

COMMENT



<https://doi.org/10.1038/s41467-022-30477-3>

OPEN

Metabolic regulation by the intestinal metformin-AMPK axis

Song-Yang Zhang¹ & Tony K. T. Lam^{1,2,3,4}  

AMP-activated protein kinase (AMPK) mediates the glucose-lowering effect of the antidiabetic agent metformin, but the sites of action remain unclear. In the March issue of *Nature Communications*, Zhang and colleagues reported that intestinal epithelium-specific AMPK α 1 knockout mice fail to respond to metformin and exhibit disruption in metabolic homeostasis secondary to changes in the gut microbiome. This highlights a therapeutic potential of targeting intestinal AMPK for diabetes.

Metformin is a first-line glucose-lowering agent for people with obesity-associated type 2 diabetes, but the underlying mechanisms remains elusive. The liver has been described to mediate metformin action through AMPK-dependent and independent pathways^{1,2}, but emerging studies highlight the gut as a target of metformin as well³. For example, oral delivery of metformin in rats and humans that targets the gut lowers plasma glucose levels independently of changes in plasma metformin levels^{4,5}, while direct delivery of metformin into the upper small intestine of rats and humans also lower plasma glucose levels in diabetic conditions^{6,7}. Thus, studies are urgently needed to identify molecular targets in the gut that are sufficient and necessary for metformin to regulate metabolic homeostasis.

Glucose homeostasis and energy balance in rodents and humans are regulated by nutrient sensing mechanisms in the small intestine⁸. AMPK is expressed in the small intestinal mucosa⁶ and, together with hepatic AMPK, is a known target of metformin. These findings raise the question of whether intestinal AMPK activation is sufficient and necessary for metformin to not only regulate glucose homeostasis, but also energy intake and expenditure as well as body weight in rodents and humans. In this regard, metformin has been documented to activate AMPK in the small intestine and to acutely and rapidly lower hepatic glucose production and plasma glucose levels in high-fat fed, obese and/or diabetic male rats⁶. But, whether small intestinal AMPK can exert metabolic control beyond glucose homeostasis such as energy intake and expenditure, and whether the gut metformin-AMPK axis has long-term effects on glucose levels and body weight in rodents and humans warrant investigation.

Phenotypes of intestinal-specific AMPK α 1 knockout mice

In March, *Nature Communications* published the work of Zhang and colleagues⁹, who generated intestinal epithelium-specific AMPK α 1 knockout mice and found that after 6 weeks of high fat feeding and in comparison to wild-type mice, these mice exhibit weight gain that is independent of hyperphagia, accompanied by impaired glucose tolerance, as assessed by a intraperitoneal glucose

¹Toronto General Hospital Research Institute, UHN, Toronto, ON, Canada. ²Department of Physiology, University of Toronto, Toronto, ON, Canada.

³Department of Medicine, University of Toronto, Toronto, ON, Canada. ⁴Banting and Best Diabetes Centre, University of Toronto, Toronto, ON, Canada.

✉email: tony.lam@uhnresearch.ca

injection test. The disruption of glucose tolerance was found to be independent of weight gain but occurred in parallel to increased expression of hepatic genes that regulate gluconeogenesis⁹. Importantly, once daily (100 mg/kg) oral metformin administration for 8 weeks lowered weight and increased glucose tolerance independent of weight changes in 6 week high-fat fed wild-type but not in intestinal AMPK α 1 knockout mice⁹. These studies highlight that metformin activates intestinal AMPK to improve glucose tolerance and lower body weight together with its lowering effect on hepatic glucose production^{6,9}, and chronic inhibition of intestinal AMPK is sufficient to dysregulate glucose homeostasis and induce obesity (Fig. 1). Although the clinical relevance remains to be directly tested, preliminary findings indicate that AMPK activity was reduced in the upper small intestine of people with obesity-associated type 2 diabetes as well⁹.

An equally important discovery was that intestinal AMPK α 1 knockout mice on chow diet develop adipocyte hypertrophy with a downregulation of the brown fat thermogenic program (i.e., reduced UCP1 expression) and a subsequent reduction in energy expenditure⁹, that likely resulted in greater obesity upon high fat feeding. The effect of intestinal AMPK α 1 knockout mice on the brown fat thermogenic program was reproduced by Zhang et al. in mice that received fecal microbiota transplant from intestinal AMPK α 1 knockout mice, altogether demonstrating a novel intestinal AMPK-microbiome-brown fat axis that regulate energy expenditure (Fig. 1). Consistent with the fact that microbiota-derived metabolite methylglyoxal is higher in people with diabetes, methylglyoxal was higher in the serum, fecal and brown fat of high-fat fed intestinal-specific AMPK α 1 knockout vs. wild-type mice⁹. In fact, methylglyoxal administration reduced brown fat UCP1 expression in vitro and in vivo⁹, but whether changes in methylglyoxal is responsible for the reduction in energy expenditure in intestinal-specific AMPK α 1 knockout mice remain unclear (Fig. 1). In addition, the specific microbes families and/or species that were involved remain unknown, although the relative abundance of the *Lachnospiraceae* family was significantly lower⁹. In summary, this elegant set of studies⁹ significantly advances and expands the understanding of the metabolic regulation by intestinal AMPK (Fig. 1).

Perspective and future directions

Although the current study illustrates metformin activates small intestinal AMPK to regulate glucose homeostasis, intestinal AMPK-independent pathways are also necessary for metformin action (Fig. 1). For example, metformin inhibits upper small intestinal mTOR to lower plasma glucose levels and hepatic glucose production independent of changes in gut AMPK¹⁰. Second, metformin induces changes in gut microbiome to inhibit small intestinal bile acid receptor FXR independent of gut AMPK to improve glucose homeostasis¹¹. Third, metformin-induced changes in gut microbiome enhances upper small intestinal glucose sensing via sodium glucose cotransporter-1 to lower hepatic glucose production¹² possibly via FXR inhibition, as direct small intestinal inhibition of FXR enhances gut glucose sensing to increase intravenous glucose tolerance¹³. If one considers the fact that gut glucose sensing is not required for upper small intestinal metformin-AMPK axis to lower glucose production⁸, it is reasonable to postulate that the upper small intestinal glucose-sodium glucose cotransporter-1 axis that facilitate metformin-mediated gut microbiota changes to lower hepatic glucose production is gut AMPK-independent as well. Collectively, we put forward a working hypothesis that metformin activates small intestinal AMPK-dependent and -independent pathways to regulate glucose and energy homeostasis (Fig. 1).

AMPK α catalytic subunit has two isoforms, α 1 and α 2, which are both activated by metformin¹. The relative contribution of AMPK α 1 and α 2 to metformin action, however, remains unclear. Interestingly, although the re-expression of either AMPK α 1 or α 2 in the liver of HF-fed liver-specific double AMPK α 1 and α 2 knockout mice still impair metformin's ability to lower glucose production as compared to wild-type mice, the loss of hepatic AMPK α 1 vs. AMPK α 2 results in higher glucose production¹⁴, suggesting that hepatic AMPK α 1 may have a dominant role for metformin action. In parallel, a recent study¹⁵ reports intestinal AMPK α 1 and 2 double knockout vs. wild-type mice on chow diet displayed comparable body weight, fat mass and glucose tolerance similar to the intestinal AMPK α 1 knockout mice⁹. In contrast, intestinal AMPK α 1 α 2 knockout mice did not show changes in energy expenditure, thermogenesis, body weight, fat mass and glucose tolerance after ten weeks on a high fat diet¹⁵. Intestinal

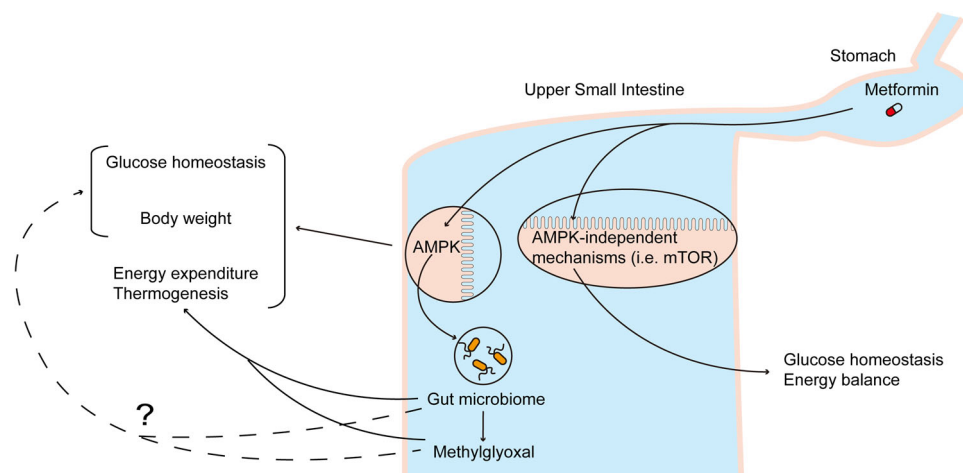


Fig. 1 Metabolic regulation by metformin through intestinal AMPK-dependent and -independent pathways. With the use of high-fat fed intestinal AMPK α 1 knockout mice, Zhang et al. demonstrated that intestinal AMPK α 1 deficiency per se altered energy expenditure and brown fat thermogenesis through changes in the gut microbiome and potentially methylglyoxal, impaired glucose tolerance and induced weight gain. Intestinal AMPK α 1 deficiency also impaired the ability of metformin to increase glucose tolerance and lower body weight. In parallel, metformin alters intestinal AMPK-independent pathways (i.e., mTOR) to regulate glucose homeostasis. We propose metformin activates small intestinal AMPK-dependent and -independent pathways to regulate glucose homeostasis and energy balance in diabetes and obesity.

AMPK α 1 α 2 knockout mice were co-housed with the wild-type mice in the same cages during the experiments that may have led to the exchange and homogenization of gut microbiome, although the gut microbiome was not assessed among the groups in the study¹⁵. A homogenization of the gut microbiome of AMPK α 1 α 2 knockout with wild-type mice may have eliminated any potential adipocyte changes since the reduction in energy expenditure and thermogenesis detected in intestinal AMPK α 1 knockout mice were gut microbiome-dependent⁹. Nonetheless, whether AMPK α 1 vs. α 2 in the intestine has additive, redundant, or even opposing metabolic roles remain to be investigated.

Metformin was equally effective in increasing glucose tolerance in high fat fed intestinal AMPK α 1 α 2 knockout and wild-type mice¹⁵. Specifically, a single oral dose of metformin was administered to the intestinal AMPK α 1 α 2 knockout mice shortly before an oral glucose tolerance test (OGTT)¹⁵, while long-term daily oral metformin administration was given to the intestinal AMPK α 1 knockout mice that undergone an intraperitoneal glucose tolerance test (IPGTT)⁹. Given that an acute constant infusion of metformin into the upper small intestine of high-fat fed rats not only activates AMPK but also inhibits mTOR in an AMPK-independent fashion to lower hepatic glucose production^{6,10}, it is possible that metformin was able to increase glucose tolerance in intestinal AMPK α 1 α 2 knockout mice¹⁵ due to a concurrent inhibition of intestinal mTOR. On the other hand, long-term metformin administration may have desensitized the effect on intestinal mTOR, leading to the inability of metformin to increase glucose tolerance in intestinal AMPK α 1 knockout mice⁹. Further, the use of OGTT that triggers gut glucose sensing in AMPK α 1 α 2 knockout mice¹⁵ may have allowed metformin to increase glucose tolerance, as metformin activates AMPK independent of glucose sensing in the upper small intestine to lower glucose production⁶, consistent with the fact that metformin failed to increase glucose tolerance (assessed by IPGTT and not OGTT) in AMPK α 1 knockout mice independent of gut glucose sensing. Nonetheless, the above speculations remain to be tested.

The study by ref. ⁹ points to the following questions for future research: are the changes in the gut microbiome and/or thermogenesis and energy expenditure incurred by intestinal AMPK-deficiency directly responsible for the induction of weight gain and/or glucose tolerance during high-fat feeding? Does metformin activate an AMPK-Reg3 γ (or α) axis to regulate glucose tolerance, hepatic glucose production and body weight? Are changes in microbiota-derived plasma methylglyoxal responsible for the metabolic benefits of metformin action? What is the relative contribution of hepatic vs. extrahepatic (i.e., brown fat) glucose metabolism in mediating the glucose-lowering effect of the small intestinal metformin-AMPK axis? To begin answering these questions would require the use of genetic and chemical tools to address the cause-and-effect in vivo relationship for the respective mechanisms.

In summary, ref. ⁹ elucidated intestinal AMPK α 1 deficiency in high-fat fed mice altered energy expenditure and thermogenesis of brown fat via changes in gut microbiome, impaired glucose tolerance and increased body weight. Further, intestinal AMPK α 1 deficiency reduced the ability of metformin to increase glucose tolerance and lower body weight in response to high-fat feeding. These studies highlight the metabolic role and therapeutic potential of activating intestinal AMPK for the treatment of diabetes and obesity.

Received: 26 December 2021; Accepted: 26 April 2022;
Published online: 23 May 2022

References

1. Zhou, G. et al. Role of AMP-activated protein kinase in mechanism of metformin action. *J. Clin. Invest.* **108**, 1167–1174 (2001).
2. Miller, R. A. et al. Biguanides suppress hepatic glucagon signalling by decreasing production of cyclic AMP. *Nature* **494**, 256–260 (2013).
3. Cherney, D. Z., & Lam, T. K. A gut feeling for metformin. *Cell Metab.* **28**, 808–810 (2018).
4. Stephensky, D., Friedman, M., Raz, I. & Hoffman, A. Pharmacokinetic-pharmacodynamic analysis of the glucose-lowering effect of metformin in diabetic rats reveals first-pass pharmacodynamic effect. *Drug Metab. Dispos.* **30**, 861–868 (2002).
5. Buse, J. B. et al. The primary glucose-lowering effect of metformin resides in the gut, not the circulation: results from short-term pharmacokinetic and 12-week dose-ranging studies. *Diabetes Care.* **39**, 198–205 (2016).
6. Duca, F. A. et al. Metformin activates a duodenal AMPK-dependent pathway to lower hepatic glucose production in rats. *Nat. Med.* **21**, 506–511 (2015).
7. Borg, M. J. et al. Comparative effects of proximal and distal small intestinal administration of metformin on plasma glucose and GLP-1, and gastric emptying after oral glucose, in type 2 diabetes. *Diabetes Obes. Metab.* **21**, 640–647 (2019).
8. Duca, F. A., Waise, T. M., Peppler, W. T. & Lam, T. K. The metabolic impact of small intestinal nutrient sensing. *Nat. Commun.* **12**, 1–12 (2021).
9. Zhang, E. et al. Intestinal AMPK modulation of microbiota mediates cross-talk with brown fat to control thermogenesis. *Nat. Commun.* **13**, 1–10 (2022).
10. Waise, T. M. et al. Inhibition of upper small intestinal mTOR lowers plasma glucose levels by inhibiting glucose production. *Nat. Commun.* **10**, 714 (2019).
11. Sun, L. et al. Gut microbiota and intestinal FXR mediate the clinical benefits of metformin. *Nat. Med.* **24**, 1919–1929 (2018).
12. Bauer, P. V. et al. Metformin alters upper small intestinal microbiota that impact a glucose-SGLT1 sensing glucoregulatory pathway. *Cell Metab.* **27**, 101–117 (2018).
13. Waise, T. Z. et al. Small intestinal taurochenodeoxycholic acid-FXR axis alters local nutrient sensing glucoregulatory pathways in rats. *Mol. Metab.* **44**, 101132 (2021).
14. Wang, Y. et al. Metformin improves mitochondrial respiratory activity through activation of AMPK. *Cell Rep.* **29**, 1511–1523 (2019).
15. Olivier, S. et al. Deletion of intestinal epithelial AMP-activated protein kinase alters distal colon permeability but not glucose homeostasis. *Mol. Metab.* **47**, 101183 (2021).

Acknowledgements

S.-Y. Z. is supported by a Canadian Institutes of Health Research (CIHR) post-doctoral fellowship. T.K.T.L. is supported by a CIHR Foundation Grant (FDN-143204) and holds the John Kitson McIvor (1915–1942) Endowed Chair in Diabetes Research & the Tier 1 Canada Research Chair in Diabetes and Obesity at the Toronto General Hospital Research Institute and the University of Toronto.

Competing interests

The authors declare no competing interests.

Additional information

Correspondence and requests for materials should be addressed to Tony K. T. Lam.

Reprints and permission information is available at <http://www.nature.com/reprints>

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2022