Localized delivery of theranostic nanoparticles and high-energy photons using microneedles-on-bioelectronics. *Adv Mater* 2021;33:2100425.
58. Lee J, Cho HR, Cha GD, Seo H, Lee S, Park CK, et al. Flexible, sticky, and biodegradable wireless device for drug delivery to brain tumors. *Nat Commun* 2019;15:10:5205.