



ELSEVIER

Contents lists available at ScienceDirect

## Data in Brief

journal homepage: [www.elsevier.com/locate/dib](http://www.elsevier.com/locate/dib)

CrossMark

# Data on the identity and myristoylation state of recombinant, purified hippocalcin

Anuradha Krishnan<sup>a,1</sup>, Jeffrey Viviano<sup>a,1</sup>, Yaroslav Morozov<sup>a</sup>, Venkat Venkataraman<sup>a,b,\*</sup>

<sup>a</sup> Graduate School of Biomedical Sciences, USA

<sup>b</sup> School of Osteopathic Medicine Rowan University, Stratford, NJ 08084, USA

## ARTICLE INFO

### Article history:

Received 18 March 2016

Received in revised form

4 April 2016

Accepted 7 April 2016

Available online 20 May 2016

### Keywords:

Hippocalcin

Calcium

NCS proteins

MALDI-TOF

ISD sequencing

Myristoylation

## ABSTRACT

In this data article we report on the purity and post translation modification of bacterially expressed and purified recombinant hippocalcin (HPCA): a member of the neuronal calcium sensor protein family, whose functions are regulated by calcium. MALDI-TOF in source decay (ISD) analysis was used to identify both the myristoylated or non-myristoylated forms of the protein. MALDI-TOF ISD data on the identity of the protein, amino acid sequence and myristoylation efficiency are provided. This data relates to the article “Single-Column Purification of the Tag-free, Recombinant Form of the Neuronal Calcium Sensor Protein, Hippocalcin Expressed in *Escherichia coli*” [1].

© 2016 Published by Elsevier Inc. This is an open access article under the CC BY license

(<http://creativecommons.org/licenses/by/4.0/>).

## Specifications Table

Subject area	Biology
More specific subject area	Protein Identification
Type of data	Table, figure
How data was acquired	In Set Decay Mass Spectrometry (MALDI-TOF ISD), DNA Sequencing

DOI of original article: <http://dx.doi.org/10.1016/j.pep.2016.03.005>

\* Corresponding author.

E-mail address: [venkatar@rowan.edu](mailto:venkatar@rowan.edu) (V. Venkataraman).

<sup>1</sup> These authors contributed equally.

<http://dx.doi.org/10.1016/j.dib.2016.04.024>

2352-3409/© 2016 Published by Elsevier Inc. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

Data format	Analyzed
Experimental factors	For MALDI-TOF ISD, Standard protocols were used
Experimental features	Purified protein was analyzed by MALDI-TOF ISD
Data source location	Stratford, New Jersey, USA
Data accessibility	Data contained within this article

Value of the data

- Confirms the identity of the bacterially expressed HPCA purified in a single step.
- Demonstrates the ability to properly myristoylate the bacterially expressed HPCA with high efficiency
- Identifies the first amino acid residue of expressed HPCA
- Provides a benchmark approach to characterizing critical aspects such as myristoylation in bacterially expressed neuronal calcium sensor proteins in particular and modified proteins in general.

1. Data

Purified HPCA was analyzed through mass spectrometry. MALDI-TOF ISD analyses were independently carried out with the myristoylated and non-myristoylated forms of HPCA. Table 1 displays the sequence of the first 8 fragments identified by ISD. The difference between non-myristoylated and myristoylated forms, as expected, is 210 Da. Data presented in Fig. 1 confirms the identity of the expressed protein, derived from the cDNA sequence as well as through MALDI-TOF ISD (underlined sequence) as HPCA [2,3]. Together, the data demonstrate the loss of the first methionine (in grey; Fig. 1) in the purified protein.

2. Experimental design, materials and methods

HPCA was purified as previously described [1]. Five µg of myristoylated or non-myristoylated HPCA was desalted using C4 ZipTip (Millipore Inc.). The sample was then mixed 1:2 with saturated 1,5-diaminonaphthalene in 50% acetonitrile and 0.1% TFA in water and spotted on the MALDI target plate. In-source decay (ISD) data was collected using Bruker MicroFlex LFR MALDI-TOF in positive linear mode. Mass range was set from 1000 to 7000 Da and the pulse ion extraction was set at 240 ns. ISD spectra were analyzed with Flex Analysis software (Bruker).

**Table 1**  
N-terminal fragments generated from myristoylated and non-myristoylated HPCA.

Fragment	Molecular Weight		
	Myr <sup>-</sup>	Myr <sup>+</sup>	Difference
GKQNSKLRP	1030.0	1240.5	210.5
GKQNSKLRPE	1159.9	1369.9	210.0
GKQNSKLRPEM	1291.1	1501.4	210.3
GKQNSKLRPEML	1404.3	1614.2	209.9
GKQNSKLRPEMLQ	1533.5	1742.3	208.8
GKQNSKLRPEMLQD	1647.7	1857.5	209.8
GKQNSKLRPEMLQDL	1760.7	1970.0	209.3
GKQNSKLRPEMLQDLR	1916.9	2126.6	209.7

```

      M G K Q N S K L R P E M L Q D L R E N T
1  atgggcaagcagaatagcaagctgcgccagagatgctgcaggacctgcagagagaacacc 60

      E F S E L E L Q E W Y K G F L K D C P T
61  gagttctctgagctggagcttcaggagtgtacaagggttcctgaaggactgcccgact 120

      G I L N V D E F K K I Y A N F F P Y G D
121  ggcatcctcaacgtggatgagttcaagaagatctacgccaacttcttcccctacggcgat 180

      A S K F A E H V F R T F D T N S D G T I
181  gcctccaagttcgcgagcatgtcttcgcacttttgacaccaacagcgacggcaccatc 240

      D F R E F I I A L S V T S R G R L E Q K
241  gacttcgggagttcatcatcgctctgagcgtgacctcgctggccgcctggagcagaag 300

      L M W A F S M Y D L D G N G Y I S R E E
301  ctcagtgtggccttcagcatgtacgacctggacggcaatggctacatcagccgggaggag 360

      M L E I V Q A I Y K M V S S V M K M P E
361  atgctagaaattgtgcaggccatttacaagatggtttcgtccgtgatgaagatgcctgag 420

      D E S T P E K R T E K I F R Q M D T N N
421  gatgagtctaccccggaagaggactgagaaaatcttcgccaaatggacacaaacaat 480

      D G K L S L E E F I R G A K S D P S I V
481  gacggcaagctgtcactggaggagttcatccgcggggccaaaagcgacccatcgatcgtg 540

      R L L Q C D P S S A S Q F *
541  cgctgtgtgcaatgcatcccgagcagcgcttccagttctga 582

```

Fig. 1. Sequence of expressed HPCA.

Rat HPCA coding region was amplified by PCR and inserted into the bacterial expression vector pET 21d between NcoI and HindIII sites. Sequencing of the construct was performed in both directions (GeneWiz Inc.).

## Acknowledgments

The work was supported by grants from the New Jersey Health (previously UMDNJ) Foundation and the Osteopathic Heritage Foundation. The support by the Rowan SOM Graduate School of Biomedical Sciences is also acknowledged. Dr. Mikhail Anikin is gratefully acknowledged for his help with the MALDI-TOF ISD measurements.

## Appendix A. Supplementary material

Supplementary data associated with this article can be found in the online version at <http://dx.doi.org/10.1016/j.dib.2016.04.024>.

## References

- [1] A. Krishnan, J.M. Viviano, Y. Morozov, V. Venkataraman, Single-column purification of the tag-free, recombinant form of the neuronal calcium sensor protein, hippocalcin expressed in *Eschericia coli*, Protein Expr. Purif. (2016), <http://dx.doi.org/10.1016/j.pep.2016.03.005>.
- [2] K. Takamatsu, M. Kobayashi, S. Saitoh, M. Fujishiro, T. Noguchi, Molecular cloning of human hippocalcin cDNA and chromosomal mapping of its gene, Biochem. Biophys. Res. Commun. 200 (1) (1994) 606–611.
- [3] A. Krishnan, T. Duda, A. Pertzev, M. Kobayashi, K. Takamatsu, R.K. Sharma, Hippocalcin, new  $\text{Ca}(2+)$  sensor of a ROS-GC subfamily member, ONE-GC, membrane guanylate cyclase transduction system, Mol. Cell. Biochem. 325 (1–2) (2009) 1–14.