

# Enrichment of heterozygous germline *RECQL4* loss-of-function variants in pediatric osteosarcoma

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Abstract Patients harboring germline pathogenic biallelic variants in genes involved in the recognition and repair of DNA damage are known to have a substantially increased cancer risk. Emerging evidence suggests that individuals harboring heterozygous variants in these same genes may also be at heightened, albeit lesser, risk for cancer. Herein, we sought to determine whether heterozygous variants in RECQL4, the gene encoding an essential DNA helicase that is defective in children with the autosomal recessive cancer-predisposing condition Rothmund-Thomson syndrome (RTS), are associated with increased risk for childhood cancer. To address this question, we interrogated germline sequence data from 4435 pediatric cancer patients at St. Jude Children's Research Hospital and 1127 from the National Cancer Institute Therapeutically Applicable Research to Generate Effective Treatment (TARGET) database and identified 24 (0.43%) who harbored loss-of-function (LOF) RECOL4 variants, including five of 249 (2.0%) with osteosarcoma (OS). These RECQL4 variants were significantly overrepresented in children with OS, the cancer most frequently observed in patients with RTS, as compared to 134,187 noncancer controls in the Genome Aggregation Database (gnomAD v2.1; P = 0.00087, odds ratio [OR] = 7.1, 95% CI, 2.9-17). Nine of the 24 (38%) individuals possessed the same c.1573delT (p.Cys525Alafs) variant located in the highly conserved DNA helicase domain, suggesting that disruption of this domain is central to oncogenesis. Altogether these data expand our understanding of the genetic factors predisposing to childhood cancer and reveal a novel association between heterozygous RECQL4 LOF variants and development of pediatric OS.

[Supplemental material is available for this article.]

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#### Ontology terms:

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## INTRODUCTION

The RecQ like helicase 4 (RECQL4) gene encodes a helicase essential for repairing DNA damage and maintaining genomic stability (Lu et al. 2016). Pathogenic homozygous or compound heterozygous variants affecting RECQL4 cause three clinically overlapping autosomal recessive (AR) disorders, namely Rothmund-Thomson syndrome (RTS), Baller-Gerold syndrome (BGS), and RAPADILINO (RAdial ray defect, PAtellar aplasia/arched or cleft PAlate, Dlarrhea/Disclosed joints, LIttle size/LImb malformation, NOse slender/NOrmal intelligence) syndrome (Wang and Plon 2019). Rothmund–Thomson syndrome is the most prevalent of these syndromes, with nononcologic features including poikiloderma; sparse hair, eyelashes, and eyebrows; short stature; dental abnormalities; dysplastic or poorly formed nails; and gastrointestinal problems in infancy (Siitonen et al. 2009). BGS is the rarest of these disorders and is characterized by craniosynostosis, radial ray defects, short stature, and malformed or missing patellae (Van Maldergem et al. 2007). Finally, RAPADILINO syndrome is characterized by short stature, radial and patellar aplasia or hypoplasia, absence of thumbs, dislocation of joints, highly arched palate, infantile diarrhea, and pigmentary changes or café au lait macules (Siitonen et al. 2003). A predisposition to cancer has been reported in all three syndromes, with the highest cancer occurrence in RTS. Patients with RTS, and to a lesser degree RAPADILINO, are at greatest risk to develop osteosarcoma (OS) (Siitonen et al. 2003; Cao et al. 2017; Wang and Plon 2019), whereas patients with all three conditions are at risk for lymphoma (Van Maldergem et al. 2007; Siitonen et al. 2009).

Through a clinical research protocol (NCT02530658), we identified a child with OS for whom germline whole-exome and whole-genome sequencing (WGS) and in-depth analysis of 156 cancer predisposition genes revealed a pathogenic alteration and a variant of uncertain significance (VUS) in *RECQL4*, with the variants present *in trans*. In light of evolving evidence that heterozygous germline variants affecting other DNA repair genes such as *ATM*, *BRCA2*, and *PALB2* confer an increased cancer risk (Thompson et al. 2005; Renwick et al. 2006; Antoniou et al. 2014; Helgason et al. 2015; Rebbeck et al. 2015; Esteban-Jurado et al. 2016; Esai Selvan et al. 2019), we questioned whether heterozygous germline *RECQL4* variants, such as the one observed in our patient, might be associated with development of pediatric cancer. To address this question, we analyzed 5562 children with cancer and identified a significant enrichment of heterozygous *RECQL4* loss-of-function (LOF) variants in patients with OS. Notably, most children carried the same *RECQL4* alteration: c.1573delT (p.Cys525Alafs), which is predicted to truncate the RECQL4 protein within its DNA helicase domain. *RECQL4* LOF variants were rarely observed in children with other cancer types, in which they were not significantly enriched compared to noncancer controls.

## RESULTS

## **Characteristics of the Index Case**

A 16-yr-old girl with previously treated OS of the left tibia developed recurrent OS of the left femur and left wrist, 5 and 8 yr after the initial diagnosis, respectively. Family history revealed a maternal first cousin with an unspecified brain tumor at 3 yr of age (Fig. 1). The maternal grandparents developed cancer after 60 yr of age, but no other family members were reported to have cancer. Germline WGS performed as part of a clinical research protocol (NCT02530658) that comprehensively analyzed 156 cancer predisposition genes (Supplemental Table 1) revealed no pathogenic or likely pathogenic (P/LP) variants in known OS genes such as *TP53*, *RB1*, the mismatch repair, and Fanconi anemia genes. However, testing did reveal two rare variants in *RECQL4*, including a frameshift c.1573delT





**Figure 1.** Three-generation pedigree of a 16-yr-old female with recurrent osteosarcoma (arrowhead). Circles and squares denote female and male family members, respectively, and shaded figures denote persons with cancer. The individuals age in years (yr), age at diagnosis (dx.), and age at death (d.) are indicated on the pedigree where applicable. (NOS) Not otherwise specified.

(p.Cys525Alafs) and an in-frame deletion of nine nucleotides that is predicted to remove three amino acids c.2412\_2420del (p.Ala805\_Arg807del) (Table 1). These variants were classified by a CLIA-certified laboratory as pathogenic and of uncertain significance, respectively, based on the American College of Medical Genetics and Genomics (ACMG) and the Association for Molecular Pathology (AMP) guidelines for sequence variant interpretation (Richards et al. 2015).

On physical examination, the patient was well-developed with no dysmorphic facial features. She had relatively thin hair and eyebrows, which by the patient's account predated chemotherapy. Her height was about 2nd percentile for age and in proportion to the calculated midparental height (height = 148 cm, midparental height = 153.5 cm). Although her head circumference was small, it was in proportion to her height and weight. Approximately 12 café au lait macules were observed on the patient's trunk and buttocks ranging in size from 4 mm to 4 cm; scattered hypopigmented areas on her left thigh and

Table 1.	Ge	rmline REC	OL4 v Exon	HGVS DNA	entified in the inde	vx case Variant type	dbSNP ID	Genotype	Parent of origin	Variant allele frequency	Coverage	Clinical significance in ClinVar	Comments
RECQL4	8	144,514,983	9	c.1573delT	p.Cys525Alafs*33	Frameshift	Rs386833845	Heterozygous	Mother	0.277	90	Р	Mother is heterozygous
RECQL4	8	144,513,269	15	c.2412_ 2420del	p.Ala805_Arg807del	In-frame deletion	Rs766312203	Heterozygous	Father	0.308	13	VUS	Father is homozygous

trunk were also observed. No axillary freckling was observed, and none of the skin lesions were consistent with poikiloderma. Based on the clinical phenotype, the patient did not meet the diagnostic criteria for RTS (Wang and Plon 2019) by the account of two clinical geneticists who examined the patient on multiple occasions and/or reviewed multiple photographs highlighting close-up views of the face, stature, and pigmented skin lesions. The patient's parents had no clinical features of RTS and no personal history of cancer. Based on parental testing, the two variants found in the patient were determined to be on opposite chromosomes (in trans). This testing revealed that the patient's father was homozygous for the c.2412\_2420del (p.Ala805\_Arg807del) VUS. The absence of RTS features in the father provided supporting evidence that this VUS was likely benign or that it is a hypomorphic allele that does not have a phenotype when found in the homozygous state but may play some role in this patient's unusual phenotype when opposite a loss-of-function variant. The patient's mother was heterozygous for the pathogenic c.1573delT (p.Cys525Alafs) variant. The patient's brother was not reported to have any clinical features of RTS and no personal history of cancer. He did not opt to pursue genetic testing and as a result, his RECQL4 genetic status remains unknown.

## Prevalence of Heterozygous Germline RECQL4 LOF Variants in Pediatric Cancer

Heterozygous pathogenic variants in DNA damage repair genes (e.g., *ATM*, *BRCA2*, *PALB2*) are associated with a moderate to high cancer risk (Thompson et al. 2005; Renwick et al. 2006; Antoniou et al. 2014; Helgason et al. 2015; Rebbeck et al. 2015; Esteban-Jurado et al. 2016; Esai Selvan et al. 2019), whereas compound heterozygous or homozygous variants confer syndromic presentations such as ataxia-telangiectasia (*ATM*) (Swift et al. 1987; Ahmed and Rahman 2006) and Fanconi anemia (*BRCA2*, *PALB2*) (Howlett et al. 2002; Reid et al. 2007), which confer an even greater cancer risk. Building upon this notion, we sought to determine whether heterozygous *RECQL4* LOF variants are more prevalent in pediatric oncology patients as compared to control individuals not selected for cancer. Among 4435 pediatric cancer patients at St. Jude Children's Research Hospital (Zhang et al. 2015; Wang et al. 2018) and 1127 in the National Cancer Institute TARGET database (dbGaP accession phs000218.v20.p7), we identified 24 of 5562 (0.43%) who carried heterozygous *RECQL4* LOF variants (Table 2), including five of 249 (2.0%) with OS (Fig. 2).

To determine whether these LOF variants are enriched in children with cancer, we examined their prevalence in the Genome Aggregation Database (gnomAD v2.1, noncancer; Lek et al. 2016), which spans 118,479 whole-exome and 15,708 whole-genome sequences from individuals who were not ascertained for having cancer. There were a total of 385 *RECQL4* LOF alleles in the gnomAD noncancer population after removal of the founder variant, c.1390+2deIT (p.Ala420\_Ala463deI), which is commonly encountered in the Finnish population at a minor allele frequency of 0.4% (Lek et al. 2016) but was absent from our cohort. Low-confidence LOF variants and variants of dubious quality were also removed as detailed in the Methods. Compared to the noncancer cohort in gnomAD, we observed a significant association between heterozygous *RECQL4* LOF variants and OS (P=0.00087; OR=7.1; 95% CI, 2.9–17.0) but no association for other tumor types. One patient with retinoblastoma was noted to harbor an additional pathogenic variant in *RB1* and was excluded from subsequent analysis. Available tumor data from 12 germline *RECQL4* LOF variant positive cases did not show loss of heterozygosity for *RECQL4* or any pathogenic or likely pathogenic variants in the remaining *RECQL4* allele.

#### Evaluation of RECQL4 Genotype and Association with Clinical Phenotype

To understand how germline *RECQL4* LOF variants impacted the encoded protein, we evaluated their location within the gene. Among the 24 variants identified, 17 (71%) resided

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ind         Description         Tammelifie         Orange         Tammelifie         No.	Age at Ethnicity diagno- of Location Cancer diagnosis sis subject Gene Chr (hg38) Exon h	Age at Ethnicity diagno- of Location sis subject Gene Chr (hg38) Exon h	Ethnicity of Location subject Gene Chr (hg38) Exon h	Location Gene Chr (hg38) Exon h	Location (hg38) Exon h	-	HGVS DNA	HGVS protein	Variant class f	WGS variant allele rrequency	WES cover- age f	Variant allele equency	Cover- age	DI ANSAb	Clinical signifi- cance in ClinVar	AF in gnomad noncancer v2.1	Ethnicity- specific AF in gnomAD noncancer v2.1	Cohor
eld         p.Ala291.au/et/d         Farmentif         NA         NA         0.687         32         ·         NA         NA         NA           ald         p.Ala291.lau/et/2         Farmentif         0.402         22         0.545         33         rs1389.447533         P         10°°°         983.°         SUGH           22A-5C         p.Ala291.Lau/et/2         Farmentif         0.402         22         0.545         3         rs1389.447533         P         10°°°         983.°         SUGH           22A-5C         p.Glu/44.E8pile         Splice         0.423         26         0.750         10°°         10°°°	Craniopharyngioma 5 NFE RECOL4 8 144,516,696 5 c.423	5 NFE RECOL4 8 144,516,696 5 c.423	NFE RECOL4 8 144,516,696 5 c.423	RECOL4 8 144,516,696 5 c.423	144,516,696 5 c.423	c.423	delG	p.Lys141Asnfs*39	Frameshift	0.294	34	NA	NA	1		AN	AN	SJCRH
delG         p.Ala271Leufs'Z         Frameshift         0.40         22         0.545         3         s138644733         P         4.28 × 10 <sup>-06</sup> 9.68 × 10 <sup>-06</sup>	Acute B- NA NFE <sup>b</sup> RECQL4 8 144,516,254 5 c.865 lymphoblastic leukemia	NA NFE <sup>b</sup> RECQL4 8 144,516,254 5 c.865	NFE <sup>b</sup> RECOL4 8 144,516,254 5 c.865	RECOL4 8 144,516,254 5 c.865	144,516,254 5 c.865	c.865	delG	p.Ala289Leufs*4	Frameshift	AN	AN	0.687	32	·		AN	AN	TARGE
32.2 2A-5C         PAIa378_E66plice         Splice         NA         NA         0.510         249         -         4.29 ×         0.70%         107%<	Acute 11 NFE RECOL4 8 144,516,248 5 c.87 lymphoblastic leukemia, NOS	11 NFE RECOL4 8 144,516,248 5 c.87	NFE RECOL4 8 144,516,248 5 c.87	RECOL4 8 144,516,248 5 c.87	144,516,248 5 c.87 <sup>.</sup>	c.87	1delG	p.Ala291Leufs*2	Frameshift	0.409	22	0.545	33 rs	1389647533	۵	4.28 × 10 <sup>-06</sup>	9.85 × 10 <sup>-06</sup>	SJCRH
P1-1C>A         p.Glu464_E8plice         Splice         0.43         2.6         0.75         0.75         0.76%	Acute B- NA SAS <sup>b</sup> RECOL4 8 144,515,890 7 c.11 lymphoblastic leukemia	NA SAS <sup>b</sup> RECOL4 8 144,515,890 7 c.11	SAS <sup>b</sup> RECOL4 8 144,515,890 7 c.11	RECQL4 8 144,515,890 7 c.11	144,515,890 7 c.11	c.11	132-2A>G	p.Ala378_E6splice	Splice	AN	AN	0.510	249			4.29 × 10 <sup>-06</sup>	3.28 × 10 <sup>-05</sup>	TARGE
$73delT$ $p.Cys2SAbleis*33$ Frameshift $0.567$ $30$ $0.538$ $31$ $0.429$ $28$ $s386333845$ $P$ $2.43 \times t_{10}^{-0.6}$ $10^{-0.6}$ $10^{-0.6}$ $10^{-0.6}$ $10^{-0.6}$ $10^{-0.6}$ $10^{-0.6}$ $10^{-0.6}$ $2.0CH$ $573delT$ $p.Cys2SAbleis*33$ Frameshift $0.35$ $31$ $0.429$ $28$ $s386333845$ $P$ $2.43 \times t_{10^{-0.6}}$ $4.21 \times t_{10^{-0.6}}$ $5.0CH$ $573delT$ $p.Cys2SAbleis*33$ Frameshift $0.31$ $6.7$ $0.316$ $5.0$ $5.0CH$ $10^{-0.6}$ $4.21 \times t_{10^{-0.6}}$ $5.0CH$ $573delT$ $p.Cys2SAbleis*33$ Frameshift $0.4$ $0.31$ $0.31$ $0.470$ $0.736$ $5.0CH$ $10^{-0.6}$ $10^{-0.6}$ $5.0CH$ $573delT$ $p.Cys2SAbleis*33$ Frameshift $0.4$ $0.4$ $0.736$ $5.023833345$ $P$ $2.43 \times t_{10^{-0.6}}$ $5.0CH$ $573delT$ $p.Cys2S2Ableis*33$ Frameshift $0.4$ $0.473$	Acute B- 3 NFE RECOL4 8 144,515,243 7 c.1: lymphoblastic leukemia	3 NFE RECOL4 8 144,515,243 7 c.1:	NFE RECOL4 8 144,515,243 7 c.1:	RECOL4 8 144,515,243 7 c.1.	144,515,243 7 c.1:	C,	391-1G>A	p.Glu464_E8splice	Splice	0.423	26	0.750	16 rs	117642173	۵	2.89 × 10 <sup>-05</sup>	6.59 × 10 <sup>-05</sup>	SJCRH
573deff $pCys52SAlafs^{-33}$ Frameshift $0.355$ $31$ $0.429$ $28$ $s38633345$ $P$ $2.43 \times t$ $421 \times t$ $5.074$ 573deff $pCys52SAlafs^{-33}$ Frameshift $NA$ $0.571$ $28$ $s38633345$ $P$ $2.43 \times t$ $4.21 \times t$ $5.0744$ 573deff $pCys52SAlafs^{-33}$ Frameshift $0.218$ $64$ $0.315$ $200$ $s38633345$ $P$ $2.43 \times t$ $4.21 \times t$ $5.0744$ 573deff $pCys52SAlafs^{-33}$ Frameshift $0.47$ $0.315$ $200$ $s38633345$ $P$ $2.43 \times t$ $4.21 \times t$ $5.0744$ 573deff $pCys52SAlafs^{-33}$ Frameshift $0.4$ $N$ $0.271$ $90$ $s38633345$ $P$ $2.43 \times t$ $10^{-64}$ $10^{-64}$ $5.0744$ 573deff $pCys52SAlafs^{-33}$ Frameshift $0.44$ $0.271$ $50$ $s38633345$ $P$ $2.43 \times t$ $10^{-64}$ $10^{-64}$ $10^{-64}$ $10^{-64}$ $10^{-64}$ <td>Acute myeloid 11 NFE RECOL4 8 144,514,983 9 c.1 leukemia</td> <td>11 NFE RECOL4 8 144,514,983 9 c.1</td> <td>NFE RECOL4 8 144,514,983 9 c.1</td> <td>RECOL4 8 144,514,983 9 c.1</td> <td>144,514,983 9 c.1</td> <td>Ū.</td> <td>573delT</td> <td>p.Cys525Alafs*33</td> <td>Frameshift</td> <td>0.567</td> <td>30</td> <td>0.550</td> <td>20 rs</td> <td>386833845</td> <td>٩</td> <td>2.43 × 10<sup>-04</sup></td> <td>4.21 × 10<sup>-04</sup></td> <td>SJCRH</td>	Acute myeloid 11 NFE RECOL4 8 144,514,983 9 c.1 leukemia	11 NFE RECOL4 8 144,514,983 9 c.1	NFE RECOL4 8 144,514,983 9 c.1	RECOL4 8 144,514,983 9 c.1	144,514,983 9 c.1	Ū.	573delT	p.Cys525Alafs*33	Frameshift	0.567	30	0.550	20 rs	386833845	٩	2.43 × 10 <sup>-04</sup>	4.21 × 10 <sup>-04</sup>	SJCRH
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573delT         p.Cys525Alafs*33         Frameshift         0.218         64         0.315         200         rs386333845         P         2.43 ×         4.21 ×         8.10 <sup>-04</sup> 9.0           573delT         p.Cys525Alafs*33         Frameshift         NA         NA         0.277         90         rs386333845         P         2.43 ×         4.21 ×         8.107 <sup>-04</sup> 107 <sup>-04</sup>	Acute 11 NFE RECOL4 8 144,514,983 9 c.1 myelogeneous leukemia	11 NFE RECOL4 8 144,514,983 9 c.1	NFE RECOL4 8 144,514,983 9 c.1	RECOL4 8 144,514,983 9 c.1	144,514,983 9 c.1	C.	573delT	p.Cys525Alafs*33	Frameshift	AN	AN	0.571	28 rs	386833845	٩	2.43 × 10 <sup>-04</sup>	4.21 × 10 <sup>-04</sup>	SJCRH
573deIT       Dc.Vy525Alafs*33       Frameshit       NA       0.277       90       rs386833845       P       2.43 ×       1.42 ×       5.107-         573deIT       p.Cy525Alafs*33       Frameshit       0.4       30       0.470       66       rs386833845       P       2.43 ×       1.42 ×       5.107-         573deIT       p.Cy5525Alafs*33       Frameshit       0.4       30       0.473       55       rs386833845       P       2.43 ×       1.07-04       107-04       5.107-         573deIT       p.Cy5525Alafs*33       Frameshit       NA       NA       0.293       58       rs386833845       P       2.43 ×       1.07-04       107-04	Acute B- 3 NFE <sup>c</sup> RECOL4 8 144,514,983 9 c.11 lymphoblastic leukemia	3 NFE <sup>c</sup> RECOL4 8 144,514,983 9 c.11	NFE <sup>c</sup> RECOL4 8 144,514,983 9 c.11	RECOL4 8 144,514,983 9 c.1!	144,514,983 9 c.1!	C.11	573deIT	p.Cys525Alafs*33	Frameshift	0.218	64	0.315	200 rs	386833845	۵	2.43 × 10 <sup>-04</sup>	4.21 × 10 <sup>-04</sup>	SJCRH
573delT         p.Cys52SAlafs*33         Frameshit         0.4         30         0.470         66         rs386333845         P         2.43 ×         2.43 ×         5.10 <sup>-04</sup> 5.0GR           573delT         p.Cys52SAlafs*33         Frameshit         0.364         44         0.473         55         rs386333845         P         2.43 ×         2.43 ×         5.10 <sup>-04</sup> 10 <sup>-04</sup>	Osteosarcoma 8 AMR <sup>e</sup> RECOL4 8 144,514,983 9 c.1	8 AMR <sup>c</sup> RECOL4 8 144,514,983 9 c.1	AMR <sup>c</sup> RECOL4 8 144,514,983 9 c.1	RECOL4 8 144,514,983 9 c.1	144,514,983 9 c.1	ίΰ	573delT	p.Cys525Alafs*33	Frameshift	AN	AN	0.277	90 rs	386833845	٩	2.43 × 10 <sup>-04</sup>	1.42 × 10 <sup>-04</sup>	SJCRF
573delT     p.Cys525Alafs*33     Frameshit     0.364     44     0.473     55     s386833845     P     2.43 ×     2.43 ×     2.43 ×     Dote       573delT     p.Cys525Alafs*33     Frameshit     NA     0.293     58     s386833845     P     2.43 ×     10 <sup>-04</sup> 10 <sup>-04</sup> 573delT     p.Cys525Alafs*33     Frameshit     NA     0.293     58     s386833845     P     2.43 ×     1.43 ×     1.4RGI       573delT     p.Cys525Alafs*33     Frameshit     NA     0.545     11     rs386833845     P     2.43 ×     1.4A ×     1ARGI       573delT     p.Cys525Alafs*33     Frameshit     NA     0.545     11     rs386833845     P     2.43 ×     1.4A ×     1ARGI       573delT     p.Cys525Alafs*33     Frameshit     NA     0.545     11     rs386833845     P     2.43 ×     1.4A ×     1ARGI       705-1delG     p.lle569_E11splice     Splice     0.364     44     0.500     10     -     -     MA     MA     SURH       717C>T     p.Gln573*     Nonsense     0.275     58     0.25     8     -     -     3.19 ×     1.15 ×     SURH	Acute B- 4 NFE RECOL4 8 144,514,983 9 c.1 lymphoblastic leukemia	4 NFE RECOL4 8 144,514,983 9 c.1	NFE RECOL4 8 144,514,983 9 c.1	RECOL4 8 144,514,983 9 c.1	144,514,983 9 c.1	ü	573delT	p.Cys525Alafs*33	Frameshift	0.4	30	0.470	66 rs	386833845	٩	2.43 × 10 <sup>-04</sup>	2.43 × 10 <sup>-04</sup>	SJCRH
573deIT     p.Cys525Alafs*33     Frameshift     NA     0.293     58     rs386833845     P     2.43 ×     2.43 ×     10 <sup>-04</sup> TaRGI       573deIT     p.Cys525Alafs*33     Frameshift     NA     0.545     11     rs386833845     P     2.43 ×     1.44 ×     TaRGI       573deIT     p.Cys525Alafs*33     Frameshift     NA     NA     0.545     11     rs386833845     P     2.43 ×     1.44 ×     TaRGI       705-1deIG     p.lle569_E11splice     Splice     0.364     44     0.500     10     -     -     NA     NA     SJCRH       717C>T     p.GIn573*     Nonsense     0.275     58     0.25     8     -     -     3.19 ×     1.15 ×     SJCRH	Germ cell tumor, 0.3 NFE RECOL4 8 144,514,983 9 c. testicular	0.3 NFE RECOL4 8 144,514,983 9 c.	NFE RECOL4 8 144,514,983 9 c.	RECOL4 8 144,514,983 9 c.	144,514,983 9 c.	U	1573delT	p.Cys525Alafs*33	Frameshift	0.364	44	0.473	55 rs	386833845	٩	2.43 × 10 <sup>-04</sup>	2.43 × 10 <sup>-04</sup>	SJCRF
573deIT     p.Cys525Alafs*33     Frameshift     NA     0.545     11     rs386833845     P     2.43 ×     144 ×     TARG       773deIT     p.Cys525Alafs*33     Frameshift     NA     0.545     44     0.500     10     04     10 <sup>-04</sup> 11 <sup>-04</sup> 51 <sup>-04</sup> 51 <sup>-04</sup> 10 <sup>-04</sup> 11 <sup>-04</sup> 51 <sup>-04</sup> <	Acute B- NA NFE <sup>b</sup> RECOL4 8 144,514,983 9 c.11 lymphoblastic leukemia	NA NFE <sup>b</sup> RECOL4 8 144,514,983 9 c.1	NFE <sup>b</sup> RECOL4 8 144,514,983 9 c.1	RECOL4 8 144,514,983 9 c.15	144,514,983 9 c.15	c.15	573deIT	p.Cys525Alafs*33	Frameshift	AN	AN	0.293	58 rs	386833845	۵	2.43 × 10 <sup>-04</sup>	2.43 × 10 <sup>-04</sup>	TARG
J5-1delG p.lle569_E11splice Splice 0.364 44 0.500 10 NA NA SJCRH 17C>T p.GIn573* Nonsense 0.275 58 0.25 8 3.19× 1.15× SJCRH 17C>T p.GIn573* 10 <sup>-05</sup> 10 <sup>-05</sup> 10 <sup>-04</sup>	Acute T- 10 AMR <sup>b</sup> RECOL4 8 144,514,983 9 c.157 lymphoblastic leukemia	10 AMR <sup>b</sup> RECOL4 8 144,514,983 9 c.15 <sup>-</sup>	AMR <sup>b</sup> RECOL4 8 144,514,983 9 c.157	RECOL4 8 144,514,983 9 c.157	144,514,983 9 c.157	c.157	'3delT	p.Cys525Alafs*33	Frameshift	AN	AN	0.545	11 rs	386833845	٩	2.43 × 10 <sup>-04</sup>	1.44 × 10 <sup>-04</sup>	TARGE
717C>T p.Gin573* Nonsense 0.275 58 0.25 8 3.19× 1.15× SJCRF 10 <sup>-05</sup> 10 <sup>-05</sup>	Hodgkin lymphoma 7 AFR RECOL4 8 144,514,363 10 c.1	7 AFR RECOL4 8 144,514,363 10 c.1	AFR RECQL4 8 144,514,363 10 c.1	RECOL4 8 144,514,363 10 c.1	144,514,363 10 c.1	с,	705-1 delG	p.Ile569_E11splice	Splice	0.364	44	0.500	10			NA	NA	SJCRF
	Spindle cell 16 AFR <sup>c</sup> RECOL4 8 144,514,350 11 c. sarcoma	16 AFR <sup>c</sup> RECOL4 8 144,514,350 11 c.	AFR <sup>c</sup> RECOL4 8 144,514,350 11 c.	RECOL4 8 144,514,350 11 c.	144,514,350 11 c.	J	1717C>T	p.Gln573*	Nonsense	0.275	58	0.25	ø	ı		3.19 × 10 <sup>-05</sup>	1.15 × 10 <sup>-04</sup>	SJCRH

Table 2. (C	ontinued)																
Subject ID	Cancer diagnosis	Age at diagno- sis	Ethnicity of subject	Gene Chr	Location (hg38) Exon	HGVS DNA	HGVS protein	Variant class fi	WGS variant allele o requency	WES :over- age fr	Variant allele C equency	over- age	D O OI ANS	Clinical signifi- ance in ClinVar	AF in gnomad noncancer v2.1	Ethnicity- specific AF in gnomAD noncancer v2.1	Cohort
SJTALL065	Acute T- lymphoblastic leukemia	8	AFR	RECOL4 8	144,513,592 13	c.2178_ 2179insCCTGGGTC	p.Ala727Profs*119	Frameshift	AN	AN	0.2	73		1	NA	NA	SJCRH
SJOS040215	Osteosarcoma	ΝA	NFE <sup>b</sup>	RECOL4 8	144,513,412 14	c.2269C>T	p.Gln757*	Nonsense	AN	AA	0.503	153 rs	137853229	٩	1.24 × 10 <sup>-04</sup>	1.45 × 10 <sup>-04</sup>	TARGET
SJOS040163	Osteosarcoma	٩N	NFE <sup>b</sup>	RECOL4 8	144,513,383 15	c.2296+1C>G	p.Arg766_E15splice	Splice	AN	AA	0.357	26 rs	199605511	,	AN	AN	TARGET
SJNHL019456	Non-Hodgkin Iymphoma	7	NFE	RECOL4 8	144,513,109 16	c.2492_2493delAT	p.His831Argfs*52	Frameshift	0.263	19	0.064	47 rs	752729755	۵.	7.02 × 10 <sup>-05</sup>	7.01 × 10 <sup>-05</sup>	SJCRH
SJHL042013	Hodgkin lymphoma	18	NFE	RECOL4 8	144,513,109 16	c.2492_2493delAT	p.His831Argfs*52	Frameshift	0.5	34	0.400	5 13	752729755	۹.	7.02 × 10 <sup>-05</sup>	7.01 × 10 <sup>-05</sup>	SJCRH
SJOS040168	Osteosarcoma	ΝA	NFE <sup>b</sup>	RECOL4 8	144,512,846 16	c.2755+1G>A	p.Ala919_E16splice	Splice	AN	AA	0.316	19 rs	373130543	1	3.16 × 10 <sup>-05</sup>	2.86 × 10 <sup>-05</sup>	TARGET
SJALL019726	Acute B- lymphoblastic leukemia	ę	NFE	RECOL4 8	144,513,109 19	c.3073_3074delAG	p.Thr1024_Glu1025fs	Frameshift	0.44	25	0.385	39	I		AN	AN	SJCRH
SJRB001130	Retinoblastoma	AN	ОТН	RECOL4 8	144,511,911 20	c.3393+2T>G	p.Arg1131_E20splice	Splice	9.0	15	AN	NA	557284122		1.52 × 10 <sup>-05</sup>	0.00 × 10 <sup>+00</sup>	SJCRH
All ethnicities	were computations	ally pred	icted unle	ss otherwise	noted. All predict	ed populations were o	consistent with self-	-reporting ur	rless othe	rwise no	oted.						





**Figure 2.** Frequency of heterozygous *RECQL4* LOF mutations across pediatric cancer types. (OS) osteosarcoma, (GCT) germ cell tumor, (HL) Hodgkin lymphoma, (ALL) acute lymphoblastic leukemia, (NHL) non-Hodgkin lymphoma, (AML) acute myeloid leukemia, (CNS) central nervous system, (LOF) loss-of-function.

within the helicase domain (residues 489–850; Fig. 3), which is critical to maintaining genome stability, specifically in the case of DNA repair (Chu and Hickson 2009; Croteau et al. 2012). All variants in the helicase domain are predicted to result in NMD or a disrupted protein product. Four of 24 (17%) patients harbored germline *RECQL4* variants amino-terminal to the helicase domain, all of which are predicted to cause nonsense-mediated decay (NMD). Of these, three were located in a region having sequence similarity to yeast Sld2 (the Sld2-like domain; residues 1–388), which is essential for the initiation of DNA replication, and the fourth was located in a nuclear targeting signal (residues 363–492) (Colombo et al. 2018). Three of 24 patients had variants located carboxy-terminal to the helicase domain, in exons 16 and 19, which are also predicted to cause NMD.

Notably, nine (38%) of the *RECQL4* LOF variant–positive individuals were noted to carry the same alteration, c.1573delT (p.Cys525Alafs), which was also present in our index case (Fig. 3). Patients with this variant carried diagnoses of acute lymphoblastic leukemia (ALL; n = 4), OS (n = 2), acute myeloid leukemia (AML; n = 2), and germ cell tumor (GCT; n = 1) and were of different racial and ethnic backgrounds (Table 2). Our analysis revealed that this particular variant is significantly enriched in the pediatric cancer population (P = 0.0024; OR = 3.3; 95% CI, 1.7–6.7) compared to the gnomAD noncancer cohort in which this variant accounts for 64 of 385 (17%) *RECQL4* LOF variants with a global allele frequency of 2.43 × 10<sup>-4</sup>. This supports the notion that the c.1573delT (p.Cys525Alafs) alteration contributes to the pathogenesis of pediatric cancer; however, its presence in a presumably healthy population suggests that it may be a lower penetrance allele.

To investigate the possible penetrance of germline *RECQL4* LOF variants, we examined the family histories of four patients from the St. Jude cohort for whom such information was available. None of these patients had first- or second-degree relatives with early onset of cancer (defined here as cancer before 50 yr of age; Fig. 4). This further supports the idea that heterozygous *RECQL4* LOF variants function as lower penetrance alleles. These data are consistent with the literature in which there are only rare reports of relatives of RTS patients who have developed OS (Siitonen et al. 2009).

#### Prevalence of Germline LOF Variants in Other Genes of the RecQ Helicase Family

Because the RecQ helicase genes are highly conserved, it remained possible that other family members might also be associated with childhood cancer development. Therefore, we examined the St. Jude and TARGET cohorts for heterozygous LOF variants in *RECQL*, *BLM*, *WRN*, and *RECQL5*. Through these studies, we identified 51 children with







heterozygous LOF variants in one of these genes, including 13 with variants in *RECQL*, 13 with variants in *BLM*, 15 with variants in *WRN*, and 10 with variants in *RECQL5* (Supplemental Table 2). Pan-cancer analyses did not reveal any significant associations between the presence of heterozygous LOF variants in other RecQ helicases and pediatric cancer (Table 3). However, cancer-specific analyses identified a significant association in which *RECQL* variants were present in 1.2% (3 out of 249) of OS cases (P = 0.037; OR = 4.2; 95% CI, 1.3–13.1). Additional nonsignificant associations of potential interest include the identification of *RECQL* variants in 1.3% (2 out of 150) of rhabdomyosarcoma cases (P = 0.071; OR = 4.6; 95% CI, 1.2–18.7) and *WRN* variants in 0.39% (9 out of 2314) of pediatric ALL patients (P = 0.061; OR = 1.9; 95% CI, 0.98–3.7).

# Population Admixture of the Childhood Cancer Cohort Compared to the gnomAD Cohort

To determine whether the enrichment of germline *RECQL4* or *RECQL* variants might be due to differences in the ethnic composition of pediatric cancer versus gnomAD noncancer cohorts, we compared the population admixtures of these two groups. Here, the ethnicity of individuals in the pediatric cancer cohort was computationally predicted (Supplemental





Figure 4. Pedigrees of families with RECQL4 LOF mutations.

Fig. 1). As shown in Supplemental Table 3, the population admixture in the pediatric cancer cohort is not equivalent to that found in gnomAD noncancer version 2.1. Nevertheless, all *RECQL4* and *RECQL* LOF variants are extremely rare with respect to both global and ethnicity-specific allele frequencies (Table 2; Supplemental Table 2). Therefore, it is unlikely that the differences in the composition of the two populations significantly impacts our results. To further examine this possibility, we evaluated for enrichment of germline heterozygous LOF variants in *RECQL4* using only non-Finnish Europeans who comprise the majority of individuals in the cancer and noncancer cohorts (Supplemental Table 3). Through this analysis, significant enrichment in pediatric OS was once again observed for variants affecting *RECQL4* (P=0.0012, OR=6.77 [2.79, 16.45]) and *RECQL* (P=0.024, OR=5.04 [1.60, 15.86]).

# DISCUSSION

As genetic testing is increasingly applied to patients with cancer, our knowledge of the germline contributions to cancer risk is dramatically expanding. Through such efforts, it has become clear that heterozygous pathogenic variants in DNA damage repair genes, such as *ATM*, *BRCA2*, and *PALB2*, are associated with a significant increase in risk to develop certain cancers. Here, we examined whether heterozygous LOF variants in *RECQL4*, the

	Podiatric	ancor pationts	gnomAl	D noncancer	Cancor ris	k
						<u> </u>
Cancer diagnosis	Carriers	Noncarriers	Carriers	Noncarriers	Odds ratio (95% CI)	<i>P</i> -value
RECQL						
Pan-cancer (all diagnoses)	13	5549	388	133,799	0.81 (0.46, 1.4)	0.52
Osteosarcoma	3	246	388	133,799	4.2 (1.3, 13.1)	0.037*
BLM						
Pan-cancer (all diagnoses)	13	5549	279	133,908	1.1 (0.64, 2.0)	0.65
Osteosarcoma	0	249	279	133,908	0	1
WRN						
Pan-cancer (all diagnoses)	15	5547	273	133,914	1.3 (0.79, 2.2)	0.29
Osteosarcoma	0	249	273	133,914	0	1
RECQL4						
Pan-cancer (all diagnoses <sup>a</sup> )	23	5539	385	133,802	1.4 (0.95, 2.2)	0.098
Osteosarcoma	5	244	385	133,802	7.1 (2.9, 17.0)	0.00087**
RECQL5						
Pan-cancer (all diagnoses)	10	5552	284	133,903	0.85 (0.45, 1.6)	0.76
Osteosarcoma	0	249	284	133,903	0	1

Table 3. Comparison of heterozygous RECQL4 LOF variants in pediatric cancer patients and the gnomAD noncancer cohort

\**P* < 0.05.

\*\**P* < 0.005.

<sup>a</sup>Patient with pathogenic *RB1* variant was excluded from statistical analysis.

gene encoding an important DNA helicase that is mutated in individuals with RTS, are more prevalent in children with cancer.

After screening 5562 pediatric cases, we identified a significant enrichment of heterozygous germline *RECQL4* LOF variants in patients with OS, the most common cancer in RTS, which is caused by homozygous or compound heterozygous variants in *RECQL4*. Although not significantly overrepresented, such variants were also identified in children with other tumor types, such as ALL, AML, craniopharyngioma, GCT, HL, and NHL. In 12 cases for whom tumor data were available, no deletions or mutations within the remaining *RECQL4* allele were detected. Although other mechanisms might account for a second hit (e.g., changes in methylation) (Mazor et al. 2015; Di Ruscio et al. 2016), it remains possible that RECQL4 haploinsufficiency, perhaps in combination with other oncogenic events, is enough to promote malignant transformation.

Our findings are consistent with recent studies documenting heterozygous germline *RECQL4* variants in adults with a variety of malignancies (Schrader et al. 2016; Jalkh et al. 2017; Mandelker et al. 2017; Tedaldi et al. 2017; AlDubayan et al. 2018; Bonache et al. 2018; Lowery et al. 2018; Na et al. 2018; Paulo et al. 2018; Penkert et al. 2018; Quezada Urban et al. 2018; Slavin et al. 2018). In addition to these studies are reports of an increased prevalence of *RECQL4* variants in patients with bladder (Na et al. 2018) or colorectal cancer (AlDubayan et al. 2018) compared to the general population. Moreover, a heterozygous germline truncating variant in *RECQL4* has been implicated as a possible cancer risk factor in an individual with prostate cancer (Paulo et al. 2018). Curiously, a previous investigation did not find enrichment of *RECQL4* mutations in sporadic cases of OS compared to the general population (Nishijo et al. 2004); however, this prior study examined a much smaller cohort including only 71 patients with OS. Thus, this association could potentially have been missed.



Among the 24 patients identified as harboring a single germline *RECQL4* LOF variant, 17 (71%) of these variants reside in the highly conserved helicase domain, with nine of these carrying the same c.1573delT (p.Cys525Alafs) alteration. Most RTS patients also harbor variants that result in a truncated protein lacking some or all of the helicase domain. Correspondingly, it has been reported that many RTS patients who developed OS harbor at least one truncating variant in *RECQL4*, whereas individuals with a clinical diagnosis of RTS and either a missense variant or no identified variant in *RECQL4* were not reported to develop OS (Wang et al. 2003). In contrast to these deleterious truncating variants, the hallmark variant in RAPADILINO syndrome, a splice site variant that causes in-frame skipping of exon 7 (Van Maldergem et al. 2006), leaves the helicase domain largely intact (Kitao et al. 1999; Siitonen et al. 2003), which may correspond with the lower incidence of cancer.

In addition to the new association of heterozygous germline RECQL4 LOF variants with pediatric OS, our data also showed evidence of an association linking RECQL variants with OS, which deserves further investigation. Germline RECQL variants have been reported in BRCA1/2-negative breast cancers (Cybulski et al. 2015; Sun et al. 2015; Kwong et al. 2016); however, this association is disputed (Kwong et al. 2016; Li et al. 2018). To date, RECQL variants have not been reported in pediatric cancer. No strong associations were found between pediatric cancer and germline heterozygous LOF variants in the other RecQ helicase genes (i.e., BLM, WRN, RECQL5). Although all RecQ helicase genes contain the highly conserved helicase domain (Hickson 2003), they each play unique roles in genome maintenance and stability (Croteau et al. 2014). Toward this end, previous work has demonstrated that only two of the five human RecQ helicases, RECQL and RECQL4, bind specifically to three well-defined DNA replication origins under native conditions (Thangavel et al. 2010), whereas the other helicases (i.e., BLM, WRN, RECQL5) can only be found at replication origins following treatment with replication inhibitors (Thangavel et al. 2010). The similar localization and putative roles of RECQL and RECQL4 may underlie how disruption of these helicases promotes the development of OS and possibly other cancers.

This study has several limitations. First, the prevalence of germline *RECQL4* variants identified in this study might not reflect the prevalence in newly diagnosed pediatric cancer patients because 50% of the patients studied were long-term survivors of childhood cancer. If the presence of heterozygous germline LOF variants is associated with poorer (or better) outcomes, it is possible that their prevalence in our cohort will be lower (or higher) than in newly diagnosed patients. A second limitation is that the population admixture of the pediatric cancer cohort examined in this study is not equivalent to that found in the gnomAD noncancer cohort (Supplemental Data). We suspect that this is due to differences in outcomes or referral patterns among different populations. Regardless, all of the LOF alleles reported in this study are extremely rare with respect to both global and ethnicity-specific allele frequencies. Last, this study is limited by the small number of individuals who harbor germline LOF variants. This factor makes it difficult to define the true spectrum of cancers associated with these germline variants. Continued evaluation of patients with cancer for the presence of heterozygous germline *RECQL4* LOF variants, and LOF variants in the other RecQ helicase genes, is warranted to validate and refine this new association.

## **METHODS**

## **Study Participants**

Study participants were from the Pediatric Cancer Genome Project (n = 1120), Genomes for Kids protocol (n = 309), St. Jude Lifetime Cohort (SJLIFE) Study (n = 3006), and Therapeutically Applicable Research to Generate Effective Treatment (TARGET) program (n = 1127).



The index case consented to participate in an IRB-approved study at St. Jude Children's Research Hospital that specifically allowed for germline interrogation and reporting.

## Variant Detection and Classification

Single-nucleotide variants, small insertions and deletions, and copy-number variations were detected from whole-exome and/or genome sequencing (WGS) of the germline as previously described (Zhang et al. 2015; Rusch et al. 2018). Genetic variants were annotated, and LOF variants were further analyzed.

## Curation of LOF Variants in gnomAD

Variants in RecQ helicase genes (*RECQL*, *BLM*, *WRN*, *RECQL4*, *RECQL5*) in the noncancer subset of the Genome Aggregation Database (gnomAD v2.1) were queried (Supplemental Tables 4–8). The putative LOF variants in gnomAD noncancer set v2.1 were downloaded first, and subsequently all variants of dubious quality via the tags "Flag" = "lc\_lof" (low confidence-loss-of-function) were excluded. Variants with the "Flag" = "lcr" or "Flag" = "segdup" were reviewed and retained if they passed the quality control and manual curation steps. In the case of *RECQL4*, the founder variant, c.1390+2delT (p.Ala420\_Ala463del), was removed as it is commonly encountered in the Finnish population at a minor allele frequency of 0.4% (Lek et al. 2016) but was absent from our cohort.

## **Statistical Analysis**

Statistical analysis of population enrichment was calculated via a 2 × 2 Fisher's exact test, and estimates of the OR were performed using the RStudio R statistical computing environment with the epiR package (https://CRAN.R-project.org/package=epiR). Statistical significance was defined by a two-sided P = 0.05.

## **Computational Prediction of Ethnicity**

Data from our pediatric cancer cohort (comprised of 4435 pediatric cancer patients at St. Jude Children's Research Hospital and 1127 from the National Cancer Institute TARGET database) was combined with data from 1000 Genomes, and the principal components (PCs) were extracted. Ethnicity data from 1000 Genomes were then used to train a random forest using the top 10 PCs which was used to predict the ethnicity of individuals in the pediatric cancer cohort. A visualization of the unsupervised ethnicity clustering is shown in Supplemental Figure 1.

## ADDITIONAL INFORMATION

#### **Data Deposition and Access**

Genomic sequence data from St. Jude Children's Research Hospital is available for request on the St. Jude Cloud Platform. The variants were submitted to ClinVar (https://www.ncbi .nlm.nih.gov/clinvar/) and can be found under accession numbers SCV000288218.6 and SCV000890968.1.

#### **Ethics Statement**

Patients reported here provided written informed consent permitting genomic analysis. Family histories have been illustrated in a de-identified manner. Participants at St. Jude Children's Research Hospital were consented to at least one of the following protocols: institutional banking protocol for Collecting, Banking and Distributing Human Tissue Samples at St. Jude Children's Research Hospital (TBANK; NCT01354002), Genomes for



Kids (G4K; NCT02530658), and/or Establishment of a Lifetime Cohort of Adults Surviving Childhood Cancer (SJLIFE; NCT00760656).

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## **Author Contributions**

J.L.M., K.V.H., K.E.N., and C.A.K. conceived the study and wrote the initial draft of the manuscript. K.V.H., R.B.M., R.N., R.M., and C.A.K. provided genetic counseling. A.S.P. and V.S. were and are involved with the clinical management of the index case. K.V.H., R.B.M., R.N., R.M., S.H.D., L.H., L.T., E.L.G., and A.O. provided participant recruitment, consent, and support. N.O., W.C., M.N.E., A.P., S.N., J.Z., Z.W., and G.W. developed software and pipelines for sequence data analysis and variant interpretation. N.O., W.C., Z.W., J.N., E.M.A., S.A.S., S.N., and G.W. analyzed sequence data and interpreted variants. J.L.M., N.O., G.W., and C.A.K. reviewed population databases and performed statistical analyses. D.W.E., J.R.D., M.M.H., L.L.R., S.N., J.Z., G.W., K.E.N., and C.A.K. provided project oversight. All coauthors reviewed the manuscript.

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