# BJC

British Journal of Cancer (2013) 109, 1031–1039 | doi: 10.1038/bjc.2013.390

Keywords: tumour-associated antigen; immune infiltration; prognosis; hepatocellular carcinoma

## Expression pattern of tumour-associated antigens in hepatocellular carcinoma: association with immune infiltration and disease progression

#### J Liang<sup>1,2,5</sup>, T Ding<sup>1,3,5</sup>, Z-W Guo<sup>2</sup>, X-J Yu<sup>1</sup>, Y-Z Hu<sup>1,4</sup>, L Zheng<sup>1,2</sup> and J Xu<sup>\*,1</sup>

<sup>1</sup>State Key Laboratory of Oncology in South China, Cancer Center, Sun Yat-sen (Zhongshan) University, Guangzhou 510 060, PR China; <sup>2</sup>State Key Laboratory of Biocontrol, School of Life Science, Sun Yat-sen (Zhongshan) University, Guangzhou 510 031, PR China; <sup>3</sup>Department of Cell Biology, Nanjing Medical University, Nanjing 210 029, PR China and <sup>4</sup>Bank of Tumor Resources, Cancer Center, Sun Yat-sen (Zhongshan) University, Guangzhou 510 060, PR China

**Background:** The distinct expression pattern of tumour-associated antigens (TAAs) might be a critical reason for the inefficacy of immunity-based treatments and heterogeneous postsurgical recovery in patients with solid tumours, including hepatocellular carcinoma (HCC). However, little is known about the clinical value of the coexpression patterns of multiple TAAs.

**Methods:** We determined the expression of multiple TAAs with identified immunogenicity (GPC3, AFP, SSX-2, NY-ESO-1, EpCAM, midkine) and the density of tumour-infiltrating immune cells by immunohistochemistry in a panel of 362 primary HCC patients. We evaluated the association between the TAAs, immune cell infiltration, clinicopathological parameters, and prognosis.

**Results:** Patients who coexpressed more TAAs had better prognosis (P < 0.00001, overall survival). The integrated pattern of TAA was associated with good differentiation and small tumour size, and with more CD57<sup>+</sup> natural killer and CD20<sup>+</sup> B-cell infiltration (P < 0.05). Multivariate Cox proportional hazards analysis identified the TAA index as an independent prognostic indicator (hazard ratio 0.625; 95% confidence interval 0.467–0.837; P = 0.002), and could further predict patient prognosis in collaboration with local immune infiltration.

**Conclusion:** Our results could provide new evidence for the improvement of prognostic molecular signatures in HCC, and a novel rationale for patient enrolment in future immunotherapeutic trials and/or clinical treatments.

Hepatocellular carcinoma (HCC) is among the world's leading cancer threats, and its incidence is increasing (Fan *et al*, 2011; Jemal *et al*, 2011). Despite improved diagnostic and treatment strategies, the overall survival (OS) of patients with HCC remains poor (Bruix and Llovet, 2009; Villanueva *et al*, 2012). Hepatocellular carcinoma is a highly complex disease that is generally resistant to commonly used chemotherapy and radiotherapy. Only a small proportion of newly diagnosed patients are eligible for

potential curative therapies, including resection and liver transplantation. However, these therapeutic procedures most often do not provide a complete cure, and half of the treated patients experience tumour recurrence within 3 years (Villanueva *et al*, 2010; Yang and Roberts, 2010; Gao *et al*, 2012).

Hepatocellular carcinoma is usually present in inflamed fibrotic and/or cirrhotic liver with extensive leukocyte infiltration. Thus, the immune status at a tumour site can largely influence the

Received 24 April 2013; revised 14 June 2013; accepted 24 June 2013; published online 18 July 2013

© 2013 Cancer Research UK. All rights reserved 0007–0920/13

<sup>\*</sup>Correspondence: Dr J Xu; E-mail: xujing@sysucc.org.cn <sup>5</sup>These authors contributed equally to this work.

biologic behaviour of HCC (Grivennikov et al, 2010; Fridman et al, 2012; Qin, 2012). Although it is commonly believed that innate and adaptive immunity would inhibit cancer growth, solid tumour cells, which often arise from chronic inflammation and survive immunoediting, can escape from immunosurveillance through different mechanisms (Disis, 2010; Vesely et al, 2011). After decades of substantial efforts, the success of several recent proof-of-concept clinical trials targeting immune regulators (e.g., ipilimumab and MDX-1106) suggests that active immunotherapy represents a path to obtain a durable and long-lasting response in cancer patients. However, their effects on the regression of human tumours remain limited, and only a relatively small fraction of patients derives clinical benefits (Mellman et al, 2011; Topalian et al, 2011). The reconstruction of immune surveillance for aberrant cells requires adequate tumour-associated antigen (TAA) expression, and the distinct expression pattern of TAAs in different tumours may be a critical reason for heterogeneous therapeutic effectiveness. Severe deficiency in TAAs is a common trick of tumour immune escape in primary cancer and/or during postsurgical relapse (Schreiber et al, 2011; Fu, 2012). However, little is currently known about the clinical significance of the coexpression pattern of multiple TAAs in solid tumours, including HCC.

In view of the present situation, we aimed to determine the expression of several TAAs with identified immunogenicity and potential therapeutic value in HCC and to evaluate their clinicopathological roles. In general, our results showed that the expression patterns of these TAAs (TAA index) were associated with local immune infiltration, disease progression, and prognosis.

#### MATERIALS AND METHODS

Patients and tissue samples. Archived formalin-fixed, paraffinembedded tissues were obtained at the Cancer Center of Sun Yat-sen University, from 362 patients who had all undergone curative resection for HCC between January 2002 and December 2005. There were 317 (87.6%) patients with hepatitis B virus infection. We defined curative resection for HCC as a resection margin of at least 1 cm, complete resection of all tumour nodules, and leaving the cut surface free of tumour based on histological examination. Intra-operative ultrasound and postsurgical contrast-enhanced computed tomography (CT) were routinely used to ensure the complete removal of the HCC (Gao et al, 2007; Kuang et al, 2011; Xu et al, 2012b). No patient received anti-cancer therapies or had distant metastasis prior to the surgery. The clinical stage of tumours was determined according to the tumor-nodes-metastasis (TNM) classification system of the International Union Against Cancer (edition 6). The Institutional Review Boards of the Cancer Center approved the study. Written informed consent was obtained from all patients. Clinicopathological characteristics are summarised in Table 1.

**Follow-up of patients and postoperative treatment.** Patients were followed postoperatively at the outpatient clinic with regular surveillance for recurrence using the serum alpha-fetoprotein (AFP) level, abdominal ultrasonography, and chest radiography at 2- to 4-month intervals (Gao *et al*, 2007; Ding *et al*, 2011; Kuang *et al*, 2011; Xu *et al*, 2012b). When tumour recurrence or metastasis was suspected, further examinations, including CT and hepatic angiography, were performed. Biopsies were obtained when necessary. Patients with confirmed recurrence received further treatment, including a second surgical resection, transcatheter arterial chemoembolisation, radiofrequency ablation or percutaneous ethanol injection. The median follow-up was 34.5 months (range 1–95 months). Of the 362 patients examined during the follow-up period, 190 patients (52.5%) died, 193 (53.3%) were

Characteristics         No. of patients (%)           Age, year         48.5           Median         48.5           Range         20–78           Gender         324 (89.5)           Male         324 (89.5)           Female         38 (10.5)           Hepatitis B virus infection         38 (10.5)           No         45 (12.4)           Yes         317 (87.6)           Hepatitis C virus infection         6 (1.7)           No         356 (98.3)           Yes         6 (1.7)           Alpha-fetoprotein         6 (1.7)           Signell <sup>-1</sup> 255 (70.4)           Child-Pugh class         23 (6.4)           A         339 (93.6)           B         23 (6.4)           Differentiation         73 (20.2)           I-II         289 (79.8)           II-IV         73 (20.2)           Single         274 (75.7)           Multiple         88 (24.3)           Tumour size         234 (64.6)           Single         309 (85.4)           > 5 cm         234 (64.6)           Absent         309 (85.4)           Present         33 (14.6)	Table 1. Patient characteristics	
Age, year           Median Range         48.5 20-78           Gender         20-78           Male Female         324 (89.5) 38 (10.5)           Hepatitis B virus infection         38 (10.5)           No Yes         45 (12.4) 317 (87.6)           Hepatitis C virus infection         107 (87.6)           No Yes         6 (1.7)           Alpha-fetoprotein         25 rg ml <sup>-1</sup> <25 rg ml <sup>-1</sup> 255 (70.4)           25 ng ml <sup>-1</sup> 255 (70.4)           /Alpha-fetoprotein         23 (64.1)            23 (64.1)           Differentiation         23 (64.1)           III-IV         73 (20.2)           Fumour number         88 (24.3)           Single Multiple         88 (24.3)           Som > 5 cm         234 (64.6)           Vascular invasion         234 (64.6)           Absent Present         309 (85.4) (301 (4.6)           Function         309 (85.4) (301 (4.6)	Characteristics	No. of patients (%)
Median Range         48.5 20-78           Gender         20-78           Male Female         324 (89.5) Semiler           Female         38 (10.5)           Hepatitis B virus infection         317 (87.6)           No Yes         45 (12.4) 317 (87.6)           Hepatitis C virus infection         356 (98.3) 6 (1.7)           No Yes         356 (98.3) 6 (1.7)           Alpha-fetoprotein         55 (70.4)           Signel <sup>-1</sup> 255 (70.4)           Child-Pugh class         23 (6.4)           A         339 (93.6) B           B         23 (6.4)           Differentiation         289 (79.8) (73 (20.2)           I-II II-IV         289 (79.8) (73 (20.2)           Single         274 (75.7) (88 (24.3)           Multiple         88 (24.3)           Tumour number         23 (6.4)           Single         274 (75.7) (83 (24.6)           Single         274 (75.7) (83 (24.6)           Vascular invasion         23 (64.6)           Vascular invasion         309 (85.4) (93 (14.6)           Present         30 (85.4) (71.6)           Present         30 (25.4) (71.5)           I-III         259 (71.5)           I-III         259 (71.5) </td <td>Age, year</td> <td></td>	Age, year	
Range         20-78           Gender         324 (89.5)           Female         38 (10.5)           Hepatitis B virus infection         338 (10.5)           No         45 (12.4)           Yes         317 (87.6)           Hepatitis C virus infection         (100)           No         356 (98.3)           Yes         6 (1.7)           Alpha-fetoprotein         (107 (29.6)           ≤ 25 ng ml <sup>-1</sup> 255 (70.4)           Child-Pugh class         23 (6.4)           Differentiation         (23 (6.4))           L-II         289 (79.8)           II-IV         73 (20.2)           Tumour number         88 (24.3)           Single         274 (75.7)           Multiple         88 (24.3)           Tumour size         (23 (6.4))           ≤ 5 cm         128 (35.4)           > 5 cm         234 (64.6)           Vascular invasion         (309 (85.4))           Present         53 (14.6)           TMM stage         11 (259 (71.5))           I -III         259 (71.5)           I -III         (259 (71.5))	Median	48.5
Gender           Male Female         324 (89.5) 38 (10.5)           Hepatitis B virus infection         317 (87.6)           No Yes         45 (12.4) 317 (87.6)           Hepatitis C virus infection $(1.7)$ No Yes         6 (1.7)           Alpha-fetoprotein $(1.7)$ $\leq 25 ng ml^{-1}$ 107 (29.6) $> 25 ng ml^{-1}$ $> 25 ng ml^{-1}$ 255 (70.4)           Child-Pugh class $(339 (93.6)$ B           A         339 (93.6) B           B         23 (6.4)           Differentiation $(1.7)$ I-II         289 (79.8) (1.8)           II-IV         73 (20.2)           Tumour number $(3.2, 1)$ Single $\leq 5 cm$ 224 (75.7) (324 (64.6)           Vescular invasion $(3.2, 4)$ $A > 5 cm$ 234 (64.6)           Vascular invasion $(3.3, 9)$ (85.4) (71.5) (1.4)           Absent Present         309 (85.4) (71.5) (1.4)           TIM stage $(1.1)$ I-III         259 (71.5) (1.5)           I-III         259 (71.5) (1.5)	Range	20–78
Male         324 (89.5)           Female         38 (10.5)           Hepatitis B virus infection         317 (87.6)           Hepatitis C virus infection         356 (98.3)           Yes         6 (1.7)           Alpha-fetoprotein         6 (1.7)           ≤ 25 ng ml <sup>-1</sup> 107 (29.6)           > 25 ng ml <sup>-1</sup> 255 (70.4)           Child-Pugh class         23 (6.4)           A         339 (93.6)           B         23 (6.4)           Differentiation         73 (20.2)           III-IV         73 (20.2)           Tumour number         88 (24.3)           Single         274 (75.7)           Multiple         88 (24.3)           Tumour size         234 (64.6)           Vascular invasion         309 (85.4)           Absent         309 (85.4)           Present         53 (14.6)           TNM stage         11           I.III         259 (71.5)           I.III         259 (71.5)	Gender	
Hepatitis B virus infection           No         45 (12.4) 317 (87.6)           Hepatitis C virus infection         107 (27.6)           No         356 (98.3) Yes           Alpha-fetoprotein         6 (1.7)           Alpha-fetoprotein         255 ng ml <sup>-1</sup> ≤ 25 ng ml <sup>-1</sup> 107 (29.6) > 25 ng ml <sup>-1</sup> > 25 ng ml <sup>-1</sup> 255 (70.4)           Child-Pugh class         23 (6.4)           Differentiation         23 (6.4)           Differentiation         73 (20.2)           Tumour number         73 (20.2)           Single         274 (75.7) 88 (24.3)           Multiple         88 (24.3)           Tumour size         25 cm           ≤ 5 cm         234 (64.6)           Vascular invasion         234 (64.6)           Vascular invasion         309 (85.4) 233 (14.6)           TNM stage         259 (71.5) 1-11           1         259 (71.5) 103 (28.5)	Male Female	324 (89.5) 38 (10.5)
No         45 (12.4)           Yes         317 (87.6)           Hepatitis C virus infection	Hepatitis B virus infection	
Yes         317 (87.6)           Hepatitis C virus infection         356 (98.3) 6 (1.7)           No Yes         6 (1.7)           Alpha-fetoprotein         6 (1.7)           ≤ 25 ng ml <sup>-1</sup> 107 (29.6) > 25 ng ml <sup>-1</sup> > 25 ng ml <sup>-1</sup> 255 (70.4)           Child-Pugh class         339 (93.6) B           A         339 (93.6) B           Differentiation         23 (6.4)           Differentiation         73 (20.2)           Tumour number         73 (20.2)           Single         274 (75.7) 88 (24.3)           Multiple         88 (24.3)           Tumour size         234 (64.6)           ✓ So cm         234 (64.6)           Vascular invasion         309 (85.4) 73 (14.6)           TNM stage         309 (85.4) 1-III           I-III         259 (71.5) 103 (28.5)	No	45 (12.4)
Hepatitis C virus infection           No         356 (98.3) 6 (1.7)           Alpha-fetoprotein         6 (1.7) $\leq 25 \text{ ng ml}^{-1}$ 107 (29.6) 255 (70.4)           Child-Pugh class         339 (93.6) B           A         339 (93.6) B           B         23 (6.4)           Differentiation         289 (79.8) 11-IV           III-IV         73 (20.2)           Tumour number         289 (79.8) 234 (64.6)           Single         274 (75.7) 88 (24.3)           Multiple         289 (274 (75.7) 234 (64.6)           Standard Standa	Yes	317 (87.6)
No         356 (98.3) 6 (1.7)           Alpha-fetoprotein         6 (1.7) $\leq 25 \text{ ng ml}^{-1}$ 107 (29.6) 255 (70.4)           Child-Pugh class         339 (93.6) 23 (6.4)           A         339 (93.6) 23 (6.4)           Differentiation         23 (6.4)           I-II         289 (79.8) 11-IV         73 (20.2)           Tumour number         88 (24.3)         23 (6.4)           Single         274 (75.7) 88 (24.3)         88 (24.3)           Tumour size         388 (24.3)         388 (24.3)           Vascular invasion         234 (64.6)         309 (85.4) 73 (14.6)         73 (14.6)           TNM stage         1         103 (28.5)         103 (28.5)	Hepatitis C virus infection	
Alpha-fetoprotein           ≤25 ng ml <sup>-1</sup> 107 (29.6)           >25 ng ml <sup>-1</sup> 255 (70.4)           Child-Pugh class         339 (93.6)           A         339 (93.6)           B         23 (6.4)           Differentiation         289 (79.8)           III-IV         73 (20.2)           Tumour number         38 (24.3)           Single         274 (75.7)           Multiple         88 (24.3)           Tumour size         234 (64.6)           Vascular invasion         309 (85.4)           Present         339 (93.6)           I         100 (28.5)	No	356 (98.3)
$\leq 25 \text{ ng ml}^{-1}$ 107 (29.6) $> 25 \text{ ng ml}^{-1}$ 255 (70.4)         Child-Pugh class         A       339 (93.6)         B       23 (6.4)         Differentiation         I-II       289 (79.8)         III-IV       73 (20.2)         Tumour number         Single       274 (75.7)         Multiple       88 (24.3)         Tumour size $\leq 5 \text{ cm}$ 128 (35.4) $> 5 \text{ cm}$ 234 (64.6)         Vascular invasion         Absent       309 (85.4)         Present       309 (85.4)         Present       53 (14.6)         TIMI stage         I       1259 (71.5)         I-III       103 (28.5)	Alpha-fetoprotein	
> 25 ng ml <sup>-1</sup> 255 (70.4)         Child-Pugh class       339 (93.6)         A       339 (93.6)         B       23 (6.4)         Differentiation         I-II       289 (79.8)         III-IV       73 (20.2)         Tumour number       73 (20.2)         Single       274 (75.7)         Multiple       88 (24.3)         Tumour size       25 cm         ≤ 5 cm       128 (35.4)         > 5 cm       234 (64.6)         Vascular invasion       309 (85.4)         Present       309 (85.4)         Present       53 (14.6)         TNM stage       1         I       259 (71.5)         I-III       103 (28.5)	$\leq 25 \mathrm{ng}\mathrm{ml}^{-1}$	107 (29.6)
Child-Pugh class         A       339 (93.6)         B       23 (6.4)         Differentiation         I-II       289 (79.8)         III-IV       73 (20.2)         Tumour number         Single       274 (75.7)         Multiple       88 (24.3)         Tumour size $\leq 5  cm$ 128 (35.4) $> 5  cm$ 234 (64.6)         Vascular invasion         Absent       309 (85.4)         Present       309 (85.4)         Present       53 (14.6)         TIMI stage         I       259 (71.5)         II-III       103 (28.5)	> 25 ng ml <sup>-1</sup>	255 (70.4)
A       339 (93.6)         B       23 (6.4)         Differentiation         I-II       289 (79.8)         III-IV       73 (20.2)         Tumour number         Single       274 (75.7)         Multiple       88 (24.3)         Tumour size          ≤ 5 cm       128 (35.4)         > 5 cm       234 (64.6)         Vascular invasion       309 (85.4)         Present       309 (85.4)         Present       53 (14.6)         TNM stage       1         I       259 (71.5)         II-III       103 (28.5)	Child-Pugh class	
B       2.5 (0.4)         Differentiation       289 (79.8)         III-IV       73 (20.2)         Tumour number       73 (20.2)         Single       274 (75.7)         Multiple       88 (24.3)         Tumour size       25 cm         ≤ 5 cm       128 (35.4)         > 5 cm       234 (64.6)         Vascular invasion       309 (85.4)         Present       53 (14.6)         TNM stage       103 (28.5)         I       103 (28.5)	A	339 (93.6) 23 (6.4)
I-II       289 (79.8)         III-IV       73 (20.2)         Tumour number       274 (75.7)         Multiple       88 (24.3)         Tumour size       25 cm         ≤ 5 cm       128 (35.4)         > 5 cm       234 (64.6)         Vascular invasion       309 (85.4)         Present       53 (14.6)         TNM stage       103 (28.5)	Differentiation	23 (0.7)
III-IV     73 (20.2)       Tumour number     73 (20.2)       Single     274 (75.7)       Multiple     88 (24.3)       Tumour size     128 (35.4)       ≤ 5 cm     128 (35.4)       > 5 cm     234 (64.6)       Vascular invasion     309 (85.4)       Present     53 (14.6)       TNM stage     103 (28.5)		280 (79 8)
Tumour number           Single         274 (75.7)           Multiple         88 (24.3)           Tumour size	III–IV	73 (20.2)
Single Multiple         274 (75.7) 88 (24.3)           Tumour size         38 (24.3)           ≤ 5 cm         128 (35.4) 234 (64.6)           Vascular invasion         309 (85.4) 7 sent           Absent Present         309 (85.4) 53 (14.6)           TNM stage         259 (71.5) 103 (28.5)	Tumour number	
Multiple         00 (24.3)           Tumour size            ≤ 5 cm         128 (35.4)           > 5 cm         234 (64.6)           Vascular invasion         309 (85.4)           Present         53 (14.6)           TNM stage         259 (71.5)           I         259 (71.5)           II-III         103 (28.5)	Single	274 (75.7)
Solution         Solution           ≤ 5 cm         128 (35.4)           > 5 cm         234 (64.6)           Vascular invasion         309 (85.4)           Absent         309 (85.4)           Present         53 (14.6)           TNM stage         259 (71.5)           I         259 (71.5)           II-III         103 (28.5)		00 (24.3)
< 5 cm	l'umour size	
Vascular invasion           Absent         309 (85.4)           Present         53 (14.6)           TNM stage         1           I         259 (71.5)           II-III         103 (28.5)	≤5 cm >5 cm	128 (35.4) 234 (64.6)
Absent         309 (85.4)           Present         53 (14.6)           TNM stage         259 (71.5)           I         259 (71.5)           II-III         103 (28.5)	Vascular invasion	
Present         53 (14.6)           TNM stage         259 (71.5)           I         259 (71.5)           II-III         103 (28.5)	Absent	309 (85.4)
I         259 (71.5)           II–III         103 (28.5)	Present	53 (14.6)
I 259 (71.5) II–III 103 (28.5)	TNM stage	
II-III 103 (20.3)		259 (71.5)
	11–111	103 (20.3)

diagnosed with tumour recurrence, and 106 (29.3%) remained alive without recurrence. Overall survival was defined as the interval between surgery and death or between surgery and the last observation for surviving patients. The time to recurrence was defined as the interval between surgery and recurrence or between surgery and the last observation for patients without recurrence.

**Tissue microarray and immunohistochemistry.** The tissue microarray (TMA) was constructed as described previously (Xu *et al*, 2012b). Briefly, blocks containing the advancing edges of tumoural and peri-tumoural HCC tissue were used for TMA construction. Haematoxylin and eosin (H&E)-stained slides were reviewed without the knowledge of the patient clinical characteristics and outcomes. Duplicate 1.0-mm tissue cores were obtained from



Figure 1. Tumour-associated antigen (TAA) expression in hepatocellular carcinoma (HCC). (A) Immunohistochemical detection of GPC3, AFP, NY-ESO-1, SSX-2, EpCAM, MDK in HCC tumour tissue (× 400 magnification). (B) Frequency of expression of the six TAAs in HCC tumour tissue determined by immunohistochemistry. (C) Frequency of coexpression of the indicated numbers of TAA (in any combination) in HCC tumour tissue.
(D) TAA expression in each sample. Each column represents the TAA expression in an individual patient. Blue bar, positive expression.
(E) The inter-relationship between each TAA in HCC tumour tissue. Values denote the Pearson correlation coefficients; values closer to 1 indicate a better correlation. \*P<0.05; \*\*P<0.001.</li>

two regions (total, four punches) in the paraffin-embedded tissue blocks. Tissue microarrays containing the tissue cores were then cut into  $5\,\mu m$  sections for immunohistochemistry (IHC) staining.

The IHC of the paraffin sections was carried out using a two-step protocol (DakoCytomation, Glostrup, Denmark). Briefly,  $5 \,\mu m$  sections were deparaffinised, and endogenous peroxidase activity was blocked with 0.3% hydrogen peroxide containing double-

distilled water for 10 min. Antigen retrieval was performed by microwave treatment in  $0.01 \text{ mol l}^{-1}$  citrate buffer, pH 6.0. Primary antibodies (Abs) were applied and incubated at 4 °C overnight. Antigen retrieval was developed with peroxidase and 3,3'-diaminobenzidine tetrahydrochloride. The sections were then counterstained with haematoxylin (Zymed Laboratories Inc.) and mounted in nonaqueous mounting medium. Antibodies information is

summarised in Supplementary Table 1. The appropriate negative controls were used, in which the primary Abs were replaced by irrelevant, isotype-matched Abs at the same concentration. If necessary, positive control tissues were applied as suggested by the manufacturers.

**Evaluation of immunohistochemical variables.** All TMA cores were screened and evaluated with a computerised image analysis platform constructed using the TMAJ Image application (http://tmaj.pathology.jhmi.edu). The expression of each antigen was scored according to the proportion of expression, as previously described (Xu *et al*, 2012b).

We initially investigated eight TAAs with identified immunogenicity and potential therapeutic function (Breous and Thimme, 2011): AFP (Thimme *et al*, 2008), GPC3 (Sawada *et al*, 2012), SSX-2 (Bricard *et al*, 2005), NY-ESO-1 (Korangy *et al*, 2004; Xu *et al*, 2012a), EpCAM (Yamashita *et al*, 2008), MDK (Jia *et al*, 2007; Kerzerho *et al*, 2010), melanoma antigen gene-A (MAGE-A) (Bricard *et al*, 2005), and telomerase reverse transcriptase (hTERT) (Mizukoshi *et al*, 2011). However, the expression of MAGE-A was below 10% and the immunodetection of hTERT was unsatisfactory due to nonspecific IHC staining in HCC (data not shown). For the six TAAs we eventually examined (AFP, GPC3, SSX-2, NY-ESO-1, EpCAM, and MDK), any proportion of any positive degree of intensity was considered positive. For the densities of lymphocytic infiltration, median cut-off points were used for the definition of subgroups.

**Statistical analysis.** Kaplan–Meier estimates were calculated and compared using the log-rank test. A multivariate Cox proportional hazard regression model was applied to estimate the adjusted hazard ratio (HR) and 95% confidence interval (CI) and to identify independent prognostic factors. The association between variables was evaluated using the  $\chi^2$  test or Fisher's exact test when appropriate. P < 0.05 was considered to indicate statistical significance. Statistical analyses were performed with SPSS 13.0 (SPSS Inc., Chicago, IL, USA).

#### RESULTS

Expression of TAAs in HCC tissue. Immunohistochemistry was performed to analyse the expression of TAAs in HCC parenchyma. Of eight initial TAAs, six (GPC3, AFP, SSX-2, NY-ESO-1, EpCAM, and MDK; Figure 1A) were eventually included and evaluated in a panel of 362 HCC patients. Most of the TAAs were frequently expressed and were detected in over 50% of the samples, except for NY-ESO-1, which was detected in only 14.6% (53/362) of the samples (Figure 1B). The frequency for coexpression of two to five TAA was 11.9% (43/362), 26.5% (96/362), 29.8% (108/362), and 20.4% (74/362), respectively. Only 1.4% (5/362) of the samples revealed no expression of any TAA, and 5% (18/362) of the samples contained coexpression of all six TAAs (Figure 1C and D). Relevance analysis showed that several antigens were significantly associated with one another, indicating that different antigens might be regulated simultaneously during immunoediting (Figure 1E).

Association between TAAs and disease progression. Patients were categorised into three groups according to the number of positive TAAs (TAA index) in the tumour tissue. As summarised in Table 2, tumour differentiation and tumour size were significantly associated with TAA expression (P = 0.009 and 0.018, respectively). In cases with less TAA expression, patients tended to have larger tumours and poor differentiation. For example, only 4.8% (14/289) of patients with grade I-II tumours expressed none or one of the six TAAs as compared with 12.3% (9/73) of patients with grade III-IV tumours, indicating that TAA

### Table 2. Association between TAA expression and patients' clinical characteristics

	Т			
Characteristics	0–1	2–3	4–6	<b>P</b> -value
Age, year				0.115
Median Range	43 25–62	50 20–77	48 21–78	
Gender				0.609
Male Female	22 1	124 15	178 22	
Hepatitis B virus infection				0.089
No Yes	4 19	23 116	18 182	
Hepatitis C virus infection				0.579
No Yes	22 1	137 2	197 3	
Alpha-fetoprotein				0.097
$\leq$ 25 ng ml <sup>-1</sup> > 25 ng ml <sup>-1</sup>	9 14	48 91	50 150	
Child-Pugh class				0.242
A B	20 3	133 6	186 14	
Differentiation				0.009
-      - V	14 9	120 19	155 45	
Tumour number				0.568
Single Multiple	18 5	101 38	155 45	
Tumour size				0.018
≤5 cm >5 cm	2 21	49 90	77 123	
Vascular invasion				0.239
Absent Present	17 6	118 21	174 26	
TNM stage				0.366
-   	15 8	95 44	149 51	
Abbreviations: TAA = tumour-associ Bold values indicate statistical signifi	ated antigen	TNM = tumo	our-lymph no	de metastasis.

deficiency might be another special feature of poorly differentiated tumours.

Kaplan-Meier survival curves were plotted to investigate the correlation between the expression of each TAA and patient survival. The log-rank statistic was used to compare survival rates. The expression of SSX-2, EpCAM, and MDK was positively correlated with better OS (P=0.024, 0.0001, and 0.017, respectively; Figure 2). Interestingly, survival was better in the patients with a higher TAA index (Figure 2G). The OS of patients who expressed more than four TAAs (median survival >60 months) was prolonged as compared with patients who expressed zero or one TAA (median survival, 18 months). The five-year OS rate in patients with 0-1 TAA expression was only 13.6% as compared with the 37.2% of those who expressed 4–6 TAAs. However, there was no significant association between TAA expression and recurrence in this cohort (Supplementary Figure 1).



Figure 2. TAA expression and overall survival (OS). Kaplan–Meier survival estimates of the OS were performed according to the expression of (A) GPC3, (B) AFP, (C) NY-ESO-1, (D) SSX-2, (E) EpCAM, (F) MDK, and (G) the TAA index in hepatocellular carcinoma (HCC) tumour tissue.

We next assessed whether the TAA index could serve as an independent predictor of OS. A multivariate Cox proportional hazards analysis was performed, and the variables that were associated with survival by univariate analysis were adopted as covariates (summarised in Table 3). Serum AFP level, tumour differentiation, tumour size, and vascular invasion remained associated with OS in the multivariate Cox proportional hazards analysis (P = 0.019, 0.002, 0.001, and < 0.0001, respectively). The TAA index could still predict OS independent of these clinical factors (HR, 0.625; 95% CI, 0.467–0.837; P = 0.002; Table 3).

Association between TAA expression and local immune cell infiltration. Recent studies by our and other groups have shown that local immune cell status could influence HCC progression (Gao *et al*, 2007; Ding *et al*, 2009; Zhou *et al*, 2009; Wu *et al*, 2013). Therefore, we also assessed the infiltration of different type of immune cells (Supplementary Figure 2) in the tumour micro-environment, and their relationship with TAA expression. As shown in Figure 3 and Supplementary Table 2, there was a positive

correlation between AFP or EpCAM and CD20<sup>+</sup> B cell or CD57<sup>+</sup> natural killer (NK) cell densities, but a negative correlation between SSX-2 and FoxP3<sup>+</sup> regulatory T (Treg) cells. However, NY-ESO-1 expression was significantly positively correlated with CD8<sup>+</sup> T, B, NK, and Treg cells. Moreover, patients who coexpressed more TAAs tended to have more B and NK cells, but not CD15<sup>+</sup> neutrophils, CD68<sup>+</sup> macrophages, or Treg cells infiltrating in the tumour. In addition, there was no correlation between the TAA index and immune cell infiltration in the peri-tumour region. Taken together, a higher TAA index could indicate a microenvironment with more anti-tumour immune cell infiltration but not tumour-educated regulatory cells in the HCC nest.

Prognostic values with combination of TAA index and immune infiltration. We next evaluated the combined influence of the TAA index and immune cell densities  $(CD3^+, CD4^+, CD8^+, CD20^+, CD57^+, and FoxP3^+$  cells; Figure 4). Patients with a high TAA index (4–6 TAA coexpression) and high density of T (including CD3<sup>+</sup>, CD4<sup>+</sup>, and CD8<sup>+</sup> cells) and CD20<sup>+</sup> B or

#### Table 3. Cox proportional Hazard regression analysis of patients' overall survival

	Univariable			Multivariable		
Variables	Hazard ratio	95% CI	<b>P</b> -value	Hazard ratio	95% CI	<b>P</b> -value
Gender (male vs female)	1.073	0.842-1.368	0.568			NA
Hepatitis B virus infection (absent vs present)	1.218	0.781-1.901	0.384			NA
Hepatitis C virus infection (absent vs present)	1.852	0.687–4.994	0.223			NA
Alpha-fetoprotein ( $\leq$ 25 ng ml <sup>-1</sup> vs > 25 ng ml <sup>-1</sup> )	1.66	1.189–2.316	0.003	1.505	1.070–2.118	0.019
Child-Pugh class (A vs B)	1.17	0.679–2.016	0.572			NA
Differentiation (I–II vs III–IV)	1.749	1.254-2.441	0.001	1.716	1.219–2.416	0.002
Tumour number (single vs multiple)	1.233	0.890-1.707	0.208			NA
Tumour size ( $\leq$ 5 cm vs > 5 cm)	2.109	1.516-2.934	<0.0001	1.796	1.266–2.547	0.001
Vascular invasion (absent vs present)	3.935	2.770-5.591	<0.0001	3.399	2.061-5.605	< 0.0001
TNM stage (I–II vs III)	2.132	1.583–2.871	<0.0001	0.926	0.595–1.442	0.735
TAA index (0–3 vs 4–6)	0.626	0.471-0.833	0.001	0.625	0.467-0.837	0.002

Abbreviations: CI = confidence interval; NA = not applicable; TAA = tumour-associated antigen; TNM = tumour-lymph node metastasis. Note: Cox proportional hazards regression model; variables associated with survival by univariate analysis were adopted as covariates in multivariate analyses. Bold values indicate statistical significance.

		AFP	GPC3	NY- ESO-1	SSX-2	EpCAM	MDK	TAA index
	CD3	-0.017	0.034	0.086	-0.063	0.017	0.104*	-0.011
	CD4	-0.076	0.056	0.086*	-0.025	-0.040	0.021	-0.033
	CD8	0.041	0.034	0.117*	-0.025	0.006	-0.035	-0.022
nour	CD20	0.099	0.147**	0.133*	-0.038	0.110*	0.077	0.122*
Tur	CD57	-0.64	0.124*	0.164**	0.038	0.121*	0.090	0.111*
	FoxP3	0.87	0.068	0.227**	-0.126*	-0.087	-0.007	0.033
	CD15	-0.29	0.011	0.102	-0.101	-0.098	-0.035	-0.022
	CD68	-0.64	-0.045	-0.023	0.025	0.098	-0.063	0.033
	CD3	0.099	0.045	0.008	-0.088	-0.017	0.035	0.033
	CD4	0.099	0.045	-0.086	-0.088	0.017	0.049	0.067
5	CD8	0.122*	0.011	0.008	-0.151**	-0.040	0.035	-0.022
Peri-tumou	CD20	-0.017	0.034	0.086	-0.113*	-0.063	0.021	-0.044
	CD57	0.017	0.034	0.086	-0.013	-0.063	-0.007	-0.022
	CD15	0.087	0.079	0.023	-0.189**	-0.133*	-0.007	-0.011
	CD68	0.006	0.068	0.133*	-0.126*	-0.063	-0.007	0.011
							Low	High

Correlation

Figure 3. The inter-relationship between tumour-associated antigen (TAA) expression and immune cell infiltration in tumoural and peri-tumoural regions. Values denote the Pearson correlation coefficients; values closer to 1 indicate a better correlation. \*P < 0.05; \*\*P < 0.001.

 $CD57^+$  NK cells all had longer survival time (median survival >60 months). Patients with a low TAA index (0–3 TAA coexpression) and low density of these effective immune cells

had shorter survival time (median survival  $\leq 28$  months; Figure 4). Supporting the general view that Treg cells suppress tumour immunity, low FoxP3<sup>+</sup> Treg cell infiltration with a high TAA index predicted better OS (median survival >60 months) as compared with any other subgroup (P<0.01, Figure 4F). Patients with high Treg density and a low TAA index had the shortest survival time (median survival, 29 months). These results demonstrated that the TAA index and immune cell infiltration could together determine tumour progression and patient survival.

#### DISCUSSION

It is commonly believed that poorly immunogenic transformed cells that escape immune surveillance would lead to the primary appearance of overt cancer and/or post-treatment relapse; however, little is known about the clinical value of the immunogenic features of malignant cells in solid tumours, including HCC. This study provides the first evidence that the expression pattern of multiple TAAs is associated with cancer progression and postsurgical prognosis in HCC patients.

Several other groups have reported the expression frequencies of single TAA, mostly on an mRNA level (Yamashita *et al*, 2008; Wang *et al*, 2009). In the present study, we further demonstrated the expression and clinical value of multiple antigens on a protein level, which should be the critical reason for heterogeneous effectiveness of immunotherapy and provide supportive evidence for 'immunoediting' in human cancers. The host immune system not only controls tumour quantity, but also shapes tumour immunogenicity. The finding that the coexpression of more TAAs was associated with better-differentiated and/or smaller tumours and the association between the expression of several antigens can aid in our understanding of the nature of immunoedited tumour progression.

Therefore, the expression patterns of TAAs in human tumours should be a result of long-term co-evolution between the immune system and malignant cells, and would be closely related to patient prognosis after surgical interventions and/or other treatments such as immunotherapies. Further analysis of the immune cell infiltration provided more evidence for the association between TAAs and tumour progression. Generally, the expression of single TAAs was positively associated with the infiltration of different lymphocyte subsets, including CD3<sup>+</sup> (Galon *et al*, 2006), CD8<sup>+</sup>







Months

TI highCD3low

0.002

0.193

0 889

TI lowCD3low

0.002

0.045

0.001

TI lowCD3lov

TI highCD3lc

TI lowCD3high

TI highCD3high



TI lowCD3high

0.045

0.193

0.160

TI highCD3high

0.001

0.889

0.160

В

Overall survival (%)

TI lowCD4low

TI highCD4low

TI lowCD4high

TI highCD4high

D

0.007

0.208

0.001



0.007

0.145

0.567



0.208

0.145

0.057

BRITISH JOURNAL OF CANCER

0.001

0.567

0.057



	TI lowCD8low	TI highCD8low	TI lowCD8high	TI highCD8high
TI lowCD8low		0.009	0.177	0.001
TI highCD8low	0.009		0.204	0.449
TI lowCD8high	0.177	0.204		0.042
TI highCD8high	0.001	0.449	0.042	





	TI lowCD20low	TI highCD20low	TI lowCD20high	TI highCD20high
TI lowCD20low		0.0004	0.084	0.004
TI highCD20low	0.0004		0.101	0.381
TI lowCD20high	0.084	0.101		0.365
TI highCD20high	0.004	0.381	0.365	



	TI lowFoxP3low	TI highFoxP3low	TI lowFoxP3 <sup>high</sup>	TI highFoxP3high
TI lowFoxP3low		0.002	0.166	0.698
TI highFoxP3low	0.002		< 0.0001	0.01
TI lowFoxP3 <sup>high</sup>	0.166	< 0.0001		0.068
TI highFoxP3high	0.698	0.01	0.068	

Figure 4. The combination of tumour-associated antigen (TAA) expression and immune cell infiltration correlates with overall survival (OS). Kaplan-Meier curves illustrate the duration of OS according to the TAA index (TI) and the density of (A) CD3<sup>+</sup>, (B) CD4<sup>+</sup>, (C) CD8<sup>+</sup>, (D) CD20<sup>+</sup>, (E) CD57<sup>+</sup>, or (F) FoxP3<sup>+</sup> cells in the tumour region. Red values indicate statistical significance.

(Gao et al, 2007),  $CD20^+$  (Nielsen et al, 2012), and  $CD57^+$  cells (Wu et al, 2013), which have all been reported to facilitate antitumour immunity, and was negatively associated with FoxP3+ Treg cells (Zhou et al, 2009). Recently, Chew et al (2012) reported that a given set of chemokines was correlated with lymphocyte infiltration and prognosis in HCC, which also support the protective role of anti-tumour immune milieu in HCC progression. Tumours coexpressing more TAAs tended to have more CD20<sup>+</sup> B

and CD57<sup>+</sup> NK cells, but not FoxP3<sup>+</sup> Treg cells or other inflammatory cells, including CD15<sup>+</sup> neutrophils (Kuang *et al*, 2011) and CD68<sup>+</sup> macrophages (Ding *et al*, 2009), which are often exploited by solid tumours to establish a tumour-promoting microenvironment. Interestingly, the unexpected result of relationship between NY-ESO-1 and FoxP3 indicated that self-TAAs might also be involved in immune homeostasis and limiting acute inflammation. Taken together, the expression of more TAAs might promote anti-tumour immune reaction or surveillance and facilitate the postsurgical recovery of HCC patients.

It is conventionally believed that the adaptive immune response mediated by tumour-infiltrating lymphocytes (TILs) influences the behaviour of human cancers (Abastado, 2012). High densities of CD3<sup>+</sup> cells in the centre of a tumour and the invasive margin of colorectal tumours predict better clinical outcomes (Galon et al, 2006). For HCC, however, results by ours (Supplementary Figure 3) and other have shown that the infiltration of total T lymphocytes was not associated with postsurgical prognosis (Gao et al, 2007). In the present study, we stratified the patients according to the TAA index, and the density of CD3<sup>+</sup> cells was positively associated with the OS in patients with a low TAA index (Figure 4A). Subsequently, apart from the type and density of immune cells, features of the tumour itself, such as TAA expression, should also be important factors of immune regulation in HCC. Therefore, the combination of the TAA index and the densities of immune cells could further predict HCC prognosis. Recently, we developed an immune-based in situ molecular classifier that could aid in the identification of patients who are at greatest risk for postsurgical recurrence of HCC (Xu et al, 2012b). The predictive values of TAAs could provide more parameters to optimise molecular classifiers for HCC outcomes. Of course, other tumour cell features (such as proliferation) should also be considered important during early cancer evolution and later progression. In tumours with weak proliferation (low Ki-67), the TAA index was closely associated with better prognosis, while all of the patients with intensive proliferation had poor prognosis (Supplementary Figure 4). In general, the coactions of immunoediting and the vital power of tumour cells could continue shaping malignancies and influence patient survival after treatments, including resections and/or biological therapies.

Although clinical trials involving immunotherapy with T-cell clones specific for a single antigen have provided a foundation for proof-of-principle studies, reduced clinical efficacy has been encountered in contrast to the substantial therapeutic impact of transfer with polyclonal TIL cultures. The outgrowth of antigenloss tumour variants in treated patients indicates the ability of rapidly adaptable tumour cells to evade narrowly focused therapies (Mellman et al, 2011; He et al, 2012). Recently, new therapies based on sophisticated knowledge of the suppressive tumour immune microenvironment were designed to overcome tolerance and reactivate anti-tumour immunity to induce potent, long-lasting responses (Mellman et al, 2011). For example, in early-phase clinical trials involving patients with advanced solid tumours such as metastatic melanoma, renal cell carcinoma, colorectal cancer, and non-small-cell lung cancer, monoclonal Abs against immune-checkpoint proteins (such as ipilimumab, tremelimumab, and MDX-1106) could induce a state of equilibrium between the immune system and cancer, resulting in prolonged disease stabilisation. Nevertheless, only a relatively small fraction of patients exhibited an objective response and derived clinical benefits (Topalian et al, 2011). In view of this, the discrepancies in the TAA profiles should be a critical reason for heterogeneous therapeutic efficacy. At present, immunotherapies that interrupt the tolerogenic pathways and reactivate endogenous immunity are being evaluated, appearing to be a promising HCC treatment option (primary or adjuvant for chemotherapy and/or surgery). To prevent

overtreatment and to achieve more convincing results, molecular classification based on TAA expression patterns should also be an important strategy in clinical trials of immunotherapy.

In brief, TAA expression patterns could serve as important prognostic factors in HCC. Tumour-associated antigen expression should be associated with anti-tumour immune infiltration, and particularly, involved in disease progression and the reconstitution of immune surveillance after surgical intervention. Moreover, our results could provide a new evidence for improvement of the prognostic molecular signatures in HCC, and a potential rational consideration for patient enrolment in future immunotherapeutic trials and/or clinical treatments.

#### ACKNOWLEDGEMENTS

This work was supported by Project Grants from the Ministry of Health of China (2012ZX10002-011).

#### CONFLICT OF INTEREST

The authors declare no conflict of interest.

#### REFERENCES

- Abastado JP (2012) The next challenge in cancer immunotherapy: controlling T-cell traffic to the tumor. *Cancer Res* **72**: 2159–2161.
- Breous E, Thimme R (2011) Potential of immunotherapy for hepatocellular carcinoma. *J Hepatol* 54: 830–834.
- Bricard G, Bouzourene H, Martinet O, Rimoldi D, Halkic N, Gillet M, Chaubert P, Macdonald HR, Romero P, Cerottini JC, Speiser DE (2005) Naturally acquired MAGE-A10- and SSX-2-specific CD8 + T cell responses in patients with hepatocellular carcinoma. *J Immunol* 174: 1709–1716.
- Bruix J, Llovet JM (2009) Major achievements in hepatocellular carcinoma. Lancet 373: 614–616.
- Chew V, Chen J, Lee D, Loh E, Lee J, Lim KH, Weber A, Slankamenac K, Poon RT, Yang H, Ooi LL, Toh HC, Heikenwalder M, Ng IO, Nardin A, Abastado JP (2012) Chemokine-driven lymphocyte infiltration: an early intratumoural event determining long-term survival in resectable hepatocellular carcinoma. *Gut* **61**: 427–438.
- Ding T, Xu J, Wang F, Shi M, Zhang Y, Li SP, Zheng L (2009) High tumorinfiltrating macrophage density predicts poor prognosis in patients with primary hepatocellular carcinoma after resection. *Hum Pathol* 40: 381–389.
- Ding T, Xu J, Zhang Y, Guo RP, Wu WC, Zhang SD, Qian CN, Zheng L (2011) Endothelium-coated tumor clusters are associated with poor prognosis and micrometastasis of hepatocellular carcinoma after resection. *Cancer* 117: 4878–4889.
- Disis ML (2010) Immune regulation of cancer. J Clin Oncol 28: 4531-4538.
- Fan ST, Mau LoC, Poon RT, Yeung C, Leung Liu C, Yuen WK, Ming Lam C, Ng KK, Ching Chan S (2011) Continuous improvement of survival outcomes of resection of hepatocellular carcinoma: a 20-year experience. *Ann Surg* 253: 745–758.
- Fridman WH, Pagès F, Sautès-Fridman C, Galon J (2012) The immune contexture in human tumours: impact on clinical outcome. *Nat Rev Cancer* **12**: 298–306.
- Fu YX (2012) New immune therapy targets tumor-associated environment: from bone marrow to tumor site. *Cell Mol Immunol* **9**: 1–2.
- Gao Q, Qiu SJ, Fan J, Zhou J, Wang XY, Xiao YS, Xu Y, Li YW, Tang ZY (2007) Intratumoral balance of regulatory and cytotoxic T cells is associated with prognosis of hepatocellular carcinoma after resection. *J Clin Oncol* 25: 2586–2593.
- Gao Q, Shi Y, Wang X, Zhou J, Qiu S, Fan J (2012) Translational medicine in hepatocellular carcinoma. *Front Med* **6**: 122–133.
- Galon J, Costes A, Sanchez-Cabo F, Kirilovsky A, Mlecnik B, Lagorce-Pagès C, Tosolini M, Camus M, Berger A, Wind P, Zinzindohoué F, Bruneval P,

Cugnenc PH, Trajanoski Z, Fridman WH, Pagès F (2006) Type, density, and location of immune cells within human colorectal tumors predict clinical outcome. *Science* **313**: 1960–1964.

- Grivennikov SI, Greten FR, Karin M (2010) Immunity, inflammation, and cancer. *Cell* **140**: 883–899.
- He J, Tang XF, Chen QY, Mai HQ, Huang ZF, Li J, Zeng YX (2012) Ex vivo expansion of tumor-infiltrating lymphocytes from nasopharyngeal carcinoma patients for adoptive immunotherapy. *Chin J Cancer* **31**: 287–294.
- Jemal A, Bray F, Center MM, Ferlay J, Ward E, Forman D (2011) Global cancer statistics. *CA Cancer J Clin* **61**: 69–90.
- Jia HL, Ye QH, Qin LX, Budhu A, Forgues M, Chen Y, Liu YK, Sun HC, Wang L, Lu HZ, Shen F, Tang ZY, Wang XW (2007) Gene expression profiling reveals potential biomarkers of human hepatocellular carcinoma. *Clin Cancer Res* 13: 1133–1139.
- Korangy F, Ormandy LA, Bleck JS, Klempnauer J, Wilkens L, Manns MP, Greten TF (2004) Spontaneous tumor-specific humoral and cellular immune responses to NY-ESO-1 in hepatocellular carcinoma. *Clin Cancer Res* 10: 4332–4341.
- Kuang DM, Zhao Q, Wu Y, Peng C, Wang J, Xu Z, Yin XY, Zheng L (2011) Peritumoral neutrophils link inflammatory response to disease progression by fostering angiogenesis in hepatocellular carcinoma. *J Hepatol* 54: 948–955.
- Kerzerho J, Adotevi O, Castelli FA, Dosset M, Bernardeau K, Szely N, Lang F, Tartour E, Maillere B (2010) The angiogenic growth factor and biomarker midkine is a tumor-shared antigen. J Immunol 185: 418–423.
- Qin LX (2012) Inflammatory immune responses in tumor microenvironment and metastasis of hepatocellular carcinoma. *Cancer Microenviron* 5: 203–209.
- Mellman I, Coukos G, Dranoff G (2011) Cancer immunotherapy comes of age. *Nature* **480**: 480–489.
- Mizukoshi E, Nakamoto Y, Arai K, Yamashita T, Sakai A, Sakai Y, Kagaya T, Yamashita T, Honda M, Kaneko S (2011) Comparative analysis of various tumor-associated antigen-specific T-cell responses in patients with hepatocellular carcinoma. *Hepatology* 53: 1206–1216.
- Nielsen JS, Sahota RA, Milne K, Kost SE, Nesslinger NJ, Watson PH, Nelson BH (2012) CD20 + tumor-infiltrating lymphocytes have an atypical CD27- memory phenotype and together with CD8 + T cells promote favorable prognosis in ovarian cancer. *Clin Cancer Res* 18: 3281–3292.
- Sawada Y, Yoshikawa T, Nobuoka D, Shirakawa H, Kuronuma T, Motomura Y, Mizuno S, Ishii H, Nakachi K, Konishi M, Nakagohri T, Takahashi S, Gotohda N, Takayama T, Yamao K, Uesaka K, Furuse J, Kinoshita T, Nakatsura T (2012) Phase I trial of a glypican-3-derived peptide vaccine for advanced hepatocellular carcinoma: immunologic evidence and potential for improving overall survival. *Clin Cancer Res* 18: 3686–3696.
- Schreiber RD, Old LJ, Smyth MJ (2011) Cancer immunoediting: integrating immunity's roles in cancer suppression and promotion. *Science* 331: 1565–1570.

- Thimme R, Neagu M, Boettler T, Neumann-Haefelin C, Kersting N, Geissler M, Makowiec F, Obermaier R, Hopt UT, Blum HE, Spangenberg HC (2008) Comprehensive analysis of the alpha-fetoprotein-specific CD8 + T cell responses in patients with hepatocellular carcinoma. *Hepatology* 48: 1821–1833.
- Topalian SL, Weiner GJ, Pardoll DM (2011) Cancer immunotherapy comes of age. J Clin Oncol 29: 4828–4836.
- Vesely MD, Kershaw MH, Schreiber RD, Smyth MJ (2011) Natural innate and adaptive immunity to cancer. Annu Rev Immunol 29: 235–271.
- Villanueva A, Hernandez-Gea V, Llovet JM (2012) Medical therapies for hepatocellular carcinoma: a critical view of the evidence. *Nat Rev Gastroenterol Hepatol* 10: 34–42.
- Villanueva A, Hoshida Y, Toffanin S, Lachenmayer A, Alsinet C, Savic R, Cornella H, Llovet JM (2010) New strategies in hepatocellular carcinoma: genomic prognostic markers. *Clin Cancer Res* 16: 4688–4694.
- Wang XY, Chen HS, Luo S, Zhang HH, Fei R, Cai J (2009) Comparison for detecting NY-ESO-1 mRNA expression levels in hepatocellular carcinoma tissues. Oncol Rep 21: 713–719.
- Wu Y, Kuang DM, Pan WD, Wan YL, Lao XM, Wang D, Li XF, Zheng L (2013) Monocyte/macrophage-elicited natural killer cell dysfunction in hepatocellular carcinoma is mediated by CD48/2B4 interactions. *Hepatology* 57: 1107–1116.
- Xu H, Gu N, Liu ZB, Zheng M, Xiong F, Wang SY, Li N, Lu J (2012a) NY-ESO-1 expression in hepatocellular carcinoma: A potential new marker for early recurrence after surgery. Oncol Lett 3: 39–44.
- Xu J, Ding T, He Q, Yu XJ, Wu WC, Jia WH, Yun JP, Zhang Y, Shi M, Shao CK, Pan WD, Yin XY, Min J, Zhuang SM, Zheng L (2012b) An in situ molecular signature to predict early recurrence in hepatitis B virus-related hepatocellular carcinoma. J Hepatol 57: 313–321.
- Yamashita T, Forgues M, Wang W, Kim JW, Ye Q, Jia H, Budhu A, Zanetti KA, Chen Y, Qin LX, Tang ZY, Wang XW (2008) EpCAM and alpha-fetoprotein expression defines novel prognostic subtypes of hepatocellular carcinoma. *Cancer Res* 68: 1451–1461.
- Yang JD, Roberts LR (2010) Hepatocellular carcinoma: A global view. Nat Rev Gastroenterol Hepatol 7: 448–458.
- Zhou J, Ding T, Pan W, Zhu LY, Li L, Zheng L (2009) Increased intratumoral regulatory T cells are related to intratumoral macrophages and poor prognosis in hepatocellular carcinoma patients. *Int J Cancer* 125: 1640–1648.

This work is published under the standard license to publish agreement. After 12 months the work will become freely available and the license terms will switch to a Creative Commons Attribution-NonCommercial-Share Alike 3.0 Unported License.

Supplementary Information accompanies this paper on British Journal of Cancer website (http://www.nature.com/bjc)