pISSN 1598-2629 · eISSN 2092-6685

# Roles of IL-33 in Resistance and Tolerance to Systemic Candida albicans Infections

Sang Jun Park<sup>1,2</sup>, Hong Rae Cho<sup>1,2</sup> and Byungsuk Kwon<sup>2,3</sup>\*

<sup>1</sup>Department of Surgery and <sup>2</sup>Biomedical Research Center, Ulsan University Hospital, School of Medicine, University of Ulsan, Ulsan 44033, <sup>3</sup>School of Biological Sciences, University of Ulsan, Ulsan 44610, Korea

IL-33 is a multifunctional cytokine that is released in response to a variety of intrinsic and extrinsic stimuli. The role of IL-33 in *Candida albicans* infections is just beginning to be revealed. This cytokine has beneficial effects on host defense against systemic *C. albicans* infections, and it promotes resistance mechanisms by which the immune system eliminates the invading fungal pathogens; and it also elevates host tolerance by reducing the inflammatory response and thereby, potentially, tissue damage. Thus, IL-33 is classified as a cytokine that has evolved functionally to protect the host from damage by pathogens and immunopathology.

[Immune Network 2016;16(3):159-164]

Keywords: IL-33, Candida albicans, Resistance, Tolerance

#### INTRODUCTION

Candida albicans is a major commensal fungus residing in the mucosal surfaces of our bodies. It is mainly found in the gastrointestinal tract, skin and genital tract. When the mucosal barrier is disturbed, C. albicans penetrates into the epithelium and causes systemic infection. Conditions associated with immunocomprimise, particularly neutropenia, are a major risk factor for invasive candidiasis (1). This deep-seated infection occurs, as hyphae spread to internal organs, among which the kidney is the major target (2). The mortality rate of patients with systemic candidiasis exceeds 30~40% even when potent antifungal agents are administered (3). Like many other pathogens, the host defense against invasive C. albicans infections is a function of immune detection and elimination of the pathogen (1). This defense strategy is referred to as resistance (4). However, mortality of the host following C. albicans infection may result largely

from immunopathology, in which the immune response against *C. albicans* inflicts tissue damage. Mice are known to tolerate high fungal burdens, even though uncontrolled fungal proliferation is lethal. Thus, it is important to understand defense mechanisms that reduce host susceptibility to tissue damage caused by fungal infection or related immunopathologies, but that do not affect the fungal burden. This defense strategy does not affect fungal burden. In this review, we discuss the role of IL-33 in host resistance and tolerance to systemic *C. albicans* infections.

### **IL-33**

IL-33 is a member of the IL-1 family of cytokines that delivers its signaling through ST2 receptors (5). Endothelial cells and epithelial cells rapidly produce IL-33 in response to a variety of intrinsic and environmental

Received on March 14, 2016. Revised on April 11, 2016. Accepted on April 12, 2016.

Abbreviations: DC, dendritic cell; HSPC, hematopoietic stem and progenitor cell; ILC2, type 2 innate lymphoid cell; iNOS, inducible nitric oxide synthase

http://immunenetwork.org

<sup>©</sup> This is an open access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/by-nc/4.0) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

<sup>\*</sup>Corresponding Author. Byungsuk Kwon, School of Biological Science, University of Ulsan, Daehak-ro 93, Nam-gu, Ulsan 44610, Korea. Tel: 82-52-259-2860; Fax: 82-52-259-2740; E-mail: bkwon@ulsan.ac.kr

Immune Network IL-33 in *C. albicans* Infections

insults (6). This is why IL-33 is referred to as an alarmin. However, there are many sources of IL-33, and the kinetics of IL-33 production vary across types of cells and stimuli (7). Interestingly, the IL-33 receptor, ST2, is expressed on virtually all types of immune cells and numerous types of nonhematopoietic cells, suggesting that it exhibits a functional diversity (8). IL-33 function is context-dependent and can have opposing effects (e.g., inflammatory or anti-inflammatory) on an immune response. For example, IL-33 initiates type 2 immune responses through secretion of IL-4, IL-5 and/or IL-13 by Th2 cells and type 2 innate lymphoid cells (ILC2s) (9). IL-33 is also associated with autoimmune and inflammatory diseases mediated by Th1 and Th17 cells (6). In addition, anticancer and antiviral effects of IL-33 require CD8<sup>+</sup> T cells, NK cells and ILC2s (10-13). Recent studies have identified anti-inflammation functions of IL-33: it plays a critical role in regulatory T cell expansion in acute colitis (14), asthma (15), and obesity (16). One effector molecule produced by regulatory T cells is amphiregulin, which plays a role in tissue repair without influencing antiviral immune responses (17). Thus, the seemingly anti-inflammatory effect of IL-33 is in fact a result of enhanced tolerance mechanisms. IL-33 also may exert its anti-inflammatory effects by suppressing Th1 or Th17-mediated immune responses. This outcome can be particularly important in inflammatory or infectious diseases in which Th1 or Th17 cells mediate immunopathology. Recent studies have begun to reveal the molecular mechanisms behind these observations. Specifically, IFN and IL-27 have been shown to counteract the activity of ILC2s mediated by IL-33 (18-20).

# IL-33 AND HOST RESISTANCE TO *C. albicans* INFECTIONS

The participation of IL-33 in host resistance to *C. albicans* infection was initially demonstrated in a fungal peritonitis model (21). IL-33 priming before infection with *C. albicans* resulted in effective fungal clearance and subsequently decreased mortality caused by fungal peritonitis (21). This priming by IL-33 acts on multiple steps of an anti-*Candida* neutrophil response. First, IL-33 priming enhances recruitment of neutrophils to the site of infection through two mechanisms: 1) It increases production of neutrophil-chemotactic CXCR2 ligands (CXCL1 and CXCL2) by peritoneal macrophages; and 2) it suppresses the internalization of surface CXCR2

in neutrophils. IL-33 does so by inhibiting TLR2-induced GRK2 expression that is required for CXCR2 internalization (22). Second, IL-33 priming of neutrophils increases their phagocytic and fungicidal activities. The increased phagocytosis of neutrophils occurs because of specific upregulation of the complement receptor CR3 through the synergistic activation of the TLR2 and Dectin-1 signaling pathways by IL-33. Increase in production of reactive oxygen species (ROS) is correlated with yeast killing of IL-33-primed neutrophils. It is not known why IL-33 specifically promotes clearance of opsonized *C. albicans*, but this occurred under physiological conditions (our unpublished data).

Mononuclear phagocytes, including resident macrophages and inflammatory monocytes, also play a critical role in *Candida* clearance (23,24). Administration of IL-33 before and during *C. albicans* infection markedly induces M2 macrophage polarization (25). IL-33-induced M2 polarization is mediated by CD4<sup>+</sup> T cell-derived IL-13 (25). Surprisingly, IL-33 stimulates bone marrowderived M2 macrophages to efficiently phagocytize and kill *C. albicans*. The mechanism of action is not known, but one possibility is that IL-33 increases killing by directly upregulating Dectin-1 and mannose receptor or by indirectly inducting IL-13 production (26,27).

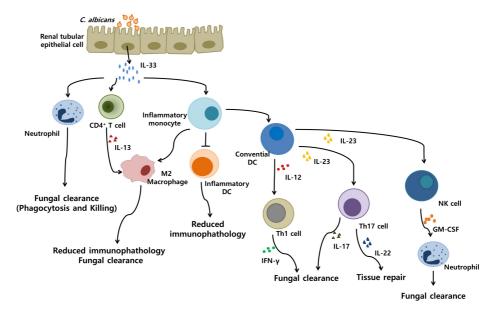
Massive emergency hematopoiesis occurs after infections to satisfy the need for myeloid cells in the periphery. Under these conditions, hematopoiesis is preferentially directed toward active granulopoiesis and suppressed lymphopoiesis in the bone marrow. In addition, hematopoietic stem and progenitor cells (HSPCs) emigrate from the bone marrow to the periphery and active extramedullary hematopoeisis occurs. Injection of IL-33 induces massive expansion of myeloid cells, particularly eosinophis, in the periphery (5). Recently, we provided an explanation for how IL-33 mediates this phenomenon (28). Vascular endothelial cells produce high levels of CCL7 in response to IL-33, and CCL7, in turn, induces emigration of HSPCs out of the bone marrow with the help of CCR2. On the other hand, IL-33 directs the differentiation of HSPCs toward myeolopoiesis in both the bone marrow and periphery by an unknown mechanism. Systemic C. albicans infections induce tubular epithelial cells of the kidney to produce IL-33 (our unpublished data) and ST2 is highly expressed on HSPCs (8). Thus, it is tempting to propose that IL-33 may be involved in HSPC differentiation during C. albicans infection. This hypothesis is substantiated by the observation that C. albicans stimulates differentiation of HSPCs toward monocytes (29). Altogether, these results strongly suggest that IL-33 may control emerIL-33 in *C. albicans* Infections

gency granulopoiesis to supply sufficient numbers of granulocytes to the site of infection, thus, indirectly increasing host resistance to *C. albicans* infections.

## IL-33 AND HOST TOLERANCE TO C. albicans INFECTIONS

Although administration of IL-33 increases host survival by promoting fungal clearance, the anti-inflammatory activity of IL-33 can also contribute to host survival during systemic C. albicans infections (25). There were significantly lower levels of proinflammatory mediators (IL-6, TNF- $\alpha$ , IL-1 $\beta$ , CXCL1, and CXCL2), relatively little neutrophil infiltrate, and M2 macrophage polarization in the kidneys of IL-33-primed mice throughout the course of infection. As mentioned previously, IL-13 mediates IL-33-induced M2 polarization. Interestingly, M2 polarization is required not only to keep fungal burden in check in the kidneys but also to maintain persistent immunosuppression associated with reduced Candida-induced immunopathology that leads to renal damage. Because hyperimmune responses are associated with more fatalities than high fungal burden in systemic C. albicans infections (29), IL-33 appears to enhance host tolerance to Candida infection via reduced immunopathology. Consistent with these results, IL-33<sup>-/-</sup> mice are more susceptible to systemic Candida infection compared to wild-type mice, showing impaired fungal clearance and severe renal immunopathology (our unpublished data). Severe renal inflammation is associated with a dysregulated differentiation of TNF- $\alpha$ - and iNOSproducing dendritic cells (Tip-DCs) in the kidney of Candida-infected IL-33<sup>-/-</sup> mice. In contrast, these mice have a lower number of IL-12- and IL-23-producing mature dendritic cells (DCs) in their infected kidneys. As reducing Tip-DCs decreases renal immunopathology without influencing fungal proliferation in IL-33 mice, IL-33 is a unique example of a factor that helps to reduce host vulnerability to damage by promoting pathogen clearance and reducing immunopathology (our unpublished data).

One important outstanding issue to explore is whether IL-33 is involved in types of tolerance other than immunopathology-related tolerance. IL-33 may control host tolerance to infections and other stresses through various means. For example, IL-33 can have direct protective effects on parenchymal cells of organs during a stress response (30), or IL-33 may exert tolerance through activation of cells that have a protective role against tissue injury. IL-33 administration expands ILC2s and regulatory T cells that can produce amphiregulin, a molecule critical



**Figure 1.** A schematic diagram demonstrating the functions of IL-33 in *C. albicans* infection. Invading *C. albicans* resides in the kidney and stimulates tubular epithelial cells to secrete IL-33. IL-33 increases phagocytosis by neutrophils and M2 macrophages either directly or indirectly, through the secretion of other cytokines such as IL-12 and IL-23 from DCs. IL-33 enhances tolerance by reducing immunopathology through increased M2 macrophage polarization and reduced inflammatory DC differentiation or presumably through promotion of tissue repair by IL-22 or amphiregulin.

http://immunenetwork.org

Immune Network IL-33 in *C. albicans* Infections

for tissue repair and host tolerance (17,31). Two other types of cells that can be targeted by IL-33 for tissue protection are M2 macrophages and Th17 cells. IL-22 secreted by these cells may be involved in tissue repair (32,33). However, dysregulation of IL-33 can result in chronic inflammation or fibrosis (34,35). Whether the tolerance mechanisms mentioned above operate in host defenses against systemic *Candida* infection needs to be tested. We propose that IL-33-induced tolerance is associated with an increased capacity for tissue repair and decreased immunopathology.

#### CONCLUDING REMARKS

Mechanisms of IL-33 action in Candida immunity are beginning to be defined. IL-33 production reaches a peak in the kidney at day 3 of systemic C. albicans infection, when maximum inflammation and active fungal proliferation are occurring. At this time, IL-33 seems to prevent overactivation of an inflammatory response on one hand and to promote fungal clearance on the other hand (Fig. 1). The former process may result mainly from blocking inflammatory DCs, while the latter may occur at multiple stages of innate and adaptive immune responses to C. albicans infection. IL-33 can directly enhance the phagocytic and fungicidal activity of phagocytes. IL-33 also promotes IL-23 production by DCs which results in activation of the NK cell-GM-CSF-neutrophil axis. Finally, DC-derived IL-12 and IL-23 may be critical in IL-33-mediated fungal clearance through the induction of Th1 and Th17 differentiation, respectively. These beneficial activities of IL-33 have therapeutic implications for patients with invasive candidiasis. The ability of IL-33 to promote Th1 and Th17 responses against C. albicans also indicates that IL-33 has merits as an anti-Candida vaccine adjuvant.

### **ACKNOWLEDGEMENT**

This work was supported by a grant from the National Research Foundation of Korea (funded by the Republic of Korea Ministry of Education, Science, and Technology Grant NRF-2013R1A1A1A01058298).

### **REFERENCES**

1. Lionakis, M. S. 2014. New insights into innate immune control

- of systemic candidiasis. Med. Mycol. 52: 555-564.
- Cassone, A. 2013. Development of vaccines for Candida albicans: fighting a skilled transformer. *Nat. Rev. Microbiol.* 11: 884-891.
- Horn, D. L., D. Neofytos, E. J. Anaissie, J. A. Fishman, W. J. Steinbach, A. J. Olyaei, K. A. Marr, M. A. Pfaller, C. H. Chang, and K. M. Webster. 2009. Epidemiology and outcomes of candidemia in 2019 patients: data from the prospective antifungal therapy alliance registry. Clin. Infect. Dis. 48: 1695-1703.
- Medzhitov, R., D. S. Schneider, and M. P. Soares. 2012. Disease tolerance as a defense strategy. Science 335: 936-941.
- Schmitz, J., A. Owyang, E. Oldham, Y. Song, E. Murphy, T. K. McClanahan, G. Zurawski, M. Moshrefi, J. Qin, X. Li, D. M. Gorman, J. F. Bazan, and R. A. Kastelein. 2005. IL-33, an interleukin-1-like cytokine that signals via the IL-1 receptor-related protein ST2 and induces T helper type 2-associated cytokines. *Immunity* 23: 479-490.
- Liew, F. Y., N. I. Pitman, and I. B. McInnes. 2010. Diseaseassociated functions of IL-33: the new kid in the IL-1 family. *Nat. Rev. Immunol*. 10: 103-110.
- Licona-Limon, P., L. K. Kim, N. W. Palm, and R. A. Flavell. 2013. TH2, allergy and group 2 innate lymphoid cells. *Nat. Immunol.* 14: 536-542.
- 8. Le, H., W. Kim, J. Kim, H. R. Cho, and B. Kwon. 2013. Interleukin-33: a mediator of inflammation targeting hematopoietic stem and progenitor cells and their progenies. *Front. Immunol.* 4: 104.
- Sonnenberg, G. F., and D. Artis. 2015. Innate lymphoid cells in the initiation, regulation and resolution of inflammation. *Nat. Med.* 21: 698-708.
- Villarreal, D. O., M. C. Wise, J. N. Walters, E. L. Reuschel, M. J. Choi, N. Obeng-Adjei, J. Yan, M. P. Morrow, and D. B. Weiner. 2014. Alarmin IL-33 acts as an immunoadjuvant to enhance antigen-specific tumor immunity. *Cancer Res.* 74: 1789-1800.
- Gao, X., X. Wang, Q. Yang, X. Zhao, W. Wen, G. Li, J. Lu, W. Qin, Y. Qi, F. Xie, J. Jiang, C. Wu, X. Zhang, X. Chen, H. Turnquist, Y. Zhu, and B. Lu. 2015. Tumoral expression of IL-33 inhibits tumor growth and modifies the tumor microenvironment through CD8<sup>+</sup> T and NK cells. *J. Immunol*. 194: 438-445.
- Kim, J., W. Kim, U. J. Moon, H. J. Kim, H. J. Choi, J. I. Sin, N. H. Park, H. R. Cho, and B. Kwon. 2016. Intratumorally establishing type 2 innate lymphoid cells blocks tumor growth. *J. Immunol.* 196: 2410-2423.
- Bonilla, W. V., A. Frohlich, K. Senn, S. Kallert, M. Fernandez,
  S. Johnson, M. Kreutzfeldt, A. N. Hegazy, C. Schrick, P.
  G. Fallon, R. Klemenz, S. Nakae, H. Adler, D. Merkler,
  M. Lohning, and D. D. Pinschewer. 2012. The alarmin

IL-33 in *C. albicans* Infections Immune Network

interleukin-33 drives protective antiviral CD8<sup>+</sup> T cell responses. *Science* 335: 984-989.

- Schiering, C., T. Krausgruber, A. Chomka, A. Frohlich, K. Adelmann, E. A. Wohlfert, J. Pott, T. Griseri, J. Bollrath, A. N. Hegazy, O. J. Harrison, B. M. Owens, M. Lohning, Y. Belkaid, P. G. Fallon, and F. Powrie. 2014. The alarmin IL-33 promotes regulatory T-cell function in the intestine. *Nature* 513: 564-568.
- 15. Morita, H., K. Arae, H. Unno, K. Miyauchi, S. Toyama, A. Nambu, K. Oboki, T. Ohno, K. Motomura, A. Matsuda, S. Yamaguchi, S. Narushima, N. Kajiwara, M. Iikura, H. Suto, A. N. McKenzie, T. Takahashi, H. Karasuyama, K. Okumura, M. Azuma, K. Moro, C. A. Akdis, S. J. Galli, S. Koyasu, M. Kubo, K. Sudo, H. Saito, K. Matsumoto, and S. Nakae. 2015. An interleukin-33-mast cell-interleukin-2 axis suppresses papain-induced allergic inflammation by promoting regulatory T cell numbers. *Immunity* 43: 175-186.
- Kolodin, D., P. N. van, C. Li, A. M. Magnuson, D. Cipolletta, C. M. Miller, A. Wagers, R. N. Germain, C. Benoist, and D. Mathis. 2015. Antigen- and cytokine-driven accumulation of regulatory T cells in visceral adipose tissue of lean mice. *Cell Metab*. 21: 543-557.
- Arpaia, N., J. A. Green, B. Moltedo, A. Arvey, S. Hemmers, S. Yuan, P. M. Treuting, and A. Y. Rudensky. 2015. A distinct function of regulatory T cells in tissue protection. *Cell* 162: 1078-1089.
- Molofsky, A. B., G. F. Van, H. E. Liang, S. J. Van Dyken, J. C. Nussbaum, J. Lee, J. A. Bluestone, and R. M. Locksley. 2015. Interleukin-33 and interferon-gamma counter-regulate group 2 innate lymphoid cell activation during immune rerturbation. *Immunity* 43: 161-174.
- Moro, K., H. Kabata, M. Tanabe, S. Koga, N. Takeno, M. Mochizuki, K. Fukunaga, K. Asano, T. Betsuyaku, and S. Koyasu. 2016. Interferon and IL-27 antagonize the function of group 2 innate lymphoid cells and type 2 innate immune responses. *Nat. Immunol.* 17: 76-86.
- Duerr, C. U., C. D. McCarthy, B. C. Mindt, M. Rubio, A. P. Meli, J. Pothlichet, M. M. Eva, J. F. Gauchat, S. T. Qureshi, B. D. Mazer, K. L. Mossman, D. Malo, A. M. Gamero, S. M. Vidal, I. L. King, M. Sarfati, and J. H. Fritz. 2016. Type I interferon restricts type 2 immunopathology through the regulation of group 2 innate lymphoid cells. *Nat. Immunol.* 17: 65-75.
- Le, H. T., V. G. Tran, W. Kim, J. Kim, H. R. Cho, and B. Kwon. 2012. IL-33 priming regulates multiple steps of the neutrophil-mediated anti-*Candida albicans* response by modulating TLR and dectin-1 signals. *J. Immunol.* 189: 287-295.
- 22. Alves-Filho, J. C., F. Sonego, F. O. Souto, A. Freitas, W. A. Verri, Jr., M. uxiliadora-Martins, A. Basile-Filho, A.

- N. McKenzie, D. Xu, F. Q. Cunha, and F. Y. Liew. 2010. Interleukin-33 attenuates sepsis by enhancing neutrophil influx to the site of infection. *Nat. Med.* 16: 708-712.
- 23. Lionakis, M. S., M. Swamydas, B. G. Fischer, T. S. Plantinga, M. D. Johnson, M. Jaeger, N. M. Green, A. Masedunskas, R. Weigert, C. Mikelis, W. Wan, C. C. Lee, J. K. Lim, A. Rivollier, J. C. Yang, G. M. Laird, R. T. Wheeler, B. D. Alexander, J. R. Perfect, J. L. Gao, B. J. Kullberg, M. G. Netea, and P. M. Murphy. 2013. CX3CR1-dependent renal macrophage survival promotes Candida control and host survival. *J. Clin. Invest* 123: 5035-5051.
- Ngo, L. Y., S. Kasahara, D. K. Kumasaka, S. E. Knoblaugh, A. Jhingran, and T. M. Hohl. 2014. Inflammatory monocytes mediate early and organ-specific innate defense during systemic candidiasis. *J. Infect. Dis.* 209: 109-119.
- Tran, V. G., H. J. Kim, J. Kim, S. W. Kang, U. J. Moon, H. R. Cho, and B. Kwon. 2015. IL-33 Enhances Host Tolerance to Candida albicans Kidney Infections through Induction of IL-13 Production by CD4<sup>+</sup> T Cells. *J. Immunol*. 194: 4871-4879.
- Coste, A., M. Dubourdeau, M. D. Linas, S. Cassaing, J. C. Lepert, P. Balard, S. Chalmeton, J. Bernad, C. Orfila, J. P. Seguela, and B. Pipy. 2003. PPARgamma promotes mannose receptor gene expression in murine macrophages and contributes to the induction of this receptor by IL-13. *Immunity* 19: 329-339.
- Gales, A., A. Conduche, J. Bernad, L. Lefevre, D. Olagnier, M. Beraud, G. Martin-Blondel, M. D. Linas, J. Auwerx, A. Coste, and B. Pipy. 2010. PPARgamma controls Dectin-1 expression required for host antifungal defense against *Candida albicans*. *PLoS Pathog*. 6: e1000714.
- Kim, J., W. Kim, H. T. Le, U. J. Moon, V. G. Tran, H. J. Kim, S. Jung, Q. T. Nguyen, B. S. Kim, J. B. Jun, H. R. Cho, and B. Kwon. 2014. IL-33-induced hematopoietic stem and progenitor cell mobilization depends upon CCR2. *J. Immunol*. 193: 3792-3802.
- 29. Majer, O., C. Bourgeois, F. Zwolanek, C. Lassnig, D. Kerjaschki, M. Mack, M. Muller, and K. Kuchler. 2012. Type I interferons promote fatal immunopathology by regulating inflammatory monocytes and neutrophils during Candida infections. *PLoS Pathog.* 8: e1002811.
- Sakai, N., H. L. Van Sweringen, R. C. Quillin, R. Schuster, J. Blanchard, J. M. Burns, A. D. Tevar, M. J. Edwards, and A. B. Lentsch. 2012. Interleukin-33 is hepatoprotective during liver ischemia/reperfusion in mice. *Hepatology* 56: 1468-1478.
- Monticelli, L. A., G. F. Sonnenberg, M. C. Abt, T. Alenghat, C. G. Ziegler, T. A. Doering, J. M. Angelosanto, B. J. Laidlaw, C. Y. Yang, T. Sathaliyawala, M. Kubota, D. Turner, J. M. Diamond, A. W. Goldrath, D. L. Farber, R. G. Collman, E. J. Wherry, and D. Artis. 2011. Innate lymphoid cells promote lung-tissue homeostasis after infection with influenza virus.

http://immunenetwork.org

Immune Network IL-33 in *C. albicans* Infections

- Nat. Immunol. 12: 1045-1054.
- 32. Kulkarni, O. P., I. Hartter, S. R. Mulay, J. Hagemann, M. N. Darisipudi, V. S. Kumar, S. Romoli, D. Thomasova, M. Ryu, S. Kobold, and H. J. Anders. 2014. Toll-like receptor 4-induced IL-22 accelerates kidney regeneration. *J. Am. Soc. Nephrol.* 25: 978-989.
- 33. Xu, M. J., D. Feng, H. Wang, Y. Guan, X. Yan, and B. Gao. 2014. IL-22 ameliorates renal ischemia-reperfusion injury by targeting proximal tubule epithelium. *J. Am. Soc. Nephrol.* 25:
- 967-977
- 34. Molofsky, A. B., A. K. Savage, and R. M. Locksley. 2015. Interleukin-33 in tissue homeostasis, injury, and inflammation. *Immunity* 42: 1005-1019.
- McHedlidze, T., M. Waldner, S. Zopf, J. Walker, A. L. Rankin, M. Schuchmann, D. Voehringer, A. N. McKenzie, M. F. Neurath, S. Pflanz, and S. Wirtz. 2013. Interleukin-33-dependent innate lymphoid cells mediate hepatic fibrosis. *Immunity* 39: 357-371.