

RESEARCH

Open Access

In-vivo quantitative T_2 mapping of carotid arteries in atherosclerotic patients: segmentation and T_2 measurement of plaque components

Luca Biasiolli^{1,2*}, Alistair C Lindsay¹, Joshua T Chai², Robin P Choudhury² and Matthew D Robson^{1,2}

Abstract

Background: Atherosclerotic plaques in carotid arteries can be characterized *in-vivo* by multicontrast cardiovascular magnetic resonance (CMR), which has been thoroughly validated with histology. However, the non-quantitative nature of multicontrast CMR and the need for extensive post-acquisition interpretation limit the widespread clinical application of *in-vivo* CMR plaque characterization. Quantitative T_2 mapping is a promising alternative since it can provide absolute physical measurements of plaque components that can be standardized among different CMR systems and widely adopted in multi-centre studies. The purpose of this study was to investigate the use of *in-vivo* T_2 mapping for atherosclerotic plaque characterization by performing American Heart Association (AHA) plaque type classification, segmenting carotid T_2 maps and measuring *in-vivo* T_2 values of plaque components.

Methods: The carotid arteries of 15 atherosclerotic patients (11 males, 71 \pm 10 years) were imaged at 3 T using the conventional multicontrast protocol and Multiple-Spin-Echo (Multi-SE). T₂ maps of carotid arteries were generated by mono-exponential fitting to the series of images acquired by Multi-SE using nonlinear least-squares regression. Two reviewers independently classified carotid plaque types following the CMR-modified AHA scheme, one using multicontrast CMR and the other using T₂ maps of carotid arteries into 4 classes: calcification, lipid-rich necrotic core (LRNC), fibrous tissue and recent IPH. Mean \pm SD of the T₂ values of voxels classified as LRNC, fibrous tissue and recent IPH were calculated.

Results: In 37 images of carotid arteries from 15 patients, AHA plaque type classified by multicontrast CMR and by T_2 maps (+ TOF) showed good agreement (76% of matching classifications and Cohen's $\kappa = 0.68$). The T_2 maps of 14 normal arteries were used to measure T_2 of tunica intima and media ($T_2 = 54 \pm 13$ ms). From 11865 voxels in the T_2 maps of 15 arteries with advanced atherosclerosis, 2394 voxels were classified by the segmentation algorithm as LRNC ($T_2 = 37 \pm 5$ ms) and 7511 voxels as fibrous tissue ($T_2 = 56 \pm 9$ ms); 192 voxels were identified as calcification and one recent IPH (236 voxels, $T_2 = 107 \pm 25$ ms) was detected on T_2 maps and confirmed by multicontrast CMR.

Conclusions: This carotid CMR study shows the potential of *in-vivo* T_2 mapping for atherosclerotic plaque characterization. Agreement between AHA plaque types classified by T_2 maps (+TOF) and by conventional multicontrast CMR was good, and T_2 measured *in-vivo* in LRNC, fibrous tissue and recent IPH demonstrated the ability to discriminate plaque components on T_2 maps.

Keywords: Atherosclerosis, Cardiovascular magnetic resonance, Carotid plaque imaging, *In-vivo* T₂ map, Plaque segmentation, Lipid-rich necrotic core, Fibrous tissue, AHA plaque type classification

* Correspondence: luca.biasiolli@cardiov.ox.ac.uk

Department of Medicine, University of Oxford, John Radcliffe Hospital, Oxford, UK

²Oxford Acute Vascular Imaging Centre (AVIC), Radcliffe Department of Medicine, University of Oxford, John Radcliffe Hospital, Oxford, UK



© 2013 Biasiolli et al.; licensee BioMed Central Ltd. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/2.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

¹Oxford Centre for Clinical Magnetic Resonance Research (OCMR), Radcliffe

Background

Acute ischemic strokes are commonly associated with unstable carotid atherosclerotic plaques that can be detected *in-vivo* by multicontrast CMR [1-3]. Morphology and composition of atherosclerotic plaques can be characterized and classified following the CMR-modified American Heart Association (AHA) scheme [4]. The strength of multicontrast CMR resides in its ability to detect the presence of different plaque components by discriminating their relative signal intensities on time-of-flight (TOF), T₁-, T₂- and PD-weighted (T₁W, T₂W, PDW) images. Identification of lipid-rich necrotic core (LRNC), calcification, intraplaque haemorrhage (IPH) and fibrous tissue by *in-vivo* multicontrast CMR has been thoroughly validated by histology in several clinical studies [4-9].

The size of LRNC and the presence of IPH were shown histologically to be strongly associated with plaque instability [10]. Relative to the adjacent sternocleidomastoid muscle, MR signal from LRNC is iso/hyper-intense on TOF, T_1W and PDW images, and hypo-intense on T_2W images [7]. IPH is an independent predictor of future cardiovascular events [11] and reflects intraplaque neovascularization and plaque vulnerability [12]. It may also stimulate the progression of atherosclerosis by fuelling the plaque core with plasma membrane lipid from extravasated, broken-down, blood cells to increase the LRNC size [8]. MR signal from fresh IPH (type I) is hyper-intense on TOF and T₁W images, and hypo/iso-intense on T₂W and PDW images, whereas MR signal from recent IPH (type II) is iso/ hyper-intense on all images [9].

Despite the advantages of multicontrast CMR, its non-quantitative nature and the need for extensive post-acquisition interpretation are barriers to the widespread clinical application of in-vivo CMR plaque characterization. Comparison of results from in-vivo CMR studies of atherosclerosis using different CMR systems and parameters can be challenging because plaque characterization is affected by variability and inconsistency in the signal intensity of plaque components relative to the reference intensity of the adjacent sternocleidomastoid muscle, which can be particularly evident in T₂W images [2]. Furthermore, image intensity inhomogeneity caused by the surface coil sensitivity can be difficult to correct and can thus affect the relative signal intensities of plaque tissues and consequently the accuracy of plaque segmentation, in particular when automated methods are applied [13]. Finally, T₁W and PDW images acquired with the Fast-Spin-Echo (FSE) sequence (used in multicontrast carotid CMR protocols) are known to suffer from blurring along the phase-encoding direction, which is inherent to the k-space acquisition strategy and causes a significant reduction of the vessel edge sharpness [14].

In-vivo quantitative mapping of relaxation times could provide an alternative method for plaque characterization. This technique measures the MR properties of plaque tissues directly, thus addressing the need for an absolute physical measure that can be standardized among different CMR systems and widely adopted in multi-centre studies, while at the same time avoiding the problems of image intensity inhomogeneity and blurring. Given these characteristics, quantitative mapping has great potential for automated plaque segmentation and classification and may provide new opportunities for quantification of plaque composition in carotid atherosclerosis that will be useful in clinical studies to evaluate emerging drugs that directly target plaque biology [15]. T₂ relaxation times of LRNC and fibrous tissue found in the literature were all measured ex-vivo [16-23], except one study that compared in-vivo and ex-vivo T2 measurements [16]. To our knowledge, no in-vivo T₂ measurements of IPH are reported in the literature.

The purpose of this study was thus to investigate the potential of quantitative T_2 mapping as an *in-vivo* technique for atherosclerotic plaque characterization. In particular, we aimed to acquire *in-vivo* high-resolution quantitative T_2 maps of carotid arteries, to study their use for AHA plaque type classification, and to measure *in-vivo* T_2 relaxation times of the main plaque components at 3 T.

Methods

In-vivo CMR

The carotid arteries of 2 healthy volunteers and 15 patients with known atherosclerosis (11 males and 4 females, 71 ± 10 years, range 54-84) were imaged on two 3T scanners (12 patients on TIM Trio and 3 on Verio, Siemens Healthcare, both running VB17 software) using the Multiple-Spin-Echo (Multi-SE) sequence, called Spin-Echo-Multi-Contrast (SE_MC) on Siemens systems. Ethics approval from the local board was obtained and all subjects gave informed written consent. Black-blood cross-sectional 2D images of carotid arteries were acquired with the Machnet 4-channel phased-array carotid coil using Double-Inversion-Recovery (DIR) preparation and cardiac gating. Chemical shift selective fat saturation (FAT SAT) was used to suppress the signal from subcutaneous and perivascular fat, mainly composed of triglycerides, without affecting the signal from cholesterol and cholesteryl esters, the main lipids in atherosclerotic plaques [21,22]. If necessary, a saturation band was positioned on the anterior region of the neck to reduce ghosting artefacts from breathing and swallowing. Low Specific-Absorption-Rate (SAR) pulses were used in order to keep the total dissipated radio-frequency energy below the SAR limit without changing the flip angle of the refocusing pulses. Multi-SE parameters were TE = 12.9, 25.8, 38.7, 51.6, 64.5, 77.4, 90.3, 103.2 ms, TR = 2 R-R intervals, field of view (FOV) = $160 \times 128 \text{ mm}^2$ and matrix size = 320×256 . Image resolution after zero-padding was 640×512 (pixel size = 0.25 mm). Slice thickness (2 mm) and pixel bandwidth (130 Hz/pix) were the same for Multi-SE and FSE. FSE T_1W (TE = 14 ms, TR = 1 R-R and ETL = 9, T_2W (TE = 89 ms, TR = 2 R-R and ETL = 15) and PDW (TE = 14 ms, TR = 2 R-R and ETL = 9) images had FOV = $150 \times 150 \text{ mm}^2$ and matrix size = 320×320 . Image resolution after zero-padding was 640×640 (pixel size = 0.234 mm). TOF angiography was acquired using 3D Fast Low Angle Shot (FLASH) with flip angle = 18° , $TR = 45 \text{ ms}, TE = 4.1 \text{ ms}, FOV = 200 \times 150 \text{ mm}^2,$ matrix size = 256×192 and slice thickness = 1 mm. 69 TOF and 11 DIR-FSE T₁W consecutive slices centred on the carotid bifurcation were first imaged in order to localize the atherosclerotic plaque. Then a series of DIR-FSE T_2W and PDW (and if necessary other T_1W) images were acquired to obtain full plaque coverage. Finally, a single-slice DIR-Multi-SE image series was acquired through the plaque centre. Given the constraints imposed by other imaging parameters, the TE series used in Multi-SE was defined considering an expected $T_2 \sim 50$ ms for the normal carotid wall, which corresponded to a transverse magnetization half-life time $T_2 \cdot ln(2) \sim 35$ ms. The last TE used in this study was approximately equal to 3 half-life times and typical SNR values measured on normal carotid wall went from ≥ 30 at the first echo to ≥ 4 at the last echo. Partial Fourier imaging was used to collect 5/8 of k-space (160 phase-encoding steps) and reduce the acquisition time of single-slice Multi-SE to circa 320 R-R intervals. The quality of Multi-SE and multicontrast images was assessed after each scan by two reviewers (A.C.L. and L.B.) who decided by consensus when carotid wall and plaque were not clearly visible.

Pulse sequence

The Multi-SE sequence design is similar to the standard FSE that is widely used in carotid imaging, the main difference being the k-space acquisition strategy. Both are Carr-Purcell-Meiboom-Gill (CPMG) based sequences in which a 90° excitation pulse is followed by a train of 180° refocusing pulses to generate multiple spin echoes at every TR [24,25]. Constant crusher gradients at the left and right of each refocusing slice selective gradient are used to select both primary and stimulated echoes, which sum up at the mid-point between two consecutive refocusing pulses, whereas all other unwanted signal pathways are filtered out. In FSE each echo is phaseencoded to a different k-space line in order to collect multiple k-space lines for a single image during every TR, whereas in Multi-SE the phase-encoding is the same for the entire echo train. In this way, each echo of the Multi-SE sequence fills a line of a series of independent k-space planes, so that a set of images with different TEs are collected (Figure 1). Accuracy and precision improvements for clinical T_2 mapping using a CPMG sequence have been demonstrated by setting the slice thickness of the refocusing pulses larger than that of the excitation pulse [26]. This simple modification stabilizes the flip angle of the refocusing pulse across each slice and minimizes refocusing profile errors. A 1.5 ratio between refocusing and excitation slice thickness was used in this study.

T₂ Mapping and plaque segmentation

Quantitative T₂ maps of manually drawn regions of interest (ROIs) including vessels and muscles were generated from the Multi-SE series of images. The first image was discarded because of the different nature of the first echo with respect to the others. Since the flip angles of refocusing pulses are not exactly 180° due to B_1 field inhomogeneities, only the signal acquired at the first TE is a pure primary echo, whereas signals acquired at the subsequent TEs are composed by primary and stimulated echoes. The signal intensity (SI) of the first echo will thus not fit in the exponential decay curve followed by the SIs of subsequent echoes, which are enhanced by stimulated echoes. For every ROI voxel the T₂ mono-exponential decay curve $SI = \beta \cdot e^{-TE/T2}$ was therefore fitted to 7 SIs collected at different TEs (β includes the effect of proton density, T_1 weighting, coil sensitivity, signal amplification and other factors).

The regression model used for T₂ mapping consisted of a nonlinear monoexponential fit (Figure 2) initialized by T_2 and β resulting from a robust linear fit of *ln(SI)*. Parameters estimated by linear least-squares regression are very sensitive to noise because the original error distribution becomes asymmetric after the logarithmic transform. To reduce this sensitivity to outliers, we used a robust regression method that iteratively minimizes the weighted sum of squares using bisquare weights, which depend on the residual of each data point [27]. To assess the statistical significance of fit parameter estimates, the two-tailed p-value of the ratio of each estimate to its standard error (derived from the diagonal elements of the estimated covariance matrix) was calculated on the t-distribution. Only linear fit parameter estimates rejecting the null hypothesis (p < 0.05) were used as initial values for the nonlinear fit in order to improve its convergence. Otherwise default initial values $\beta = 500$ and $T_2 = 50$ ms were used. The nonlinear least-squares regression used the Levenberg-Marquardt algorithm [28]; data points at long TEs with SNR < 2 were discarded; 95% confidence intervals for the exponential curve (Figure 2) and for the fit parameters were estimated. Finally, only voxels with significant T_2 and β estimates (p < 0.05) were accepted for generating T_2 maps.



The carotid arteries were subsequently segmented using a semi-automated method [29] that detected inner and outer wall boundaries on the second image (TE = 25.8 ms, TR = 2 R-R) of the Multi-SE series.

After this procedure, T_2 maps of atherosclerotic arteries were segmented into 3 tissue types using a Bayes classifier [30] defined by the probability model $P(C_i|T_2) = P(C_i)P(T_2|C_i)$ combined with the maximum a posteriori (MAP) decision rule. $P(C_i|T_2)$ with i = 1,2,3 is the posterior conditional probability for a voxel with a specific T_2 value to belong to one of 3 classes: LRNC, fibrous tissue or recent IPH. The MAP rule then assigned a particular voxel to the tissue type with the highest posterior probability. $P(C_i|T_2)$ is determined by the prior probability $P(C_i)$, which was assumed to be equal for every class, and the normal probability distribution $P(T_2|C_i)$, which was estimated from a set of approximately 100 voxels for each class. These voxels were manually selected from all the T₂ maps to represent a specific tissue using multicontrast CMR as a guide, i.e. they were chosen inside a plaque component identified on multicontrast CMR [4,9]. In order to help matching the corresponding plaque features, multicontrast images were co-registered [31], so that inner and outer vessel boundaries detected on T₁W images [29] could then be superimposed on PDW and T₂W images. Calcification was identified by voxels inside the vessel wall boundaries with SNR < 2 at TE = 14 ms on the T₂ decay curve (synthetic PDW image). Finally, mean ± SD of the T₂ values of



voxels classified as LRNC, fibrous tissue and recent IPH were calculated. The algorithms for quantitative T_2 mapping, vessel wall segmentation and plaque classification were implemented in Matlab (Mathworks).

To compare atherosclerotic plaque characterization by T_2 maps with conventional multicontrast CMR, two reviewers (A.C.L. and L.B.) independently classified the carotid arteries of 15 patients as normal or diseased (plaque type III, IV-V, VI, VII or VIII) following the CMR-modified AHA scheme [4,9]. One reviewer analysed multicontrast CMR (T_1W , T_2W and PDW image intensity inhomogeneity was corrected [13]) and TOF, while the other relied on T_2 maps and TOF at matching slice locations. Both were blinded to the identification and the clinical data of each patient.

Results

Out of 42 image series (including common, internal and external carotid arteries) acquired with single-slice Multi-SE in 15 patients, 5 were rejected due to poor image quality. The corresponding multicontrast (TOF, T_1W , T_2W and PDW) images were all accepted and the AHA plaque type classification performed on them by one reviewer found 14 normal and 23 diseased arteries (7 type III, 10 type IV-V, 2 type VI, 3 type VII and 1 type VIII plaques). AHA classification performed by the other reviewer on T_2 maps and TOF found 11 normal and 26 diseased arteries (11 type III, 8 type IV-V, 3 type VI, 2 type VII and 2 type VIII plaques). The overall agreement on AHA plaque type between the reviewers was good (76% of matching classifications and Cohen's $\kappa = 0.68$, Table 1).

The T₂ maps of the 10 arteries classified as normal by both reviewers, together with 4 common carotid arteries from 2 healthy volunteers, were used to measure T_2 mean \pm SD of normal carotid wall (T₂ = 54 \pm 13 ms, Table 2). The T_2 maps of the 15 atherosclerotic arteries with advanced plaques (graded as type IV-V, VI, VII or VIII by both reviewers) were segmented by the Bayes classifier into 4 plaque components: calcification, LRNC, fibrous tissue and recent IPH. From a total of 11865 voxels, the T₂ measurements of 668 voxels were rejected because not statistically significant (p > 0.05). 192 voxels were identified as calcification in one type IV-V, one VI and two VII plaques. 2394 voxels with $T_2 = 37 \pm 5$ ms were classified as LRNC by the segmentation algorithm, mostly detected in type IV-V and VI plaques (Table 2). 7511 voxels with $T_2 = 56 \pm 9$ ms were classified as fibrous tissue and normal intima/media (Table 2). Figure 3 shows an atherosclerotic artery with LRNC and a thick fibrous cap separating it from the lumen (type IV-V). Figure 4 shows a large complex plaque (type VI) at the carotid bifurcation composed of lipid and necrotic material mixed together with fibrous tissue. These features

Table 1 AHA plaque type classification by multicontrast CMR vs. T_2 maps + TOF

Multicontrast	T ₂ maps + TOF								
CMR	Normal	Ш	IV-V	VI	VII	VIII	Total		
Normal	10	4					14		
III	1	6					7		
IV-V		1	7	1		1	10		
VI				2			2		
VII			1		2		3		
VIII						1	1		
Total	11	11	8	3	2	2	37		

agreed with the multicontrast CMR classification, which revealed the presence of fresh IPH (hyper-intensity on TOF and T_1W images).

Finally, large areas of recent IPH were detected on T_2 maps in two type VI plaques for a total of 528 voxels measuring $T_2 = 105 \pm 25$ ms. However, only the plaque with recent IPH in Figure 5 (236 voxels with $T_2 = 107 \pm 25$ ms) was confirmed as type VI by multicontrast CMR. In the other 13 plaques, sparse voxels and small regions (for a total of 572 voxels with $T_2 = 86 \pm 9$ ms) were classified as recent IPH by the segmentation algorithm. Additionally, since most clinical CMR studies of atherosclerosis used the sternocleidomastoid muscle as a signal intensity reference for plaque tissue classification in T_2W images [4,7,9], we generated T_2 maps of sternocleidomastoid muscles in volunteers and patients and measured $T_2 = 39 \pm 6$ ms.

Discussion

The objective of this study was to investigate the use of T_2 mapping for plaque characterization. We acquired in-vivo high-resolution T₂ maps of carotid arteries in 15 atherosclerotic patients using the Multi-SE sequence and demonstrated good agreement between AHA plaque type classified by T₂ maps and TOF images and by conventional multicontrast CMR. The present study measured the T₂ of LRNC, fibrous tissue and recent IPH and showed the potential of T_2 mapping for plaque segmentation and classification. Our in-vivo T₂ measurements (Table 2) were slightly longer but broadly in agreement with the only in-vivo T₂ study of carotid atherosclerosis available in the literature [16]. The T_2 values of tunica intima and media measured in 14 normal carotid arteries fell within the T₂ range reported by *ex-vivo* studies of carotid and coronary atherosclerosis [16-20]. With the exception of one study that reported much longer values [18], the T₂ measurements of fibrous tissue and LRNC in 15 advanced plaques were comparable with those obtained ex-vivo [16,17,19,20]. Recent IPH was characterized by long T₂, consistent with signal hyper-

Study	Field strength	Num. of TEs	Num. of plaques	Location	LRNC [ms]	Fibrous tissue [ms]	Intima & media [ms]	Ref.
in-vivo	3 T	7	15	carotid	37 ± 5	56 ± 9	54 ± 13	*This study
in-vivo	1.5 T	2	7	carotid	28 ± 6	51 ± 10	48 ± 7	[16]
ex-vivo	1.5 T	2	7	carotid	31 ± 5	51 ± 9	52 ± 7	[16]
ex-vivo	1.5 T	8	10	carotid	47 ± 14	60 ± 13	67 ± 23	[17]
ex-vivo	1.5 T	4	10	carotid	84 ± 41	95 ± 32	60 ± 6	[18]
ex-vivo	9.4 T	7	3	carotid	35-49	48-60	72-76	[19]
ex-vivo	4.7 T	4	7	coronary	31 ± 7	55 ± 11	50 ± 10	[20]
ex-vivo	1.5 T	10	8	various	55 ± 3	79 ± 4	81 ± 3	[21]
ex-vivo	4.7 T	10	8	various	50 ± 3	63 ± 1	65 ± 2	[21]
ex-vivo	9.4 T	20	8	various	20 ± 3	30 ± 2	30 ± 3	[21]
ex-vivo	1.5 T	32	13	various	< 20	100-200	68 ± 10	[22]
ex-vivo	3 T	7	14	aorta/iliac	54 ± 3	89 ± 6	76 ± 9	[23]

Table 2 T_2 measurements (mean \pm SD) of arterial wall and plaque tissues

*Fibrous tissue and normal arterial wall cannot be discriminated *in-vivo* by T_2 alone. This study measured the T_2 values of tunica intima and media in 14 normal arteries, while both fibrous tissue and normal arterial wall of the 15 atherosclerotic arteries were categorized together as fibrous tissue.

intensity on T_2W images [9], and was detected in one complex (type VI) plaque (confirmed by multicontrast CMR). In other plaques the classification of some sparse voxels and small regions as recent IPH was most likely caused by T_2 overestimation errors: their T_2 values were longer than their neighbouring voxels but shorter than the complex plaque and TOF images showed no signs of IPH. The measured T_2 of sternocleidomastoid muscles agreed with the range of values reported in literature for muscle tissue (30-50 ms) at 1.5 T and 3 T [32]. It is slightly longer than the T_2 of LRNC but shorter than that of normal carotid wall and fibrous tissue, confirming it as a suitable intensity reference in T_2W images [4,7,9]. However, intensity inhomogeneity introduced by the sensitivity profile of



Figure 3 Multicontrast CMR and I_2 map of a type IV-V plaque in the right common carotid artery. Multicontrast CMR shows a hypo-intense region compared to the sternocleidomastoid muscle (SM) on the T_2 W image, indicative of LRNC (arrow). The T_2 map and its segmentation show a thick fibrous cap separating the LRNC, characterized by shorter T_2 values, from the lumen.



of fresh IPH (arrow), characterized by hyper-intensity on TOF and T_1W images compared to the sternocleidomastoid muscle (SM). The T_2 map and its segmentation illustrate that the plaque is composed by fibrous tissue, pools of lipid and necrotic material (LRNC) and a small calcification. Some sparse voxels were classified as recent IPH due to T_2 overestimation.

surface coils, which strongly affects the sternocleidomastoid muscle due to its skin proximity, must be corrected before it can be used as reference.

Table 2 shows the *ex-vivo* studies of atherosclerosis that were found in literature [16-23]. Despite their methodological differences (i.e. field strength, imaging parameters and arterial locations from which the plaques had been excised), the common feature of these *ex-vivo* studies is a shorter T_2 in LRNC than in fibrous tissue and normal intima and media, which was confirmed by our *in-vivo* T_2 measurements. However, care must be taken when comparing *in-vivo* and *ex-vivo* T_2 values, because the effect of temperature and fixatives on tissue specimens can be significant [18]. Five *ex-vivo* CMR studies were performed at body temperature [16,18,20,21,23] and three at room temperature [17,19,22].

Our *in-vivo* T_2 measurement method is more accurate than the previous *in-vivo* study [16], which calculated T_2 of 7 carotid arteries by collecting and fitting only 2 echoes with relatively short TE (20 and 55 ms). In that study [16] the high sensitivity to noise due to the limited number of echoes constraining the T_2 exponential decay curve was aggravated by the use of short TR (1 R-R),which causes more T_1 weighting, and very thick slices (5 mm), which can mix T_2 values of different tissues (partial volume averaging). We used a CPMG sequence (Multi-SE) to acquire more echoes (7 TEs ranging from 25.8 to 103.2 ms) designed to obtain the best possible sampling of the T_2 relaxation decay curve of target tissues. The Multi-SE acquisition parameters were more appropriate for T_2 quantitation than those used by the previous *in-vivo* study, since TR was longer (2 R-R) and slices thinner (2 mm). We also implemented a nonlinear regression method that is more robust to outliers, and we ensured that the T_2 estimates were statistically significant by accepting only results with p < 0.05. Finally, the present study imaged 14 normal and 15 atherosclerotic carotid arteries, more than those analysed previously (Table 2).

In some cases the first image of the Multi-SE series showed artefacts due to residual signal from slowly flowing blood not suppressed by DIR. We speculated that the main cause was inaccurate shimming of B_0 inhomogeneities. Flow artefacts did not produce T_2 errors because the first contrast image was never used to estimate T_2 , as explained in the methods.

Limitations

The main limitation of this study is the absence of plaque histology, since only two of the participating patients underwent surgery and their endarterectomy specimens were of insufficient quality for detailed analysis. However, plaque characterization performed on *in-vivo*



manually placed to indicate the occluded artery.

 T_2 maps agreed with results of multicontrast CMR, which has been previously validated by histology [4-9]. A physical limitation of T_2 mapping is that the presence of fresh IPH (type I) cannot be detected by T_2 alone. Fresh IPH infiltrating the LRNC is characterized by short T_2 [8,9], so it can only be detected using T_1 information, e.g. independent identification on TOF images (Figure 4).

Finally, imperfections of the refocusing pulses represent the main source of errors in T_2 quantitation [33], although the CPMG sequence used in this study partially compensates for them. Stimulated echoes produced by non-ideal refocusing pulses are acquired together with the primary echoes, thus introducing T_1 weighting into the signal and altering the pure T₂ relaxation decay. After discarding the first echo, which is purely primary, the contribution of stimulated echoes to the remaining 7 echoes may have caused T₂ overestimation. These errors depend on the amount of T_1 weighting introduced by the use of TR = 2 R-R. A very long TR ($\sim 5 \cdot T_1$ of target tissue) would limit the T₁ effect, but it would be absolutely impractical under in-vivo high-resolution imaging conditions. Finally, the elimination of the first echo may have affected the estimation of short T₂ components in the LRNC (~35 ms), which have a transverse magnetization half-lifetime (~24 ms) as long as the second TE. In general, this problem can be alleviated by the use of shorter echo spacing (ESP). This study used the shortest ESP possible, given the requirements for matrix size, receiver bandwidth and the duration of low-SAR refocusing pulses. Future developments will aim at improving *in-vivo* T_2 measurement accuracy, decreasing acquisition time and validating plaque characterization with histology.

Conclusions

This study draws attention to the potential use of *in-vivo* T_2 mapping quantitative for carotid plaque characterization. On 37 carotid arteries imaged in-vivo in 15 atherosclerotic patients, AHA plaque type classification using high-resolution T_2 maps and TOF images showed good agreement with conventional multicontrast CMR. T₂ relaxation times measured in-vivo at 3 T for tunica intima and media in 14 normal arteries and for LRNC, fibrous tissue and recent IPH in 15 arteries with advanced plaques demonstrated the ability to discriminate 4 major plaque components including calcification, thus supporting the application of plaque segmentation and classification based on T₂ maps.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

LB, RPC and MDR provided the concepts. LB, ACL and MDR developed the CMR protocol. ACL and JTC recruited the patients. LB, ACL, JTC and MDR collected data. LB and ACL reviewed data. LB developed algorithms,

analysed data and drafted the manuscript. RPC and MDR supervised the project and acted in last author capacity. All authors read and revised the manuscript critically for important intellectual content, and approved the final version.

Acknowledgements

This study was funded by the Engineering and Physical Sciences Research Council (EPSRC) and the Oxford Biomedical Research Centre, National Institute for Health Research funding scheme. LB was supported by the EPSRC Knowledge Transfer Secondment Award. RPC is a Wellcome Trust Senior Fellow and JTC is a MRC training Fellow. The authors wish to acknowledge the facilities provided by the University of Oxford Centre for Clinical Magnetic Resonance Research (OCMR) and the Oxford Acute Vascular Imaging Centre (AVIC).

Received: 13 March 2013 Accepted: 8 August 2013 Published: 16 August 2013

References

- Dong L, Kerwin WS, Ferguson MS, Li R, Wang J, Chen H, Canton G, Hatsukami TS, Yuan C. Cardiovascular magnetic resonance in carotid atherosclerotic disease. J Cardiovasc Magn Reson. 2009; 11:53.
- Watanabe Y, Nagayama M. MR plaque imaging of the carotid artery. Neuroradiology. 2010; 52:253–74.
- Lindsay AC, Biasiolli L, Lee JMS, Kylintireas I, MacIntosh BJ, Watt H, Jezzard P, Robson MD, Neubauer S, Handa A, Kennedy J, Choudhury RP. Plaque features associated with increased cerebral infarction after minor stroke and TIA: a prospective, case–control, 3-T carotid artery MR imaging study. J Am Coll Cardiol Img. 2012; 5:388–96.
- Cai J-M, Hatsukami TS, Ferguson MS, Small R, Polissar NL, Yuan C. Classification of human carotid atherosclerotic lesions with in vivo multicontrast magnetic resonance imaging. *Circulation*. 2002; 106:1368–73.
- Yuan C, Mitsumori LM, Ferguson MS, Polissar NL, Echelard D, Ortiz G, Small R, Davies JW, Kerwin WS, Hatsukami TS. In vivo accuracy of multispectral magnetic resonance imaging for identifying lipid-rich necrotic cores and intraplaque hemorrhage in advanced human carotid plaques. *Circulation*. 2001; 104:2051–56.
- Yuan C, Zhang S, Polissar NL, Echelard D, Ortiz G, Davis JW, Ellington E, Ferguson MS, Hatsukami TS. Identification of fibrous cap rupture with magnetic resonance imaging is highly associated with recent transient ischemic attack or stroke. *Circulation*. 2002; 105:181–85.
- Saam T, Ferguson MS, Yarnykh VL, Takaya N, Xu D, Polissar NL, Hatsukami TS, Yuan C. Quantitative evaluation of carotid plaque composition by in vivo MRI. Arterioscler Thromb Vasc Biol. 2005; 25:234–39.
- Takaya N, Yuan C, Chu B, Saam T, Polissar NL, Jarvik GP, Isaac C, McDonough J, Natiello C, Small R, Ferguson MS, Hatsukami TS. Presence of intraplaque hemorrhage stimulates progression of carotid atherosclerotic plaques: a high-resolution magnetic resonance imaging study. *Circulation*. 2005; 111:2768–75.
- Chu B, Kampschulte A, Ferguson MS, Kerwin WS, Yarnykh VL, O'Brien KD, Polissar NL, Hatsukami TS, Yuan C. Hemorrhage in the atherosclerotic carotid plaque: a high-resolution MRI study. Stroke. 2004; 35:1079–84.
- Redgrave JNE, Lovett JK, Gallagher PJ, Rothwell PM. Histological assessment of 526 symptomatic carotid plaques in relation to the nature and timing of ischemic symptoms: the Oxford plaque study. *Circulation*. 2006; 113:2320–28.
- Hellings WE, Peeters W, Moll FL, Piers SRD, van Setten J, der Spek PJV, de Vries J-PPM, Seldenrijk KA, Bruin PCD, Vink A, Velema E, de Kleijn DPV, Pasterkamp G. Composition of carotid atherosclerotic plaque is associated with cardiovascular outcome: a prognostic study. *Circulation*. 2010; 121:1941–50.
- Michel J-B, Virmani R, Arbustini E, Pasterkamp G. Intraplaque haemorrhages as the trigger of plaque vulnerability. *Eur Heart J.* 2011; 32:1977–85.
- Salvado O, Hillenbrand C, Zhang S, Wilson DL. Method to correct intensity inhomogeneity in MR images for atherosclerosis characterization. *IEEE Trans Med Imaging*. 2006; 25:539–52.
- 14. Biasiolli L, Lindsay AC, Choudhury RP, Robson MD. Loss of fine structure and edge sharpness in fast-spin-echo carotid wall imaging:

measurements and comparison with multiple-spin-echo in normal and atherosclerotic subjects. *J Magn Reson Imaging*. 2011; **33**:1136–43.

- Lindsay AC, Choudhury RP. Form to function: current and future roles for atherosclerosis imaging in drug development. *Nat Rev Drug Discov.* 2008; 7:517–29.
- Toussaint J-F, LaMuraglia GM, Southern JF, Fuster V, Kantor HL. Magnetic resonance images lipid, fibrous, calcified, hemorrhagic, and thrombotic components of human atherosclerosis in vivo. *Circulation*. 1996; 94:932–38.
- Degnan AJ, Young VE, Tang TY, Gill AB, Graves MJ, Gillard JH, Patterson AJ. Ex vivo study of carotid endarterectomy specimens: quantitative relaxation times within atherosclerotic plaque tissues. *Magn Reson Imaging.* 2012; 30:1017–21.
- Dalager-Pedersen S, Falk E, Ringgaard S, Kristensen IB, Pedersen EM. Effects of temperature and histopathologic preparation on the size and morphology of atherosclerotic carotid arteries as imaged by MRI. J Magn Reson Imaging. 1999; 10:876–85.
- Morrisett J, Vick W, Sharma R, Lawrie G, Reardon M, Ezell E, Schwartz J, Hunter G, Gorenstein D. Discrimination of components in atherosclerotic plaques from human carotid endarterectomy specimens by magnetic resonance imaging ex vivo. *Magn Reson Imaging*. 2003; 21:465–74.
- Sun B, Giddens DP, Long R, Taylor WR, Weiss D, Joseph G, Vega D, Oshinski JN. Automatic plaque characterization employing quantitative and multicontrast MRI. *Magn Reson Med.* 2008; 59:174–80.
- Toussaint JF, Southern JF, Fuster V, Kantor HL. T2-weighted contrast for NMR characterization of human atherosclerosis. Arterioscler Thromb Vasc Biol. 1995; 15:1533–42.
- Martin AJ, Gotlieb AI, Henkelman RM. High-resolution MR imaging of human arteries. J Magn Reson Imaging. 1995; 5:93–100.
- Raynaud J-S, Bridal SL, Toussaint J-F, Fornès P, Lebon V, Berger G, Leroy-Willig A. Characterization of atherosclerotic plaque components by high resolution quantitative MR and US imaging. J Magn Reson Imaging. 1998; 8:622–29.
- Carr HY, Purcell EM. Effects of diffusion on free precession in nuclear magnetic resonance experiments. *Phys Rev.* 1954; 94:630.
- 25. Meiboom S, Gill D. Modified spin-echo method for measuring nuclear relaxation times. *Rev Sci Instrum.* 1958; 29:688–91.
- Pell GS, Briellmann RS, Waites AB, Abbott DF, Lewis DP, Jackson GD. Optimized clinical T2 relaxometry with a standard CPMG sequence. J Magn Reson Imaging. 2006; 23:248–52.
- Huber PJ, Ronchetti E. Robust statistics. 2nd ed. Hoboken: John Wiley & Sons; 2009.
- Seber GAF, Wild CJ. Nonlinear regression. Hoboken: John Wiley & Sons; 2003.
- Wang Q, Robson MD, Francis JM, Petersen SE, Channon KM, Neubauer S, Wiesmann F. Accuracy of quantitative MR vessel wall imaging applying a semi-automated gradient detection algorithm - a validation study. J Cardiovasc Magn Reson. 2004; 6:895–907.
- Bishop CM. Pattern Recognition and Machine Learning. New York: Springer; 2006.
- Biasiolli L, Noble JA, Robson MD. Multicontrast MRI registration of carotid arteries in atherosclerotic and normal subjects, Proceedings of SPIE, Medical Imaging 2010: Image Processing. San Diego; 2010: p. 7623. 76232N.
- 32. De Graaf RA. In vivo NMR spectroscopy: principles and techniques. 2nd ed. Chichester: John Wiley & Sons; 2007.
- Poon CS, Henkelman RM. Practical T2 quantitation for clinical applications. J Magn Reson Imaging. 1992; 2:541–53.

doi:10.1186/1532-429X-15-69

Cite this article as: Biasiolli *et al.*: **In-vivo quantitative** T_2 mapping of carotid arteries in atherosclerotic patients: segmentation and T_2 measurement of plaque components. *Journal of Cardiovascular Magnetic Resonance* 2013 **15**:69.