International Harmonization of Nomenclature and Diagnostic Criteria (INHAND): Nonproliferative and Proliferative Lesions of the Rabbit

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The INHAND (International Harmonization of Nomenclature and Diagnostic Criteria for Lesions Project (www.toxpath.org/inhand. asp) is a joint initiative of the Societies of Toxicologic Pathology from Europe (ESTP), Great Britain (BSTP), Japan (JSTP) and North America (STP) to develop an internationally accepted nomenclature for proliferative and non-proliferative lesions in laboratory animals. The purpose of this publication is to provide a standardized nomenclature for classifying microscopic lesions observed in most tissues and organs from the laboratory rabbit used in nonclinical safety studies. Some of the lesions are illustrated by color photomicrographs. The standardized nomenclature presented in this document is also available electronically on the internet (http://www.goreni.org/). Sources of material included histopathology databases from government, academia, and industrial laboratories throughout the world. Content includes spontaneous lesions as well as lesions induced by exposure to test materials. Relevant infectious and parasitic lesions are included as well. A widely accepted and utilized international harmonization of nomenclature for lesions in laboratory animals will provide a common language among regulatory and scientific research organizations in different countries and increase and enrich international exchanges of information among toxicologists and pathologists. (DOI: 10.1293/tox.2021-I001; J Toxicol Pathol 2021; 34: 1838–292S)

Keywords: rabbit, pathology, toxicopathology, nomenclature, background findings, INHAND, New Zealand

SECTION 1: INTRODUCTION

The INHAND Project (<u>International Harmonization</u> of <u>Nomenclature and Diagnostic Criteria for Lesions</u>) is a joint initiative of the societies of toxicologic pathology from Europe (European Society of Toxicologic Pathology - ESTP), UK

(British Society of Toxicological Pathologists - BSTP), Japan (Japanese Society of Toxicologic Pathology - JSTP), and North America (Society of Toxicologic Pathology - STP) to unify, update and complete the existing WHO/IARC and STP/SSNDC nomenclature systems. The INHAND nomenclature and the related diagnostic criteria should represent the future international standard in toxicologic pathology. They represent a consensus of senior toxicologic pathologists and were reviewed by the INHAND-GESC (INHAND-Global Editorial and Steering Committee) for compliance with INHAND principles. The initial series of nomenclature publications were focused on lesions in rats and mice. With the decision of the SEND initiative (Standard for the Exchange of Non-clinical Data) to model the

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controlled terminology (CT) based on the INHAND nomenclature and the decision of the Federal Drug Administration (FDA) to make the use of the SEND CT mandatory for electronic submissions of nonclinical studies, the INHAND project was extended to other laboratory animal species including the monkey, rabbit, mini-pig, dog and fish.

However, the recommendations for diagnostic criteria and preferred terminology may not be applicable in all situations. Purposes of specific experiments or the specific context of a given study may require deviation from this standardized nomenclature and diagnostic criteria. The appropriate diagnoses are ultimately based upon on the discretion of the senior toxicologic Study Pathologist.

The present publication provides a set of standardized terms and diagnostic criteria to be used in toxicologic pathologic studies on the most commonly used strains of laboratory rabbits - New Zealand White (NZW) representative of nonpigmented and Dutch Belted representative of a pigmented rabbit. Throughout this publication, lesions applicable for use in toxicologic-pathology studies in rabbits are tabulated. As rabbits have been most frequently used for tissue specific studies in young animals (e.g. ocular, dermal, and intramuscular), compilation of a broad listing of the incidence of spontaneous or background findings, and a tabulation of the incidence of subacute to chronic responses to chemicals, drugs and biomaterials are limited. The terms and thus, the tabulations, build on the existing rodent nomenclature. In most instances, the description and definition of the rodent lesion applies to the rabbit and is not further described. This publication focuses on lesions that are unique to the rabbit and are not observed in rodents, and lesions in rabbit that share the same terminology with a rodent lesion but display different morphologic features. Lesions that are unique to rats or mice and are not to be used in rabbit are denoted accordingly in the tabulation. The tabulated lesions are categorized according to the following characteristics: "Common", "Uncommon", "Not Observed but Potentially Relevant" and "Not Applicable". The distinction between common and uncommon lesions is based on the occurrence in untreated laboratory rabbits in the authors' experience and is not based on published references. Also, it should be kept in mind that the rabbit used in toxicologic studies are usually of young age and are only for a relative short time on study, a fraction of the normal life span of a rabbit. In addition, references to lesions seen in older pet and breeding animals are mentioned in the text where relevant. Before entering a study, the health status of individual animals is checked carefully, and the individual rabbits selected for a toxicologic study are in excellent condition. For these reasons, the spectrum and frequency of changes are different from those in diagnostic laboratories, and, therefore, common age-related lesions including neoplasms are rarely seen in these animals. Thus the vast majority of neoplastic lesions have been categorized as "Uncommon". The category "Not Applicable" refers to rodent specific lesions and terms as the use of these terms in rabbits is considered not appropriate. Examples are chronic progressive nephropathy in the kidney or fibro-osseous lesion of bones. "Not Observed but Potentially Relevant" are changes that have not been described or observed in laboratory rabbits, however, the use of these terms has been considered permissible should a lesion meet the diagnostic criteria.

Like all other INHAND publications, the nomenclature and diagnostic criteria for the rabbit are also available online (www. goreni.org). The online version contains any change controls, additional images and useful links to differential diagnoses characterizing it as a practical tool for diagnostic work.

The recommended nomenclature is generally descriptive rather than diagnostic. The diagnostic criteria used require standard hematoxylin and eosin stained paraffin sections only. Histochemical or immunohistochemical staining characteristics may be addressed in the comments section of the respective lesion. Such special techniques may be required in some situations, but a comprehensive discussion of these methods is outside the scope of this publication. Systemic non-proliferative lesions that occur across organ systems and are not specific to an organ are reviewed in the section on systemic pathology. Although the rodent publications provide "synonyms" for each term, the non-rodent publications have used the notation "Other term(s)". While these "synonyms" or "other terms" have been used historically, the primary listed term is the preferred term and will link to the controlled terminology in SEND. These "other terms" are listed with some of the entries to aid the pathologist when comparing current study findings with archival material. These other terms are archaic terms and should not be used because they are no longer preferred diagnostic entities.

Lesions included in this nomenclature system may be further specified by modifiers. Criteria are given for modifiers that are considered to be of particular relevance. These modifiers should be consistently applied. It is upon the discretion of the pathologist to use additional modifiers not suggested in this nomenclature system. Such modifiers may describe the location, tissue type, or duration among others. Further principles of the INHAND nomenclature have been published separately¹.

SECTION 2: SYSTEMIC PATHOLOGY

There are a number of microscopic findings that may be seen across several organs and/or tissues and are not specific to just one organ system. There are also a number of different findings that are present across several organs and/or tissues that together constitute a syndrome. Those findings that occur in multiple tissues are listed here for convenience, and they are also described under the organ systems in which they occur if they have unique features. Syndromes specific to the rabbit are mentioned in individual chapters, but their definitive descriptions are presented here.

Rabbits used in general toxicology studies are bred under barrier conditions, which are microbiologically defined, and are kept in strictly controlled/biosecure facilities when on study, so infectious disease (parasitism, bacterial, fungal and viral diseases) is unlikely. *Pasteurella, Encephalitozoon* and *Eimeria* spp. have been reported in the past but are rarely observed today^{2, 3}.

The tables below give an indication of how frequently the changes may be observed in the laboratory rabbit, associated diseases/conditions, etiologies or inducing agents, and a list of tissues where they may be found. Where further explanation is deemed useful, selected lesions are discussed in more detail below the table (Table 1).

Apoptosis

Comments: For a full discussion see Elmore, S. (2007). Apoptosis: A Review of Programmed Cell Death. *Toxicologic Pathology*, *35*(4), 495–516

Infiltrate, Inflammatory Cell

Comments: NZW rabbits on surgical studies may develop granulomas if orthopedic sutures are implanted into dorsal fascia⁵. NZW Rabbits are commonly used to test biocompatibility by implantation of novel medical materials into intramuscular and other tissue locations. Tissue reaction is

scored using ISO 10993-6:2016⁶ by characterizing inflammatory cells, necrosis, granulation tissue and fibrosis. Injured and regenerating skeletal muscle may also stimulate adipogenesis resulting in fatty infiltration⁷. Fatty metaplasia needs to be differentiated from the normal fat pads containing blood vessels and nerves that occur between muscle bundles when implants are incorrectly implanted or medical materials migrate into intermuscular sites.

Metaplasia, Osseous or Cartilaginous

Comments: Cartilage and bone formation, with or without intraosseous bone marrow is a common finding with intramuscular but not subcutaneous implantation tests of bone substitution biomaterials. Osseous metaplasia is readily induced in dogs and baboons, and to a lesser extent in rabbits and mice with calcium phosphates (CP) or hydroxyapatite/ calcium phosphate (HCP). Rabbits form bone and bone marrow with HCP⁸⁻¹⁰. Rarely, cartilage, bone and bone marrow may form as a sequela of intramuscular implantation of novel polymers and other biomaterials.

Mineralization

Comments: Rabbits do not require Vitamin D to regulate calcium absorption from the gut. Excess calcium in the diet is therefore more likely to cause metastatic calcification than in other species¹¹.

Vacuolation, Macrophages

Other term(s): Phospholipidosis

Pathogenesis/cell of origin: Macrophage

Differential diagnoses: accumulation adipocyte, phagocytic vesicle, lysosome.

Comments: Phospholipid vacuoles may be positive for LAMP2. Similar to the other laboratory species. Tissues affected by phospholipidosis vary by drug.

Table 1.	1. Microscopic Findings of Systemic Pathology (General	lly Used Preferred Terms): Rabbit
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Finding	Common	Uncommon	Associated Diseases/ Conditions	Tissues commonly reported in:
Non-proliferative				
Congenital				
Agenesis/hypoplasia		Х		
Malformation syn. congenital malformation		Х		Gall bladder (bifurcation)
Multisystemic				
Abscess		Х		
Accumulation, adipocytes	Х			
Amyloid		Х		
Apoptosis *+		Х		
Atrophy		Х		
Basophilic granules		Х		
Congestion		Х		Nasal cavity, lung, liver, vagina
Edema		Х		
Extramedullary hematopoiesis		Х		Spleen, liver, adrenal
Fracture		Х		Lumbar spine, usually associated with handling
Hemorrhage	Х			Larynx, trachea, thyroid, thymus, bronchi, bronchioles
Infiltrate, inflammatory cell * [insert appropriate cell type]		Х	Mononuclear, lymphocyte, plasma cell, macrophage/ monocyte, neutrophil, eosinophil, heterophil, mixed	Multiple tissues; differentiate form MALT
Inflammation		х	Acute, chronic, chronic active, granulomatous, granuloma	Multiple tissues; may be due to bacterial, viral or parasitic diseases e.g. <i>Pasteurella</i> sp. in the lungs; foreign body inflammatory reactions with multinucleated giant cells are common with implantation of biomaterials and medical devices
Metaplasia		Х		
Metaplasia, Osseous/cartilaginous *		Х		Lung, eye and skeletal muscle (implant associated)
Mineralization *	Х			Ovary, kidney, cerebral & cerebellar leptomeninges, blood vessels, lung, skeletal muscle
Necrosis		Х		
Parasite		Х	Coccidiosis (<i>Eimeria spp</i>); Microsporidiosis (<i>Encephalitozoon cuniculi</i>)	Adrenal gland, brain, eye, intestine, kidney, liver
Pigment		Х	Hemosiderin	Spleen, liver, bone marrow, adrenal, glomeruli, lymph node sinuses
Pigment		Х	Melanin, hemosiderin, lipofuscin	Skin, leptomeninges of pigmented strains
Pigment, macrophage	Х		Tattoo ink, inhaled particulate matter	(cervical) lymph node (from ear tattoos), lung
Serous atrophy of fat		Х		
Single cell necrosis [‡]		Х		
Tissue, ectopic	Х			Accessory adrenal cortical tissue, accessory spleen, bifurcate gall bladder; ectopic thyroid, ectopic thymus
Vacuolation		Х		Spleen, lymph node, lungs, choroid plexus
Vacuolation, macrophages *#		Х		
Proliferative Neoplastic				
Lymphoma		x		Liver small intestine multiple tissues

* Terminology with diagnostic criteria or comments described in the text. # Inducible lesion. ‡ Refer to ⁴ for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

SECTION 3: CARDIOVASCULAR SYSTEM

A. Anatomy of the Heart

Detailed anatomy and physiology of the rabbit heart has been described by various authors^{12, 13} and is not within the scope of this text. However, one difference to note is that the right atrioventricular valve of the heart is bicuspid instead of tricuspid as occurs in the rodent.

Histopathology of the heart should include all relevant compartments and structures of the heart, including the ventricular, atrial and interventricular septal wall, the valves and the coronary vessels.

Xenobiotics may cause myocardial changes due to exaggerated pharmacodynamic activity or as a result of a direct toxic effect on cardiomyocytes. Severe acute toxic insults can cause acute cardiomyocyte death, and the regenerative potential of cardiomyocytes is generally insufficient to replace significant myocardial loss. Biochemical changes such as alterations in calcium homeostasis can occur if the insult is of mild severity, and these generally result in reversible cardiac arrhythmia. Cardiomyocytes are generally replaced by fibroblasts, with collagenous deposits leading to loss of cardiac contractility. Changes in the heart are known to be induced by "stress" i.e. catecholamine (CA) induced cardiomyopathy. Study-related procedures can elevate serum stress biomarkers and exacerbate the frequency and severity of myocardial inflammatory cell infiltrates¹⁴.

Rabbit models of heart disease have been comprehensively reviewed¹⁵ (Table 2).

Malformation

Other terms: Congenital malformation

Comments: Congenital lesions of the heart and blood vessels are rare and are reported infrequently in rabbits. Consequently, the lesions are usually only seen in Developmental and Reproduction Toxicity (DART) studies, in which fetuses are carefully dissected. Ventricular septal defect has been reported in a 10-month-old female NZW rabbit¹⁶. Other conditions occasionally seen in rabbits are: right sided aortic arch and patent ductus arteriosus.

Fibrosis, Myocardium (Figure 1)

Comments: Myocardial fibrosis may be induced in rabbits after anesthesia with the α 2-agonist detomidine, alone and in combination with ketamine or diazepam^{17, 18}. The presence of myocardial fibrosis does not always result in clinical signs and adversity should be judged on a case by case basis. There is also an age-related increased fibrosis in the ventricles and interventricular septum. Ventricular stiffness and wall thickness increase in the aging rabbit heart¹⁹.

Infiltrate, Inflammatory Cell, Myocardium (Figure 2)

Comments: Mononuclear inflammatory cell infiltrates are recorded infrequently in the myocardium. The foci are usu-

ally at the base of the interventricular septum but have been reported in the atrial and ventricular free walls. There is no accompanying myocardial necrosis or fibrosis associated with this lesion.

Inflammation, Myocardium

Comments: Inflammatory cells may be associated with cardiomyocyte necrosis, interstitial edema and early fibrosis (see Necrosis). This finding can be induced by catecholamines secondary to stress^{14, 20}. Severity and incidence are increased in rabbits subjected to more handling and procedures. Increase in circulating catecholamines act on adrenergic receptors expressed in the heart and stimulate contractility. Heart lesions are primarily in the left ventricle and papillary muscle^{20, 21}. In severe cases, the inflammatory cell foci may resolve as focal fibrosis. This tends to be an idiosyncratic reaction and may affect one or two animals in a study, suggesting a subset of animals may be more vulnerable to stress responses and/or do not habituate to stressors¹⁴.

Mineralization, Cardiomyocyte/Myocardium (Figure 3)

Comments: Generally a background lesion but may be exacerbated by some xenobiotics; common in left atrial appendage. Mineralization may be seen at necropsy in older animals, e.g. ex-breeding colony animals.

Necrosis, Cardiomyocyte (Figure 4)

Other terms: Degeneration/necrosis, cardiomyocyte

Comments: Myocardial inflammation with/without minimal necrosis and/or fibrosis may be seen as a stress-induced finding in occasional animals on toxicity studies. These foci are minimal to moderate in severity grade and most affect one part of the myocardium, usually papillary muscles, left ventricular free wall, or may be multifocal throughout intraventricular septum, right ventricular free wall and atria. They are more commonly seen in animals subject to multiple procedures or handling events and thought to be catecholamine induced necrosis. Animals may be found in extremis or dead without previously showing any clinical signs. Although only an occasional occurrence in young rabbits used in toxicology studies, this sudden death syndrome is recognized as a stress induced event, caused by handling/ invasive procedures (injections, blood sampling) in pet rabbits (Bradley, unpublished data). A recent study showed that the incidence, composition and severity of these foci may be exacerbated by handling and procedures that occur in toxicology studies, mediated by a stress response¹⁴.

Myocardial necrosis and fibrosis can be induced in rabbits after anesthesia with the α 2-agonist detomidine, alone and in combination with ketamine or diazepam^{17, 18}. There is reduction in coronary flow reserve as a consequence of the hypoxemia associated with ketamine/xylazine administration due to xylazine interaction with α 2-receptors in coronary vessels. Impairment of coronary blood flow causes myocardial ischemia with subsequent necrosis. The rabbit is a species with limited collat-

Table 2. Microscopic Findings of the Heart: Rabbit

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Malformation *		Х		
Non-proliferative				
Accumulation, adipocyte, myocardium	Х			
Amyloid		Х		
Apoptosis ‡		Х		
Atrophy		Х		
Cardiomegaly			Х	
Degeneration		Х		
Degeneration/necrosis		Х		
Edema			Х	
Fibrosis, myocardium *		Х		
Fibrosis			Х	
Hypertrophy		Х		
Infarct		Х		
Infiltrate, inflammatory cell, [insert appropriate cell type] myocardium *	Х			
Inflammation, myocardium *		Х		
Karyomegaly/Karyocytomegaly			Х	
Mineralization *	Х			
Necrosis, cardiomyocyte *		Х		
Necrosis/infiltrate		Х		
Parasite		Х		
Pigment		Х		
Single cell necrosis #		Х		
Thrombus, atrium			Х	
Tissue, ectopic, thyroid		Х		
Vacuolation, cardiomyocyte		Х		
Proliferative Non-neoplastic				
Hyperplasia, subendocardium			Х	
Hyperplasia, mesothelium			Х	
Proliferative Neoplastic Lesions				
Schwannoma, endocardium			Х	
Schwannoma, intramural			Х	
Mesothelioma, pericardium			Х	
Mesothelioma, atriocaval			X	

* Terminology with diagnostic criteria or comments described in the text. + Refer to 4 for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

eral circulation in the myocardium and is therefore predisposed to ischemia induced by coronary vasoconstriction^{22, 23}.

B. Anatomy of the Blood Vessels

Rabbit blood vessels are generally thin-walled and prone to collapse and may form hematomata on puncture – a feature important for studies where test items are given by intravenous administration. The exception to this is the pulmonary arteries which are enveloped in a prominent smooth muscle layer, which may be misinterpreted as hypertrophy^{24, 25}.

Specific large vessels may be required on safety assessment studies in which animals have been dosed via intravascular catheter through bolus injection or slow infusion. Large vessels that can be easily sampled in rabbits include the thoracic and abdominal aorta/vena cava, carotid arteries, femoral arteries/veins and iliac arteries/veins. Some lesions occur in vesselspecific progression (e.g. atherosclerosis) (Table 3).

Malformation

Other terms: Congenital malformation

Comments: Congenital lesions of the blood vessels are rare and are reported infrequently in rabbits. Consequently, the lesions are usually only seen macroscopically in Developmental and Reproduction Toxicity (DART) studies, in which fetuses are carefully dissected. Conditions occasionally seen in rabbits are right sided aortic arch and patent ductus arteriosus.

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Malformation *		Х		
Non-proliferative				
Abscess		Х		
Amyloid		Х		
Aneurysm, artery or aortic			Х	
Angiectasis			Х	
Apoptosis [‡]		Х		
Degeneration/necrosis, media or wall		Х		
Dilatation		Х		
Embolus		Х		
Fibrosis, perivascular		Х		
Hemorrhage, media or wall		Х		
Hypertrophy, endothelium/media or wall, artery		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation, media or wall, artery		Х		
Intimal thickening, acellular *#		Х		
Intramural plaque, artery			Х	
Metaplasia			Х	
Mineralization *	Х			
Necrosis		Х		
Necrosis/inflammation, media or wall, artery		Х		
Single cell necrosis [‡]		Х		
Thrombus			Х	
Vacuolation, media or adventitia, artery			Х	
Proliferative Non-Neoplastic Lesions				
Hyperplasia, intima		Х		
Proliferative Neoplastic Lesions				
Hemangioma			Х	
Hemangiosarcoma			Х	

Table 3. Microscopic Findings of the Vessels and Valves: Rabbit

* Terminology with diagnostic criteria or comments described in the text. # Inducible lesion. + Refer to 4 for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

Intimal Thickening, Acellular

Other terms: Atherosclerosis

Diagnostic Features: Atherosclerotic plaques primarily composed of macrophage-derived foam cells.

Comments: In rabbits, the presence of atherosclerosis-like lesions was first described in 1913²⁶. High plasma cholesterol concentrations, especially of low-density lipoprotein (LDL) cholesterol, result in atherosclerotic lesion formation. The accumulation of foam cells results in the formation of fatty streaks, the earliest observable abnormality of the vessel wall. The rabbit exhibits hypercholesterolemia within a few days of an administration of a high cholesterol diet, and so can be used as an animal model in efficacy studies to see if drugs affect inducement of atheromatous lesions. The basal release of nitric oxide (NO), is greater with endothelium-intact aortic rings from female rabbits than from male rabbits, and so this protective effect of circulating estradiol

means female animals are less prone to diet-induced atherosclerotic lesions than male animals. Cholesterol-rich diets have been used to induce widespread atheromatous lesions within a short time period (3 months). Genetically altered strains of the NZW rabbit are also used extensively. The Watanabe rabbit (Watanabe heritable hyperlipidaemia rabbit, WHHL) is used to study the pathology of type IIa human familial hypercholesterolemia. The WHHL rabbit has a genetic deprivation of functional LDL receptors. In these animals, the atherosclerotic process begins in utero, and the lesions progress with age27. Rinke and Hartmann characterized atherosclerosis in NZW and WHHL rabbits²⁸. The lesions were extremely pronounced in the vessels of the cardiac atria, including the valves and extended with degenerative changes into the myocardium. Some nearly occlusive arteries, without surrounding myocardial change, were also seen. Inflammatory response and areas of regeneration, occasionally accompanied by dystrophic mineralization, were

frequently observed. Rabbit models for the study of human atherosclerosis have been reviewed previously²⁹.

Mineralization, Aorta/Medial or Mural Artery (Figure 5)

Comments: Generally, a background lesion but may be exacerbated by some xenobiotics. Occasionally evidence of the closure of the ductus arteriosus may be seen as foci of mineralization in the media of major blood vessels of young rabbits, depending on plane of section. Calcification of the

aorta, pulmonary artery, and femoral artery may be seen microscopically in young animals as an incidental finding. These lesions can be exacerbated by an increased calcium supply or vitamin D overdosage¹¹ or where fresh pellets are given (freshly opened packets of standard rabbit chow contain more degradable vitamins as the declaration of ingredient contents are corrected to be those present at the expiry date of the food). Mineralization may be seen at necropsy in older animals, e.g. ex-breeding colony animals.

SECTION 4: DIGESTIVE SYSTEM B. A (ORAL CAVITY, SALIVARY GLANDS, ESOPHAGUS, SAL

STOMACH, INTESTINES, AND EXOCRINE PANCREAS)

A. Anatomy of the Oral Cavity and Esophagus

The mouth opening of rabbits is small so that a thorough examination of the buccal cavity is difficult or in some animals almost impossible. The oral cavity is long and curved. Erosions of the mucosa may occur due to irregular growth or sharp edges of broken teeth.

Teeth

The dental formula of the rabbit is 26 or 28 teeth. The maxilla contains 4 incisors, no canines, six premolars, and 4-6molars. The second set of maxillary incisors are small, caudal to the main incisors, and are known as the peg teeth. The mandible contains 2 incisors, no canines, 4 premolars, and 6 molars. Rabbits are hypsodonts and have a long crown without a true tooth root³⁰. Rabbits chew their food using their tongue elaborately, moving their jaw more than 120 times per minute.

Tongue

The tongue is relatively large in comparison to the overall body size of rabbits and has the standard 4 papillae types, namely vallate, foliate, fungiform, filiform³⁰.

Esophagus

The esophagus comprises three layers of semi-involuntary striated muscle; the rabbit esophagus lacks mucus glands³⁰ (Table 4).

Cleft Palate

Comments: Cleft Palate is a common finding in rabbit teratogenicity studies. Congenital alveolar cleft is a malformation occurring as a result of non-fusion of primary palate during weeks 4–12 of gestation and may be induced by glucocorticoids³¹.

Hyperplasia

Comments: Gingival hyperplasia has been recorded in NZW rabbits administered cyclosporine A chronically (L. Himmel, personal communication).

Papilloma, Squamous Cell

Comments: Caused by oral papilloma virus (papovavirus). Small grayish nodules present on the ventral buccal cavity and/or the underside of the tongue. Lesions are seen in animals 2–18 months old. The infection is self-limiting and uncommon in young animals but may occur in older breeding stock.

B. Anatomy of the Salivary Glands

Salivary Glands

There are 4 pairs of salivary glands in the rabbit: parotid, submaxillary (also called mandibular), sublingual, and zygomatic. The parotid gland is the largest and runs from below the ear base to the front of the ear base and is bounded by the skin and masseter muscle. The parotid gland duct runs rostrally along the lateral surface of the masseter muscle and is adjacent to the branches of the facial nerve. The parotid gland duct empties into the oral cavity opposite the last upper molar. The submaxillary gland is oval-shaped and located at the angle of the mandible. The sublingual gland is small in the rabbit. The zygomatic salivary gland rests just ventral to the lacrimal gland adjacent to the anteroventral angle of the orbit (see also Lacrimal Glands subsection in Special Senses section). Rabbit saliva contains amylase but has only trace amounts of lipase and urea. The rabbit submaxillary gland has continuous secretion of saliva³⁰ (Table 5).

Accumulation

Comments: This diagnosis should be used to describe the microscopic correlates with a macroscopic sialocele.

Depletion, Secretory, Acinar Cell

Comments: This is seen as a generalized process secondary to reduced food intake in rabbits as in many other species, and also in animals suffering from mucoid enteropathy. There may be diffuse depletion of parotid salivary gland zymogen, with associated vacuolar degeneration of exocrine cell cytoplasm³².

Necrosis, Glandular

Other terms: Infarct; Metaplasia, Squamous Cell

Unilateral minimal focal/multifocal mandibular salivary gland necrosis with acute inflammation has been reported in rabbits after auricular artery catheterization. Although thrombi were not identified microscopically, necrosis/acute inflammation was consistent with recent infarction³³. Extensive coagulative necrosis and associated squamous metaplasia (so-called "necrotizing sialometaplasia") have been reported in the mandibular salivary as sequelae of mandibular fracture³⁴ and photodynamic therapy (di-sulfonated phthalaocyanine and laser irradiation)³⁵ and were likewise attributed to infarction³⁵.

C. Anatomy of the Stomach

Stomach

The stomach of a healthy rabbit is never empty, due to the practice of coprophagy, and like the rat and horse, a rabbit cannot vomit³⁶. In new-born rabbits, an empty stomach indicates agalactia of the mother. The stomach is large, thin walled, and unlike rodents, there is no non-glandular region

 Table 4.
 Microscopic Findings of the Oral Cavity, Pharynx, Tongue and Esophagus: Rabbit

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Non-proliferative				
Congenital				
Cleft palate *	Х			
Malformation			Х	
Non-proliferative				
Amyloid		Х		
Apoptosis [‡]		Х		
Cyst	Х			
Degeneration/necrosis, muscle		Х		
Edema		Х		
Erosion/ulcer	Х			
Hemorrhage		Х		
Hyperkeratosis		Х		
Infarct		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]	Х			
Inflammation, foreign body		Х		
Inflammation, mixed cell	Х			
Inflammation, mononuclear cell	Х			
Inflammation, vessel		Х		
Metaplasia			Х	
Mineralization (+ locator)	Х			
Necrosis (+ locator)	Х			
Pigment		Х		
Single cell necrosis [‡]		Х		
Syncytia, epithelium	Х			
Tissue, ectopic		Х		
Tissue, ectopic, sebaceous gland		Х		
Yeast		Х		
Proliferative Non-Neoplastic				
Hyperplasia *#		Х		
Proliferative Neoplastic				
Adenoma			Х	
Adenocarcinoma			Х	
Carcinoma, squamous cell			Х	
Leiomyoma			Х	
Leiomyosarcoma			Х	
Papilloma, squamous cell *		Х		
Tumor, granular cell, benign			Х	
Tumor, neuroendocrine cell			Х	
		-		

* Terminology with diagnostic criteria or comments described in the text. # Inducible lesion. * Refer to 4 for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

in the rabbit stomach (Figures 6, 7). Occasionally, a bezoar may be found acting as a foreign body and eventually obstructing the pyloric region. Free ingesta in the abdominal cavity may mimic rupture of the gastric wall. *Helicobacter* spp. have been identified in the stomach of rabbits but not associated with inflammation or ulceration³⁷. The stomach is prone to very fast autolysis and in premature decedent toxicologic study animals it should be sampled as soon as possible (Table 6).

Parasite

Comments: Parasites of the gastrointestinal system include nematodes, cestodes and protozoans, but are rarely a problem in well-managed barriered facilities. Protozoal parasites cause the most common and significant disease. Numerous species of *Eimeria* are capable of infecting rabbits: *Eimeria stiedai* (frequently referred to as *E. stiedae*), *E. magna, E irresidua* and *E. intestinalis.*

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Ectopic tissue	Х			
Non-proliferative				
Abscess		Х		
Accumulation *		Х		
Accumulation, adipocytes		Х		
Amyloid		Х		
Angiectasis		Х		
Apoptosis ⁺		Х		
Atrophy	Х			
Calculus, duct		Х		
Cyst	Х			
Degeneration/necrosis		Х		
Degranulation, (+ locator)		Х		
Depletion, secretory *		Х		
Dilatation, duct		Х		
Ectasia, duct		Х		
Edema		Х		
Focus, basophilic		Х		
Fibrosis		Х		
Granules increased		Х		
Hemorrhage		Х		
Hypertrophy		Х		
Infarct		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]	Х			
Inflammation, mixed cell	Х			
Inflammation, mononuclear cell	Х			
Inflammation, vessel		Х		
Metaplasia, acinar cell		Х		
Metaplasia, osseous		Х		
Metaplasia, squamous cell	Х			
Mineralization (+ locator)	Х			
Multinucleated giant cells		Х		
Necrosis, glandular *	Х			
Pigment		Х		
Secretion, deceased, acinar cell		Х		
Single cell necrosis [‡]		X		
Tissue. ectopic		X		
Vacuoles, autophagic, acinar cell		X		
Vacuolation		X		
Yeast	Х			
Proliferative Non-neoplastic				
Hyperplasia #		х		
Proliferative Neoplastic				
Adenoma			х	
Adenocarcinoma			X	
Myoepithelioma, malignant			Х	
Tumor, mixed, benign			Х	
Tumor, mixed, malignant			Х	

 Table 5.
 Microscopic Findings of the Salivary Glands: Rabbit

* Terminology with diagnostic criteria or comments described in the text. # Inducible lesion. + Refer to 4 for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Ectopic tissue	Х			
Malformation			Х	
Non-proliferative				
Amyloid		Х		
Apoptosis ‡		Х		
Atrophy	Х			
Cyst	Х			
Degeneration/necrosis		Х		
Dilatation, glands	Х			
Diverticulum	Х			
Edema		Х		
Erosion/ulcer	Х			
Globules, eosinophilic		Х		
Helicobacter sp.		Х		
Hemorrhage		Х		
Infarct		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]	Х			
Inflammation (+ cell type)		Х		
Metaplasia		Х		
Mineralization (+ locator)	Х			
Necrosis, mucosa	Х			
Parasite *		Х		
Pigment		Х		
Single cell necrosis [‡]		Х		
Syncytia, epithelium	Х			
Yeast	Х			
Proliferative Non-neoplastic				
Hyperplasia #		Х		
Proliferative Neoplastic				
Adenoma			Х	
Adenocarcinoma			Х	
Gastrointestinal stromal tumor (GIST), benign			Х	
Gastrointestinal stromal tumor (GIST), malignant			Х	
Leiomyoma			Х	
Leiomyosarcoma			Х	
Tumor, basal cell, benign			Х	
Tumor, neuroendocrine cell, benign			Х	

 Table 6.
 Microscopic Findings of the Stomach: Rabbit

* Terminology with diagnostic criteria or comments described in the text. # Inducible lesion. + Refer to 4 for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

D. Anatomy of the Small and Large Intestines

Intestine

The intestinal tract of the rabbit is anatomically distinct from that of the other laboratory species. Locations from which samples for histopathological examinations should be ideally taken are illustrated.

Duodenum

The length of the duodenum is relatively great in rabbits

and the proximal part of the duodenum is characterized by a wide layer composed of Brunner's glands. The sample for histology therefore should be taken close to the pyloric region. In the rabbit two distinct cell types are present within the glands. Serous cells, which occur in small groups in the blind endings to the tubules, contain a concentration of rough endoplasmic reticulum in the basal cytoplasm, and the apical cytoplasm is occupied principally by discrete secretory droplets. Intercellular secretory canaliculi occur between opposed serous cells and between serous and

195S

mucous cells. The latter cells possess little rough endoplasmic reticulum but exhibit an extensive development of the Golgi apparatus in the supranuclear region. Secretory droplets are pale and show a tendency to fuse into complexes. No intercellular canaliculi occur between mucous cells³⁸. In contrast to other species, Paneth cells are visible in the duodenum at the base of the duodenal crypts and are easily recognized by the large eosinophilic granules that occupy most of the cytoplasm.

Jejunum

The jejunum is located along the left side of the cecum. It is very long and its junction with the ileum is indicated by the ileocecal fold, which attaches to the apex of the vermiform appendix of the rabbit. The jejunum should be sampled distal to the junction with the pancreatic ducts. The villi of the jejunum are thin and longer than in the duodenum and display a tall columnar epithelium with a low number of intermingled goblet cells. Paneth cells are easily to detect at the base of the crypts. Peyer's patches can be found only in the distal region of the jejunum.

Ileum

The ileum is also located along the left side of the cecum. The terminal part of the ileum is enlarged to form the round, expanded muscular sac – the sacculus rotundus. The sacculus rotundus is a common site for foreign body impaction. Both the sacculus rotundus and vermiform appendix appear pale coloured at necropsy due to the large amount of lymphoid tissue in their walls. These two lymphoid organs contain more than 50% of the total lymphoid tissue of the rabbit, accounting for the relatively small size of the spleen. The surface of the sacculus rotundus is covered by short villi, and the lamina propria is similar to that of the vermiform appendix. The thickness of the lymphoid tissue is variable. The tunica muscularis of the sacculus rotundus is thicker than that seen in the vermiform appendix.

Cecum

The cecum occupies the major portion of the middle to lower abdomen ventrally, being coiled around itself into three major turns or gyri and is freely movable in the peritoneal cavity. The cecal wall consists of epithelium-lined lamina propria, with short indented crypts giving an irregular contour to the surface. Both tunica muscularis and submucosa are thin. Scattered goblet cells are located between tall columnar cells³⁹. The haustrated cecum is tightly coiled and tapers to form the light-colored vermiform appendix. The rabbit is a hindgut fermenter, and so the cecum is much larger than the stomach (about 10 times the size). There is a small area of lymphoid tissue approximately 2 mm in diameter on the inner wall of the cecum adjacent to the ileocecal orifice. The lymphoid follicles at this site have direct contact with the lumen of the cecum (unlike the follicles in the vermiform appendix and sacculus rotundus). The lymphoid tissue in this ileocecal plaque is more loosely arranged and

more diffuse in character than that of the other two intestinal lymphoid structures. A tall columnar epithelium covers the surface of the follicle and goblet cells are rare. The tunica submucosa and muscularis are thin³⁹.

The vermiform appendix consists of a thick continuous layer of lymph follicles, the apical portions of which extend as protrusions above the lymph follicle proper. These protrusions are covered by columnar epithelium which is strongly infiltrated with lymphoid cells. At the base of the protruding portion of the follicle a thin column of lamina propria, emerging from the reticular connective tissue surrounding the lymph follicles, extends between the protruding portions of neighbouring follicles. This column of lamina propria is covered by a high columnar epithelium which contains numerous goblet cells and is continuous with the epithelium overlying the lymph follicle at the base of the apical protrusion. Above the bulging portion of the follicle the columns of lamina propria combine to form a covering over the follicle with a slightly lower columnar epithelium. Oval and slit like openings indicate areas which are not covered by the lamina propria. Occasional goblet cells are present in this columnar epithelium; the lamina muscularis mucosae is dispersed and difficult to define. The lymph follicles constitute 70% of the entire thickness of the intestinal wall. The epithelium adjacent to the lymphoid follicles is of a higher columnar cell type. The nuclei are more elongated, microvilli are short and sparse, and vesicles and mitochondria are abundant on the apical portions when examined by SEM. Goblet cells are numerous within this epithelium³⁹.

Colon

The proximal part of the ascending colon is closely associated with the cecum but can be identified by its characteristic tight haustrations and prominent teniae. The transverse colon is divided into proximal and distal portions and is separated by the fusi coli - a muscular spindle-shaped organ with a greatly thickened mucosa. The fusi coli is heavily supplied with ganglion cell aggregates and is under the influence of prostaglandins. The fusi coli, along with the muscular contractions of the sacculations and haustra, is responsible for directing the separation of fiber from nonfiber components of feeds. This speeds the fiber components through the colon where it is excreted as hard feces. Antiperistaltic action moves fluids and small particles in a retrograde manner through the colon to the cecum, where it is retained for fermentation⁴⁰. Cecal contents are selectively passed as cecotrophs, also referred to as "soft feces", to be consumed directly from the rectum. Cecotrophs are covered with mucus to protect them from the acid pH of the stomach (pH 1.2–1.5) and composed of water, nitrogen, electrolytes and vitamins. The arrival of cecotrophs at the anus triggers a neural response, resulting in licking the anal area and consumption of the cecotrophs. This is usually 4-8 hours after feeding, generally in the evenings, therefore cecotrophs are also known as "night feces". These soft pellets contain twice the protein and half the fibre of the

daytime fecal pellets. Coprophagy improves the utilisation of nitrogen, provides an abundance of certain B vitamins and conserves water. The excretion of hard daytime feces is related to feeding. Daytime fecal pellets are firm and dry, are excreted during the first four hours after feeding and are not ingested.

The most proximal part of the colon, adjacent to the sacculus rotundus, is expanded to form the bulb like structure – the ampulla cecalis coli, which is the most muscular portion. It consists of epithelial lined lamina propria (tunica mucosa) possessing wide open crypts. The walls of the crypts often show irregular contours, especially near the lumen, and give the impression of villi. The tunica muscularis is thicker than in other parts. Goblet cells are scarce³⁹. The ampulla coli is entirely free of mesenteric connections. Distal to the ampulla, the ascending colon spirals around the cecum producing several flexures before joining the transverse colon. The distal ascending colon, and transverse colon are small in diameter, and are not haustrated.

Peri-rectal tissue

There are focal apocrine-type glands in the submucosa near the anorectal junction, often called "anal glands" (further discussed in the Integument section). Their relationship to the "inguinal" (apocrine/sebaceous) gland complex (Integument section) is not clear (Table 7).

Degeneration, Neuron, Myenteric Plexus

Diagnostic features: Chromatolytic degeneration of preand postganglionic sympathetic and parasympathetic neurons (enteric neurons in the myenteric and submucosal plexus in the small intestine), as well as chromatolysis of somatic and autonomic lower motor neurons in the brain stem and spinal cord.

Comments: This is part of a "syndrome" or constellation of findings that includes both plexus and brain stem. The dysautonomia has been compared to similar lesions in horses, cats and hares. It is assumed that the causative toxin is present in hay and grass, causing the same lesion in horses, rabbits and hares. Animals affected also show impaction of the large intestine with dry food material.

Dilatation, Duct

Comments: In animals suffering from mucoid enteropathy, the ducts of the Brunner's glands in the duodenum become dilated, with cuboidal to low columnar acinar epithelium, rather than tall mucus-filled epithelial cells³².

Hyperplasia, Goblet Cell

Comments: Mucoid enteropathy is usually seen in young

animals, 2-3 months old, but can be seen in adults. In animals suffering from mucoid enteropathy there is an increase in the size and number of goblet cells in the intestinal epithelium, affecting duodenum, jejunum and ileum, but most apparent in sections of the ileum. Colonic crypts may be irregularly dilated due to mucous plugs. Goblet cell hyperplasia may also occur in the cecum due to cecal stasis and impaction, and in the hepatic bile ducts. Staining with Alcian blue-Periodic Acid Schiff indicates depletion of richly stained acidic colonic mucus and replacement with weakly staining light green-blue foamy mucus. Mucoid "enteritis" is a misnomer, for there is no hyperemia, congestion, local leukocytic response, necrosis, or fever, with goblet cell hyperplasia and a mucus hypersecretory state instead being the pathognomonic finding. Mucoid "enteropathy" is the correct term for this condition³². The etiology is unknown, but an enterotoxin-induced secretory diarrhea caused by Escherichia coli or Clostridium spiriforme is suspected. Antibiotic induced enterotoxemia can also be a factor, especially with lincomycin, clindamycin or erythromycin. The finding may be induced in toxicology studies assessing antibiotics.

Parasite (Figures 8, 9)

Comments: Parasites of the gastrointestinal system include nematodes, cestodes and protozoans, but are rarely a problem in well-managed barriered facilities. Protozoal parasites cause the most common and significant disease. Numerous species of *Eimeria* are capable of infecting rabbits: *Eimeria stiedai* (frequently referred to as *E. stiedae*), *E. magna, E irresidua* and *E. intestinalis*.

E. Anatomy of the Exocrine Pancreas

The pancreas of the rabbit is small and diffuse and located in a pocket formed by the transverse colon, the stomach and the duodenum. It may be difficult to locate in the abundant adipose tissue of the omentum, and retains only the accessory pancreatic duct, which enters the ascending duodenum distal to the entrance of the biliary duct. The right lobe of the pancreas is situated in the mesoduodenum of the duodenal loop. The left lobe lies between the stomach and transverse colon. There is a single pancreatic duct that opens at the junction of the transverse and ascending loops of the duodenum. The duct drains both pancreatic lobes. Technically, this is the accessory pancreatic duct as the main pancreatic duct connection to the duodenum disappears during embryonic development (Table 8).

Tissue, Ectopic

Comments: Ectopic spleen has been recorded in the pancreas of a NZW⁴¹.

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Ectopic tissue	Х			
Malformation			Х	
Non-proliferative				
Abscess		Х		
Amyloid		Х		
Apoptosis [‡]		Х		
Atrophy	Х			
Cyst	Х			
Degeneration		Х		
Degeneration/necrosis		Х		
Degeneration, neuron, myenteric plexus *	Х			
Dilatation, (+ locator) *	Х			
Edema		Х		
Erosion/ulcer	Х			
Hemorrhage		Х		
Hypertrophy, Paneth cell	Х			
Infarct		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]	Х			
Inflammation, mixed cell	Х			
Inflammation, mononuclear cell	Х			
Inflammation, vessel		Х		
Intussusception	Х			
Lymphangiectasis		x		
Metaplasia, squamous cell/Paneth cell/osseous	x			
Mineralization (+ locator)	X			
Necrosis mucosa	X			
Paneth cell decreased		x		
Parasite *		X		
Pigment		X		
Prolance		x		
Single cell pecrocis [‡]		X X		
Svincytia, enithelium	x	Л		
Vacualation mucosa	X			
Vacuolation, mucosa	X X			
Proliferativa Non noonlastia	Λ			
Homenal size *		V		
Hyperplasia				
Dualiforativa Nacarlastia		Λ		
			V	
Adenoma			<u></u> Х	
Adenocarcinoma			X	
Carcinoma, Brunner's glands			<u>X</u>	
Gastrointestinal stromal tumor (GIST), benign			<u>X</u>	
Gastrointestinal stromal tumor (GIST), malignant			<u>X</u>	
Leiomyoma			<u>X</u>	
Leiomyosarcoma			<u>X</u>	
Tumor, neuroendocrine cell, benign			Х	

Table 7. Microscopic Findings of the Small and Large Intestines: Rabbit

* Terminology with diagnostic criteria or comments described in the text. # Inducible lesion. + Refer to 4 for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Ectopic tissue *		Х		
Non-proliferative				
Abscess		Х		
Accumulation, adipocytes	Х			
Amyloid		Х		
Apoptosis ⁺		Х		
Atrophy	Х			
Autophagic vacuoles, acinar cell		Х		
Cyst	Х			
Degeneration/necrosis		Х		
Degranulation, (+ locator)		Х		
Dilatation, duct		Х		
Ectasia, duct		Х		
Edema		Х		
Focus, basophilic		Х		
Fibrosis		Х		
Halos, peri-insular, decreased		Х		
Halos, peri-insular, increased		Х		
Hemorrhage		Х		
Infarct		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]	Х			
Inflammation, mixed cell	Х			
Inflammation, mononuclear cell	Х			
Inflammation, vessel		Х		
Metaplasia, ductular		Х		
Metaplasia, hepatocyte	Х			
Mineralization (+ locator)	Х			
Necrosis		Х		
Pigment		Х		
Secretion, decreased		Х		
Single cell necrosis [‡]		Х		
Tissue, ectopic		Х		
Proliferative Non-neoplastic				
Hyperplasia #		Х		
Proliferative Neoplastic				
Adenoma, acinar cell			Х	
Adenoma, ductal cell			Х	
Adenocarcinoma, acinar cell			X	
Adenocarcinoma, ductal cell			X	

Table 8. Microscopic Findings of the Exocrine Pancreas: Rabbit

* Terminology with diagnostic criteria or comments described in the text. # Inducible lesion. + Refer to 4 for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

Section 5: Endocrine System: (Pituitary, Pineal, Thyroid, Parathyroid, Adrenal Glands and Endocrine Pancreas)

For detailed general considerations on the endocrine system, please refer to the rodent INHAND publication⁴².

The endocrine system of the rabbit is made up of the pituitary, pineal, thyroid, parathyroid, adrenal, and islets of Langerhans of the pancreas as well as parts of the male and female gonads and the epithelial lining of the duodenum⁴³.

A. Anatomy of the Pituitary Gland

In the rabbit the pituitary has three major divisions: lobus glandularis (adenohypophysis), lobus nervosus (neurohypophysis) and the infundibular stalk which attaches the pituitary to the median eminence of the hypothalamus^{43, 44}. Strong muscarinic receptor protein-like (mAChRp-L) immunoreactivity is associated with the blood vessels of the anterior and intermediate lobes of the rabbit pituitary⁴⁵. Sensitive and specific autoregulatory control systems for thyrotropin (TSH), luteinizing hormone (LH) and follicle stimulating hormone (FSH) exist in the rabbit pituitary⁴⁶ (Table 9).

B. Anatomy of the Pineal Gland

Pineal Gland

Calcareous concretions are common, which increase with age and apparently do not affect function of the gland. The rabbit pineal gland has an inhibitory effect on gonadotropin release⁴⁷ (Table 10).

Table 9. Microscopic Findings of the Pituitary Gland: Rabbit

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Aberrant craniopharyngeal structures		Х		
Aplasia/hypoplasia		Х		
Persistent Rathke's pouch			Х	
Non-proliferative				
Angiectasis		Х		
Apoptosis [‡]		Х		
Atrophy			Х	
Cyst		Х		
Fibrosis		Х		
Gliosis, pars nervosa			Х	
Hemorrhage		Х		
Hypertrophy, pars distalis		Х		
Hypertrophy, pars intermedia		Х		
Infarct		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Metaplasia, osseous		Х		
Necrosis		Х		
Pigment		Х		
Pseudocyst		Х		
Single cell necrosis [‡]		Х		
Thrombus		Х		
Vacuolation		Х		
Proliferative Non-neoplastic				
Hyperplasia, pars distalis/intermedia			Х	
Proliferative Neoplastic				
Adenoma, pars distalis/intermedia			Х	
Carcinoma, pars distalis/intermedia			Х	
Craniopharyngioma, benign			Х	
Pituicytoma, benign			Х	
Craniopharyngioma, malignant			Х	
Pituicytoma, malignant			Х	

⁺Refer to ⁴ for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

Finding	Common	Uncommon	Not Observed but potentially Relevant	Not Applicable
Congenital				
Aplasia/hypoplasia			Х	
Non-proliferative				
Amyloid		Х		
Angiectasis		Х		
Apoptosis ⁺		Х		
Cyst		Х		
Fibrosis		Х		
Fibers, striated muscle				Х
Hemorrhage		Х		
Infarct		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Mineralization		Х		
Necrosis		Х		
Pigment		Х		
Single cell necrosis ⁺		Х		
Striated muscle fibers		Х		
Thrombus		Х		
Vacuolation		Х		
Proliferative Non-neoplastic				
Hyperplasia		Х		
Proliferative Neoplastic				
Pinealoma, benign			Х	
Pinealoma, malignant			X	

 Table 10.
 Microscopic Findings of the Pineal Gland: Rabbit

Since the pineal gland is not routinely evaluated there is limited experience in incidences of these lesions in the rabbit. ⁺Refer to ⁴ for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

C. Anatomy of the Thyroid Gland

The two lobes of the thyroid gland in the rabbit are less clearly circumscribed compared to other laboratory species and are flattened, with the isthmus barely visible (Table 11).

D. Anatomy of the Parathyroid Gland

The paired parathyroid glands are usually located on the anterior and lateral aspect of the thyroid lobes, located in or immediately outside of the thyroid gland in the rabbit, and are separated from the thyroid by a thin capsule of fibrous connective tissue. The rabbit possesses two parathyroid glands within the thyroid and two located in the fascial plan between the sternohyoid and sternothyroid muscles and the carotid artery⁴⁸ (Table 12).

Tissue, Ectopic

Comments: Ectopic parathyroid tissue can occur in the thymus or dorsolateral to the esophagus near the larynx.

E. Anatomy of the Adrenal Gland

Adrenal Cortex and Medulla

In the rabbit, each of the pair of suprarenal adrenal glands is composed of an inner medulla and outer cortex. The adrenal cortex is voluminous in the rabbit and is derived from the interrenal gland associated with the mesonephros in lower vertebrates, which is involved in the maintenance of normal functioning kidneys43. The ultrastructure of the capsule of the rabbit adrenal gland is made up of three layers with the outermost layer consisting of collagen and elastic fibrillae with cytoplasmic processes of fibroblasts in between⁴⁹. Myofibroblasts are present in the middle layer as well as unmyelinated nerves, indicating a contractile function. The basal laminae of the fenestrated capillaries in the inner vascular layer is occasionally fused with that of the outer zona glomerulosa, suggesting a probable route for blood supply and secretion⁴⁹. The cortex is made up of three layers: the zona glomerulosa, zona fasciculata, zona reticularis⁴⁴. Fazekas and Sandor have demonstrated that there is an unusual pathway of aldosterone biosynthesis in the rabbit adrenal whereby aldosterone is formed mainly from corticosterone via 18-hydroxy-corticosterone⁵⁰. The adrenal medulla contains chromaffin cells and ganglion cells

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Aplasia/hypoplasia		Х		
Cyst, ultimobranchial		Х		
Duct, thyroglossal, persistent		Х		
Thyroid dysplasia			Х	
Non-proliferative				
Alteration, colloid		Х		
Amyloid		Х		
Angiectasis		Х		
Apoptosis [‡]				
Atrophy		Х		
Cyst		Х		
Fibrosis		Х		
Follicle, cystic		Х		
Hemorrhage		Х		
Hypertrophy, follicular cell		Х		
Infarct		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Mineralization		Х		
Necrosis		Х		
Pigment		Х		
Single cell necrosis ⁺		Х		
Tissue, ectopic	Х			
Thrombus		Х		
Vacuolation		Х		
Proliferative Non-neoplastic				
Hyperplasia, C-cell			Х	
Hyperplasia, follicular cell			Х	
Proliferative Neoplastic				
Adenoma, C-cell			Х	
Adenoma, follicular cell			Х	
Carcinoma, C-cell			Х	
Carcinoma, follicular cell			Х	

Table 11. Microscopic Findings of the Thyroid Gland: Rabbit

[#] Refer to ⁴ for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

arranged into trabeculae⁴⁴. The adrenal glands are the most commonly affected endocrine organs secondary to chemical exposure⁵¹. In the adrenal glands, chemically induced lesions are found most frequently in the zona fasciculata and reticularis and to a lesser extent in either the zona glomerulosa or the medulla.

Paraganglia

The paraganglia including the carotid and aortic bodies (made up of neuroendocrine cells) are also considered to be endocrine tissues of the rabbit⁴³ (Table 13).

F. Anatomy of the Endocrine Pancreas (Islets of Langerhans)

Extensive deep connections between the capillary beds of the islets and the exocrine tissue form a highly developed portal system in the rabbit which allows the islet hormones of insulin, glucagon and somatostatin to influence exocrine pancreatic cells⁵². Nearly all of the efferent islet blood flow goes to the acinar capillaries before leaving the pancreas. Thus, the flow to the islets is large enough to permit significant local actions of the islet hormones on the exocrine pancreas, confirming of the existence of an insuloacinar portal system⁵³ (Table 14).

Hyperplasia, Islet Cell

Pathogenesis/cell of origin: islet cells

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Aplasia/hypoplasia		Х		
Cyst, ultimobranchial		Х		
Duct, thyroglossal, persistent		Х		
Non-proliferative				
Amyloid		Х		
Angiectasis		Х		
Apoptosis [‡]				
Atrophy		Х		
Cyst		Х		
Fibrosis		Х		
Hemorrhage		Х		
Hypertrophy		Х		
Infarct		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Multinucleated giant cells			Х	
Necrosis		Х		
Pigment		Х		
Single cell necrosis [‡]		Х		
Tissue, ectopic *	Х			
Thrombus		Х		
Vacuolation		Х		
Proliferative Non-neoplastic				
Hyperplasia			Х	
Proliferative Neoplastic				
Adenoma			Х	
Carcinoma			X	

Table 12. Microscopic Findings of the Parathyroid Gland: Rabbit

* Terminology with diagnostic criteria or comments described in the text. + Refer to 4 for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

Diagnostic Features: Diffuse islet cell hyperplasia, characterized by increased numbers of otherwise unremarkable islet cells, resulting in variably increased cellularity and

overall diameter of islet profiles

Comments: may be seen as an induced treatment related effect by test articles that cause hyperglycemia.

Finding	Common	Uncommon	Not Observed but	Not Applicable
Congenital			Totentially Relevant	
Aplasia/hypoplasia		x		
Non-proliferative		A	-	
Amyloid		x		
Angiectasis		X		-
Apontosis [‡]		X		
Atrophy	Х			
Cyst		Х		
Degeneration. cvstic		Х		
Fibrosis		Х		
Hematopoiesis, extramedullary		Х		
Hemorrhage		Х		
Hypertrophy, cortical, diffuse/focal		Х		
Infarct		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Metaplasia, osseous		Х		
Mineralization		Х		
Necrosis		Х		
Persistent X-zone				Х
Pigment		Х		
Single cell necrosis [‡]		Х		
Tissue, ectopic		Х		
Thrombus		Х		
Vacuolation cortex decreased diffuse		Х		
Vacuolation cortex decreased focal		Х		
Vacuolation cortex increased diffuse		Х		
Vacuolation cortex increased focal		Х		
Proliferative Non-Neoplastic				
Hyperplasia, cortical/medullary			Х	
Hyperplasia, subcapsular cell				Х
Proliferative Neoplastic				
Adenoma, cortical cell			Х	
Adenoma, subcapsular cell				Х
Ganglioneuroma, benign			Х	
Myelolipoma			Х	
Pheochromocytoma, complex, benign			Х	
Pheochromocytoma, benign			Х	
Carcinoma, cortical cell			Х	
Carcinoma, subcapsular cell				Х
Neuroblastoma, malignant			Х	
Pheochromocytoma, complex, malignant (adrenal gland)			Х	
Pheochromocytoma, malignant			Х	

 Table 13.
 Microscopic Findings of the Adrenal Gland: Rabbit

⁺Refer to ⁴ for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Aplasia/hypoplasia		Х		
Non-proliferative				
Amyloid, islet		Х		
Angiectasis		Х		
Apoptosis, islet cell ⁺		Х		
Atrophy, islet cell		Х		
Cyst		Х		
Degranulation, islet cell		Х		
Fibrosis, islet		Х		
Hemorrhage, islet		Х		
Hypertrophy, islet cell		Х		
Infarct		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Metaplasia, hepatocyte			Х	
Necrosis		Х		
Pigment. islet		Х		
Single cell necrosis		Х		
Vacuolation, islet cell		Х		
Proliferative Non-neoplastic				
Hyperplasia, islet cell *#		Х		
Proliferative Neoplastic				
Adenoma, islet cell			Х	
Adenoma, acinar-islet cell			Х	
Carcinoma, islet cell			X	
Carcinoma, acinar-islet cell			X	

 Table 14.
 Microscopic Findings of the Endocrine Pancreas: Rabbit

 \ast Terminology with diagnostic criteria or comments described in the text. ${}^{\#}$ Inducible lesion.

⁺Refer to ⁴ for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

SECTION 6: HEMATOPOIETIC AND LYMPHOID SYSTEM

A. Anatomy of the Bone Marrow

During embryogenesis in rabbits, hematopoietic progenitors (Hematopoietic stem cells - HSCs) arise within several sites including the extraembryonic yolk sac and within the placenta. HSCs are also present in the fetal liver. Shortly before birth, HSCs and hematopoietic cells are both present in the bone marrow. Morphological studies by King and Ackerman conclusively indicated that erythrocytes develop extravascularly, arising from mesenchymal or reticular cells in the fetal bone marrow⁵⁴. Mature erythrocytes enter the circulation through discontinuities in the sinusoidal walls. Neither endothelial cells nor blood-borne lymphocytes make an apparent contribution to erythropoiesis. The first hematopoietic cells to form in the fetal marrow are determined and develop along the erythrocytic line. These proerythroblasts initially arise randomly in the marrow parenchyma and are not in obvious association with the sinusoids. Subsequent maturation and proliferation of the primitive erythrocytic cells result in the formation of colonies of erythrocytic cells at all stages of development. As these colonies enlarge, the erythrocytic elements come in close association with the sinusoids. In later stages of marrow development, developing erythrocytic and granulocytic cells become intermixed and more randomly associated in the extravascular space of the marrow. In rabbits, extramedullary hematopoiesis (EMH) occurs primarily in the spleen⁵⁵.

Adipose cells begin to develop at 2 weeks of age and proceed so that the adult pattern of red and yellow marrow is fully established by 4 months of age. Adipose cell development occurs in both trunk and limb bones; the magnitude of the process, however, being considerably greater in the limb bones. Adipocyte precursors may be present in the marrow at birth with a differential distribution in the areas of prospective red and yellow marrow. Thus, fatty involution of marrow appears to be a programmed developmental event⁵⁶.

Bone marrow is variably distributed within the medullary cavity of long and flat bones. Bone marrow for microscopic evaluation in rabbits is typically collected from the femur. Tissue is processed by standard techniques for hematoxylin and eosin stained formalin fixed paraffin embedded decalcified bone. Additionally, marrow casts may be collected from femoral bone marrow and processed for histology. A general guidance for histopathology assessment of bone marrow tissue sections is available ⁵⁷. Romanowsky stained bone marrow smears may be made for cytology. Rabbit neutrophils (heterophils) have intracytoplasmic granules that cause them to resemble eosinophils. True rabbit eosinophils have larger darker granules. Lymphocytes are the predominant leukocyte. Basophils are more common than in other mammals, making up 2–7% of the leucocyte population of the rabbit (Table 15).

Leukemia

Comments: Leukemia is reported sporadically in research rabbits^{58–61}.

Lymphoma

Comments: Lymphoma is the most common neoplasm of juvenile and young adult rabbits. It has been reported in rabbits as young as 4 months of age⁶² and appears to be more common overall in younger rabbits⁶³. Reports have included tumors of both B-cell⁶⁴ and T-cell origin^{61, 65}.

B. Anatomy of the Thymus

The rabbit thymus develops bilaterally from the endoderm of the third pharyngeal pouch and the surrounding mesenchyme. It is recognized as the pacemaker of lymphopoiesis in that if it fails to develop prenatally, then the immune system cannot be established⁶⁶. The rabbit thymus develops very late, at Embryonic Day (ED) 10 (about the third of gestation period). At ED29, the demarcation between the cortex and the medulla becomes easily distinct in all lobules. At this age, Hassall's corpuscles can be observed within the medulla. They are few in number, small in size and show different stages of their formation. Some Hassall's corpuscles are represented by collection of swollen epithelial cells, other corpuscles consist of few layers of concentrically arranged epithelial cells with centrally located keratin substance. At 1 and 2 weeks postnatally, the Hassall's corpuscles increase in size and number to be large, acidophilic, rounded bodies consisting of a central degenerated hyaline mass surrounded by concentrically arranged epithelialreticular cells⁶⁷. They are unique to the thymus⁶⁸. The Hassall's bodies are structurally organized from medullary reticuloepithelial cells, which usually undergo hypertrophy prior to their inclusion in the outer cell layer of the corpuscles69.

In rabbits, the thymus has two parts; thoracic and cervical. The rabbit thymus acquires the lobulated appearance at ED14. In the thorax, the thymus is separated into three lobes: the right dorsal thoracic lobe, the right ventral thoracic lobe, and the left thoracic lobe70. Grossly, the shape of the left lobe of the thymus is quadrilateral in outline with extended narrow craniomedial angle in the neck, while the right lobe is triangular in outline with its base cranially directed and extended narrow craniomedial angle in the neck. The dorsal aspect of both lobes is concave showing the cardiac impression which is larger on the left lobe. In addition, the right lobe shows a pulmonary impression laterally. Medially, the left lobe slightly overlaps the right one and both lobes are connected only by small amount of interlobar connective tissue. The ventral aspect is convex and related to the sternum. While the thymus remains relatively large in adult rabbits13, with increasing age the thymus undergoes a proportional decrease in both cortical and medullary size71 (Table 16).

C. Anatomy of the Spleen

There are no significant variations macroscopic structures or microarchitecture between the rabbit and rodent spleens⁷². Duplication of the spleen has been observed sporadically⁷³ (Rinke, pers. observation) (Table 17).

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Agenesis/hypoplasia			Х	
Non-proliferative				
Abscess		Х		
Amyloid		Х		
Apoptosis [‡] , increased, [insert appropriate cell type]		Х		
Angiectasis		Х		
Atrophy, serous, of fat #		Х		
Cellularity, decreased, adipocyte		Х		
Cellularity, decreased, bone marrow		Х		
Dyshematopoiesis		Х		
Fibrosis		Х		
Hypersegmentation, granulocyte		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Metaplasia, osseous		Х		
Mineralization		Х		
Necrosis, lymphocyte		Х		
Pigment, macrophage		Х		
Pigment		Х		
Serous atrophy of fat		Х		
Single cell necrosis ⁺		Х		
Proliferative Non-neoplastic				
Cellularity, increased, [insert appropriate cell type]		Х		
Proliferative Neoplastic				
Eosinophil Granulocytic Sarcoma			Х	
Histiocytic sarcoma			Х	
Leukemia, erythroid/myeloid/megakaryocytic/mast cell/NOS *		Х		
Lymphoma*		Х		
Tumor, mast cell, benign			Х	
Tumor, mast cell, malignant			Х	

 Table 15.
 Microscopic Findings of the Bone Marrow: Rabbit

* Terminology with diagnostic criteria or comments described in the text. # Inducible lesion. * Refer to 4 for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

D. Anatomy of the Lymph Nodes

Lymph nodes are present at birth and develop upon antigenic stimulation. Knowledge about the area(s) drained by the lymph nodes is needed to interpret their morphology^{74–77}. The mesenteric lymph nodes form a compact unit overlying the left side of the superior mesenteric artery¹³. Other intra-abdominal nodes include those near the posterior end of the mesoduodenum, adjacent to the portal vein near the lesser curvature of the stomach, and near the junction of the splenic and superior mesenteric veins¹³. There are no species-specific terms for lymph nodes in rabbits (Table 18).

E. Anatomy of the Mucosa-Associated Lymphoid Tissue (MALT)

The mucosal associated lymphoid tissue (MALT) is organized into congenital and acquired organized submucosal lymphoid accumulations associated with the mucosal epithelium. This can include loose clusters of lymphocytes or wellorganized lymphoid follicles. Gut-associated, nasal-associated MALT and tonsils are congenital and lymphoid aggregates in other tissues are acquired with antigenic exposure (e.g. conjunctiva, oral, pharyngeal and nasal cavities, upper (larynx and trachea) and lower (bronchus/bronchial) respiratory tract, Eustachian tube, middle ear, stomach, gall bladder, cecum, colon, rectum, reproductive and urinary tracts). The ileocecal MALT is located at the ileocecal junction as an enlargement of the large intestine. The ileocecal MALT account for 50% of the total lymphoid tissue of the rabbit and are the reason for the small size of the spleen³⁰. For detailed descriptions of the vermiform appendix and sacculus rotundus, please refer to the intestinal tract section.

The gut associated lymphoid tissue (GALT) should be examined in studies where the test article is administered orally, and is usually examined in a routine section of jejunum or il-

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Agenesis/hypoplasia			Х	
Ectopic tissue		Х		
Non-proliferative				
Abscess		Х		
Amyloid		Х		
Apoptosis [‡] , increased, lymphocyte		Х		
Cellularity, decreased, lymphocyte		Х		
Corticomedullary distinction, loss of		Х		
Corticomedullary ratio, decreased		Х		
Corticomedullary ratio, increased		Х		
Cyst, epithelial		Х		
Epithelial cell free zones, increased		Х		
Hypoplasia		Х		
Involution, age-related	Х			
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Metaplasia, osseous		Х		
Mineralization		Х		
Necrosis		Х		
Pigment, macrophage		Х		
Pigment		Х		
Single cell necrosis [‡]		Х	-	
Tissue, ectopic (specify tissue)		Х		
Tissue, ectopic, parathyroid		Х		
Tissue, ectopic, thyroid		Х		
Thymic corpuscles, increased		Х		
Tingible body macrophage, increased		Х		
Vacuolation, macrophage		Х		
Proliferative Non-neoplastic				
Cellularity, increased, [insert appropriate cell type]		Х		
Proliferative Neoplastic				
Histiocytic sarcoma			Х	
Lymphoma		Х		
Thymoma, benign			Х	
Thymoma, malignant			Х	

Table 16. Microscopic Findings of the Thymus: Rabbit

⁺ Refer to ⁴ for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

eum that contains Peyer's Patches, however one of the other MALTs - sacculus rotundus (ileocecal tonsils), vermiform appendix, colonic and rectal lymphoid aggregates, cryptopatches (CPs) and isolated lymphoid follicles (ILFs) - would suffice.

Rabbits develop a primary antibody repertoire through somatic diversification of Ig genes (dependent on intestinal microbial flora). Rabbits generate their antibody repertoire in three stages: (a) neonatal repertoire is generated by B lymphopoiesis in fetal liver and bone marrow (limited by preferential V(H) gene segment usage); (b) between 4 and 8 weeks after birth, gut-associated lymphoid tissue (GALT) develops a complex primary antibody repertoire, (c) the primary antibody repertoire is subsequently modified during antigen-dependent immune responses (the secondary repertoire)78.

Most MALT is present at birth (gut, nasal and tonsils), and is acquired in other sites thereafter⁷⁹. Age-related functional decline of the mucosal immune response and such age-related involution is not described below as a separate item; comparison with concurrent controls is needed to decide whether or not age-related changes have occurred in a study (Table 19).

Cellularity, Decreased, Lymphocyte

Comments: Rabbits are especially sensitive to lymphoid atrophy caused by glucocorticoid administration.

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Agenesis/hypoplasia			Х	
Tissue, ectopic			Х	
Non-proliferative				
Abscess		Х		
Amyloid		Х		
Angiectasis		Х		
Apoptosis ⁺ , increased, lymphocyte		Х		
Cellularity, decreased, white pulp/red pulp		Х		
Congestion	Х			
Erythrophagocytosis		Х		
Fibrosis		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Mineralization		Х		
Necrosis, lymphocyte		Х		
Pigment, macrophage		Х		
Pigment		Х		
Single cell necrosis ⁺		Х		
Tingible body macrophages, increased		Х		
Vacuolation, macrophage		Х		
Proliferative Non-neoplastic				
Accumulation, adipocyte		Х		
Aggregates, macrophage, increased		Х		
Cellularity, increased, [insert appropriate cell type]		Х		
Extramedullary hematopoiesis, increased		Х		
Hyperplasia, nodular		Х		
Hyperplasia, mast cell			Х	
Proliferative Neoplastic				
Eosinophil Granulocytic Sarcoma			Х	
Histiocytic sarcoma			Х	
Leukemia, erythroid/myeloid/megakaryocytic/mast cell/ NOS		Х		
Lymphoma		X		

 Table 17.
 Microscopic Findings of the Spleen: Rabbit

⁺Refer to ⁴ for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Aplasia/hypoplasia			Х	
Non-proliferative				
Abscess		Х		
Amyloid		Х		
Apoptosis [‡] , increased, lymphoid		Х		
Cellularity, decreased, lymphocyte		Х		-
Dilatation, sinus		Х		
Erythrocytes, intrasinusoidal		Х		
Extramedullary hematopoiesis		Х		
Fibrosis, (+ locator)		Х		-
Granuloma		Х		
Hypertrophy/hyperplasia, high endothelial venule (HEV)		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Lymphangiectasis		Х		
Metaplasia, osseous		Х		
Mineralization		Х		
Necrosis		Х		
Pigment, macrophage		Х		
Pigment		Х		-
Single cell necrosis [‡]		Х		
Tingible body macrophages increased		Х		
Vacuolation, macrophage		Х		
Proliferative Non-neoplastic				
Accumulation, adipocyte		Х		
Aggregates, increased, macrophage		Х		
Cellularity, increased, (+ cell type)		Х		
Extramedullary hematopoiesis, increased		Х		
Hyperplasia, angiomatous			Х	
Proliferative Neoplastic				
Eosinophil Granulocytic Sarcoma			Х	
Histiocytic sarcoma			Х	-
Leukemia, erythroid/myeloid/megakaryocytic/mast cell/NOS		Х		
Lymphoma		Х		
Tumor, mast cell, benign			Х	
Tumor, mast cell, malignant			Х	

Table 18.	Microscopic Findings of the Lymph Nodes: Rabbit

⁺Refer to ⁴ for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Aplasia/hypoplasia			Х	
Non-proliferative				
Abscess		Х		
Amyloid		Х		
Apoptosis [‡] , lymphoid, increased		Х		
Cellularity, decreased, lymphocyte *#		Х		
Degeneration, follicle associated epithelium		Х		
Hyaline material		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Lymphangiectasis		Х		
Material, hyaline		Х		
Mineralization		Х		
Necrosis, (+ cell type)		Х		
Pigment, macrophage		Х		
Pigment		Х		
Single cell necrosis [‡]		Х		
Tingible body macrophages, increased		Х		
Vacuolation, macrophage		Х		
Proliferative Non-neoplastic				
Aggregates, macrophage, increased		Х		
Cellularity, increased, (+ cell type)		Х		
Extramedullary hematopoiesis		Х		
Hyperplasia, follicle-associated epithelium		Х		
Hypertrophy/hyperplasia, high endothelial venule (HEV)		Х		
Metaplasia, squamous, follicle-associated epithelium		Х		
Proliferative Neoplastic				
Eosinophil Granulocytic Sarcoma			Х	
Histiocytic sarcoma			Х	
Leukemia, erythroid/myeloid/megakaryocytic/mast cell/NOS		X		
Lymphoma		X		

 Table 19.
 Microscopic Findings of the MALT: Rabbit

* Terminology with diagnostic criteria or comments described in the text. # Inducible lesion. * Refer to 4 for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

Section 7: Hepatobiliary System (Liver and Gall Bladder)

A. Anatomy of the Liver

The liver is situated in the epigastric region reaching the level of the right 7th and left 9th ribs. The right hepatic, caudate and quadrate lobes are single, and the left lobe is divided into medial and lateral parts⁸⁰. The caudate process of the caudate lobe is highly developed and overlies the right kidney (Table 20).

Cytoplasmic Alteration (Figure 12)

Other terms: Rarefaction, Glycogen accumulation.

Historical terms: Granular degeneration, hyaline degeneration, ground glass change.

Diagnostic features: Rabbit livers are often pale at necropsy. This cytoplasmic rarefaction is associated with glycogen accumulation and varies both between the sexes and diurnally.

Comments: Generally, rabbits not fasted before euthanasia show high levels of glycogen accumulation. This decreases during the day, as it is usual practice to remove access to food on the day of necropsy. Therefore, animals euthanized in the morning will have more glycogen in the liver than those euthanized in the afternoon. The Study Pathologist should be aware of this diurnal change and interpret any changes in glycogen accumulation with knowledge of whether the animal was fasted prior to euthanasia, for how long and at what time of day the necropsy occurred⁸¹. Electron microscopy or special stains are needed for a definitive diagnosis to distinguish between lipidosis (macrovesicular and/or microvesicular steatosis), phospholipidosis, and the gestational lipidosis seen in pregnant animals.

Infarct

Comments: Excess pressure from occlusive dressings, wrapping, sleeves or jackets used to enhance exposure to dermal therapeutics or to protect wound dressings and medical devices has been found to induce necrosis ("corset liver") in the liver⁸² and regional infarction in the liver and spleen (Schuh, unpublished data) of rabbits.

Inflammation (Figure 13)

Comments: An autoimmune hepatitis, similar to that seen in man, has been reported in a 5-year-old rabbit. Portal areas were characterized by a marked infiltrate of plasma cells, and some lymphocytes that extended into the surrounding parenchyma with destruction of the limiting plate architecture. In some areas the lymphocytic infiltrate was more extensive and surrounded the bile duct. There were many macrophages with engulfed pigment and apoptotic bodies. Periportal hepatocytes were swollen with karyomegaly and prominent nucleoli⁸³.

Mineralization

Comments: Slight or advanced dystrophic mineralization of the liver may occur due to a calcium and Vitamin D imbalance, for example in cases of over-supplementation. Usually the change occurs first in the aortic arch and heart before it reaches other organs¹¹.

Necrosis

Comments: Multifocal hepatocellular necrosis may occur as a stress-related disease in young (6–12 weeks-old) animals caused by Gram-negative Clostridium (Bacillus) piliformis (Tyzzer's disease). Clinical symptoms are diarrhea, dehydration and death within 12-48 h. Bacterial detection is mainly difficult due to autolysis and Escherichia coli overgrowth; microbes can be found in living cells only (Periodic Acid Schiff, Giemsa, Warthin-Starry staining). Hepatic copper toxicosis is seen occasionally in laboratory rabbits that are fed a copper-rich diet, leading to hepatocellular copper storage. Animals that then undergo a stressful event (often coupled with anorexia) may acutely release the copper from the hepatocytes. Affected livers have an accentuated lobular pattern macroscopically and centrilobular to midzonal hepatocellular necrosis with mild periportal fibrosis and biliary hyperplasia⁸⁴.

Parasite

Comments: Parasites include nematodes, cestodes and protozoans, but are rarely a problem in well-managed barriered facilities. Protozoal parasites cause the most common and significant disease. Numerous species of *Eimeria* are capable of infecting rabbits (Figures 14, 15). Hepatic lesions caused by the microsporidian *Encephalitozoon cuniculi* are recognized in rabbits (Figure 16). Hepatic lesions often include granulomatous foci along with periportal infiltration by macrophages, multinucleated giant cells, lymphocytes, and plasma cells. Inflammation can extend into the hepatic portal veins and branches of the hepatic artery². Parasite presence can be confirmed by real-time PCR⁸⁵.

Pigment (Figure 17)

Comment: This term should be used for material within hepatocytes or Kupffer cells only. Pigment in canaliculi should be termed 'Plug, bile'. Periportal and centrilobular storage of iron positive pigment has been observed, but rarely. The pigment deposition may be restricted to single lobes, but also within an entire lobe the distribution may vary significantly. Pseudomelanosis has been described previously⁷³. The livers are grossly discolored from dark brown to black. Histologically, intracytoplasmic pigment granules are observed that contain iron and lipofuscins. This alteration is not known to be associated with lesions in other organs.

Vacuolation, Cytoplasm

Other terms(s): Gestational lipidosis

Comments: May be seen in rabbits on reproductive toxicity

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Non-proliferative				
Abscess		Х		
Amyloid		Х		
Angiectasis		Х		
Apoptosis [‡]		Х		
Atrophy, hepatocyte		Х		
Cholangiofibrosis		Х		
Congestion	Х			
Crystals		Х		
Cyst, (+ locator)	Х			
Cytoplasmic alteration *	Х			
Degeneration, cystic		Х		
Degeneration, hydropic	Х			
Erythrocytes, intrahepatocellular		Х		
Extramedullary hematopoiesis	Х			
Fatty change	Х			
Fibrosis		Х		
Focus of cellular alteration				Х
Hemorrhage		Х		
Hepatocytes, intravascular		X		
Hepatodiaphragmatic nodule				Х
Hypertrophy, hepatocyte/endothelial cell		X		
Hypertrophy/hyperplasia, Kupffer cell		X		
Hypertrophy/karyomegaly, endothelial cell		X		
Inclusions, intranuclear and cytoplasmic		X		
Infarct *		X		
Infiltrate inflammatory cell [insert appropriate cell type]	x			
Inflammation *	71	x		
Intrahenatocellular erythrocytes		X		
Intravascular henatocytes		1	X	
Karvocytomegaly and/or multinucleated henatocytes		x	1	
Metaplasia glandular		X		
Metaplasia, gundului Metaplasia, pancreatic acinar cell		X		
Mineralization *		X	-	
Necrosis focal/multifocal *		X X		
Necrosis, rocal		X X		
Darasite *^		X		
Phospholipidosis		Λ	v	-
Digment * henotocyte/Kunffer cell		v	Λ	
Dlug bilo		X V		
Single cell perrorie t		<u></u> У		
Thrombus				
Tingue estaria		<u></u> 		
Verseletion extendence *		<u></u> 		
		Λ		
Proliferative Non-Neoplastic		v		
Hyperplasia, bile duct		<u> </u>		
Hyperplasia, angiomatous		<u>X</u>		
Hyperplasia, hepatocyte, non-regenerative		<u>X</u>		
Hyperplasia, hepatocyte, regenerative		X		
Hyperplasia, Ito cell			<u> </u>	
Hyperplasia, oval cell			X	
Proliferative Neoplastic				
Adenoma, hepatocyte		-	X	-
Adenoma, hepatocholangiocellular			X	
Carcinoma, hepatocyte			Х	
Carcinoma, hepatocholangiocellular			X	
Hemangioma			Х	
Hemangiosarcoma			Х	
Hepatoblastoma			Х	
Histiocytic sarcoma			Х	
Tumor. Ito cell, benign			х	

 Table 20.
 Microscopic Findings of the Liver: Rabbit

* Terminology with diagnostic criteria or comments described in the text. ^ Described in systemic pathology section. ⁺Refer to ⁴ for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

studies. Pregnancy toxemia is a metabolic disease primarily during late pregnancy (less frequently in postpartum, pseudopregnant, and nonpregnant females), characterized by low morbidity and high mortality, and exacerbated by an inability to consume adequate energy to match metabolic demands. Risk factors include: obesity, improper nutrition such as a diet too low in fiber, sudden stress, and anorexia. Fat deposits become rapidly mobilized, resulting in hepatic lipidosis and ketosis.

B. Anatomy of the Gall Bladder

A gall bladder is present. The gall bladder has a cylindrical shape and does not reach the livers ventral edge. The bile duct enters the duodenum very close to the pylorus. Rabbits produce very large amounts of bile, about seven times as much as a dog on a weight basis³⁶. Approximately 20–50% of rabbits produce atropinase in the bile³⁰. Thus, atropine is not recommended during anaesthesia (Table 21).

Malformation

Comments: Bifurcate gall bladder or duplicated gall bladder is occasionally seen in rabbits⁸⁶. It is clinically silent and an incidental finding at necropsy.

Calculus

Comments: Rabbits fed a diet containing 0.75% dihydrocholesterol for 7 days develop bile acid allodeoxycholic

Finding	Common	Uncommon	Not Observed but	Not Applicable
Congenital			Totentiany Relevant	
Cvst			х	
Malformation *		X		
Non-proliferative				
Abscess		X		
Amyloid		Х		
Apoptosis [‡]		Х		
Calculus *		Х		
Congestion	Х			
Crystals		Х		
Fibrosis		Х		
Hemorrhage		Х		
Hyalinosis		Х		
Inclusions, intranuclear and cytoplasmic		X		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation *		Х		
Metaplasia		Х		
Mineralization		Х		
Necrosis, focal/multifocal		X		
Necrosis, single cell		Х		
Parasite *		Х		
Pigment		Х		
Thrombus		Х		
Tissue, ectopic, pancreas		Х		
Single cell necrosis [‡]		Х		
Vacuolation, macrophage		Х		
Proliferative Non-Neoplastic				
Hyperplasia		Х		
Metaplasia, glandular		Х		
Proliferative Neoplastic				
Adenoma			Х	
Adenocarcinoma			X	
Cholangioma			Х	
Cholangiocarcinoma			X	

 Table 21.
 Microscopic Findings of the Gall Bladder: Rabbit

* Terminology with diagnostic criteria or comments described in the text. * Refer to ⁴ for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

(ADCA) and deoxycholic acid (DCA) stones in the gall bladder. In this model, inflammatory changes in the gall bladder mucosa are often observed even before stones are formed. Within 3 days of the lithogenic diet, abnormalities of platelet function are detectable. Platelet aggregation upon addition of adenosine diphosphate (ADP) is impaired. At the same time the red cells become crenated and develop thorny spicules (echinocytes). This morphological change is associated with intracellular dehydration and excessive loss of potassium. These changes coincide with a rise in serum ADCA and DCA and precede a slow rise in serum cholesterol. *In vitro* incubation studies also suggest that the bile acids probably caused membrane injury to the platelets and red cells. It is concluded that changes in the bile ADCA and DCA probably induce gall bladder epithelial injury in this model of experimental cholelithiasis⁸⁷.

Inflammation

Comments: Rabbits have well defined mucosa associated

lymphoid tissue in the gall bladder and this should not be confused with inflammatory changes.

Parasite

Comments: Parasites are rarely a problem in well-managed barriered facilities. Protozoal parasites cause the most common and significant disease. *Eimeria stiedai (stiedae)* primarily damages the biliary system. After oral uptake of sporulated oocysts, the sporozoites penetrate the intestinal wall, reach the liver via blood vessels, and enter the epithelial cells of the bile ducts. The prepatent period is 15–18 days. Severe infections may produce abscessation, and bile duct swelling macroscopically, occasionally with hepatomegaly. Microscopically, multifocal granulomatous lesions surround bile ducts and often there is marked papillary ductule epithelial hyperplasia with intraluminal schizonts and zygotes⁸¹. Cured hepatic coccidiosis may be seen as areas of peribiliary fibrosis.

SECTION 8: SKIN

Introduction

Rabbit skin is commonly used for *in vivo* and *in vitro* investigation and testing of many chemical agents, ranging from cosmetics to carcinogens^{88–90}. The pathology of both spontaneous and induced conditions of the skin are similar in rabbits and humans, hence their use in vaccine studies. Careful recording of the nature, intensity and duration of the inflammatory response of the skin adjacent to implanted or injected substances is important in the assessment of the local tolerability of agents intended for contact with human tissues. The chemical and physical properties of injected chemicals or vaccines and their adjuvants as well as size, shape and surface texture of implanted biomaterials may modify the histological features and temporal pattern of the inflammatory and reparative responses^{91–94}. Such studies in rabbits are conducted with descriptors following the ISO-10993 guidelines.

A. Anatomy

Rabbits are born hairless. The adult rabbit body is well furred except for a few sites which are covered with sparse or no hair: around external nares; mammary gland nipples; scrotum; inguinal folds; pinnae^{12, 95}. Foot pads are absent, though the palmar and planter surfaces are thickly furred²⁵.

Mature rabbits have a large, fat-filled skin fold on the ventral surface of the neck called the dewlap^{24, 25}. The dewlap is particularly well developed in older females. The upper lip is divided by the philtrum, a distinct cleft, confluent with the external nares, and the lips have multiple prominent vibrissae.

Normal rabbit fur is composed of three hair types, produced by different follicles^{88, 96} that are arranged in clusters. A typical cluster consists of a single large central primary follicle (producing 50–60 μ m diameter, 3–4 cm long guard hairs) surrounded by 2–4 somewhat smaller lateral primary follicles (which produce 25–30 μ m diameter, 3.0–3.5 cm long awn hairs) and 20–50 much smaller secondary follicles (producing 15 μ m diameter, 2.5–3.0 cm long down hairs). Down hairs are the most abundant type (90–95%) and form the dense "inner coat", while the coarser guard and awn hairs constitute the protective "outer coat"^{88, 96}.

The large, elongated pinnae (external ears) can represent up to 12% of the rabbit body surface area²⁴. The pinna possesses a large central artery as well as marginal (lateral) veins that are frequently used for blood collection and intravenous injections^{24, 97}. The density of hair follicles in the pinna (80/cm²) is much lower than in many other body sites⁹⁰. Due to the large surface area, well-developed vascularization (arteriovenous anastomoses), and relatively sparse hair coat, the pinna dissipates body heat well, and plays a major role in overall thermoregulation in rabbits^{98, 99}. The epidermis of both pinna and body skin is thin^{90, 100}. Rabbit pinna epidermis is about 16–18 µm thick, compared to about 58–64 µm thickness of pig ear skin⁹⁰.

Abundant dermal sebaceous glands are associated with hair follicles in pinna and body skin, but sudoriferous (sweat) glands appear to be absent or vestigial in rabbit skin^{95, 100}. The rabbit is classified as a functionally "non-sweating" species⁹⁸.

Several specialized glands in or around the skin of rabbits produce externally released odiferous secretions that are important in social and sexual behavior modulation^{101–103}. These include the submandibular (chin or mental) skin gland; the inguinal gland complex (brown and white glands); and the anal (rectal, perirectal) glands. These glands have also been collectively or singularly referred to as "apocrine" glands¹⁰⁴, though the actual mode of secretion remains unclear^{105, 106}. As a result of this overlapping and sometimes contradictory terminology, when reviewing the literature, it is important to determine which gland(s) are being discussed, especially in older references.

The submandibular (mental) skin gland (Figures 18, 19) is located in the subcutis of the ventral mandible^{102, 104}. Submandibular gland secretions are thought to function primarily as territorial markers. Rabbits distribute the secretions by rubbing their chins against inanimate objects and even other rabbits; this "chinning" behavior is more common in males^{101, 103}. The submandibular gland is unpaired but multilobed, with a large central and two smaller lateral lobes; each lobe has an excretory duct opening onto the external skin¹⁰². The epithelium is cuboidal to low columnar, with epithelium of intact, sexually mature males being taller and more vacuolated than that of mature females^{102, 105}.

The submandibular skin gland is sexually dimorphic, with development function, and morphology strongly dependent on sex hormones. Intact males have much larger and heavier submandibular glands, with larger acini lined by taller epithelial cells^{102, 104, 107}. In males, the oily submandibular secretions may coat the skin of the chin. In females, the gland is much less developed and may be difficult to find and sample in young animals. Gonadectomy of males reduces the size, weight, and secretory epithelium height of submandibular glands, while testosterone administered to castrated males reverses these effects^{102, 107}. In females, ovariectomy has opposite effects, resulting in increased gland size and weight, as well as increased acinar diameter and epithelial height^{102, 107}.

Rabbits also have specialized glands in the nictitating membrane (see Special Senses section).

The paired inguinal gland complexes are located on either side of the penis or clitoris, and thus are sometimes referred to as "preputial" and "clitoral" glands (Figure 20) (see also Male Reproductive System and Female Reproductive System sections). Each gland complex is composed of two adjacent but morphologically different lobulated glands: the dorsolateral "white" gland and the medioventral "brown" gland. The white gland has typical sebaceous histomorphology, (Figure 21) while the brown gland is composed of branching tubules lined by simple cuboidal to columnar epithelium¹⁰⁶ (Figure 22).

Although closely apposed, the white and brown glands are separate structures with separate ducts (lined by keratinized stratified squamous epithelium) which open into the hairless skin fold at the base of the penis and clitoris.

The strong-smelling combined inguinal gland secretion is considered to function in individual identification¹⁰³. Compared to the submandibular skin gland, the brown and white inguinal glands respond similarly to gonadectomy and sex hormone administration, but the extent of response (sensitivity) is overall less pronounced^{104, 105}.

The inguinal glands (preputial and clitoral) are further discussed in the Male Reproductive System and Female Reproductive System sections, respectively.

Anal (or perirectal) glands, the third type of odiferous gland in rabbits, produce secretions considered important in territorial marking¹⁰³ (Figure 23). These glands are located in the anorectal submucosa (see also Peri-rectal tissue subsection in Digestive section).

B. Common Diseases of the Rabbit Skin

Based on its direct interaction with the environment, the skin is subject to many spontaneous and housing-related diseases. Morphological lesions should be described using the nomenclature listed below, however spontaneous diseases of the skin should be diagnosed in combination with clinical data. In a pathology report, the disease terms listed below could be used as "syndromes" to summarize and interpret morphological lesions.

Acariasis: Mite infestations of the skin can occur in laboratory rabbits^{24, 108–110}. Ear mite (*Psoroptes cuniculi*) infestations can result in prominent epidermal hyperkeratosis and dermal inflammation of the external ear canal and pinna, often exacerbated by secondary bacterial and fungal infections. Fur mites (*Cheyletiella parasitovorax*, other *Cheyletiella spp., Leporacarus gibbus;* and other species), and mange mites such as *Sarcoptes scabei* (sarcoptic mange; scabies), and *Notoedres cati* (notedric mange) can also cause skin lesions in various sites, including alopecia, epidermal hyperkeratosis and ulceration, and dermal inflammation^{24, 108, 109}.

Aural hematoma: Hematoma of the pinna can occur due to various causes¹¹¹.

Ulcerative pododermatitis in rabbits: Also known as "sore hocks", ulcerative pododermatitis most commonly affects the plantar aspects of the tarsus and metatarsus. Inflammation (often with bacterial infection) is considered a sequel to a primary inciting cause of pressure necrosis of the skin (due to obesity, wire-bottomed cages, general lack of sanitation etc.) and is seldom observed with current husbandry standards ^{24, 109, 110}.

Moist dermatitis: So-called moist dermatitis of the facial and neck (dewlap) skin can result from trapped moisture in large dewlaps or underlying dental disease with hypersalivation, often complicated by secondary bacterial infection^{109, 112, 113}. Similar inflammatory changes can occur in other locations such as the perineal skin.

Virus-related skin and subcutaneous proliferative lesions of

rabbits (such as those induced by myxoma virus, Shope fibroma virus, and various papillomaviruses) are beyond the scope of this paper, and the reader is referred to recent reviews^{114, 115}.

C. Dermatotoxicology

Testing of topically applied chemicals for acute dermal irritation/corrosion is typically performed in rabbits. However, as outlined in the supplement to the Organisation for Economic Co-operation and Development (OECD) test guideline 404 on dermal irritation/corrosion testing, consideration of existing data, structure activity relationships, physiochemical properties of the test item and testing in validated *in vitro* and ex vivo systems are recommended, before an *in vivo* study is conducted (Table 22).

Hamartoma, Collagenous

Comments: Collagenous hamartomas in the skin of rabbits are benign, solitary, poorly demarcated dermal nodules composed of dense collagen with interspersed fibrocytes^{116, 117}.

Inflammation

Comments: Generalized "exfoliative dermatitis" or "sebaceous adenitis" characterized by sebaceous gland atrophy, perifollicular, dermal, and/or epidermal lymphocytic infiltrates, and hyperkeratosis has been reported in rabbits^{65, 83, 110, 118–120}. An association with concurrent thymoma or thymic lymphoma was noted in several cases^{110, 118, 120}. Rabbits have been noted to be more sensitive to petroleum jelly-induced dermal irritation than rodents and minipigs, while rabbits and rats are more sensitive than minipigs to a topical antibiotic applied to abraded skin¹²¹. Conversely, rabbits exhibited decreased subcutaneous inflammation and biodegradation of polyurethane implants compared to rodents¹²².

Xanthomatous Alteration

Comments: Xanthoma or xanthomatous alteration refers to non-neoplastic accumulations of lipid-laden macrophages ("foam cells") in the dermis. Due to their altered metabolism and genetic background, Watanabe heritable hyperlipidemic (WHHL) rabbits are prone to such xanthomatous lesions. In WHHL rabbits, xanthomatous alteration (xanthomas) most commonly occur in plantar tendons of the distal limbs and digits with occasional extension into adjacent subcutis or dermis^{123–125}. Dermal xanthomatous alteration has also been reported in the hereditary high triglyceridemia (TGH) rabbit, an inbred rabbit strain which like the WHHL rabbit, has high blood cholesterol and lipids and a propensity to develop atherosclerosis¹²⁶. It can be induced also by feeding high-fat diets to NZW rabbits²⁸.

Hyperplasia, Keratinocyte

Other terms: actinic keratosis

Pathogenesis/cell of origin: keratinocyte

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Non-proliferative				
Abscess		Х		
Accumulation, macrophage		Х		
Adnexal dysplasia		Х		
Amyloid		Х		
Apoptosis [‡]		Х		
Atrophy, epidermis/dermis/adnexa		Х		
Congestion		Х		
Cyst, squamous		Х		
Degeneration		Х		
Dilatation		Х		
Edema, epidermis, intracellular/intercellular	Х			
Elastosis		Х		
Erosion/ulcer		Х		
Fibrosis	Х			
Hamartoma, collagenous *		Х		
Hemorrhage		Х		
Hyperkeratosis, epidermis/adnexa		Х		
Infiltrate, inflammatory cell, [insert appropriate cell type]		Х		
Inflammation, (+ locator) *		Х		
Mineralization	Х			
Necrosis, (+ locator)	Х			
Pigment	Х			
Pustule		Х		
Single cell necrosis [‡]		Х		
Thrombus	Х			
Vesicle		Х		
Xanthomatous alteration *#		Х		
Proliferative Non-neoplastic				
Hyperplasia, epidermis/adnexa/melanocyte		Х		
Hyperplasia, keratinocyte *		Х		
Proliferative Neoplastic				
Adenoma, sebaceous cell			Х	
Fibroma			Х	
Keratoacanthoma			Х	
Melanoma, benign			Х	
Papilloma, squamous cell			Х	
Tumor, basal cell, benign			Х	
Tumor, hair follicle, benign			Х	
Tumor, mixed, benign			Х	
Adenocarcinoma			Х	
Carcinoma, basal cell			Х	
Carcinoma, eccrine gland			Х	
Carcinoma, sebaceous cell			Х	
Carcinoma, squamous cell			Х	
Carcinosarcoma			Х	
Histiocytic sarcoma			Х	
Lymphoma, cutaneous			Х	
Melanoma, malignant			Х	

Table 22. Microscopic Findings of the Skin: Rabbit

* Terminology with diagnostic criteria or comments described in the text. # Inducible lesion. + Refer to 4 for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

Comments: Actinic keratosis has been reported in the pinnae of a pet rabbit¹²⁷. The affected animal, a Flemish male, spent considerable time exposed to sunlight. Clinically, there was pinnal erythema and scaling, with histopathologic findings considered consistent with early actinic keratosis: epidermal hyperkeratosis, epidermal hyperplasia with dysplasia, and dermal fibrosis. Actinic (solar) keratosis is an intraepidermal manifestation of abnormal keratinocyte proliferation and can progress to squamous cell carcinoma. Ultraviolet (UV) light is the causative agent. Although exposure to sunlight is not a factor in laboratory studies, actinic keratosis and drug-induced pruritis should be considered as a differential diagnosis in laboratory rabbits showing scaling or pruritis of the ear pinna in studies with test items that cause exacerbated photosensitivity.
SECTION 9: MAMMARY GLAND

A. Anatomy of the Mammary Gland

Similar to humans, each mammary gland in the rabbit has several ducts (milk canals) which open independently in the nipple (rather than a single duct as in some other species such as mice)95, 128, 129. This important anatomical difference observed when comparing rabbits and humans to rodents occurs because when division of the mammary bud starts at rabbit ED26, each sprout which arises forms a primary milk canal¹²⁹ (Propper et al., 2013). Rabbit mammary glands are paired, with usually four (sometimes five) pairs of mammae, each with a nipple, distributed along the ventral thorax and abdomen²⁵. The presence of nipples in male rabbits has been documented in detail previously⁹⁵. Newborns do not require colostrum as all immunity is acquired through the placenta. Rabbits have a unique pattern of nursing: it occurs only once a day, with circadian periodicity, throughout lactation¹³⁰. Explants of pseudopregnant rabbit mammary glands in organ culture are used to investigate hormonal changes during pregnancy. Proliferative lesions in laboratory rabbits may arise from genotoxic test items, infectious agents and changes in hormonal homeostasis as part of the aging process or pseudopregnancy (Table 23).

Inflammation

Comments: Mammary gland inflammation (mastitis) is rarely encountered in modern laboratory facilities due to biosecurity and animal husbandry procedures operations^{131, 132}. More commonly, histological changes are limited to infiltrates of foci of lymphocytes, neutrophils, macrophages, or plasma cells, alone or in combination.

Hyperplasia

Comments: In cases of pseudopregnancy, the entire mammary gland may be diffusely enlarged and hyperplastic and resemble feline fibroadenomatosis (M. Rinke, personal communication). Descriptions of mammary gland "cystic dilatation" in one cohort of untreated rabbits appeared consistent with mammary gland hyperplasia (with increased secretory activity)¹³³. Findings consistent with mammary gland hyperplasia (described as mammary "cystic dilatation" with "epithelial infolding") were noted in female NZW rabbits with elevated serum prolactin levels and pituitary adenomas¹³⁴. Mammary gland hyperplasia and dysplasia were noted in another female NZW rabbit with concurrent mammary gland adenocarcinoma and a prolactin-secreting pituitary adenoma¹³⁵. These results suggested that mammary gland hyperplasia may have been related to elevated prolactin levels.

Experimental reversible mammary gland hyperplasia in rabbits has been induced by systemic administration of cyclosporine A. In one study, intravenous administration of cyclosporine A to nulliparous NZW female rabbits resulted in diffuse mammary hyperplasia characterized by increased numbers of alveoli, increased alveolar branching, increased secretion in alveolar lumens, and decreased fibrous stroma compared to control glands¹³⁶. In another study, intravenous administration of cyclosporine A to male and female NZW rabbits resulted in diffuse mammary gland hyperplasia characterized by abundant ectatic, well-differentiated glands, but with increased fibrous stroma, and the change was termed "fibroadenomatous hyperplasia"¹³⁷. In both studies, the mammary gland hyperplasia regressed after cessation of cyclosporine A administration.

Elevated plasma prolactin levels attributed to cyclosporine A were noted in one of these studies¹³⁶. Thus, a possible role of prolactin in development of mammary gland hyperplasia was postulated^{136, 137}.

Gynecomastia has been found in a male rabbit that concurrently had a testicular Leydig cell tumor¹³⁸.

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Non-proliferative				
Abscess		Х		
Amyloid		X		
Angiectasis		Х		
Apoptosis [‡]		X		
Atrophy		Х		
Basophilia		Х		
Congestion		Х		
Corpora amylacea			Х	
Cyst		Х		
Degeneration		Х		
Dilatation		Х		
Edema		Х		
Fibrosis	Х			
Hemorrhage	Х			
Hypertrophy, alveolar and/or ductal epithelial cell	Х			
Infarct		X		
Infiltrate, inflammatory cell [insert appropriate cell type]	Х			
Inflammation *		Х		
Metaplasia			Х	
Mineralization	Х			
Necrosis		Х		
Pigment		Х		
Single-cell necrosis ⁺		Х		
Thrombus		X		
Proliferative Non-Neoplastic				
Hyperplasia, lobuloalveolar *	Х			
Proliferative Neoplastic				
Adenoma			Х	
Adenomyoepithelioma			Х	
Fibroadenoma			Х	
Tumor, mixed, benign			Х	
Adenocarcinoma			Х	
Adenocarcinoma arising in fibroadenoma			Х	
Carcinoma, adenosquamous			X	
Carcinosarcoma			X	
Sarcoma arising in fibroadenoma			Х	

Table 23. Microscopic Findings of the Mammary Gland: Rabbit

* Terminology with diagnostic criteria or comments described in the text. ⁺Refer to ⁴ for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

SECTION 10: NERVOUS SYSTEM

The neuroanatomy of the rabbit is covered in detail elsewhere¹³⁹. The large brain size of the rabbit compared to rodents necessitates a different approach to trimming and embedding the brain. Pardo *et al.* have published a recommended trimming schemes for routine and neurotoxicity studies¹⁴⁰. Although mature when on study, nervous system findings are not common in laboratory rabbits due to their young age.

This lexicon has been organized in tiers based on cell type (e.g., neuron, glia), and sometimes by cellular target (e.g., cell body, axon, myelin). A final set of terms has been included to demonstrate common artifacts which have been misidentified as neurodegenerative or neurotoxic lesions by inexperienced diagnosticians. Neural neoplasms are an important but very infrequent finding in non-rodents including rabbits evaluated on standard toxicity studies, due to the age group concerned. Conventional toxicity studies do not permit repeated assessment for lesion progression over time, so an appreciation of the true future biological behavior of neural neoplasms is unclear. Thus, in this lexicon we predict the impact on host function as "benign" or "malignant" based only on morphologic characteristics such as cellular differentiation, invasiveness, and proliferative rate. Certain lesions that have well-differentiated features but are biologically aggressive over time (e.g. glial neoplasms) have been termed "malignant" and "low grade" rather than "benign" to better address the usual clinical outcome of this category of neural tumors. Innovations in noninvasive imaging should help resolve questions regarding the biological behavior of these lesions in the future.

The most common non-proliferative anomalies in the CNS include damaged or dying cells (especially neurons, their axonal processes, and the myelin sheaths that insulate the axons) and the various reparative changes designed to minimize (in the CNS) or reverse (in the PNS) the damage. At the cellular level, the most important processes will involve neurons, various glial lineages, and blood vessels (Table 24).

Hydrocephalus

Comments: Cases of hydrocephalus in rabbits have been presumed to be related to a single autosomal recessive gene (hy/hy); however, occurrence with other abnormalities suggests that inheritance may be more complicated. In some cases, the condition appears to be inherited along with various ocular anomalies as an autosomal gene with incomplete dominance. Hydrocephalus may also occur in rabbits as a congenital condition related to hypovitaminosis A in pregnant females¹⁴¹. Hydrocephalus is also seen secondary to craniosynostosis. A strain of NZW rabbits with congenital, nonsyndromic coronal suture synostosis has been developed previously Mooney *et al.*, 1996, which mimics the pathology of human craniosynostosis¹⁴².

Parasite (Figure 24)

Other terms: Encephalitis;

Comments: Encephalitozoon cuniculi is a common cause of neurological disease in pet rabbits, but nowadays is a rare occurrence in barriered colonies. Encephalitozoon cuniculi is a microsporidian pathogen of rabbits and other mammals that usually produces granulomatous encephalitis and interstitial nephritis. Severe systemic disease is rare except in immunosuppressed animals. Primary diagnosis is based on the presence of granulomatous encephalitis together with chronic interstitial nephritis and occasionally hepatitis but can be confirmed by serology or a combination of special staining methods, immunohistochemistry (IHC), and polymerase chain reaction (PCR). Real-time PCR is the most sensitive method for the confirmation of E. cuniculi infection. Differential diagnosis for E. cuniculi is Toxoplasma gondii infection. E. cuniculi infection is characterized by granuloma(s) composed of macrophages with large cytoplasm (epithelioid cells), which may be surrounded by inflammatory cells such as lymphocytes, plasma cells, and eosinophils, sometimes accompanied by fibrosis. The macrophages may form multinucleated giant cells. Focal lesions are largely restricted to the gray matter and primarily spare the white matter, with inner layers of the cerebral cortex more commonly affected than the middle or the outer layers. Lymphoplasmacytic meningoencephalitis, characterized by perivascular cuffs and leptomeningeal infiltrates consisting primarily of lymphocytes and plasma cells, is also usually present in affected animals. Spores are located in parasitophorous vacuoles in the parenchyma of the brain, either without inflammation or close to focal inflammatory lesions. Lesions can be categorized into 6 histopathological subtypes depending on the characteristics of the inflammatory response⁸⁵. Rabbits affected with encephalitozoonosis most frequently exhibit multiple neurologic signs (head tilt, ataxia, circling, nystagmus, rotational movements around the body length axis, seizures, paresis, head tremors, swaying or nodding at rest, and behavior changes), kidney dysfunction (azotemia), and phacoclastic uveitis. Brain lesions may lead to altered cholinesterase values.

Vacuolation, Choroid Plexus (Figure 25)

Comments: Spontaneously occurring choroid plexus vacuolation in rabbits takes two forms, discrete microvacuoles within the epithelium of the choroid plexus or large macrovacuoles (adipocytes) within the stroma. Neither form is associated with clinical signs, both are considered incidental findings at histopathology and should not be recorded as a finding in preclinical studies unless the adipocytes are altered spontaneously (i.e. lipoma) or after xenobiotic treatment¹⁴³. Vacuolated macrophages can be seen in the choroid plexus of animals dosed with PEGylated compounds. These are discrete macrovacuoles and are clearly within either macrophages and/or endothelial cells. Since removal of foreign bodies or materials from circulation is a normal function of macrophages, the resulting vacuolation is considered a normal physiological response in phagocytic cells. The inner core of the choroid plexus, containing the vacuolated

Congenial X Extopic issue, neuron X Abscess X Abscess X Accumulation, mainar, Schwann cell X Ascumulation, mainar, Schwann cell X Astrocyte welling/vacuolation X Astrophysic actions X Astrophysic actions X Astrophysic actions X Coledestroi Cleffs X Cyst. squamous X Degeneration, nerve fiber X Degeneration, nerve fiber X Degeneration in prevensition X Edema, intranyolinic X Edema, intranyolinic X Infarrot X Infarrot X Infarrot X Infarrot X Infarrot X	Finding	Common	Uncommon	Not Recorded but Potentially Relevant	Not Applicable
Ectopic tissue, neuron X Hydrocephaliss* X Absess X Accumulation, laminar, Schwan cell X Accumulation, matrix X Accumulation, matrix X Accumulation, matrix X Astrocyt swelling/vaculation X Chubits/ty, fouron X Chubits/ty, fouron X Chubits/ty, neuron X Degeneration, axon X Degeneration, axon X Degeneration, axon X Edema, intranspointie X Estimation X Destroply, axon X Edema, intranspointie X Estimation X Destroply, axon X Infartet X Infartet, inflammator, cell [incert appropriate cell type] X Infartet X Infartet, inflammator, cell [incert appropriate cell type] X Infartet, inflammator, cell [incert appropri	Congenital				
Hydrocphilabs* X Abscess X Accumulation, Inminar, Schwann cell X Accumulation, Inminar, Schwann cell X Accumulation, Inminar, Schwann cell X Antrophy control X Chularity, feerescel, neuron X Chularity, feerescel, neuron X Cyst, squancous X Degeneration, axon X Degeneration, axon X Degeneration, axon X Ederan, intramyelinic X Detreased elluarity, neuron X Electrophy, ason X Electrophy, ason X Electrophy, ason X Influent	Ectopic tissue, neuron		Х		
Non-poliferative X Abscess X Accumulation, luminar, Schwann cell X Accumulation, matrix X Antrologio, matrix X Antropied X Astrocytic swelling/vacuolation X Astropyte swelling/vacuolation X Astropyte swelling/vacuolation X Astrophagy, neuron, ganglion X Cellularity, decreased, neuron X Cholesterici Lefth X Decreased cellularity, neuron X Degeneration, neuro fiber X Descreased cellularity, neuron X Descreased cellularity neuroprists, chorod plecus X Extramedullary bernatopolicsis, chorod plecus X Infarct X Infarct Infannatory cell [insert appropriate cell type] X Infarcts, infannatory cell [insert appropriate cell type] X	Hydrocephalus *		Х		
Abscess X Accumulation, harinar, Schwann cell X Accumulation, marins, Schwann cell X Amydolg X Amydolg X Astrocyte swelling/raucolation X Astrocyte swelling/raucolation X Astrocyte swelling/raucolation X Astrocyte swelling/raucolation X Cellularity, decreased, neuron X Cellularity, decreased, neuron X Cholesterol effs Cholesterol X Cholesterol effs X Cholesterol effs X Cholesterol effs X Cholesterol effs X Ch	Non-proliferative				
Accumulation, Jaminar, Schwann cell X Accumulation, matrix X Antropy reseveling/vacuolation X Astrocy to seveling/vacuolation X Astrocy reseveling/vacuolation X Astrocy resons X Astrocy resons X Autophagy, neuron, ganglion X Cellularity, decreased, neuron X Chotesteel defa X Chronatolysis X Cyst, squamosu X Decreased cellularity, neuron X Degeneration, acon X Despectation, acon X Detrocylination X Dystrophy, axon X Estramedullary hematopoiesis, choroid plexus X Idiosis, not otherwise specified (NOS) X Hemorrhage X Infiltrate, inflammatory cell [insert appropriate cell type] X Infiltrate, inflammaton X Microplia, neuronal X Microplia, standor X Microplia, neuronal X Infiltrate, inflammation	Abscess		Х		
Accumulation, matrix X Amyloid X Astracytes welling/accolation X Astracytosis X Autophy, scons X Autophagy, actron, ganglion X Cholesterol defts X Chonatolysis X Chonatolysis X Cyst, squamoas X Decremed cellularity, neuron X Degeneration, nerve fiber X Degeneration, nerve fiber X Degeneration, nerve fiber X Degeneration, nerve fiber X Edoma, intramyelinis X Edoma, intramyelinis X Edoma, intramyelinis X Infartet X Infartet, inflammatory cell [insert appropriate cell type] X Inflartet, inflammatory cell [insert appropriate cell type] X Microgliosis X Microgliosis X Microgliosis X Microgliosis X Microgliosis X Stradelliosis <td< td=""><td>Accumulation, laminar, Schwann cell</td><td></td><td>Х</td><td></td><td></td></td<>	Accumulation, laminar, Schwann cell		Х		
Amyolid X Astrocytosis X Astrocytosis X Astrophagy, neuron, ganglion X Cellularity, decreased, neuron X Commatolysis X Chonsteriol defts X Chromatolysis X Operational defts X Decreased cellularity, neuron X Degeneration, axon X Degeneration, nerve fiber X Dernyelinaria, axon X Despresentation, nerve fiber X Edema, intramyolinic X Edema, intramyolinic X Estraneedullary hematopolesis, choroid plexus X Hettorotopia, neuronal X Infart X Infart X Infart X Mineralization X Mineralization X Mineralization X Necrosis, neuron X Mineralization X Necrosis, neuron X Pigment, lipofuscin X Necrosis, neuron X Necrosis, neuron X Mineralization X Necrosis, neuron X Necrosis, neuron X	Accumulation, matrix		Х		
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Autophagy, neuron, ganglion X Cellularity, decreased, neuron X Chromatolysis X Chromatolysis X Cereased cellularity, neuron X Degeneration, acon X Description, acon X Description, acon X Detroyelination X Edema, intramyelinic X Edema, intramyelinic X Extramedultary hematopolesis, choroid plexus X Heterotopia, neuronal X Heterotopia, neuronal X Inflartat X Infiltrate, inflammatory cell [insert appropriate cell type] X Infiltrate, inflammatory cell [insert appropriate cell type] X Mincreditosis X Mincreditosis X Mincreditosis X Necrosis, neuron X	Atrophy, axons		Х		
Cellularity, decreased, neuron X Choostrol elefts X Choostrol elefts X Decreased cellularity, neuron X Degeneration, axon X Degeneration, nerve fiber X Degeneration, nerve fiber X Destrophy, axon X Edema, intramyclinitic X Extramedulary hematopoiesis, choroid plexus X Cilosis, not otherwise specified (NOS) X Hemorrhage X Hetroropia, neuronal X Infitrate, inflammation X Miscogliosis X Statellitosis X	Autophagy, neuron, ganglion		X		
Cholesterol clefts X Chromatolysis X Ocereased cellularity, neuron X Degeneration, axon X Degeneration, axon X Dernychnation X Dernychnation X Dystrophy, axon X Estramedullary hematopoiesis, chooid plexus X Eldema, intramyclinic X Gilosis, not otherwise specified (NOS) X Hemorrhage X Inflart X Inflartate, inflammatory cell [insert appropriate cell type] X Mitorogliosis X Mitorogliosis X Mineralization X Necrosis, neuron X Parasite* X Parasite* X Single cell necrosis' X Single cell necrosis ' X Tissue, ectopic, neuron X Vacuolation, neuron X Vacuolat	Cellularity, decreased, neuron		X		
Chromatolysis X Cyst, squamous X Deceneration, axon X Degeneration, axon X Degeneration, nerve fiber X Demyelination X Dystrophy, axon X Edema, intramyelinic X Extramedullary hematopoiesis, choroid plexus X Gliosis, not otherwise specified (NOS) X Hemorrhage X Infart X Infarter, inflammatory cell [insert appropriate cell type] X Infartazion X Microgliosis X Microgliosis X Microgliosis X Necrosis/inflammation, media or wall, artery X Parasite * X Pignent, lipofuscin X Statellitosis X Single cell necrosis ! X Thrombus X Tissue, ectopic, neuron X Vacuolation, white matter X Vacuolation, cheroid plexus * X Vacuolation, cheroid plexus * X Thrombus X	Cholesterol clefts		X		
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Astrocytoma, malignant X Carcinoma, choroid plexus X Ependymoma, benign X Ependymoma, malignant X Glioma, mixed, malignant X Glioma, not otherwise specified (NOS) X Medulloblastoma X Neuromyoblastoma, malignant X Oligodendroglioma, malignant X Panilloma, eboroid plexus X				V	
Carcinoma, choroid piexusXEpendymoma, benignXEpendymoma, malignantXGlioma, mixed, malignantXGlioma, not otherwise specified (NOS)XMedulloblastomaXNeuromyoblastoma, malignantXOligodendroglioma, malignantXPanilloma, choroid plexusX	Astrocytoma, malignant			<u>Λ</u> ν	
Ependymona, bengn X Ependymoma, malignant X Glioma, mixed, malignant X Glioma, not otherwise specified (NOS) X Medulloblastoma X Neuromyoblastoma, malignant X Oligodendroglioma, malignant X Panilloma, choroid playus X	Carcinoma, chorida piexus			<u> </u>	
Ependymoma, malignant X Glioma, mixed, malignant X Glioma, not otherwise specified (NOS) X Medulloblastoma X Neuromyoblastoma, malignant X Oligodendroglioma, malignant X Panilloma, choroid playus X	Ependymoma, benign			<u> </u>	
Glioma, mixed, malignant X Glioma, not otherwise specified (NOS) X Medulloblastoma X Neuromyoblastoma, malignant X Oligodendroglioma, malignant X Panilloma, choroid playus X	Ependymoma, malignant			<u> </u>	
Medulloblastoma X Medulloblastoma X Neuromyoblastoma, malignant X Oligodendroglioma, malignant X Panilloma choroid playus X	Clieme not otherwise specific 1 (NOS)			Λ V	
International X Neuromyoblastoma, malignant X Oligodendroglioma, malignant X Panillama, choroid playus X	Madullablastama			<u>Λ</u> ν	
Neuromyoolastoma, malignant X Oligodendroglioma, malignant X Panilloma, choroid plexus X	Neuromychlastoma			<u>Λ</u> ν	
Ongoucharlognoma, mangnant A Panilloma, choroid plexus V	Olicodondroaliomo, malignant			Λ V	-
	Panilloma, choroid nlexus			<u>л</u> Х	

 Table 24.
 Microscopic Findings of the Brain: Rabbit

* Terminology with diagnostic criteria or comments described in the text. ⁺Refer to ⁴ for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

Finding	Common	Uncommon	Not Recorded but Potentially Relevant	Not Applicable
Congenital			· ·	
Ectopic tissue, neuron		Х		
Hydromyelia		Х		
Syringomyelia		Х		
Syringomyelia/hydromyelia		Х		
Non-proliferative				
Abscess		Х		
Accumulation, matrix		Х		-
Accumulation, laminar, Schwann cell		Х		-
Astrocyte swelling		Х		
Astrocyte vacuolation		Х		
Astrocytosis		Х		
Atrophy, axon		Х		
Autophagy, neuron, dorsal root ganglion		Х		
Cellularity decreased, neuron		Х		
Cholesterol clefts		Х		-
Chromatolysis		Х		
Cyst. squamous		Х		
Degeneration, axon		х		
Degeneration, nerve fiber		X		
Demvelination *		Х		
Dystrophy, axon		х		
Ectopic tissue			X	
Gliosis, not otherwise specified (NOS)		Х		
Hemorrhage		Х		
Heterotopia, neuronal			X	
Infarct		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		х		
Inflammation		Х		
Intramvelinic edema		X		
Microgliosis		Х		
Mineralization		X		
Necrosis, neuron		X		
Necrosis/inflammation. media or wall. artery		X		
Neuronophagia		X		
Pigment, lipofuscin		X		
Radiculoneuropathy				X
Satellitosis		X		
Single cell necrosis [†]		X		
Swelling, astrocyte		X		
Thrombus		X		
Type II astrocytes		X		
Vacuolation, neuron		X		
Proliferative Non-Neoplastic				
Hyperplasia glial cell not otherwise specified (NOS)			x	
Proliferative Neoplastic				
Astrocytoma, malignant			Х	
Glioma, mixed, malignant			X	
Glioma, not otherwise specified (NOS)			X	
Oligodendroglioma, malignant			X	

* Terminology with diagnostic criteria or comments described in the text. ⁺Refer to ⁴ for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

macrophages, is outside the CNS. There are two theories as to how the vacuolated macrophages arise: (i) hydrophilic PEG filters through the vasculature into the choroid plexus core and is kept there by the choroidal epithelial tight junctions and subsequently phagocytosed by resident macrophages, or (ii) circulating macrophages containing PEG filter out in this high blood flow area¹⁴⁴ (Table 25).

Demyelination, Nerve Fiber, Spinal Cord

Comments: New-born rabbits show an area in the dorsal part of the lateral funiculus of the first cervical segment of the spinal cord, from which myelinated nerves are few/absent¹⁴⁵. Localised/focal unmyelinated axons are occasionally seen in the sections of lateral funiculus of the lumbar spinal cord of young adult rabbits. This is a normal anatomical difference in rabbits and there are no clinical signs (Table 26).

Pigment

Other terms: Melanosis

Comments: Spontaneously occurring melanosis of the meninges (especially those of the brain) may be seen in pigmented rabbit strains such as the NZW x New Zealand Red F1 Cross and Dutch Belted. It may be seen grossly at necropsy (Table 27).

Common Artifacts

Certain incidental changes are commonly misidentified as neuropathologic lesions by inexperienced researchers. The changes listed here are the most common of such findings – dark neurons and white matter vacuolation. Identified artifacts should be not reported in the pathology dataset. However, systematic artifacts where one dose group only is involved may be recognized and noted as a comment to the organ/tissue at the discretion of the study pathologist (Table 28).

Table 26. Microscopic Findings of the Meninges: Rabbit

Finding	Common	Uncommon	Not Recorded but Potentially Relevant	Not Applicable
Non-proliferative				
Abscess		Х		
Aggregate, granular cell		Х		
Angiectasis		Х		
Cholesterol clefts		Х		
Cyst, squamous		Х		
Fibrosis		Х		
Hemorrhage		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Mineralization		Х		
Necrosis		Х		
Necrosis/inflammation, media or wall, artery		Х		
Pigment *		Х		
Pigment, lipofuscin		Х		
Single cell necrosis [‡]		Х		
Thrombus		Х		
Proliferative Non-neoplastic				
Meningioangiomatosis			Х	
Proliferative Neoplastic				
Meningioma, benign			Х	
Meningioma, malignant			Х	
Tumor, granular cell, benign			Х	
Tumor, granular cell, malignant			Х	

* Terminology with diagnostic criteria or comments described in the text. + Refer to 4 for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

Finding	Common	Uncommon	Not Recorded but Potentially Relevant	Not Applicable
Non-proliferative				
Accumulation, laminar, Schwann cell		Х		
Accumulation, matrix			Х	
Atrophy, axon		Х		
Autophagy, neuron, dorsal root ganglion		Х		
Cellularity, decreased, neuron		Х		
Chromatolysis		Х		
Cholesterol clefts		Х		
Cyst, squamous		Х		
Degeneration, nerve fiber		Х		
Degeneration, axon		Х		
Demyelination		Х		
Dystrophy, axon		Х		
Ectopic tissue		Х		
Hemorrhage		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Intramyelinic edema		Х		
Mineralization		Х		
Necrosis/inflammation, media or wall, artery		Х		
Pigment, lipofuscin		Х		
Radiculoneuropathy				Х
Renaut Body		Х		
Single cell necrosis [‡]		Х		
Proliferative Non-Neoplastic				
Hyperplasia, (+ cell type)			Х	
Proliferative Neoplastic				
Schwannoma			Х	

 Table 27.
 Microscopic Findings of the Peripheral Nerves: Rabbit

Table 28.	Artifactual	Microscopic	Findings of	of the Nervo	us System: l	Rabbit
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Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Artifact, dark neuron	Х			
Bubbles, myelin		Х		
Vacuolation, white matter	Х			

SECTION 11: FEMALE REPRODUCTIVE SYSTEM

Introduction

Standard INHAND nomenclature for non-proliferative and proliferative female reproductive system findings in rats and mice has previously been published¹⁴⁶ and should be used for the rabbit as appropriate. Generally, very few spontaneous or treatment induced microscopic changes are reported in the female reproductive tract of rabbits. Female rabbits become sexually mature at 5 months of age, and so rabbits on routine tox studies are fully mature.

A. Anatomy and Physiology of the Ovary

Rabbits are non-seasonally polyestrous. Female rabbits are induced ovulators and so decreased/absent corpora lutea and atretic follicles are normal. Ovulation can be induced in studies where vaginal swabs/dosing is a frequent event, and corpora hemorrhagica may be seen. Ovulation is triggered about 10 hours following copulation. Some animals are triggered into pseudopregnancy lasting 15–17 days, with or without galactopoiesis and endometrial hyperplasia.

Ovarian interstitial tissue is well developed in rabbits and was termed an "interstitial gland" in 1902¹⁴⁷. This "gland" secretes progestogens. At 3 months of age, follicular atresia begins and the theca interna cells differentiate into the primary and then secondary interstitial gland¹⁴⁸. The primary interstitial gland is composed of small polygonal cells with hyperchromatic nuclei and scant basophilic cytoplasm. It is highly vascularized. The ovary interstitium is almost completely composed of large polygonal cells with round vesicular nuclei and abundant eosinophilic cytoplasm containing numerous lipid droplets. There is high 3- β -hydroxysteroid dehydrogenase activity within the interstitial cells of secondary interstitial gland, which increases in activity with increase in size of the gland.

The ovaries of normal rabbits often have surface epithelial structures resembling ovarian papillomas¹⁴⁹. The ovarian mesothelium (surface epithelium) may form short, broad papilla or papillae with slender villous processes. They have an inner core with few fibroblasts, small blood vessels, and loose or hyalinized connective tissues and are lined by pseudostratified or multi-layered epithelium. These structures should not be mistaken for neoplasms and occur only on the ovary and not on the surface of other abdominal organs or structures (Table 29, 30).

B. Anatomy of the Uterus and Cervix

Rabbits have a true bicornuate uterus, with no uterine corpus present. There are two separate cervices. Each uterine horn has a separate cervical canal and the tract is classified as uterus duplex, vagina simplex. The broad ligaments typically contain a large amount of adipose tissue. The light, transmission and scanning microscopic anatomy of the rabbit vagina, cervix and uterus have been comprehensively described by^{150, 151}. Placentation is discoid labyrinthine hemodischorial with countercurrent fetomaternal blood flow; gestation lasts 25-30 days. Females give birth to a litter of 3-10. Neonates are weaned at about 6-8 weeks (Table 31).

Aneurysm

Comments: Endometrial venous aneurysms have been reported in three adult NZW rabbits with a history of intermittent severe urogenital bleeding (hematuria). The endometrium of all three rabbits had multiple blood-filled vesicles which projected into the uterine lumen, which were considered to be congenital or acquired defects in the vessel walls, as has occurred in humans¹⁵².

Angiectasis

Comments: A focus of multiple, dilated thin walled blood vessels is occasionally seen as a congenital defect. They may occasionally rupture and bleed into the uterine lumen.

Decidual Reaction

Comments: Decidual reactions have been reported in nonpregnant rabbits given estrogens and progesterone as an early lesion in the development of deciduosarcoma^{153, 154}.

Hyperplasia

Comments: Spontaneous endometrial hyperplasia (polypoid and/or cystic) can be seen at 1 year of age¹⁵⁵, although it is more common in rabbits 4–5 years old.

Deciduosarcoma

Pathogenesis/cell of origin: Uterine stromal cells and uterine metrial gland cells.

Differential Diagnosis: Sarcoma, endometrial stromal; Mesenchymal proliferative lesion

Diagnostic Features: Rare malignant tumor of hypertrophied stromal cells with abundant PAS positive rarefied cytoplasm intermingled with numerous globular lymphocytes. Hypertrophied blood vessels characterize the lesion.

Comments: Deciduosarcomas are neoplasms unique to rabbits, and metastasis may occur. They are hormone-dependent and therefore may not actually be true neoplasms. Deciduosarcomas have been reported as induced tumors in rabbits on toxicity studies involving estrogen and progestin administration^{153, 154, 156}, and so because of this, the rabbit is a poor model for evaluating the effects of contraceptive steroids. In addition, a spontaneous deciduosarcoma has also been reported in a 6-year-old Dutch Belted rabbit¹⁵⁷. Deciduosarcomas may appear after as little as 30 days of treatment of non-pregnant rabbits with estrogens and progesterone¹⁵⁴. Exogenous estrogens are necessary for decidualization of the endometrium and to produce deciduosarcoma; exogenous progesterone promotes the process¹⁵⁶. Withdrawal of the estrogen/progesterone treatment results in atrophy of decidual cells and tumors and disappearance

Table 29. Microscopic Findings of the Ovary: Rabbit

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Hypoplasia		Х		
Non-proliferative				
Abscess		Х		
Age related atrophy	Х			
Amyloid		Х		
Angiectasis		Х		
Apoptosis [‡]		Х		
Atrophy	Х			
Atrophy, corpora lutea		Х		
Corpora lutea, increased number		Х		
Cyst, bursal/epithelial/follicular/luteal/paraovarian/rete ovarii, not otherwise specified (NOS)	Х			
Decreased number/absent follicles/corpora lutea		Х		
Degeneration, oocyte/corpora lutea		Х		
Edema		Х		
Follicle, luteinized/polyovular		Х		
Fibrosis		Х		
Granuloma		Х		
Hemorrhage		Х		
Hypertrophy, corpora lutea/ interstitial cell		X		
Immature		X		
Increased number, atretic follicles/corpora lutea		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Mineralization		X		
Necrosis		Х		
Ovotestis			Х	
Pigment		X		
Single cell necrosis [†]		X		
Tissue, ectopic		X		
Vacuolation, corpora lutea/granulosa cell/theca cell/interstitial cell		X		
Proliferative Non-neoplastic				
Hyperplasia, epithelium/granular cell		Х		
Adenosis		Х		
Proliferative Neoplastic				
Adenoma			Х	
Cystadenoma			Х	
Leiomyoma		Х		
Luteoma, benign			Х	
Schwannoma, benign			Х	
Teratoma, benign			Х	
Thecoma, benign			Х	
Tumor. Sertoli cell/granulosa cell, benign			Х	
Tumor, sex cord stromal, mixed, benign			Х	
Carcinoma, embryonal/tubulostromal/volk sac			Х	
Choriocarcinoma			Х	
Cystadenocarcinoma			X	
Dysgerminoma			X	
Leiomyosarcoma			X	
Schwannoma, malignant			X	
Teratoma, malignant			X	
Thecoma malignant			X	
Tumor. Sertoli cell/granulosa cell. malignant			X	
Tumor, sex cord stromal, mixed, malignant			X	
,,,,				

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Hypoplasia		Х		
Non-proliferative				
Abscess		Х		
Apoptosis [‡]				
Atrophy		Х		
Cyst	Х			
Dilatation		Х		
Edema		Х		
Fibrosis		Х		
Granuloma		Х		
Hemorrhage		Х		
Immature		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation, oviduct		Х		
Mineralization		Х		
Necrosis		Х		
Salpingitis isthmica nodosa			Х	
Single cell necrosis [‡]		Х		
Proliferative Non-neoplastic				
Hyperplasia, epithelium		Х		
Proliferative Neoplastic				
Leiomyoma			Х	
Schwannoma, benign			Х	
Schwannoma, malignant			Х	

Table 30. Microscopic Findings of the Oviduct: Rabbit

* Refer to 4 for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

of decidual tumors¹⁵⁴. Transplantation of deciduosarcomas into nude mice as well as primary cell culture (in the absence of hormones) results in failure of cells to grow. Deciduosarcomas will invade the uterine walls and the uterine lymphatics and will metastasize to the lungs. Deciduosarcomas occurring in the spleen and other abdominal viscera may not be metastases, but instead are primary tumors, because treatment of castrated male rabbits with estrogen and progesterone will result in deciduosarcoma in the spleen at a high incidence. Induced deciduosarcomas are composed of large anaplastic decidual cells with large hyperchromatic nuclei. Multiple nuclei as well as bizarre giant nuclei may be observed. Most cells are vacuolated, containing multiple small or a few large vacuoles. Necrosis in the tumor is common. The spontaneously occurring deciduosarcoma is described as an invasive mass originating in the mesometrium of the endometrial stroma and invading the myometrium and mesometrium to form a large nodular mass. Cells are arranged in sheets and streaming bundles dissecting between the smooth muscle cells or arranged concentrically around large dilated blood vessels, with multiple foci of necrosis. Cells are anaplastic and consisted of two intermingling populations – spindloid cells with scant eosinophilic

cytoplasm and cigar shaped nuclei with a single nucleolus, or epithelioid cells with abundant vacuolated eosinophilic cytoplasm, large oval eccentric nuclei with 1–2 nucleoli. Binucleate and multinucleate cells are common, as well as frequent giant cells, anisocytosis and nuclear pleomorphism. There are multiple mitotic figures per high power field.

Special Techniques: The neoplastic cells show positive cytoplasmic staining for vimentin, and nuclear staining for estrogen and progesterone receptors, and are negative for desmin, α smooth muscle actin (α SMA), pancytokeratin, and CD10.

C. Anatomy of the Vagina

The vulva lies between the two inguinal sinuses; the vagina joins the urethra at the vestibule. The vestibule is surrounded by erectile tissue. The crura of the clitoris are present caudal to this erectile tissue, with the body and glans extending caudally to the vulva. The paired clitoral (inguinal) glands are located in the subcutis on either side of the base of the clitoris¹⁰⁶ (see Integument section) (Table 32, 33).

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Hypoplasia			Х	
Mesonephric duct remnants			Х	
Non-proliferative				
Abscess		Х		
Adenomyosis		Х		
Aggregate, granular cell		Х		
Amyloid		Х		
Aneurysm *		Х		
Angiectasis *		Х		
Apoptosis ⁺		Х		
Atrophy		Х		
Basophilia			Х	
Cyst, NOS	Х			
Decidual reaction *		Х		
Decidualization, focal			Х	
Degeneration, epithelial		Х		
Dilatation, luminal/glandular/cystic		Х		
Edema		Х		
Fibrosis		Х		
Granuloma		Х		
Hemorrhage		Х		
Hypertrophy, epithelium/myometrium		Х		
Immature		Х		
Infarct		X		
Infiltrate, inflammatory cell [insert appropriate cell type]		X		
Inflammation		X		
Metaplasia, squamous cell		Х		
Mineralization		Х		
Necrosis		Х		
Pigment		Х		
Prolapse		X		
Pvometra		Х		
Single cell necrosis [‡]		Х		
Tissue, ectopic		Х		
Vacuolation, epithelium		Х		
Proliferative Non-neoplastic				
Hyperplasia, epithelium/granular cell *	х			
Hyperplasia segmental cystic			X	
Neonlastic Lesions				
Adenoma, endometrial			х	
Keratoacanthoma			X	
Leiomyoma			X	
Panilloma squamous cell			X	
Polyn			X	
Schwannoma henian			X	
Tumor granular cell benign			X	
Adenocarcinoma endometrial			X	
Carcinoma, squamous cell			X	
Deciduosarcoma #*		v	Α	
Leiomyosarcoma		Λ	v	
Histioevtic sarcoma			<u>л</u> V	
Schwannoma malignant			<u>л</u> V	
Tumor, gronulor coll moligrant			<u>л</u> v	
Tumor, granular cell, malignant			<u>л</u> v	
iunor, mixed Mullerian, malignant			Λ	

 Table 31.
 Microscopic Findings of the Uterus and Cervix: Rabbit

* Terminology with diagnostic criteria or comments described in the text. # Inducible lesion. + Refer to 4 for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Cyst (mesonephric duct remnants)			Х	
Non-proliferative				
Abscess		Х		
Adenosis		Х		
Aggregate, granular cell		Х		
Angiectasis		Х		
Apoptosis ⁺		Х		
Atrophy, epithelial	Х			
Cyst, NOS	Х			
Degeneration, epithelial		Х		
Dilatation		Х		
Edema		Х		
Erosion/ulcer		Х		
Fibrosis		Х		
Granuloma		Х		
Hemorrhage		Х		
Hypertrophy		X		
Immature		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Keratinization, increased		Х		
Metaplasia, squamous cell		Х		
Mineralization		Х		
Mucification, increased		Х		
Necrosis		Х		
Pigment		Х		
Prolapse		Х		
Rudiment, prostatic		Х		
Single cell necrosis ⁺		Х		
Vacuolation, epithelial		Х		
Vagina, imperforate		Х		
Proliferative Non-neoplastic				
Adenosis				Х
Hyperplasia, epithelium/granular cell		Х		
Proliferative Neoplastic				
Keratoacanthoma			Х	
Leiomyoma		Х		
Papilloma, squamous cell			Х	
Polyp, vaginal			Х	
Schwannoma, benign			Х	
Tumor, granular cell, benign			Х	
Carcinoma, squamous cell/adenosquamous			Х	
Leiomyosarcoma		X		
Sarcoma, endometrial stromal			Х	
Schwannoma, malignant			Х	
Tumor, granular cell, malignant		Х		

 Table 32.
 Microscopic Findings of the Vagina: Rabbit

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Non-proliferative				
Abscess		Х		
Angiectasis		Х		
Apoptosis [‡]			Х	
Atrophy		Х		
Basophilia			Х	
Degeneration		Х		
Dilatation		Х		
Edema		Х		
Fibrosis		Х		
Granuloma		Х		
Hemorrhage		Х		
Immature		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Necrosis		Х		
Pigment		Х		
Single cell necrosis [‡]		Х		
Proliferative Non-neoplastic				
Hyperplasia, squamous cell		Х		
Hyperplasia		Х		
Proliferative Neoplastic				
Adenoma			Х	
Papilloma, squamous cell			Х	
Tumor, basal cell, benign			Х	
Adenocarcinoma			Х	
Carcinoma, squamous cell			Х	
Tumor, basal cell, malignant			Х	

 Table 33.
 Microscopic Findings of the Clitoral (Inguinal) gland: Rabbit

SECTION 12: MALE REPODUCTIVE SYSTEM

Male rabbits become sexually mature at 6–7 months of age and so rabbits used on routine safety assessment studies are usually fully mature (Table 34).

Atrophy, Tubule

Comments: Tubular atrophy is an infrequent finding in the testis. It may be unilateral or bilateral but is generally localized affecting only a few tubules and has not been reported as affecting the whole testis. There is an accompanying oligospermia and sloughing of ductal epithelium in the epididymides¹⁵⁸.

Adenoma, (+ cell type)

Comments: Interstitial (Leydig) cell^{138, 159} and granular cell tumors¹⁶⁰ are reported in adult/young adult rabbits (Table 35).

Cell Debris, Lumen

Comments: There is often more cellular debris in the lumen of rabbit epididymis than in other species (Table 36).

A. Anatomy of the Prostate, Proprostate and Paraprostate

In the male rabbit, two structures (termed "prostate" and "proprostate" in this review) analogous to the prostate of other species are located dorsal to the seminal vesicle, urethra, and urinary bladder (Figures 27, 28). These structures are two closely apposed bilobed glands which are considered separate organs with independent excretory ducts (rather than parts of a single multilobed gland)^{161, 162}. The more caudally situated gland is generally designated as the "prostate"^{162–164}, and is composed of a cini lined by uniform, very tall columnar epithelium with abundant pale eosinophilic cytoplasm and basally located oval nuclei. The more cranially situated gland has been most commonly designated as the "proprostate"^{162, 163}. Proprostate acini are lined by low columnar epithelium with bright eosinophilic cytoplasm and centrally located oval to round nuclei (Figures 29, 30).

Male rabbits also have "paraprostate" glands associated with the urethra^{161–163}. The paraprostate is comprised of small, club-shaped structures that lie on the dorsolateral aspect of the prostate and are embryologically derived from the urethral wall (Figure 31). These glands are histologically identical to the prostate (Figure 32). These are not routinely specifically sampled in toxicology studies but can appear fortuitously in cross-sections of the accessory sex-organ/urethral region as small clusters of prostate-like acini embedded in the periure-thral connective tissue (Figure 33) (Table 37).

Metaplasia, Squamous Cell

Comments: Spontaneous focal keratinized squamous metaplasia of the prostate and proprostate epithelium is an infrequent finding peculiar to the rabbit and is clinically silent. It is important to recognize this as a spontaneous le-

sion when evaluating test articles with androgenic or estrogenic actions. The finding is well described in Dutch Belted and NZW rabbits ¹⁶⁴. It is characterized by generally small foci of increased numbers of lamellated squamous-like cells with or without superficial keratinization and sloughing of cornified debris into the gland alveolar lumen; the squamous metaplastic foci are located within the gland epithelium or can also extend into the underlying lamina propria.

B. Anatomy of the Seminal Vesicles

In most species, the paired seminal vesicles (vesicular glands) are dorsal to the ampullae. For consistency with nomenclature in rats, mice, and other species, this review uses the term "seminal vesicle" for the analogous structure in the male rabbit. Rather than separate paired organs as in other species, the rabbit seminal vesicle appears grossly as a single sac-like structure located dorsal to the urinary bladder and ventral to the prostate complex^{162, 163}. The cranial (blind) end of the seminal vesicle has a distinctive grossly visible "dimpled" indentation, which corresponds a microscopic appearance of two sideby-side glands with separate lumens in the proximal portion (appreciable in cross-sections). In the more distal main body of the seminal vesicle, the central separating wall disappears to create a conjoined single lumen^{161, 165}. The histologic appearance of seminal vesicle epithelium (basophilic cuboidal to low columnar cells) is similar to that in other species (Table 38).

C. Anatomy of the Bulbourethral Gland

The bilobed bulbourethral gland is located dorsal to the urethra and caudal to the prostate/proprostate^{161–163, 166}. The histologic appearance of the bulbourethral gland epithelium is similar to that in other species. The two lobes of bulbourethral gland are subdivided into lobules by thick connective tissue septa and embedded entirely in the bulboglandularis muscle, portions of which penetrate the interlobar and interlobular septa of the gland. Excretory ducts of the bulbourethral gland exit ventrally to empty into the urethra. Each lobule of the bulbourethral gland has a substantial lamina propria with large central lumina lined by a simple cuboidal to columnar to pseudostratified epithelium (Table 39).

D. Anatomy of the Preputial Gland

So-called "preputial" glands (inguinal gland complex) are paired structures in the subcutis on either side to the base of the penis and prepuce¹⁰⁶, which are further described in the Skin section. Descent of the testes into the scrotum occurs at 2.5 to 3 months of age. However, males can retract their testes into the abdominal cavity via the inguinal ring (Table 40).

E. Anatomy of the Vas Deferens (Ampulla)

As the deferent ducts converge, their walls become thickened to form ampullae, connected to each other by the genital

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Agenesis		Х		
Aplasia		Х		
Cryptorchidism		Х		
Hypoplasia		Х		
Non-proliferative				
Abscess		Х		
Amyloid		Х		
Angiectasis		Х		
Apoptosis [‡]		Х		
Atrophy, Leydig cell/tubular *	Х			
Degeneration/atrophy, tubule		Х		
Degeneration, germ cell/tubular		Х		
Depletion, germ cell		Х		
Dilatation, rete testis/tubule		Х		
Edema		Х		
Exfoliation, germ cell		Х		
Fibrosis		X		
Hemorrhage		X		
Infiltrate, inflammatory cell [insert appropriate cell type]		X		
Inflammation		X		
Mineralization		X		
Multinucleated giant cell	x	11		
Necrosis	A	x		
Necrosis/inflammation_vascular/nerivascular		X		
Pigment		X		
Residual hodies, atvnical		X		
Single cell necrosis ‡		x		
Sherm granuloma		X V		
Sperm stasis		X X		
Spermetid retention		x v		
Spermatic retention				
		X		
Vacuolation, macrophage	V	Λ		
Partification, Leydig cell/tubule	Λ			
Proliferative Non-heoplastic		V		
Hyperplasia, (+ cell type)		X		
Proliferative Neoplastic		37		
Adenoma, (+ cell type) *		X		
Gonadoblastoma			X	
Seminoma, benign			<u>X</u>	
Teratoma, benign			X	
Tumor, granulosa cell, benign				X
Tumor, granular cell, benign			X	
Tumor, mixed Sertoli-Leydig cell, benign			X	
Tumor, Sertoli cell, benign			Х	
Carcinoma, embryonal/Leydig cell/rete testis/yolk sac			Х	
Choriocarcinoma			Х	
Mesothelioma, malignant			Х	
Seminoma, malignant			Х	
Teratoma, malignant			Х	
Tumor, Sertoli cell, malignant			Х	

 Table 34.
 Microscopic Findings of the Testis: Rabbit

* Terminology with diagnostic criteria or comments described in the text. + Refer to 4 for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Aplasia		Х		
Hypoplasia		X		
Non-proliferative				
Abscess		Х		
Amyloid		Х		
Apoptosis [‡]		Х		
Atrophy, duct	Х			
Cell debris, lumen *	Х			
Cribriform change		Х		
Degeneration, epithelium		Х		
Dilatation, duct		Х		
Edema		Х		
Fibrosis		Х		
Granuloma, sperm		Х		
Hemorrhage		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Karyomegaly		Х		
Metaplasia, squamous cell	Х			
Mineralization		Х		
Necrosis		Х		
Necrosis/inflammation, vascular/perivascular		Х		
Single cell necrosis [‡]		X		
Sperm stasis		X		
Sperm, decreased, lumen		Х		
Spermatocele		Х		
Vacuolation, epithelium	Х			
Proliferative Non-neoplastic				
Adenosis			Х	
Proliferative Neoplastic				
Adenoma, Leydig cell			Х	
Histiocytic sarcoma			Х	

 Table 35.
 Microscopic Findings of the Epididymis: Rabbit

* Terminology with diagnostic criteria or comments described in the text. * Refer to 4 for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

fold. In contrast to rats and mice, the rabbit distal vas deferens has a very wide ampullary region^{162, 163}. The ampullae can be so large that they have been mistaken for "paired" seminal vesicles at gross necropsy. Microscopically, the ampulla exhibits abundant, large lamina propria glands, which result in a distinctive "honeycomb" appearance (Figure 33) (Table 41).

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Aplasia		Х		
Hypoplasia		Х		
Non-proliferative				
Amyloid		Х		
Angiectasis		Х		
Apoptosis [‡]		Х		
Atrophy, duct	Х			
Cell debris, lumen	Х			
Cribriform change		Х		
Degeneration, epithelium		Х		
Dilatation, duct		Х		
Edema		Х		
Fibrosis		Х		
Granuloma, sperm		Х		
Hemorrhage		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Metaplasia, squamous cell	Х			
Mineralization		Х		
Necrosis		Х		
Necrosis/inflammation, vascular/perivascular		Х		
Single cell necrosis [‡]		Х		
Sperm stasis		Х		
Spermatocele		Х		
Vacuolation, epithelium	Х			
Proliferative Non-neoplastic				
Hyperplasia			Х	

Table 36. Microscopic Findings of the Efferent ducts: Rabbit

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Aplasia		Х		
Hypoplasia		X		
Non-proliferative				
Abscess		Х		
Amyloid		Х		
Angiectasis		X		
Apoptosis [‡]		X		
Atrophy		X		
Basophilia		X		
Corpora amylacea		Х		
Degeneration		Х		
Dilatation, acinar/acinus/vesicle		Х		
Edema		X		
Fibrosis		X		
Hemorrhage		X		
Infiltrate, inflammatory cell [insert appropriate cell type]		X		
Inflammation		X		
Metaplasia, squamous cell *	Х			
Mineralization		Х		
Necrosis		X		
Necrosis/inflammation, vascular/perivascular		Х		
Pigment		Х		
Single cell necrosis [‡]		Х		
Vacuolation, epithelial		X		
Proliferative Non-neoplastic				
Hyperplasia, atypical/diffuse/reactive		Х		
Proliferative lesion, mesenchymal			Х	
Proliferative Neoplastic				
Adenoma			Х	
Papilloma, squamous cell			Х	
Tumor, granular cell, benign			Х	
Adenocarcinoma			Х	
Carcinoma, squamous cell			Х	
Carcinosarcoma			X	
Tumor, granular cell, malignant			Х	
Tumor, neuroendocrine, malignant			Х	

 Table 37.
 Microscopic Findings of the Prostate, Proprostate and Paraprostate: Rabbit

* Terminology with diagnostic criteria or comments described in the text. + Refer to 4 for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Aplasia		Х		
Hypoplasia		Х		
Non-proliferative				
Abscess		Х		
Amyloid		Х		
Angiectasis		Х		
Apoptosis ⁺		Х		
Corpora amylacea		Х		
Degeneration		Х		
Dilatation, acinar/vesicle		Х		
Edema		Х		
Fibrosis		Х		
Hemorrhage		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Metaplasia			Х	
Mineralization		Х		
Necrosis		Х		
Necrosis/inflammation, vascular/perivascular		Х		
Pigment		Х		
Single cell necrosis ⁺		Х		
Vacuolation, epithelial		Х		
Proliferative Non-neoplastic				
Hyperplasia, atypical/diffuse/reactive			Х	
Proliferative lesion, mesenchymal			Х	
Proliferative Neoplastic				
Adenoma			Х	
Tumor, epithelial-stromal, benign			Х	
Tumor, granular cell, benign			X	
Adenocarcinoma			X	
Tumor, granular cell, malignant			X	

Table 38. Microscopic Findings of the Seminal Vesicle: Rabbit

 † Refer to 4 for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Aplasia		Х		
Hypoplasia		Х		
Non-proliferative				
Abscess		Х		
Amyloid		Х		
Angiectasis		Х		
Apoptosis [‡]		Х		
Atrophy		Х		
Corpora amylacea		Х		
Degeneration		Х		
Dilatation, acinar/vesicle		Х		
Edema		Х		
Fibrosis		Х		
Hemorrhage		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Metaplasia		Х		
Mineralization		Х		
Necrosis		Х		
Necrosis/inflammation, vascular/perivascular		Х		
Single cell necrosis ⁺		Х		
Vacuolation, epithelial		Х		
Proliferative Non-neoplastic				
Hyperplasia, atypical/diffuse/reactive			Х	
Proliferative Neoplastic				
Adenoma			Х	
Adenocarcinoma			X	

 Table 39.
 Microscopic Findings of the Bulbourethral gland: Rabbit

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Aplasia			Х	
Hypoplasia			Х	
Non-proliferative				
Abscess		Х		
Apoptosis [‡]		Х		
Atrophy		Х		
Basophilia		Х		
Degeneration		Х		
Dilatation		Х		
Edema		Х		
Hemorrhage		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Necrosis		Х		
Single cell necrosis ⁺		Х		
Vacuolation, epithelial		Х		
Proliferative Non-neoplastic				
Hyperplasia, atypical/diffuse/reactive/squamous cell			Х	
Proliferative Neoplastic				
Adenoma			Х	
Papilloma, squamous cell			Х	
Adenocarcinoma			Х	
Carcinoma, squamous cell, malignant			Х	
Tumor, basal cell, malignant			X	

Table 40. Microscopic Findings of the Preputial (Inguinal) gland: Rabbit

Table 41.	Microscopic	Findings of	f the Ampullary	gland: Rabbit
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Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Aplasia			Х	
Hypoplasia			Х	
Non-proliferative				
Abscess		Х		
Angiectasis		Х		
Apoptosis ‡		Х		
Degeneration		Х		
Edema		Х		
Hemorrhage		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Necrosis		Х		
Necrosis/inflammation, vascular/perivascular		Х		
Single cell necrosis [‡]		Х		
Vacuolation, epithelial		Х		
Proliferative Non-neoplastic				
Hyperplasia, atypical/diffuse/reactive			Х	
Proliferative Neoplastic				
Adenoma			Х	
Adenocarcinoma			Х	

SECTION 13: RESPIRATORY SYSTEM

Adult and juvenile rabbits are occasionally used for inhalation and intranasal safety assessment studies to test local tolerability and potential toxicity for drugs, chemicals and particulates. Rabbits are prone to atherosclerotic lesions in a variety of organs including the lung. For detailed description, see the cardiovascular section. Rabbits can be used as animal models for tuberculosis research, and they can be infected via snout only inhalation of the bacteria¹⁶⁷. In addition, rabbits are used as animal models for induced hypertension and aspiration pneumonia, and for sinus research, i.e. study of surgical packing material or surgical techniques. Modern laboratory animal management practices within rabbit facilities are such that spontaneous infectious processes should only be infrequently encountered; thus, the lesions related to infectious respiratory tract diseases are not described in detail in this document.

A. Anatomy of the Nasal Cavity

The nostrils of rabbits contain sensory pads at the entrance, making the nose very sensitive to touch¹⁶⁸. The nostrils are still when the rabbit is relaxed but can twitch at up to 150 twitches per minute. Rabbits are obligate nose breathers because the epiglottis is positioned rostrally to the soft palate, resulting in direct continuity of the nasopharynx, larynx, and trachea^{25, 168}.

The anatomy and histology of rabbit nasal cavity tissues have been described previously¹⁶⁹. Four types of epithelia line the nasal cavity: squamous, transitional, respiratory and olfactory, and their distribution at various levels of the nasal cavity is described¹⁶⁹, along with recommended sectioning planes to be used for inhalation studies: Level I is sectioned immediately posterior to the incisors, Level II at the first palatal ridge, Level III immediately anterior to the first upper premolar teeth, and Level IV immediately anterior to the first upper molar. Level I is lined predominantly by squamous epithelium with small amounts of thick transitional epithelium, and examination is recommended only for studies involving test article administration via instillation. Level II is lined primarily with transitional and respiratory epithelia, whereas Levels III and IV are lined with respiratory and olfactory epithelia, and often contain nasal-associated lymphoid tissue. The vomeronasal organs are evident only in Level II (Table 42).

Cleft Palate

Comments: Cleft Palate is a common finding in rabbit teratogenicity studies. Congenital alveolar cleft is a malformation occurring as a result of non-fusion of primary palate during weeks 4–12 of gestation, and may be induced by glucocorticoids³¹ (Table 43).

B. Anatomy of the Larynx

Iatrogenic damage may be noted after intubation, and so care should be taken in interpreting short term studies where surgical preparations have taken place. Intubation damage has been described previously¹⁷⁰. Submucosal glands are scanty or absent in the larynx^{171–173} (Table 44).

C. Anatomy of the Trachea and Bronchi

The upper (tracheobronchial) airways have very scanty mucous cells and relatively abundant club cells. For example, in rabbits, mucous cells constitute less than 2% of tracheal epithelium, while club cells constitute 17–25% of the upper airway epithelium¹⁷². Submucosal glands are scanty or absent in the trachea, and bronchi^{171–173} (Table 45).

D. Anatomy of the Lungs (Alveoli and Bronchioles)

The left lung consists of only two lobes (cranial and caudal), whereas the much larger right lung has four lobes (cranial, middle, caudal, and accessory)174. Multiple generations of distal intrapulmonary airways (bronchioles) branch to the level of terminal bronchioles (respiratory bronchioles are not present)175, 176. Rabbit blood vessels are generally thin-walled and prone to collapse and hematoma formation on puncture a feature important to remember for studies where test items have been given by intravenous administration. The exceptions to this are the pulmonary arteries which are enveloped in a prominent smooth muscle layer, which may be misinterpreted as hypertrophy^{24, 25}. There is a relative paucity of bronchus associated lymphoid tissue (BALT) in laboratory rabbits compared to other species. BALT is acquired and increases with age and environmental antigen exposure. Due to modern husbandry systems, relatively little BALT is seen in the young animals on toxicology studies.

In rabbits (like other laboratory species and humans), ciliated cells constitute a high percentage (40–55%) of airway lining epithelium at all levels from the trachea to the distal bronchioles^{172, 176}. In rabbits club cells (formerly known as Clara cells) are the predominant secretory cell of the distal airways (club cells and ciliated cells constitute about 50% each of the distal airway epithelium of rabbits)¹⁷². However, the upper (tracheobronchial) airways have very scanty mucous cells and relatively abundant club cells. For example, in rabbits, mucous cells constitute less than 2% of tracheal epithelium, while club cells constitute 17–25% of the upper airway epithelium¹⁷². Submucosal glands are scanty or absent in the trachea, larynx, and bronchi, and absent in bronchioles^{171–173} (Table 46).

Atherosclerosis

Other term(s): Plaque, atheromatous

Comments: Cholesterol-rich diets have been used to induce wide-spread atheromatous lesions within a short time period (3 months) in NZW rabbits. Early foam cell accumulation to partly occlusive atheromatous plaques were observed in the larger lung arteries²⁸. Unique proliferations are seen as age related changes in the pulmonary arteries of both sexes and can resemble iatrogenic lesions¹⁷⁷. Genetically altered strains of the NZW rabbit such as the Watanabe rab-

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Cleft palate *		Х		
Non-proliferative				
Abscess		Х		
Amyloid		Х		
Angiectasis		Х		
Apoptosis [‡]		Х		
Atrophy		Х		
Congestion	Х			
Corpora amylacea		Х		
Degeneration		Х		
Deviation, nasal septum		Х		
Edema		Х		
Embolus		Х		
Eosinophilic globules		Х		
Erosion/ulcer		Х		
Hemorrhage		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation [insert appropriate cell type]		Х		
Metaplasia		Х		
Necrosis		Х		
Perforation, septum		Х		
Regeneration		Х		
Single cell necrosis [‡]		Х		
Thrombus		Х		
Proliferative Non-neoplastic				
Hyperplasia, (+ cell type)		Х		
Proliferative Neoplastic				
Adenoma			Х	
Papilloma, squamous cell			Х	
Tumor, neuroendocrine cell, benign			Х	
Adenocarcinoma			Х	
Carcinoma, adenosquamous			Х	
Carcinoma, neuroepithelium			Х	
Carcinoma, squamous cell			Х	

Table 42. Microscopic Findings of the Nasal Cavity: Rabbit

bit (Watanabe heritable hyperlipidemia rabbit, WHHL) are also used extensively.

Fibrosis

Comments: A rabbit model of pulmonary fibrosis due to pneumoconiosis induced by a single instillation of a known amount of silica dust into the right lung, is available^{178–180}. Affected lungs show diffuse areas of increased alveolar macrophages containing dark pigment (dust particles) and/ or vacuoles, with few associated areas of alveolar wall thickening and increased reticular fibers. In this model, severity/distribution of unilateral fibrosis, is scored for each lung lobe.

Hernia, Diaphragmatic

Comments: Fox and Crary reported 55 cases of left diaphragmatic hernia in three related strains of rabbits kept at the Jackson Laboratory under normal colony conditions: strains AX, AX_{bubu} and III_c^{181} . Associated abnormalities included hypoplasia of the ipsilateral lung and an increased incidence of ventricular septal defects. Death was attributable to respiratory insufficiency. Genetic analysis suggests recessive inheritance. The condition is neither sex limited nor sex linked. The authors believe that two autosomal recessive genes are involved and have proposed the symbols *dh 1* and *dh 2* for the two genes that must both be present in homozygous condition for the development of diaphragmatic hernia in these rabbit strains.

 Table 43.
 Microscopic Findings of the Paranasal Sinuses and Nasopharynx: Rabbit

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Non-proliferative				
Apoptosis [‡]		Х		
Congestion		Х		
Edema		Х		
Erosion/ulcer		Х		
Hemorrhage		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Necrosis		X		
Single cell necrosis ⁺		X		
Proliferative Non-neoplastic				
Hyperplasia, (+ cell type)		Х		
Metaplasia, squamous cell		Х		
Proliferative Neoplastic				
Adenoma			Х	
Papilloma, squamous cell			Х	
Adenocarcinoma			Х	
Carcinoma, adenosquamous			Х	
Carcinoma, neuroepithelium			Х	
Carcinoma, squamous cell			X	

Table 44.	Microscop	ic Findings	of the Lar	ynx: Rabbit
		0		

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Non-proliferative				
Abscess		Х		
Apoptosis [‡]		Х		
Congestion	Х			
Degeneration		Х		
Ectasia, submucosal glands		Х		
Edema		Х		
Epithelial alteration			Х	
Erosion/ulcer		Х		
Hemorrhage		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Necrosis		Х		
Regeneration		Х		
Single cell necrosis [‡]		Х		
Proliferative Non-neoplastic				
Hyperplasia, (+ cell type)		Х		
Metaplasia, squamous cell		Х		
Proliferative Neoplastic				
Papilloma			Х	
Tumor, neuroendocrine cell, benign			Х	
Adenocarcinoma			Х	
Carcinoma, squamous cell			Х	
Tumor, neuroendocrine cell, malignant			Х	

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Non-proliferative				
Apoptosis ⁺		Х		
Bronchiectasis		Х		
Congestion	Х			
Degeneration		Х		
Ectasia, submucosal glands		Х		
Edema		Х		
Erosion/ulcer		Х		
Fibrosis		Х		
Hemorrhage		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Aggregates, macrophage, increased		Х		
Metaplasia		Х		
Necrosis		Х		
Regeneration		Х		
Single cell necrosis		Х		
Proliferative Non-neoplastic				
Hyperplasia, (+ cell type)		Х		
Metaplasia, squamous cell		Х		
Proliferative Neoplastic				
Papilloma			Х	
Tumor, neuroendocrine cell, benign			Х	
Adenocarcinoma			Х	
Carcinoma, squamous cell			Х	
Tumor, neuroendocrine cell, benign/malignant			Х	

Table 45. Microscopic Findings of the Trachea and Bronchi: Rabbit

Infiltrate, Inflammatory Cell (Figures 34, 35)

Comments: Peribronchial/perivascular infiltrates of granulocytes and mononuclear cells are commonly seen around larger airways and their associated vessels as an incidental background finding in rabbits on routine toxicology studies⁸¹. Should be differentiated from Bronchus associated lymphoid tissue (BALT).

Inflammation

Comments: Cooper *et al.* reported a congenital surfactant pneumonia in the audiogenic (EIII/JC) strain of rabbits¹⁷⁷. This pneumonia was frequently grossly visible as irregular firm tan nodules in the cranioventral portions of lung lobes. Histologically the lesions consisted of multifocal to coalescing intra-alveolar aggregates of large numbers of multinucleate giant cells, predominantly foreign body type, with epithelioid and foamy macrophages and few lymphocytes, plasma cells and heterophils. Lesions occasionally extended into smaller bronchioles. There was frequent type II pneumocyte hyperplasia (adenomatosis) and alveolar septal fibrosis. "Billups bodies" – globular to ring-like brown to gray acellular material – was free within the alveoli and cytoplasm of giant cells. This acellular material was Alcian

blue and PAS positive. The material was immunoreactive for surfactant protein-A and had the ultrastructural appearance of multilamellar vesicles, suggesting a genetic defect in surfactant metabolism.

Macrophages Increased, Alveolar (Figure 36)

Comments: Tissue resident macrophages are the first responders to insults to tissues, and the "aggregates" are generally proliferation or hyperplasia of these responding macrophages. Barros *et al.* reported an infiltration of the mucosa and submucosa of the trachea and bronchi by macrophages, multinucleated giant cells, lymphocytes, and mast cells with associated basal lamina calcium deposits, in six rabbits after intoxication by the calcinogenic plant *Solanum glaucophyllum*¹⁸². Increased diffusely distributed macrophages may also occur in Dutch Belted rabbits that relatively frequently show cardiomyopathy of unclear origin.

Metaplasia, Osseous (Figure 37)

Comments: Osseous metaplasia is an incidental spontaneous background finding in rabbit lungs, which has a differing appearance depending on the stage of the lesion. In early stages, they are composed of a dense knot of small

 Table 46.
 Microscopic Findings of the Lungs (Alveoli and Bronchioles): Rabbit

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Cyst, congenital			Х	
Diaphragmatic hernia *		X		
Hypoplasia			Х	
Non-proliferative				
Abscess		Х		
Alveolar emphysema		_	Х	
Alveolar lipoproteinosis			Х	
Apoptosis [‡]		X		
Atelectasis		X		
Atherosclerosis *		X		
Bronchiectasis		Х		
Congestion	Х			
Degeneration		X		
Ectasia, acinus/submucosal glands		X		
Edema		X		
Embolus		X		
Erosion/ulcer		X		
Fibrosis *		X		
Hemorrhage		X		
Hypertrophy, media, artery		X		
Infarct		X		
Infiltrate, inflammatory cell [insert appropriate cell type] *		X		
Inflammation *		X		
Linoproteinosis alveolus		X		
Macrophages increased, alveolar *		X		
Material extracellular [insert morphology/color]		X		
Metaplasia mucous cell/squamous cell		X		
Metaplasia osseous *	x			
Mineralization		x		
Necrosis		X		
Pigment		X		
Pigment/foreign material		X		
Pyothorax		X		
Regeneration		X		
Single cell necrosis [†]		X		
Thrombus *		X		
Proliferative Non-neonlastic		A		
Hypernlasia (+ cell type)		x		
Metanlasia mucous/squamous cell		X		
Proliferative Neoplastic		A		
Adenoma bronchioloalveolar			x	
Panilloma			X	
Tumor neuroendocrine cell henign			X X	
Adenocarcinoma			X	
Carcinoma acinar			 V	
Carcinoma, adenosquamous			X X	
Carcinoma, aucnosquamous				
Carcinoma, solamous cell			X	
Histiocytic sarcoma			X	
1115100 jule bareonia			2 1	

* Terminology with diagnostic criteria or comments described in the text. [‡] Refer to ⁴ for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

cells with hyperchromatic nuclei and scant eosinophilic cytoplasm within the wall of an alveolus. This progresses to a larger, sometimes papillomatous mass with a fibrous core and a dark, low spindle to cuboidal epithelial-like covering. As the lesion progresses, it changes into eosinophilic to basophilic, osteoid to mineralized bone, resembling the lesion classically seen in rodent lungs.

Thrombus (Figure 38)

Comments: Small thrombi, with an accompanying arteritis/periarteritis, are often seen as incidental findings in the lungs of rabbits on routine toxicity studies (Table 47).

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Cyst, congenital			Х	
Non-proliferative				
Abscess		Х		
Apoptosis [‡]		Х		
Effusions, non-inflammatory		Х		
Fibrosis		Х		
Hemorrhage		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Macrophages increased		Х		
Metaplasia		Х		
Mineralization		Х		
Pigment		Х		
Pigment/foreign material		Х		
Pyothorax		Х		
Single cell necrosis ⁺		Х		
Proliferative Non-neoplastic				
Hyperplasia, mesothelium			Х	

Table 47. Microscopic Findings of the Pleura: Rabbit

Section 14: Skeletal System (Bone, Joint, Tooth)

A. Anatomy of Bone

Compared to rodents, rabbit bone has a larger proportion of osteonal structures, with dense osteons within the center of cortical bone and a primary vascular structure running longitudinally¹⁸³. Rabbits also have more cancellous bone than rodents and this cancellous bone shows a higher degree of remodelling into secondary bone¹⁸⁴. The secondary bone retains the primary vascular structure with vascular canals of osteons on the periosteal and endosteal surfaces running parallel to the long axis of the medullary cavity^{183, 185}. Compared to rodents, rabbits have relatively more trabecular bone mass that undergoes Haversian remodelling¹⁸⁴. Rabbits do not have secondary centers of ossification present at birth¹⁸⁶. NZW rabbits typically reach skeletal maturity at 8-11 months¹⁸⁷, with cartilage developing a mature appearance prior to epiphyseal closure¹⁸⁸. Rabbits have significant cartilage healing ability at an early age¹⁸⁹, but periosteal support of cartilage declines markedly from 6-12 months and is very variable after one year¹⁹⁰. Rabbit femoral condyle cartilage is relatively thin with 0.25-0.75 mm compared to 2.2-2.4 mm in humans¹⁸⁸.

Skull

Rabbit tympanic bullae are relatively larger than those of most mammals and rabbits have a longer and more tubular external auditory meatus. The orbits are very large, and joined by an interorbital foramen. The zygomatic arch has a prominent protuberance, called the zygomatic process that is at the caudal aspect of the arch³⁰. The maxilla is fenestrated giving a lacy appearance on radiographs. Radiographic anatomy of the rabbit skull has been well described^{191, 192}.

Vertebrae and ribs

The most common vertebral formula is C7 T12 L7 S4 Ca16, but this can vary widely in the thoracolumbar region. A recent report found that 44% of rabbits had 12T/7L, 33% had 13T/6L, and 23% had 13T/7L. Additionally, the spinal cord terminated with S2 in 79% of rabbits but in S1 in 19% and in L3 in 2% of animals¹⁹³. The lumbar vertebrae have prominent mammillary processes on the cranial articular process. Rabbits are unique among domestic animals for having the dorsal aspect of the lumbar vertebral mammillary processes level with and slightly ventral to the spinous process. S1–S3 are routinely fused in rabbits, while S3–S4 fusion is more variable³⁰. The first 7 ribs articulate with the sternum and the final 5 are free. The costal cartilage sections of the 7th– 9th ribs are attached. The sternum includes 6 sternebrae³⁰.

Limbs

Rabbits are digitigrade.

Forelimb

The rabbit has paired clavicles and the only direct attach-

ment between the forelimb and the axial skeleton is the sternoclavicular ligament. The carpus includes two rows of carpal bones. In the 5 digits in the forelimb, P1 has two phalanges and P2–P5 have three phalanges³⁰.

Hindlimb

The femur articulates only with the tibia. The fibula fuses distally with the tibia for approximately half of its length. Six tarsal bones are arranged in three rows. P1–P5 all have three phalanges³⁰. The rabbit stifle joint is considered to have minimal to no resemblance to the human stifle anatomy¹⁹⁴.

Non-hematopoietic marrow

Rabbits have relatively fatty bone marrow which is not an ideal for autogenous bone and marrow harvesting or transplantation¹⁸⁴ (Table 48).

Malformation, Skeletal

Comments: Syringomyelia, hypoplasia pelvis, femoral luxation, and distal foreleg curvature are occasionally seen together as a hereditary defect in fetuses in Developmental and Reproductive Toxicology (DART) studies.

Bone Increased, Trabeculae

Comments: Osteopetrosis has been recorded in Dutch Belted rabbits; feed formulation errors have historically caused vitamin A toxicity-related increased bone deposition, sometimes leading to hydrocephalus¹⁹⁵.

Necrosis (Figures 39, 40)

Comments: Osteonecrosis can be induced with steroid therapy, as there may be infarction from fat or lipid-laden fibrin and platelet-containing emboli^{196, 197}.

Fracture

Comments: The combination of a light skeleton (6–8% of body weight ^{24, 30}) and ample skeletal muscle (>50% of body weight) predisposes rabbits to vertebral, sometimes even spontaneous, fracture^{24, 195}, often at the 7th lumbar vertebrae³⁰. Lumbosacral fractures may also be seen as sequelae to convulsions/seizures induced by CNS active test articles. Avulsion of the tuberositas tibiae has been reported¹⁹⁸.

Hyperplasia, Chondrocyte

Comments: Chondrocyte hyperplasia, cartilage degeneration and necrosis are seen with fluoroquinolone antibiotics¹⁹⁹.

B. Anatomy of Joints

Spontaneous lesions of the joints and synovium are rare in rabbits used in nonclinical toxicology studies. The relative lack of changes in the joints of the rabbit is most likely related to the young age of the animals used. The most frequently used trimming plane to prepare femorotibial joints for microscopic eval-

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Malformation, skeletal *		Х		
Non-proliferative				
Apoptosis [‡]		Х		
Bone decreased, trabeculae and/or cortex		Х		
Bone increased, trabeculae and/or cortex *		Х		
Callus		Х		
Cyst, bone		Х		
Eroded surface, increased		Х		
Fibro-osseous lesion (FOL)				Х
Fibrous osteodystrophy (FOD)		Х		
Fracture *		Х		
Fracture/Callus		Х		
Growth plate closed		Х		
Growth plate partially closed		Х		
Growth plate open		Х		
Hemorrhage		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Necrosis *		Х		
Osteoblastic surface increased		Х		
Osteoclasts increased		Х		
Osteoid increased		Х		
Physeal dysplasia		Х		
Physis, decreased thickness		Х		
Physis, increased thickness		Х		
Single cell necrosis ⁺		Х		
Proliferative Non-neoplastic				
Hyperplasia, chondrocyte *		Х		
Hyperplasia, osteoblast, focal		Х		
Proliferative Neoplastic				
Chordoma			Х	
Chondroma			Х	
Osteoma			Х	
Osteoblastoma			Х	
Osteofibroma			Х	
Chondrosarcoma			X	
Fibrosarcoma, osteogenic			Х	
Osteochondroma			Х	
Osteosarcoma			X	

Table 48. Microscopic Findings of Bone: Rabbit

* Terminology with diagnostic criteria or comments described in the text. + Refer to 4 for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

uation is parasagittal orientation (taken off-center through one femoral condyle), a portion of meniscus, and the corresponding tibial plateau.

Typical scoring systems used in toxicology studies are the International Cartilage Research Society (ICRS) visual Histological Assessment Scale²⁰⁰ and the Osteoarthritis Research Society International (OARSI) score²⁰¹. These are more appropriate for use in animal models than the Mankin scoring system used in man (Table 49).

Degenerative Joint Disease (DJD)

Comments: Induced models are used for research i.e. partial lateral meniscectomy²⁰².

Table 49. Microscopic Findings of t	the Joint and Synovium: Rabbit
-------------------------------------	--------------------------------

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Non-proliferative				
Abscess		Х		
Apoptosis [‡]		Х		
Degeneration, articular cartilage		Х		
Degeneration, chondromucinous		Х		
Degenerative Joint Disease (DJD) *		Х		
Hemorrhage		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Metaplasia		Х		
Necrosis		Х		
Osteophyte		Х		
Single cell necrosis [‡]		Х		
Proliferative Non-neoplastic				
Hyperplasia, synovial cell		Х		
Proliferative Neoplastic				
Sarcoma, synovial			Х	

* Terminology with diagnostic criteria or comments described in the text. + Refer to 4 for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

C. Anatomy of Teeth

The maxilla contains 4 incisors, no canines, six premolars, and 4–6 molars. The mandible contains 2 incisors, no canines, 4 premolars, and 6 molars. Unlike rodents, rabbits have two pairs of maxillary incisors caudal to the main incisors ("peg teeth") and have a total of 26–28 teeth. A slight degree of brachygnathism is normal in the rabbit so that the large pair of inferior incisors usually contact the small superior pair during occlusion. The large incisor teeth are adapted for gnawing and continue to erupt throughout life. There are no canine teeth and a large diastema exists between the incisors and premolars. Incisors consist of crown only, with extra-alveolar and intraalveolar parts. Rabbits are hypsodonts and have a long crown without a true tooth root³⁰.

The labial or convex side of the incisors is covered by a layer of enamel. The lingual or concave side of the incisors is enamel-free but does have a very thin layer of cementum into which fibers of the periodontal ligament are embedded. Enamel is not formed over the top of the incisors. Before eruption, the tip is filled with dentin produced by odontoblasts of the pulp. As the tip wears away with use, the odontoblasts form more dentin (secondary dentin) so that the pulp is never exposed. Incisors have a widely open apical foramen (Table 50).

Malocclusion

Comments: Erosions of the mucosa may occur due to irregular growth or sharp edges of broken teeth.

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Malocclusion *	Х			
Non-proliferative				
Abscess		Х		
Alteration, dentin matrix			Х	
Apoptosis [‡]		Х		
Concretion, pulp			Х	
Cyst			Х	
Degeneration		Х		
Dental dysplasia		Х		
Denticle			Х	
Dentin matrix alteration			Х	
Dentin, decreased			Х	
Dentin, niches			Х	
Fracture		Х		
Necrosis		Х		
Periodontal pocket			Х	
Pulp concretion			Х	
Resorption			Х	
Single cell necrosis ⁺		Х		
Thrombus		Х		
Proliferative Neoplastic				
Ameloblastoma			Х	
Ameloblastic odontoma			Х	
Fibroma, cementifying/ossifying			Х	
Fibroma, odontogenic			Х	
Odontoma			Х	
Tumor, odontogenic, benign			Х	
Tumor, odontogenic, malignant			X	

Table 50. Microscopic Findings of the Tooth: Rabbit

* Terminology with diagnostic criteria or comments described in the text. ⁺ Refer to ⁴ for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

Section 15: Soft Tissue (Soft Tissue, Mesothelium, Adipose, Skeletal and Smooth Muscle)

The pathology of both spontaneous and induced conditions of soft tissues are similar in rabbits and humans, hence their use in vaccine studies. Careful recording of the nature, intensity and duration of the inflammatory response of the soft tissues to implanted or injected substances is important in the assessment of the local tolerability of agents intended for contact with human tissues. The chemical and physical properties of injected chemicals or vaccines and their adjuvants as well as size, shape and surface texture of implanted biomaterials may modify the histological features and temporal pattern of the inflammatory and reparative responses^{91–94}. Such studies in rabbits are conducted with descriptors following the ISO-10993 guidelines (Table 51).

A. Anatomy of Adipose Tissue

Brown fat is converted to white fat in the interscapular region of the rabbit as it ages, which correlates with the disappearance of catecholamines from the sympathetic nerve fibers in arterial blood vessels in both brown and white fat²⁰³. The peritoneal mesothelium of the anterior abdominal wall of the rabbit is characterized by flattened mesothelial cells with tight junctions, desmosomes, cytoplasmic pinocytic vesicles and microvilli²⁰⁴. It has been shown that adipose tissue may have an important paracrine function in smooth muscle cell proliferation in blood vessels²⁰⁵ (Table 52).

Hyperplasia

Comments: Hyperplasia of adipose tissue (obesity) has been reported associated with pregnancy toxemia, but also with ketosis in non-pregnant NZW rabbits²⁴.

B. Anatomy of Muscle

The distribution of lesions of smooth muscle generally parallels the normal distribution of smooth muscle in the body so that lesions occur mostly in the female genital tract, the gastrointestinal tract and skin but only rarely in deep soft tissue. However, it has been shown that adipose tissue may have an important paracrine function in smooth muscle cell proliferation in blood vessels²⁰⁵. In soft tissues, particularly in inflammatory processes, smooth muscle cells may be difficult to distinguish from myofibroblasts. They both express smooth muscle actin but smooth muscle cells usually contain desmin²⁰⁶ (Table 53).

Atrophy

Comments: Destruction and denervation atrophy in skeletal muscle caused by injection of local anesthetics i.e. lidocaine or bupivacaine is reported in the muscles adjacent to the facial nerve in rabbits²⁰⁷.

Degeneration

Comments: Vitamin E deficiency causes skeletal muscle hyaline degeneration and mineralization, as does ketamine and xylazine intramuscular injection in Dutch Belted rabbits. The preferred site for intramuscular injection is the dorsal lumbar muscle. Alum granulomas or minimal focal myofiber degeneration are often seen at vaccination sites^{81, 208}.

Mineralization

Comments: Hypervitaminosis D as well as vitamin E deficiency cause widespread mineralization²⁰⁹.

Necrosis

Comments: Afifi *et al.* reported muscle necrosis seen with 1,1'-methylenebis[4-[(hydroxyimino)methyl]-pyridinium] dimethanesulfonate intramuscular injection to NZW rabbits²¹⁰. Sequestration of non-viable fat tissue (fat necrosis) transplanted into NZW rabbits has been reported²¹¹. Abscesses are common in the skin of domestic rabbits¹¹⁰. Rabbits restrained for six hours a day for 35 days developed focal, small necrotic areas in the skeletal muscle²¹².

Parasite

Comments: Sarcocystosis, presumably caused by *Sarcocystis cuniculi*, was recently reported in 2 purpose-bred, SPF Dutch Belted laboratory rabbits²¹³. Sarcocysts were found in the eyelid of one rabbit and the tongue of the other (Table 54).

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Non-proliferative				
Abscess		Х		
Amyloid		Х		
Apoptosis [‡]		Х		
Atrophy		Х		
Degeneration		Х		
Fibroplasia		Х		
Fibrosis		Х		
Hemorrhage		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Metaplasia, osseous/cartilaginous		Х		
Mineralization		Х		
Necrosis		Х		
Single cell necrosis [‡]		Х		
Proliferative Non-neoplastic				
Hyperplasia		Х		
Proliferative Neoplastic				
Fibroma			Х	
Hibernoma			Х	
Leiomyoma			Х	
Lipoma			Х	
Rhabdomyoma			Х	
Fibrosarcoma			Х	
Leiomyosarcoma			Х	
Liposarcoma			Х	
Mesenchymoma, malignant			Х	
Rhabdomyosarcoma			Х	
Sarcoma, NOS (Not otherwise specified)			X	

Table 51. Microscopic Findings of the Soft Tissue: Rabbit

Table 52.	Microscopic	Findings of	the Adipose	Tissue: Rabbit
		2		

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Non-proliferative				
Abscess		Х		
Apoptosis ⁺		Х		
Atrophy		Х		
Degeneration		Х		
Fibrosis		Х		
Hemorrhage		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Inflammation, lipogranulomatous		Х		
Mineralization		Х		
Necrosis		Х		
Single cell necrosis [‡]		Х		
Proliferative Non-neoplastic				
Hyperplasia *		Х		
Proliferative Neoplastic				
Hibernoma			Х	
Lipoma			Х	
Liposarcoma			Х	
Sarcoma, NOS (Not otherwise specified)			Х	

* Terminology with diagnostic criteria or comments described in the text. [‡] Refer to ⁴ for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

 Table 53.
 Microscopic Findings of the Smooth and Skeletal Muscle: Rabbit

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Non-proliferative				
Abscess		Х		
Apoptosis [‡]		Х		
Atrophy *		Х		
Degeneration *		Х		
Fibrosis		Х		
Hemorrhage		Х		
Hypertrophy		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Metaplasia, osseous/cartilaginous		Х		
Mineralization *		Х		
Necrosis *		Х		
Parasite *		Х		
Single cell necrosis ⁺		Х		
Vacuolation, myocyte		Х		
Proliferative Non-neoplastic				
Hyperplasia		Х		
Proliferative Neoplastic				
Rhabdomyoma			Х	
Rhabdomyosarcoma			Х	
Sarcoma, NOS (Not otherwise specified)			Х	

* Terminology with diagnostic criteria or comments described in the text. ⁺Refer to ⁴ for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Non-proliferative				
Abscess		Х		
Apoptosis [‡]		Х		
Fibrosis		Х		
Fibroplasia		X		
Hemorrhage		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Metaplasia		Х		
Single cell necrosis ⁺		Х		
Proliferative Non-neoplastic				
Hyperplasia		Х		
Proliferative Neoplastic				
Mesothelioma, malignant			Х	

 Table 54.
 Microscopic Findings of the Mesothelium: Rabbit

SECTION 16: SPECIAL SENSES

The otic (ear) and olfactory systems are not routinely evaluated in toxicity studies in the rabbit. The ocular system is subdivided into the eye and the ocular adnexa (glands and other eye-associated tissues). At necropsy, Davidson's fixative is usually the fixative of choice for the eyes and optic nerves. For a thorough examination of the ocular system, at a minimum, the eyes, optic nerves, and major ocular adnexa (eyelids, lacrimal glands, nictitating membrane with nictitans gland, and Harderian glands) should be examined.

A. Anatomy of the Eye

At birth, the eye is developmentally immature, and differentiation continues and is substantially complete when the eyelids open at about post-natal days $10-11^{214-218}$.

The rabbit eye is relatively large in proportion to body size and slightly flattened along the antero-posterior plane²¹⁷. The circular, rather shallow orbits are at an approximately right angle to the sagittal plane of the head, so the eyes protrude prominently^{217, 219–222}. Each of the eyes of the rabbit has approximately 190-degree field of view, but they also work together to have a small amount of binocular vision. Because of their lateral placement, the angle between the eyes is 150–175 degrees, and the overall visual field is extremely wide (330 – to almost 360 degrees)^{217, 219–224}. In contrast, the binocular field of vision is correspondingly quite narrow (10–35 degrees wide)²²¹.

B. Anatomy of the Eyelids

Rabbits are born blind with closed eyelids, which open at about post-natal days 10–11. The superior eyelid (palpebra) of the rabbit is shorter and thicker and has larger and more abundant eyelashes than the inferior eyelid²¹⁹. The outer surfaces of

the eyelids are lined by typical keratinized stratified squamous epithelium. As in other skin regions of the rabbit, the eyelid skin lacks sweat glands.

Meibomian (tarsal) glands are present in the superior and inferior eyelids, embedded as linear arrays in the tarsal plate (Figure 41). In one study of NZW rabbits, Meibomian gland numbers/eyelid averaged from 32.30 +/- 3.43 and 27.15 +/- 2.35 in the superior and inferior eyelids, respectively²²⁵. In each Meibomian gland, the sebaceous holocrine acini are aligned along and empty via short ductules into a single central duct via short ductules. The central duct in turn opens at the conjunctival margin^{219, 226}. The central excretory duct is normally lined by keratinized stratified squamous epithelium^{226, 227}.

Small clusters of accessory lacrimal gland acini (glands of Wolfring) have been described in the superior eyelid of rabbits²²⁸ (Figure 42). These are located submucosally, and anterior to the Meibomian gland arrays (Figures 43, 44).

Extraocular muscles involved in movement of the eyelids include the levator superior palpebrae and orbicularis oculi^{219, 229–233}, as well as the depressor palpebrae inferioris (present in rabbits but not in most other mammalian species)^{220, 234, 235} (Table 55).

C. Anatomy of the Conjunctiva

Compared to humans and many other species, the precorneal tear film of rabbits is thicker, more stable, and has a substantially different composition (i.e., types and proportions of lipid and mucins)^{236–239}.

Conjunctival submucosal lymphoid cell aggregates, with or without germinal centers (conjunctiva-associated lymphoid tissue [CALT]) are normally present in rabbits^{240–242}. CALT aggregates are more abundant in the palpebral and fornical conjunctivae versus the bulbar conjunctiva. They are also more abundant in the inferior than superior eyelid^{219, 240, 242}. CALT

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Non-proliferative				
Abscess		Х		
Apoptosis ⁺		Х		
Atrophy, Meibomian gland		Х		
Congestion		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation (+ locator)		Х		
Metaplasia		Х		
Single cell necrosis ŧ		Х		
Proliferative Non-neoplastic				
Hyperplasia, squamous cell			Х	
Proliferative Neoplastic				
Papilloma, squamous cell			Х	
Carcinoma, squamous cell			Х	

Table 55. Microscopic Findings of the Eyelid: Rabbit

aggregates are absent in neonatal rabbits, develop rapidly after the eyes open (at about post-natal day 10–11), reach maximum numbers and size in mature rabbits (7–20 months), and then decline in aging animals²⁴⁰ (Table 56).

Hyperplasia, Conjunctiva

Other terms: *Pseudopterygium; aberrant conjunctival overgrowth; ankyloblepharon; circumferential conjunctival hyperplasia; conjunctival centripetalization; conjunctival hyperplasia; conjunctival stricture; corneal occlusion; epicorneal conjunctival membrane; precorneal membranous occlusion; pinguecula bilateralis; pseudosymblepharon; pterygium; pterygium conjunctivae.*

Comments: Infrequently, the rabbit cornea is progressively covered by hyperplastic conjunctiva around the entire perimeter. Of unknown etiology, the condition appears to be unique to rabbits^{222, 243–248}. Conjunctival hyperplasia can be unilateral or bilateral, and is characterized clinically by a circumferential, nonadherent membranous flap which arises from the perilimbal bulbar conjunctiva and grows centripetally and symmetrically across the anterior corneal surface. Microscopically, the membranes consist of a collagenous central stroma lined by conjunctival mucosa²⁴⁷.

D. Anatomy of the Cornea

The rabbit cornea is relatively large and slightly elliptical (slightly longer along the horizontal axis)^{219–221, 243}. Although there are breed and age variations, central corneal thickness in

the living rabbit ranges from about $0.35-0.44 \text{ mm}^{249-256}$, which is thinner than the human cornea $(0.53-0.58 \text{ mm})^{257}$.

Most authors concur that the rabbit corneal stroma lacks a Bowman's layer, a subepithelial condensation of the collagenous stroma present in humans and certain other species^{219, 258–261}. Differently from rats and mice, the nuclei of rabbit corneal keratocytes are visible as hyperreflective structures. Thus, their density can be easily evaluated. Endothelial cell density is higher than in rats and mice, and the endothelium regenerates in response to injury or loss^{262, 263} (Table 57).

Attenuation, Endothelium

Comments: Unlike many other species, the rabbit corneal endothelium has rather robust capacity to proliferate to cover defects or loss of individual endothelial cells. Following insult, the rabbit corneal endothelium generally undergoes endothelial proliferation and regenerates, with occasional multinucleate cell formation, rather than endothelial attenuation²⁶³.

Dermoid, Cornea

Comments: A case of a corneal dermoid in a dwarf rabbit²⁶⁴, and a limbic dermoid in a NZW rabbit²⁶⁵ have been reported. Although uncommon, dermoids should be on the list of differential diagnoses for corneal masses in rabbits. Animals exhibiting these changes on pre-study examination should be removed from the study cohort before dosing commences.

Table 56. Microscopic Findings of the Conjunctiva: Rabbit

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Non-proliferative				
Abscess		Х		
Apoptosis [‡]		Х		
Atrophy, epithelium		Х		
Cyst, inclusion		Х		
Dermoid		Х		
Edema		Х		
Erosion/ulcer		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]	Х			
Inflammation	Х			
Metaplasia		Х		
Pigment		Х		
Single cell necrosis ⁺		Х		
Proliferative Non-neoplastic				
Hyperplasia, conjunctival *		Х		
Hyperplasia, squamous cell			Х	
Proliferative Neoplastic				
Papilloma, squamous cell			Х	
Carcinoma, squamous cell			Х	

* Terminology with diagnostic criteria or comments described in the text. + Refer to 4 for diagnostic criteria and use of the terms apoptosis and single cell necrosis.
| Finding | Common | Uncommon | Not Observed but
Potentially Relevant | Not Applicable |
|--|--------|----------|--|----------------|
| Congenital | | | | |
| Dermoid, corneal * | | Х | | |
| Non-proliferative | | | | |
| Abscess | | Х | | |
| Apoptosis ‡ | | Х | | |
| Atrophy | | Х | | |
| Attenuation endothelium * | | Х | | |
| Cyst, inclusion | | Х | | |
| Degeneration | | Х | | |
| Descemetocele | | Х | | |
| Dystrophy, corneal * | | Х | | |
| Edema | | Х | | |
| Erosion/ulcer | | Х | | |
| Fibroplasia | | Х | | |
| Fibrosis | | Х | | |
| Hypertrophy, Descemet's membrane | | Х | | |
| Infiltrate, inflammatory cell [insert appropriate cell type] | | Х | | |
| Inflammation | | Х | | |
| Keratinization | | Х | | |
| Metaplasia | | Х | | |
| Mineralization * | | Х | | |
| Neovascularization | | Х | | |
| Necrosis | | Х | | |
| Single cell necrosis [‡] | | Х | | |
| Vacuolation, lipid, cornea *# | | Х | | |
| Vacuolation, epithelium or endothelium | | Х | | |
| Proliferative Non-neoplastic | | | | |
| Hyperplasia, endothelium | | Х | | |
| Hyperplasia, squamous cell | | Х | | |
| Proliferative Neoplastic | | | | |
| Papilloma, squamous cell | | | Х | |
| Carcinoma, squamous cell | | | Х | |

Table 57.	Microscopic	Findings	of the	Cornea:	Rabbit
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* Terminology with diagnostic criteria or comments described in the text. # Inducible lesion. + Refer to 4 for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

Dystrophy, Cornea

Pathogenesis/cell of origin: Corneal epithelium and associated basement membrane and stroma.

Differential Diagnosis: Corneal dysplasia

Diagnostic Features: Thickened, elevated epithelium interspersed with areas of abnormally thin epithelium.

Comments: When precisely used, the term "corneal dystrophy" describes a disease process of mineralization in the cornea (especially the epithelial basement membrane), with the lesions being spontaneously occurring, non-inflammatory, usually involving the central cornea, and are often bilateral and symmetrical²⁶⁶. The Dutch Belted rabbit can exhibit such a corneal dystrophy²⁶⁶, sometimes referred to as "anterior corneal dystrophy". This is a spontaneous, possibly inherited condition characterized by clinically observed central to peripheral corneal opacities which correspond microscopically to the focal areas of epithelial basement thickening and irregularity^{266, 267}. This true corneal dystrophy is comparable to Thiel-Behnke corneal dystrophy (TBCS) caused by defects in transforming growth factor beta (TGF β) in humans²⁶⁷.

However, most reports of rabbit "corneal dystrophy" do not describe such true corneal dystrophy, because in rabbits, the term "corneal dystrophy" has been used to describe several apparently unrelated non-inflammatory conditions of uncertain etiology, which are characterized by various changes in the corneal epithelium, stroma, and/or endothelium.

For example, a morphologically different condition termed "corneal epithelial dystrophy" has been described in two

4-month-old NZW rabbits²⁶⁸. The affected animals had clinically observed unilateral circumferential corneal opacities extending from the limbus to the central cornea. Microscopically, the opacities consisted of alternating areas of corneal epithelial thinning and thickening (hyperplasia) with unremarkable basement membranes. It is unclear if this is a true corneal dystrophy or not²⁶⁸.

Another apparently distinct condition termed "pre-Descemet's membrane corneal dystrophy" has been described in adult NZW rabbits²⁶⁹. Microscopically, focal peripheral to central accumulations of ectopic corneal endothelial cells were present subjacent to Descemet's membrane, and corresponded to clinically observed corneal opacities. By electron microscopy these cells were found to be secreting matrix material based on the cells' intracytoplasmic content, presence of a dense and homogenous material associated with the outer cell membrane, location of these cells close to Descemet's membrane, and the cells' linear organization. The change was assumed to be present at birth because it was seen in 2-week-old rabbits. The nomenclature of dystrophy for this lesion reflects the nomenclature of a similar lesion in humans. No mineralization has been reported with this lesion in rabbits.

Vacuolation, Lipid, Cornea

Other terms: corneal lipidosis; lipid keratopathy

Pathogenesis/cell of origin: lipid-laden keratocytes, foamy macrophages, sterol clefts, and/or multinucleated giant cells in the corneal stroma.

Differential Diagnosis: vacuolation, mucopolysaccharides; vacuolation, NOS; edema.

Diagnostic Features: Microscopic changes in the cornea are primarily in the stroma (especially anteriorly), and include lipid-containing keratocytes and foamy macrophages, sterol clefts, and/or multinucleated giant cells, often associated with secondary neovascularizaton and inflammatory cell infiltrates^{270–273}. Similar lesions have also been described in the nictitating membrane²⁷³.

Comments: Corneal lipid vacuolation has been described in various rabbit breeds with high serum cholesterol levels due to familial predisposition (in Watanabe heritable hyperlipidemic [WHHL] rabbits ²⁷⁰ or resulting from high dietary cholesterol in various other rabbit breeds^{271, 273}. In one study of "New Zealand" rabbits fed high-cholesterol diets, the typical corneal stromal changes were present but other lesions were also observed: lipid accumulation in corneal epithelium and endothelium, as well as lipid-laden macrophages in the iris stroma, ciliary body, ciliary processes, and choroid, and increased lipid staining in the retina²⁷¹. Other studies with rabbits fed high-cholesterol diets have also demonstrated light and transmission electron microscopic effects in other regions of the globe including: lipidladen macrophages in the choroid and suprachoroid; lipid accumulation in the retinal pigment epithelium (RPE) and astrocytes; Bruch's membrane thickening; RPE hypertrophy; Müller cell and astrocyte activation; and degeneration and/or necrosis of choroidal vessel endothelium and retinal neurons^{274–277}. Instillation of cationic amphiphilic drugs in juvenile white rabbits induce corneal phospholipidosis²⁷⁸.

Mineralization

Comments: Albino and pigmented rabbits can exhibit "calcific" or "band keratopathy"-like subepithelial and stromal mineralization following corneal injury from various causes^{220, 279–281}. Hypervitaminosis D can also result in corneal mineralization, but in rabbits seems to occur only in eyes compromised by concurrent ocular inflammation^{282, 283}.

E. Anatomy of the Anterior Chamber and Aqueous Humor

Many features of the rabbit uvea (iris, ciliary body, and choroid), aqueous filtration system, and anterior chamber are related to the low accommodative ability. The rabbit anterior chamber is large, being 2.3 - fold larger than that of the cynomolgus macaque even though the eyes of both species are of similar size²⁸⁴ and having a slightly greater diameter than the much larger human eye²⁸⁵. Compared to humans, the rabbit anterior chamber is shallower and more curved due to the displacement of the iris anteriorly by the large lens^{220, 285}. Yüksel *et al.*²⁵⁶ reported that mean anterior chamber depth of young "New Zealand" rabbits is 2.08 +/- 0.16 mm, which is very similar to the 2.161 +/- 0.11 mm noted by Werner *et al.*²⁸⁵ in male NZW rabbits (Table 58).

F. Anatomy of the Filtration Angle

The rabbit iridocorneal (filtration) angle is relatively large and deep, partly because of the small ciliary muscle. Rabbits have multiple slit-like venous collector channels known as the angular aqueous plexus, rather than a singular canal of Schlemm^{233, 286–291} (Table 59).

Malformation, Filtration Angle

Other terms: Buphthalmos; buphthalmia

Comments: A type of developmental glaucoma, inherited as an autosomal recessive trait with incomplete penetrance, has long been recognized in rabbits^{24, 292–298}. This hereditary glaucoma has been most commonly recognized in NZW albino rabbits^{295, 297, 298}, but can also occur in other albino strains such as AXBU/J²⁹⁶ and in "pigmented" rabbits²⁹³. The fundamental phenotypic defect is incomplete and/or abnormal development of iridocorneal angle structures (i.e., goniodysgenesis), resulting in impaired drainage of aqueous humor from the eye²⁹⁵. Clinical signs generally become evident early in life, around 2–3 months of age or even earlier, and include elevated intraocular pressure (IOP), corneal edema, increased corneal diameter, and eventually

Table 58. Microscopic Findings of the Anterior Chamber and Aqueous Humor: Rabbit

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Non-proliferative				
Proteinaceous fluid			Х	
Inflammation			Х	
Proliferative Non-neoplastic and Neoplastic				
-				

Table 59.	Microscopic	Findings of th	e Filtration Angle:	Rabbit
			• /	

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Malformation, filtration angle *		Х		
Non-proliferative				
Narrowed filtration angle *		Х		
Single cell necrosis		Х		
Proliferative Non-neoplastic				
Proliferation, trabecular meshwork			Х	

* Terminology with diagnostic criteria or comments described in the text.

grossly detectable enlargement and excessive protrusion of the globe^{243, 292–295, 297, 298}. Most cases are bilateral, but unilateral involvement has been reported^{294, 295, 297}. Microscopically, aqueous outflow structural abnormalities include a narrowed, truncated, or absent ciliary cleft, shrunken or compressed trabecular meshwork, absent or poorly developed pectinate ligaments, and posterior displacement of the angular aqueous plexus^{292, 293, 295–299}. The ciliary body can also be hypoplastic. Associated changes can include pathologic optic nerve head cupping, optic nerve atrophy, and retinal changes ranging from decreased or degenerate ganglion cells to extensive full thickness retinal atrophy^{292, 294}.

Narrowed Filtration Angle

Comments: Experimentally induced glaucoma in rabbits has been studied as an animal model of human disease^{220, 292, 300}. Glaucomatous changes can also be secondary to ocular inflammation, trauma, and other causes²⁴⁸, and would therefore be considered acquired and having a normally formed, but possibly obstructed, filtration angle.

G. Anatomy of the Uvea (Iris, Ciliary Body and Choroid)

Many features of the rabbit uvea (iris, ciliary body, and choroid), are related to the low accommodative ability. The ciliary body is divided into the anterior pars plicata (ciliary processes and ciliary muscle) and the posterior pars plana. In keeping with the low accommodative ability, the rabbit ciliary body muscle (smooth muscle) is small and poorly developed^{219, 235, 243, 287, 291}. The pars plicata ciliary processes are radially arranged leaflike folds which arise from the anterior ciliary body and extend along the posterior iris surface. In the rabbit, long and short ciliary processes alternate, with the longer processes often interconnected by lateral, epithelium-covered stromal bridges to each other and to the posterior iris, forming the so-called "ciliary web", "circular ledge", or "sims" (Figure 45)^{220, 235, 301–304}.

The pars plana of rabbits is relatively narrow, so the lens and pars plana are in closer proximity in the rabbit compared to the cynomolgus macaque^{284, 304, 305}. The junction of the pars plana and the peripheral retina in rabbits and many non-primate species is unindented and smooth, and thus more appropriately referred to as the ora ciliaris retinae rather than as the ora serrata. The ora ciliaris retinae of rabbits is situated relatively more anteriorly than is the ora serrata of the cynomolgus macaque²⁸⁴.

The rabbit choroid exhibits increased thickness in a horizontal nasotemporal band inferior to the optic nerve head (along the retinal visual streak; see Retina section below)^{217, 220}. A choroidal tapetum lucidum is not present in rabbits²⁴³ (Table 60).

Metaplasia, Osseous

Comments: Ciliary body osseous metaplasia consists of small, irregular, non-mineralized osteoid masses in the ciliary body of otherwise unremarkable eyes. This has been noted as a rare incidental change in NZW rabbits³⁰⁶; whether it also occurs in other strains is unknown. Although the etiology is unknown, these may be minor developmental anomalies, similar to the uveal "heterotopic ossification" in other species such as guinea pigs and dogs^{307–309}. Scleral osseous metaplasia was also reported as a reaction to intraocular osteoinductive hydroxyapatite and polyethylene polymer implants in experimentally manipulated (eviscerated) eyes of NZW rabbits³¹⁰.

 Table 60.
 Microscopic Findings of the Uvea (Iris, Ciliary Body and Choroid): Rabbit

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Adhesion, iris			Х	
Hypoplasia, choroid		Х		
Hypoplasia, ciliary body		Х		
Malformation, iris			Х	
Persistent pupillary membrane		Х		
Non-proliferative				
Apoptosis [‡]		Х		
Atrophy		Х		
Congestion		Х		
Edema		Х		
Hemorrhage		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Metaplasia, osseous *		Х		
Neovascularization		Х		
Pigment, increased/decreased, iris		Х		
Single cell necrosis [‡]		Х		
Vacuolation, lipid *		Х		
Vacuolation, cytoplasm, epithelium		Х		
Proliferative Non-neoplastic				
Hyperplasia, melanocyte			Х	
Proliferative Neoplastic				
Adenoma, ciliary body, iris			Х	
Leiomyoma, uvea			Х	
Melanoma, uvea, benign			Х	
Schwannoma, intraocular/optic nerve, benign			Х	
Melanoma, uvea, malignant			Х	
Adenocarcinoma, ciliary body, iris			Х	
Meningioma, malignant, optic nerve			Х	
Schwannoma, intraocular/optic nerve, malignant			Х	

* indicates terminology with diagnostic criteria or comments described in the text. + Refer to 4 for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

Vacuolation, Lipid

Comments: Accumulations of foam cells, extracellular lipid and occasional cholesterol clefts have been found in the iris and ciliary body of the eyes in about 50% of NZW rabbits fed a cholesterol-rich diet over three months²⁸. Similar ocular lesions were described for WHHL rabbits³¹¹.

H. Anatomy of the Lens

The rabbit lens is large compared to that of haplorhine primates^{219, 234}. The rabbit lens is 3.9- fold larger than that of cynomolgus macaques, even though the eyes of both species are of similar size²⁸⁴. The rabbit lens is also larger and thicker than the human lens, even though the human eye is overall much larger^{217, 220}. NZW rabbits have mean lens thickness and diameter of 6.36 +/- 0.13 mm and 10.47 +/- 0.31 mm, respectively, while humans have mean lens thickness and diameter of 4.24 +/-0.46 mm and 9.58 +/-0.27 mm, respectively²⁸⁵. Rabbits are distinctive in having linear shaped sutures, with the anterior and posterior sutures oriented vertically and horizontally, respectively^{219, 312} (Table 61).

Degeneration, Lens Fiber (Figure 46)

Comments: Lens fiber degeneration is a microscopic finding that is usually correlated with ophthalmic examination findings of lens opacity. Spontaneous lens opacities can occur at different ages in rabbits and can exhibit breed or strain specificity²⁴, ^{313–317}. Congenital lenticular opacities of uncertain etiology have been reported in neonates, but most spontaneous cataracts occur in adult or even aged animals²²². In a study involving rabbits ranging in age from about 2.7 to 9.6 months, Munger *et al.* reported lens opacity incidences of 5.7% in albino NZW and 1.1% in pigmented NZW × NZ Red F1 hybrids³¹⁴. In two NZW inbred strains

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Non-proliferative				
Apoptosis [‡]		Х		
Degeneration, lens fiber *		Х		
Dislocation, lens, anterior or posterior			Х	
Fibroplasia, lens epithelium		Х		
Hypertrophy, lens capsule/epithelium/fiber		Х		
Inflammation		Х		
Mineralization, lens fiber		Х		
Necrosis, lens epithelium		Х		
Parasite *		Х		
Rupture, lens capsule		Х		
Single cell necrosis ⁺		Х		
Vacuolation, lens epithelium/fiber		Х		
Proliferative Non-neoplastic				
Hyperplasia, lens epithelium		Х		

Table 61. Microscopic Findings of the Lens: Rabbit

* Terminology with diagnostic criteria or comments described in the text. + Refer to 4 for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

(EII/JC and EIIJC-HLA-A2), spontaneous lens opacities had an overall much later onset, being noted only in animals 37 months or older³¹⁵. Nuclear sclerosis (increased compaction of inner cortical lens fibers) was noted only in older animals (average age 6.0 +/- 2.9 years in one series²²² and 3.5–4.0 years in another series)³¹⁸. Lens fiber degeneration and more extensive lesions can develop as sequelae of many non-heritable spontaneous or experimentally induced conditions that secondarily affect the lens (i.e., diabetes, glaucoma, posterior synechia, and proliferative vitreoretinopathy) ^{24, 248, 300, 319–321}.

Many cases of spontaneous lenticular opacities in rabbits are suspected to be of heritable (genetic) origin^{314, 315}. A recent study has demonstrated that experimentally induced mutations of the α -crystallin gene result in heritable opacities in knockout rabbits and their offspring³²².

Experimentally induced lens fiber degeneration in rabbits occurs with administration of various chemical test articles; exposure to microwaves, UVA radiation, or electric current; hyperoxia; and dietary imbalances^{220, 294, 320, 321, 323–325}. Lens fiber degeneration in rabbits in toxicity studies can also result from extraneous physical trauma to the lens (i.e., from misplaced hypodermic needles during intravitreal injections or from cage accidents or other misadventure).

Parasite

Comments: The common microsporidian parasite *Encephalitozoon cuniculi* can infect many tissues of rabbits, including the eye. Ocular infections can result in multiple pathologic changes including corneal ulceration, edema, and epithelial and endothelial necrosis; iris and ciliary body edema and epithelial degeneration; retinal atrophy; and generalized inflammatory cell infiltrates^{326, 327}. In the lens, infections can also

result in degeneration of the lens, which in turn sometimes ruptures, resulting in secondary uveal inflammation (phacoclastic uveitis)^{85, 222, 326, 328–331}. The route of infection for the lens is unclear, though transplacental vertical transmission has been proposed^{328, 331}.

I. Anatomy of the Vitreous

The vitreous cavity of the rabbit eye is relatively small, with a vitreous-to-globe area ratio of 0.4 compared to a 0.7 ratio in the similarly sized eye of the cynomolgus macaque²⁸⁴. In rabbits, strong vitreal attachment occurs along the retinal medullary rays³³².

The collagen fibrils of the rabbit vitreous are condensed into funnel-like lamellae which are more prominent and uniform than those in humans. The lamellae generally extend anteroposteriorly from the vitreous base to the optic nerve head³³³. Cloquet's canal, a conduit for hyaloid vessels during fetal development, is retained consistently in adult rabbits³³³, even though most authors agree that this structure does not routinely persist in adult human eyes³³⁴ (Table 62).

J. Anatomy of the Retina

Further maturation of the retina continues post eyelid opening for several weeks more or even to adulthood^{215, 216, 335, 336}. Regression of ocular fetal vessels (hyaloid vessels and vasculosa lentis) has been reported to be complete by about postnatal days 14–20^{337, 338}, but persistence of embryonic vessel remnants is common in rabbits^{339–341}.

The rabbit retina exhibits many anatomical and histologic differences compared to that of humans and other primates and non-primate mammalian species. The medullary rays are

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Aplasia, vitreous		Х		
Persistent hyperplastic primary vitreous		Х		
Persistent hyaloid vessels	Х			
Non-proliferative				
Fibroplasia		Х		
Hemorrhage		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Metaplasia, bone or cartilage		Х		
Mineralization, vitreous		Х		
Pigment, macrophage, hemosiderin		Х		
Proliferative Non-neoplastic and Neoplastic				

Table 62. Microscopic Findings of the Vitreous: Rabbit

easily visible ophthalmoscopically as two broad, pale, winglike bands that emanate from the optic nerve head and extend horizontally across the temporal and nasal fundus to just posterior to the equator^{219, 220, 222}. Histologically, the medullary rays consist of myelinated nerve fibers (axons of the retinal ganglion cells)^{336, 342–347}. In typical sagittal histologic sections, the medullary rays appear as focal elevations of the inner retinal surface due to the collectively increased thickness of their myelinated nerve fibers. Unlike most species, the rabbit retina has oligodendrocytes, which are localized in the medullary rays^{343, 344}. Rabbit retinal astrocytes are confined to the medullary rays, especially the nerve fiber and ganglion cell layers, with some astrocytes closely associated with the blood vessels overlying the medullary rays^{345, 347}.

Although not visible ophthalmoscopically, the visual streak is well developed in the rabbit^{220, 335, 343, 347–352}. Histologically, in typical sagittal sections, the rabbit visual streak is a discrete nasotemporal linear zone of increased ganglion cell density, located inferior to the optic nerve head and medullary rays^{220, 348, 350–353}.

The area centralis is a specialized focus of increased ganglion cell density distinct from the linear visual streak which is present in the retina of many mammals^{335, 354}. Whether the rabbit possesses an area centralis is a subject of some controversy, with some authors^{348, 351, 353}, but not others^{217, 350} reporting its presence.

Rabbits are presumed to have dichromatic color vision^{355, 356}. Density gradients are present in the rabbit retina, with the "blue" S-cones concentrated in small zones in the inferior retina^{335, 355, 356}. Dual-opsin cones (which co-express both "blue" and "green" opsins) have been demonstrated in rabbits³⁵⁷. The intrinsically photosensitive retinal ganglion cells (ipRGC) are a recently discovered third class of nonciliary retinal photoreceptor³⁵⁸. The recent identification of melanopsin gene expression in the rabbit retina suggests the possibility that iprGC may also occur in this species³⁵⁹. Compared to many other species, the Müller macroglial cells of the rabbit retina are especially prominent^{220, 360}, with the entire transretinal span of individual Müller cells and their processes often easily traceable on routine H&E sections. The Müller cells are distributed throughout the rabbit retina, including the medullary rays^{215, 345, 347, 361}.

Bruch's membrane is a laminated extracellular matrix located between the retinal pigmented epithelium (RPE) and the adjacent choriocapillaris and is composed of the RPE and choriocapillaris capillary basement membranes and associated collagen and elastin layers^{362, 363}. Bruch's membrane in rabbits is thinner than that of humans³⁶⁴. The RPE contain melanosomes, which lack melanin granules in albino rabbits such as the NZW strain. RPE melanin granules tend to diminish in number (due to fusion) and accumulate lipofuscin as agingrelated changes. The Dutch Belted rabbit and NZW F1 cross rabbits offer the ability to test potential effects of xenobiotics on the eye of pigmented rabbits.

Retinal and choroidal arterial supply is dual and variable among rabbits, arising from the external carotid artery as it feeds the external ophthalmic artery or from the internal carotid artery as it feeds the internal ophthalmic artery³⁶⁵. The rabbit retina is partially vascularized (merangiotic vascularization), which is unique among the domestic and common laboratory species^{342, 366, 367}. The central retinal artery enters the optic nerve and then passes into the globe, where it divides into nasal and temporal arteries that extend horizontally across the fundus (parallel to and overlying the medullary rays; other regions of the retina are avascular)^{342, 366, 367}.

In rabbits, the exit point of the optic nerve from the globe is superior to the antero-posterior pole axis and horizontal-most meridian plane of the globe^{219, 234, 333, 360, 367, 368}. Thus, the rabbit optic nerve head is located in the superior rather than central fundus.

The rabbit optic nerve head is horizontally oval and normally deeply indented (physiologic cupping), which is appreciable both ophthalmoscopically and in histologic preparations^{217, 219, 220, 234, 368}. In many species such as humans, macaques, horses, and pigs, the scleral exit channel at the optic nerve head is multi-fenestrated due to a well-developed lamina cribrosa, a sieve-like network of collagenous plates^{366, 369–371}. Rabbits have a poorly developed lamina cribrosa, and the optic nerve head contains neuronal tissue and astrocytes in addition to oligodendroglia^{219, 234, 360, 366, 368, 369}. This connective tissue meshwork also contains elastin and lends support for the nerve tissue.

In rabbits, a central retinal artery enters the optic nerve ventrally, just posterior to the optic nerve head^{219, 366, 368}. Main veins from the two medullary rays also exit via the optic nerve³⁷² (Table 63).

Deposits, Extracellular Matrix, Subretinal Pathogenesis/cell of origin: RPE

Diagnostic Features: Discrete, focal, rounded eosinophilic deposits located between the RPE and Bruch's membrane, sometimes resulting in a focal "dome"-like inward elevation of the RPE.

Comments: These deposits between the RPE and Bruch's membrane are not common but can occur as spontaneous findings in otherwise unremarkable rabbit eyes³⁷³, and morphologically resemble drusen though they have not been biochemically characterized in rabbits.

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Arteriolar loop, pre-retinal		Х		
Non-proliferative				
Apoptosis ⁺		Х		
Atrophy, retinal, global/inner/outer		Х		
Congestion		Х		
Deposits, extracellular matrix, subretinal *		Х		
Detachment, retina		Х		
Displacement, photoreceptor nuclei		Х		
Edema		Х		
Eosinophilic bodies, retina *		Х		
Fibroplasia, retinal, subretinal or epiretinal		Х		
Folds, retina	Х			
Glial cells, increased		Х		
Hemorrhage		Х		
Hypertrophy, retinal pigment epithelium (RPE) *	Х			
Inclusions (intracytoplasmic accumulation), RPE		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Mineralization		Х		
Myelin, increased		Х		
Necrosis		Х		
Necrosis, single cell		Х		
Neovascularization		Х		
Pigment, increased		Х		
Pigment, decreased		Х		
Polarity loss, RPE		Х		
Retinal rosettes		Х		
Single cell necrosis [‡]		Х		
Vacuolation, cytoplasmic/extracellular		Х		
Proliferative Non-neoplastic				
Hyperplasia, retinal pigmented epithelium		Х		

 Table 63.
 Microscopic Findings of the Retina: Rabbit

Proliferative Neoplastic

* Terminology with diagnostic criteria or comments described in the text. * Refer to 4 for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

Eosinophilic Bodies, Retina (Figure 47)

Pathogenesis/cell of origin: retinal neurons (horizontal cells)

Diagnostic Features: Occasional, round to oval, pale pink, circular structures with indistinct outlines, located in the inner nuclear layer (INL) and outer plexiform layer (OPL).

Comments: The pale eosinophilic round structures in the rabbit retina termed "eosinophilic bodies" have been demonstrated by immunohistochemistry and transmission electron microscopy to be neurofilament accumulations in horizontal cell neurites (dendrites) ³⁷⁴. Affected retinas were otherwise histologically unremarkable. The retinal eosinophilic bodies occur in several rabbit breeds including NZW, Japanese White, and Dutch Belted, with higher incidences in older animals, suggesting that they are an aging-related change.

Hypertrophy, Retinal Pigment Epithelium (RPE) (Figure 48)

Diagnostic Features: Small foci of enlarged and often displaced RPE in the subretinal space. The RPE aggregates are frequently located near the optic nerve head (peripapillary) or at the peripheral margins of the retina but can also be found less frequently in other retinal regions. The swollen RPE cells have abundant pale basophilic, granular cytoplasm, which often has a brownish tinge attributed to lipofuscin³⁷³. Pigment granules for pigmented breeds are concentrated around these vacuoles³⁷⁵.

Comments: Young adult rabbits sometimes exhibit this spontaneous RPE change unassociated with any concurrent

ocular pathology³⁷³. The exact etiology is undetermined. This spontaneous RPE hypertrophy was first recognized in a pigmented rabbit breed (Dutch Belted) but can also occur in other pigmented breeds (e.g. New Zealand Red) as well as in albino breeds (Table 64).

K. Anatomy of the Sclera

The rabbit sclera is thicker at the limbus (0.5 mm) and thinner at the posterior pole (approximately 0.18-0.2 mm) $^{219, 234}$. In comparison, human sclera thickness is similar at the limbus (approximately 0.5-0.53 mm), but much greater at the posterior pole and near the optic nerve (0.86 mm-1.0 mm)^{376, 377}.

In rabbits, a large orbital venous sinus (also called the orbital "venous plexus") is located posterior to the globe and receives the venous drainage from the eye and orbital contents²¹⁹, ²²⁰, ²³³, ³⁷⁸, ³⁷⁹.

Arteries supplying the rabbit eye (such as the long and short posterior ciliary arteries) generally enter the sclera in the posterior globe near the optic nerve, while the central retinal artery enters within the optic nerve itself^{233, 367, 378} (Table 65).

Metaplasia Osseous

Comments: Scleral osseous metaplasia has been reported as a reaction to intraocular osteoinductive hydroxyapatite and polyethylene polymer implants in experimentally manipulated (eviscerated) eyes of NZW rabbits³¹⁰.

Table 64. Microscopic Findings of the Optic Nerve: Rabbit

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Non-proliferative				
Atrophy		Х		
Degeneration, axon		Х		
Demyelination		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Necrosis		Х		
Single cell necrosis ⁺		Х		
Vacuolation		Х		
Proliferative Non-neoplastic				
Glial cells, increased number		Х		
Proliferative Neoplastic				
Glioma, benign			Х	
Meningioma, benign			Х	
Meningioma, malignant			Х	
Schwannoma, benign			Х	
Schwannoma, malignant			X	

⁺Refer to ⁴ for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Non-proliferative				
Apoptosis [‡]		Х		
Atrophy		Х		
Hemorrhage		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Metaplasia, osseous *		Х		
Single cell necrosis [‡]		Х		
Proliferative Non-neoplastic and Neoplastic				

Table 65. Microscopic Findings of the Sclera: Rabbit

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* Terminology with diagnostic criteria or comments described in the text. * Refer to 4 for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

L. Anatomy of the Ocular Adnexa - Glands and Ducts

The gross anatomical terminology of the rabbit ocular adnexal glands has been the subject of some confusion in the scientific literature, with many different, often inconsistent, and even contradictory nomenclature schemes promulgated by various authors. Thus, caution is warranted when reviewing published terminology schemes and descriptions for these structures.

The rabbit zygomatic salivary gland is located along the zygomatic arch near the inferior portion of the lacrimal gland^{234, 380, 381}. Portions of the zygomatic salivary are sometimes sampled along with the adjacent lacrimal gland, appearing in routine histologic preparations as clusters of deeply basophilic mucous-cell acini with tiny lumens.

The ocular adnexa, especially the Harderian gland and most of the extraocular muscles are closely associated with the large venous sinus (or plexus) present in the posterior orbit of rabbits^{219, 233, 378}.

Harderian Gland

The rabbit Harderian gland is large, located in the medial ventral orbit, and attached to the base of the nictitating membrane^{219, 220, 234, 382, 383}. The rabbit Harderian gland is composed of two closely apposed but grossly and histologically distinct lobes, surrounded by a thin fibrous capsule^{219, 234, 384}. A single excretory duct opens on the concave surface of the base of the nictitating membrane^{219, 379, 385}.

Histologically, the Harderian gland is classified as tubuloacinar (tubuloalveolar). Acinar epithelial cells of both lobes are characterized by abundant cytoplasmic lipid vacuoles, and the secretions of both lobes are predominantly lipid^{383, 385–390}.

Based on their grossly visible coloration, the two lobes are known as the "white" lobe and the "pink" (or "red") lobes^{219, 383–385, 391} (Figure 49). The more inferiorly situated pink lobe (Figure 50) is about twice as large as the more superiorly located white lobe^{219, 383, 384, 391, 392}. In the white lobe (Figure 51), acinar cell lipid vacuoles are small, more uniform and very densely packed, imparting a typically darker, more eosinophilic microscopic appearance in routine H&E sections. In contrast, in the grossly pink lobe acinar cells contain larger, clear vacuoles, and thus appear paler microscopical-ly²³⁴, 382–384, 390–392.

Unlike rats, mice, and hamsters, the rabbit Harderian glands does not appear to exhibit histologic sexual dimorphism and/ or porphyrin secretion^{390, 393}. However, intact male rabbits have particularly large Harderian glands, which increases further in size during the breeding season.

Lacrimal Glands

The rabbit lacrimal gland is large, long, thin, and multilobulated, with pink to pale red macroscopic coloration^{217, 220} (Figure 52). Its lobes fill most of the inferior orbit and extend into the posterior superior orbit²⁹⁰. Although most of the lacrimal gland is intraorbital, portions may extend onto the lateral zygomatic arch and even extraorbitally onto the zygomatic bone^{234, 382}. Most authors describe a single excretory duct, which opens into the superior eyelid conjunctiva at the lateral canthus^{382, 394, 395}.

As in several other species, the rabbit lacrimal gland exhibits a degree of sexual dimorphism, with adult males exhibiting larger acini and greater numbers of acinar cells than females³⁹⁶.

Nictitating Membrane and Nictitans Gland

The rabbit has a well-developed nictitating membrane (third eyelid)^{217, 219, 382}. It consists of a crescentic conjunctival fold reflected from the medial canthus molded to the contours of the globe, and adjacent to the Harderian gland. A central hyaline cartilage plate, embedded in fibrous loose connective tissue, reinforces the nictitating membrane^{217, 219, 234, 382}.

The inner surface of the nictitating membrane is lined by goblet-cell containing conjunctival mucosa, while the more external surface is lined by non-keratinized stratified squamous epithelium (Figure 53).

The nictitans gland (gland of the third eyelid) is thin and flat, and surrounds most of the length of the central cartilage. It is a modified lacrimal-type gland with serous acini³⁸². Several ducts extend through the cartilage plate to open on the inner surface of the nictitating membrane^{219, 382, 397} (Table 66).

Cytoplasmic Alteration, Harderian Gland or Lacrimal Gland

Other terms: Metaplasia, Harderization; Ectopic gland.

Comments: Cytoplasmic alteration of the lacrimal gland or of the Harderian gland occur relatively commonly in the rabbit³⁹⁸. In the Harderian gland, it is possible to observe small islands of normal lacrimal gland, while less frequently, islands of normal Harderian gland acini may be observed in the lacrimal gland (Figure 54). The lacrimal gland alteration in the Harderian gland has been reported as early as 3 weeks of age and increases in incidence with age in the lacrimal glands of males and females but occurs with a greater incidence and extent in males. Harderian gland alteration in the lacrimal gland is recognized less commonly and is not well characterized.

Necrosis, Harderian Gland or Lacrimal Gland

Comments: Necrosis and acute to subacute inflammation in the Harderian and/or lacrimal glands consistent with infarction due to embolism of microthrombi associated with medial auricular artery catheterization has been reported³³.

Nasolacrimal Duct

In adult rabbits, the nasolacrimal duct (NDL) has the same three regions as described in other mammals (orbital lacrimal canaliculi and sac, bony NDL canal, and non-bony NDL canal in nasal cavity).

In the rabbit, a single lacrimal punctum is located in the inner surface of the inferior eyelid near the medial canthus about 3 mm from the lid margin^{219, 222, 382, 399}. This opens into a poorly developed lacrimal sac-like dilatation which widens into the nasolacrimal duct^{222, 399–402}. The nasolacrimal duct terminates just caudal to the mucocutaneous junction of the nares^{18, 220, 222}.

According to most authors, the rabbit nasolacrimal sac and duct are lined by stratified to pseudo-stratified columnar epithelium, which in many areas has a conspicuously bilayered appearance^{399–402}. Some of the epithelial lining cells

Table 66. Microscopic Findings of the Harderian, Lacrimal, and Nictitans Glands: Rabbit

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Non-proliferative				
Apoptosis [‡]		Х		
Atrophy		Х		
Congestion		Х		
Cyst		Х		
Cytoplasmic alteration, acinar, Harderian gland or lacri- mal gland *		Х		
Degeneration		Х		
Dilatation		Х		
Edema		Х		
Hemorrhage		Х		
Hypertrophy		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Karyomegaly		Х		
Necrosis *		Х		
Necrosis, single cell		Х		
Porphyrin increased				Х
Regeneration		Х		
Single cell necrosis [‡]		Х		
Proliferative Non-neoplastic				
Hyperplasia, acinar		Х		
Neoplastic				
Adenoma			X	
Adenocarcinoma			Х	

* Terminology with diagnostic criteria or comments described in the text. + Refer to 4 for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Non-proliferative				
Apoptosis [‡]		Х		
Atrophy		Х		
Congestion		Х		
Cyst, inclusion		Х		
Degeneration		Х		
Edema		Х		
Erosion/ulceration		Х		
Fibrosis		Х		
Hemorrhage		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Metaplasia		Х		
Necrosis		Х		
Single cell necrosis [‡]		Х		
Proliferative Non-neoplastic				
Hyperplasia, epithelial		Х		
Proliferative Neoplastic				
Papilloma, squamous cell			Х	
Carcinoma, squamous cell			Х	

Table 67. Microscopic Findings of the Nasolacrimal Duct: Rabbit

⁺Refer to ⁴ for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

secrete mucins^{399, 402}. In rabbits, portions of the nasolacrimal duct are surrounded by a prominent cavernous body with large blood vessels in a loose stroma⁴⁰² (Table 67).

M. Anatomy of the External Ear

The lesions of the convex surface of the external ear (pinna) are largely those of haired skin and are discussed with the Integumentary section. The external ear (pinna) is easily assessed during macroscopic examination. The large veins in the rabbit pinna are often used for blood collection or Test Article administration, and if the latter are sampled for histopathology as the "Injection/Treatment Site". The concave external areas of the pinnae have a thin epidermis and dermis with a paucity of hair follicles. In the external ear canal just outside of the bony collar (i.e. at about the level of the obtuse-angle turn), the ear canal retains the thin epidermis but has a circumferential zone containing sebaceous glands (ceruminous glands) but lacking hair follicles. These glands are absent at this site in rodents. Within the internal acoustic meatus, defined by the presence of the bony collar, the dermis is very thin to non-existent with the epidermis almost lying upon the temporal bone. Rabbits have multiple small ceruminous glands (simple sebaceous glands that secrete cerumen, or "ear wax") (Table 68, 69, 70).

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Non-proliferative				
Apoptosis [‡]		Х		
Congestion		Х		
Cyst. tympanic membrane		Х		
Debris, external ear canal		Х		
Edema		Х		
Erosion/ulceration		Х		
Fibrosis		Х		
Hemorrhage		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation, auricular cartilage		Х		
Inflammation, external ear canal		Х		
Metaplasia		Х		
Perforation, tympanic membrane		Х		
Single cell necrosis ⁺		Х		
Proliferative (non-neoplastic)				
Hyperplasia, epithelium		Х		
Neoplastic				
Papilloma, squamous cell			Х	
Carcinoma, squamous cell			X	

 Table 68.
 Microscopic Findings of the External Ear: Rabbit

⁺ Refer to ⁴ for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Non-Proliferative				
Apoptosis [‡]		Х		
Atrophy, bone		Х		
Cholesteatoma		Х		
Congestion		Х		
Cyst		Х		
Edema		Х		
Fibrosis		Х		
Hemorrhage		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation, granulomatous/catarrhal		Х		
Metaplasia, squamous cell		Х		
Mineralization		Х		
Necrosis, bone		Х		
Necrosis, tympanic membrane		Х		
New bone formation		Х		
Perforation, tympanic membrane		Х		
Single cell necrosis [‡]		Х		
Tissue, granulation		Х		
Ulcer		Х		
Proliferative Non-neoplastic				
Fibrosis		Х		
Proliferative Neoplastic				

Table 69. Microscopic Findings of the Middle Ear: Rabbit

⁺Refer to ⁴ for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

Table 70.	Microscopic Findings of the Inner Ear: Rabbit

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Non-proliferative				
Apoptosis ⁺		Х		
Cellularity decreased, spiral ganglion/spiral limbus/spiral ligament, and/or stria vascularis		Х		
Congestion		Х		
Degeneration, axon/hair cells and/or epithelium		Х		
Edema		Х		
Erosion/ulceration		Х		
Fibrosis		Х		
Hair cell, decreased number		Х		
Hemorrhage		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation, inner ear		Х		
Loss, disorganization or disruption, otolith		Х		
Metaplasia		Х		
Necrosis cartilage/hair cell		Х		
Necrosis neuronal/vestibular organ		Х		
New bone formation		Х		
Otolith loss, disorganization or disruption		Х		
Single cell necrosis ⁺		Х		
Vacuolation, hair cell/supporting cell		Х		
Vacuolation, stria vascularis		Х		
Proliferative Non-neoplastic and Neoplastic				
_				

⁺Refer to ⁴ for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

SECTION 17: URINARY SYSTEM

Calcium absorption and metabolism in the rabbit is poorly understood. Rabbits normally have a higher blood calcium range than other laboratory animal species and are predisposed to cystic, urethral, ureteral and renal calculi. Rabbits excrete 45–60% excess calcium though the urine as calcium carbonate. Mineralized foci are commonly seen throughout the urinary tract. Urine is normally alkaline, and cloudy to pigmented, caused by the presence of albumin, fine calcium carbonate and ammonium magnesium phosphate crystals. The fact that rabbits are horizontal quadrupeds may also predispose the anterior wall to retention of microcrystals and other particles as compared to humans. Rabbit urine varies in color from creamy yellow to dark red depending on the presence of porphyrin pigments derived from the diet or xenobiotics e.g. antibiotics. Care must be taken to differentiate between red urine caused by porphyrin excretion and hematuria. While modern laboratory animal management practices have limited the incidence of infectious processes in the kidney, inflammatory conditions related to infectious disease may still occur (Table 71).

Cyst

Comments: Renal lesions resembling human polycystic kidney disease were reported in a retrospective evaluation of NZW rabbit kidney tissue⁴⁰³.

Glomerulopathy

Comments: Rabbits are susceptible to a glomerulopathy induced by corticosteroids⁴⁰⁴⁻⁴⁰⁶. The initiating lesion is a glomerulopathy characterized by aneurysmal capillary dilatation with nodular changes of eosinophilia and cellular loss in the glomeruli. Bowman's space often contains erythrocytes or eosinophilic material. Bowman's capsule may be necrotic. Ultrastructurally, glomerular capillaries may be occluded with a proteinaceous coagulum, endothelium may be swollen, and podocytes have loss of foot processes. Basement membranes may be thickened or tortuous. Epithelial cells of the glomerular tufts may have hyaline globules, vacuoles or be intensely osmophilic. Renal tubules may have fatty infiltration, hyaline droplets, and cellular necrosis. Tubules contain erythrocytes and protein casts. Clinical pathology parameters may demonstrate increased BUN, glycosuria, albuminuria, and hematuria with clinically red urine. Kidney weights may be increased. It is important to recognize the susceptibility of rabbits to corticosteroid glomerulopathy as it may be confused with a test article-related effect if animals are given corticosteroids for palliative purposes during a toxicology study.

Mineralization (Figure 55)

Comments: Mineralized foci are commonly seen in the collecting ducts and medulla of rabbit kidneys. Mineralized foci in the tubules or interstitial areas of the cortex are present in 60% of male and female rabbits evaluated^{81, 407}. Focal mineralization is also occasionally recorded in the urothelium of the urinary bladder.

Nephropathy, Spontaneous

Pathogenesis/cell of origin: Proximal and distal tubules

Diagnostic Features: The histological findings are generally recorded separately (basophilia tubules, dilated/cystic tubules, pigmented tubules, interstitial inflammatory cell infiltrate), and the term nephropathy is only used when at least three of the aforementioned components are present. A spontaneous nephropathy syndrome is commonly seen in clinically normal apparently healthy rabbits from colonies free from Encephalitozoon cuniculi. There are no clinical signs accompanying these lesions and no evidence of progression/greater severity of the findings on longer term studies i.e. the lesion is not thought to be progressive unlike the nephropathy in rats. It is considered to be a syndrome particular to the NZW rabbit as similar findings have not been recorded in mixed breed pet rabbits. The renal findings are observed in 80% of apparently healthy young rabbits, more frequently in females than males⁸¹. Spontaneous findings of mineralization, tubular basophilia and dilatation have been reported previously in young laboratory rabbits less than 1 year of age^{408, 409}, but the incidence of this lesion seems to be increasing in the NZW population. Basophilic tubules, dilated/cystic tubules, pigmented tubules, interstitial inflammatory cell infiltration, and mineralization have been reported in juvenile NZW rabbits as little as 8 weeks old (Bradley, unpublished data). Study pathologists should exercise caution when interpreting kidney findings in apparently healthy rabbits as these renal nephropathy findings may mask nephrotoxic effects.

Vacuolation

Comments: Vacuolation of the proximal convoluted tubule epithelium is a regular finding in young non-pregnant females (35%). These vacuoles stain positively with Oil red O stain for neutral lipids. Vacuolation of the proximal convoluted tubule epithelium has been recorded in juvenile NZW rabbits as little as 8 weeks old (Bradley, unpublished data) (Table 72, 73, 74).

Table 71.	Microscopic Findings of the Kidney: Rabbit	
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Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
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* Terminology with diagnostic criteria or comments described in the text. ⁺ Refer to ⁴ for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Aplasia, ureter			Х	
Non-proliferative				
Abscess		Х		
Apoptosis [‡]		Х		
Calculus		X		
Crystals		X		
Dilatation		X		
Edema		X		
Erosion/ulcer		X		
Fibrosis		X		
Hemorrhage		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		X		
Single cell necrosis ⁺		X		
Proliferative Non-neoplastic				
Hyperplasia, urothelium		Х		
Metaplasia, glandular/squamous cell			Х	
Proliferative Neoplastic				
Papilloma, squamous cell/urothelial			Х	
Adenocarcinoma			Х	
Carcinoma, squamous cell/urothelial		-	Х	

 Table 72.
 Microscopic Findings of Ureter: Rabbit

⁺Refer to ⁴ for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

Ethical Practices: All procedures used to prepare macroscopic and microscopic images of animal specimens for this article were performed in accordance with regulations and established guidelines for humane treatment of research animals and were reviewed and approved in advance by the relevant Institutional Animal Care and Use/Ethics Committee.

Declaration of Conflicting Interests: The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Funding: The author(s) received no financial support for the research, authorship and/or publication of this article.

Acknowledgements: The authors wish to express their thanks to the INHAND GESC, and the BSTP, ESTP, JSTP and STP membership for comprehensive reviews, excellent comments and helpful edits. We also thank Dr Rupert Kellner for manuscript review and Ms Beth Mahler, Ms. Emily Singletary, and Ms. Maureen Puccini from EPL Inc., for image editing. Photographs used in this document were either provided from the coauthors or are as acknowledged in the figure legends.

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Congenital				
Diverticulum			Х	
Non-proliferative				
Abscess		Х		
Angiectasis		Х		
Apoptosis [‡]		Х		
Calculus		Х		
Crystals	Х			
Dilatation		Х		
Edema		Х		
Erosion		Х		
Fibrosis		Х		
Hemorrhage		Х		
Hypertrophy, urothelium		Х		
Inclusions, urothelium				Х
Infiltrate, inflammatory cell [insert appropriate cell type]		Х		
Inflammation		Х		
Metaplasia		Х		
Mineralization, urothelium		Х		
Necrosis		Х		
Parasite		Х		
Proteinaceous plug				Х
Single cell necrosis ⁺		Х		
Ulcer		Х		
Uropathy, obstructive		Х		
Vacuolation, urothelium		Х		
Proliferative Non-neoplastic				
Hyperplasia, urothelium			Х	
Metaplasia, glandular/squamous cell			Х	
Proliferative Neoplastic				
Proliferative lesion, mesenchymal				Х
Papilloma, squamous cell/urothelial			X	
Adenocarcinoma			X	
Carcinoma, squamous cell/urothelial			X	

 Table 73.
 Microscopic Findings of the Urinary Bladder: Rabbit

 $^{\scriptscriptstyle 4}$ Refer to 4 for diagnostic criteria and use of the terms apoptosis and single cell necrosis.

Finding	Common	Uncommon	Not Observed but Potentially Relevant	Not Applicable
Non-proliferative				
Abscess		Х		
Apoptosis [‡]		Х		
Congestion		Х		
Edema		Х		
Erosion		Х		
Fibrosis		Х		
Hemorrhage		Х		
Hypertrophy, urothelium		Х		
Infiltrate, inflammatory cell [insert appropriate cell type]		X		
Inflammation		Х		
Metaplasia		Х		
Obstruction		Х		
Plug, proteinaceous				Х
Single cell necrosis [‡]		X		
Proliferative Non-neoplastic				
Hyperplasia, urothelium		Х		
Metaplasia, glandular/squamous cell		Х		
Proliferative Neoplastic				
Proliferative lesion, mesenchymal				Х
Papilloma, squamous cell/urothelial			Х	
Adenocarcinoma			Х	
Carcinoma, squamous cell/urothelial			Х	

 Table 74.
 Microscopic Findings of the Urethra: Rabbit

⁺Refer to ⁴ for diagnostic criteria and use of the terms apoptosis and single cell necrosis.



- FIGURE 1.—Heart, Fibrosis Myocardium, H&E.
- FIGURE 2.—Heart, Infiltrate, inflammatory cells, H&E.
- FIGURE 3.—Heart, Mineralization myocardium H&E.
- FIGURE 4.—Heart, Necrosis myocardium, H&E.
- FIGURE 5.—Aorta, Mineralization, H&E.
- FIGURE 6.—Stomach, Cardioesophageal junction, H&E.



11

- FIGURE 7.—Stomach, Cardioesophageal junction, H&E. FIGURE 8.—Gastrointestinal tract, Parasite (*Eimeria*), H&E. FIGURE 9.—Gastrointestinal tract, Parasite (*Eimeria*), H&E. FIGURE 10.—Thyroid, Diffuse Hyperplasia, H&E. FIGURE 11.—Thyroid, Diffuse Hyperplasia, H&E (high mag). FIGURE 12.—Liver, Cytoplasmic alteration, H&E.



- FIGURE 13.—Liver, Inflammation, liver periportal, H&E. FIGURE 14.—Liver, Parasites (*Eimeria*), H&E.
- FIGURE 15.—Liver, Parasite (Eimeria), H&E.
- FIGURE 16.-Liver, Parasite (Encephalitozoon), H&E (Courtesy of Aaron Sargeant).
- FIGURE 17.—Liver, Pigment (Liver toxicosis), H&E.
- FIGURE 18.—Submandibular skin (mental) gland, H&E.



- FIGURE 19.—Submandibular skin (mental gland), H&E (high mag).
 FIGURE 20.—Inguinal gland complex: sebaceous (left), apocrine (right), H&E.
 FIGURE 21.—Inguinal gland complex: sebaceous (left), apocrine (right), H&E (high mag).
 FIGURE 22.—Female, Inguinal gland complex (apocrine), vagina, H&E (high mag).
 FIGURE 23.—Rectum, Anal gland, H&E.
 FIGURE 24.—Brain, Parasitic encephalitis, H&E.



FIGURE 25.—Brain, Vacuolation, choroid plexus, H&E.

- FIGURE 26.—Peripheral Nerve, Demyelinization fibres, H&E.
- FIGURE 27.—Prostate, and associated glands, H&E.
- FIGURE 28.—Prostate (left) and Proprostate (right), H&E.
- FIGURE 29.—Proprostate, H&E.
- FIGURE 30.—Proprostate, H&E.



- FIGURE 31.—Paraprostate, H&E. FIGURE 32.—Prostate, H&E. FIGURE 33.—from left to right: Seminal vesicle, Vas deferens, Ampulla, Urethra, H&E. FIGURE 34.—Lung, Infiltrate, inflammatory cell, H&E. FIGURE 35.—Lung, Infiltrate, heterophil, H&E. FIGURE 36.—Lung, Macrophages increased, alveolar H&E.



- FIGURE 37.—Lung, Metaplasia, osseous, H&E. FIGURE 38.—Lung, Thrombus, H&E. FIGURE 39.—Bone, necrosis, H&E.

- FIGURE 40.—Bone, Physeal necrosis, H&E.



- FIGURE 41.—Eyelid with Meibomian gland, H&E. FIGURE 42.—Eyelid, accessory lacrimal gland (gland of Wolfring), H&E.
- FIGURE 43.—Eyelid, accessory lacrimal gland (gland of Wolfring) Meibomian gland, H&E. FIGURE 44.—Eyelid, accessory lacrimal gland (gland of Wolfring) and Meibomian gland, H&E (high mag).
- FIGURE 45.—Eye, Ciliary Body, Ciliary Web, H&E.
- FIGURE 46.—Eye, Degeneration lens fibre, H&E.



FIGURE 47.—Eye, Retina pigment epithelium, hypertrophy H&E. FIGURE 48.—Eye, Eosinophilic bodies, retina, H&E.

- FIGURE 49.—Harderian Gland, pink lobe (left) and white lobe (right), H&E.
- FIGURE 50.—Harderian Gland, white lobe, H&E (high mag).
- FIGURE 51.—Harderian Gland, pink lobe, H&E (high mag).
- FIGURE 52.—Zygomatic salivary gland (left) and Lacrimal gland (right), H&E.



- FIGURE 53.—Nictitating membrane, H&E. FIGURE 54.—Harderian gland, Cytoplasmic alteration, H&E. FIGURE 55.—Kidney, Renal mineralisation, H&E. FIGURE 56.—Kidney, Nephroblastoma, H&E.

References

- Mann PC, Vahle J, Keenan CM, Baker JF, Bradley AE, Goodman DG, Harada T, Herbert R, Kaufmann W, Kellner R, Nolte T, Rittinghausen S, and Tanaka T. International harmonization of toxicologic pathology nomenclature: an overview and review of basic principles. *Toxicol Pathol.* 40(Suppl): 7S–13S. 2012. [Medline] [CrossRef]
- Fuentealba IC, Mahoney NT, Shadduck JA, Harvill J, Wicher V, and Wicher K. Hepatic lesions in rabbits infected with Encephalitozoon cuniculi administered per rectum. *Vet Pathol.* 29: 536–540. 1992. [Medline] [CrossRef]
- Wasson K, and Peper RL. Mammalian microsporidiosis. *Vet Pathol.* 37: 113–128. 2000. [Medline] [CrossRef]
- Elmore SA, Dixon D, Hailey JR, Harada T, Herbert RA, Maronpot RR, Nolte T, Rehg JE, Rittinghausen S, Rosol TJ, Satoh H, Vidal JD, Willard-Mack CL, and Creasy DM. Recommendations from the INHAND Apoptosis/Necrosis Working Group. *Toxicol Pathol.* 44: 173–188. 2016. [Medline] [CrossRef]
- Carr BJ, Ochoa L, Rankin D, and Owens BD. Biologic response to orthopedic sutures: a histologic study in a rabbit model. *Orthopedics*. 32: 828. 2009. [Medline]
- International Organization for S. 10993: Biological evaluation of medical devices —Part 6 Tests for local effects after implantation. Switzerland. 2016.
- Hosoyama T, Ishiguro N, Yamanouchi K, and Nishihara M. Degenerative muscle fiber accelerates adipogenesis of intramuscular cells via RhoA signaling pathway. *Differentiation*. **77**: 350–359. 2009. [Medline] [CrossRef]
- Cheng L, Wang T, Zhu J, and Cai P. Osteoinduction of calcium phosphate ceramics in four kinds of animals for 1 year: dog, rabbit, rat, and mouse. *Transplant Proc.* 48: 1309–1314. 2016. [Medline] [CrossRef]
- Ripamonti U. Osteoinduction in porous hydroxyapatite implanted in heterotopic sites of different animal models. *Biomaterials*. 17: 31–35. 1996. [Medline] [CrossRef]
- Yuan H, van Blitterswijk CA, de Groot K, and de Bruijn JD. Crossspecies comparison of ectopic bone formation in biphasic calcium phosphate (BCP) and hydroxyapatite (HA) scaffolds. *Tissue Eng.* 12: 1607–1615. 2006. [Medline] [CrossRef]
- Kamphues J, Carstensen P, Schroeder D, Meyer H, Schoon HA, and Rosenbruch M. Effekte einer steigenden Calcium- und Vitamin D-Zufuhr auf den Calciumstoffwechsel von Kaninchen (Effects of increasing calcium- and vitamin D supply on calcium metabolism of rabbits). *J Anim Physiol Anim Nutr (Berl).* 56: 191–208. 1986. [CrossRef]
- Cruise LJ, and Brewer NR. Anatomy. In: The Biology of the Laboratory Rabbit, 2nd ed. PJ Manning, DH Ringler, and CE Newcomer (eds). Academic Press, San Diego. 47–61. 1994.
- Kozma C, Macklin W, Cummins LM, and Mauer R. The anatomy, physiology and biochemistry of the rabbit. In: The Biology of the Laboratory Rabbit, 1st ed. SH Weisbroth, RE Flatt, and AL Kraus (eds). Academic Press, New York. 50–69. 1974.
- Sellers RS, Pardo I, Hu G, Khan KN, Perry R, Markiewicz V, Rohde C, Colangelo J, Reagan W, and Clarke D. Inflammatory cell findings in the female rabbit heart and stress-associated exacerbation with handling and procedures used in nonclinical studies. *Toxicol Pathol.* 45: 416–426. 2017. [Medline] [CrossRef]
- Pogwizd SM, and Bers DM. Rabbit models of heart disease. Drug Discov Today Dis Models. 5: 185–193. 2008. [Medline] [CrossRef]
- Vörös K, Seehusen F, Hungerbühler S, Meyer-Lindenberg A, and von der Hoeh N. Ventricular septal defect with aortic valve insufficiency in a New Zealand White rabbit. *J Am Anim Hosp Assoc.* 47: e42–e49. 2011. [Medline] [CrossRef]
- Hurley RJ, Marini RP, Avison DL, Murphy JC, Olin JM, and Lipman NS. Evaluation of detomidine anesthetic combinations in the rabbit. *Lab Anim Sci.* 44: 472–478. 1994. [Medline]
- Marini RP, Li X, Harpster NK, and Dangler C. Cardiovascular pathology possibly associated with ketamine/xylazine anesthesia in Dutch belted rabbits. *Lab Anim Sci.* 49: 153–160. 1999. [Medline]

- Cooper LL, Odening KE, Hwang MS, Chaves L, Schofield L, Taylor CA, Gemignani AS, Mitchell GF, Forder JR, Choi BR, and Koren G. Electromechanical and structural alterations in the aging rabbit heart and aorta. *Am J Physiol Heart Circ Physiol.* **302**: H1625–H1635. 2012. [Medline] [CrossRef]
- Downing SE, and Chen V. Myocardial injury following endogenous catecholamine release in rabbits. *J Mol Cell Cardiol.* 17: 377–387. 1985. [Medline] [CrossRef]
- Weber HW, and Van Der Walt JJ. Cardiomyopathy in crowded rabbits. Recent Adv Stud Cardiac Struct Metab. 6: 471–477. 1975. [Medline]
- Flores NA, Davies RL, Penny WJ, and Sheridan DJ. Coronary microangiography in the guinea pig, rabbit and ferret. *Int J Cardiol.* 6: 459–471. 1984. [Medline] [CrossRef]
- Maxwell MP, Hearse DJ, and Yellon DM. Species variation in the coronary collateral circulation during regional myocardial ischaemia: a critical determinant of the rate of evolution and extent of myocardial infarction. *Cardiovasc Res.* 21: 737–746. 1987. [Medline] [CrossRef]
- 24. Barthold SW, Griffey SM, and Percy DH. Pathology of Laboratory Rodents and Rabbits, 4th ed. Blackwell Publishing, Iowa. 2016.
- Lossi L, D'Angelo L, De Girolamo P, and Merighi A. Anatomical features for an adequate choice of experimental animal model in biomedicine: II. Small laboratory rodents, rabbit, and pig. *Ann Anat.* 204: 11–28. 2016. [Medline] [CrossRef]
- 26. Anitschkow N, and Chalatow S. Classics in arteriosclerosis research: On experimental cholesterin steatosis and its significance in the origin of some pathological processes arteriosclerosis: An Official Journal of the American Heart Association. *Inc.* **3**: 178–182. 1913.
- 27. Yanni AE. The laboratory rabbit: an animal model of atherosclerosis research. *Lab Anim.* **38**: 246–256. 2004. [Medline] [CrossRef]
- Rinke M, and Hartmann E. Dietary lipid excess in mice and rabbits. In: Classic Examples in Toxicologic Pathology, 5th ed. E Karbe, W Drommer, PG Germann, G Morawietz, and R Kellner (eds). European Society of Toxicologic Pathology. 2013.
- Fan J, Kitajima S, Watanabe T, Xu J, Zhang J, Liu E, and Chen YE. Rabbit models for the study of human atherosclerosis: from pathophysiological mechanisms to translational medicine. *Pharmacol Ther.* 146: 104–119. 2015. [Medline] [CrossRef]
- Sohn J, and Couto MA. Chapter 8: Anatomy, Physiology, and Behavior. In: The Laboratory Rabbit, Guinea Pig, Hamster, and Other Rodents, 1st ed. MA Suckow, KA Stevens, and RP Wilson (eds). Academic Press (Elsevier), Amsterdam. 195–217. 2012.
- Walker BE. Induction of cleft palate in rabbits by several glucocorticoids. Proceedings of the Society for Experimental Biology and Medicine Society for Experimental Biology and Medicine [(New York, NY)]. 125:1281–1284. 1967.
- van Kruiningen HJ, and Williams CB. Mucoid enteritis of rabbits. Comparison to cholera and cystic fibrosis. *Vet Pathol.* 9: 53–77. 1972. [Medline] [CrossRef]
- Lee S, Sorden S, Dunn D, Dwyer A, and Sonnentag P. The effects of blood collection techniques on Ocular lesions in Rabbits. *Invest Ophthalmol Vis Sci.* 54: 3039. 2013.
- 34. Villano JS, and Cooper TK. Mandibular fracture and necrotizing sialometaplasia in a rabbit. *Comp Med.* **63**: 67–70. 2013. [Medline]
- Meyer M, Speight P, and Bown SG. A study of the effects of photodynamic therapy on the normal tissues of the rabbit jaw. *Br J Cancer.* 64: 1093–1097. 1991. [Medline] [CrossRef]
- Smallwood J. Anatomy of the Laboratory Rabbit. In: A Guided Tour of Veterinary Anatomy: Domestic Ungulates and Laboratory Mammals. Saunders, Philadelphia. 1992.
- Van den Bulck K, Decostere A, Baele M, Marechal M, Ducatelle R, and Haesebrouck F. Low frequency of Helicobacter species in the stomachs of experimental rabbits. *Lab Anim.* 40: 282–287. 2006. [Medline] [CrossRef]
- 38. Leeson CR, and Leeson TS. The fine structure of Brunner's glands in the rabbit. *Anat Rec.* **159**: 409–419. 1967. [Medline] [CrossRef]
- Snipes RL. Anatomy of the rabbit cecum. Anat Embryol (Berl). 155: 57–80. 1978. [Medline] [CrossRef]

- Ruckebusch Y, and Fioramonti J. The Fusus coli of the rabbit as a pacemaker area. *Experientia*. 32: 1023–1024. 1976. [Medline] [CrossRef]
- Adak A, Prasad MC, Lonkar PS, Kapurkar UM, Brahmankar MG, and Patel MV. Ectopic spleen in the pancreas of New Zealand White rabbits. *Vet World*. 6: 360–362. 2013. [CrossRef]
- Brändli-Baiocco A, Balme E, Bruder M, Chandra S, Hellmann J, Hoenerhoff MJ, Kambara T, Landes C, Lenz B, Mense M, Rittinghausen S, Satoh H, Schorsch F, Seeliger F, Tanaka T, Tsuchitani M, Wojcinski Z, and Rosol TJ. Nonproliferative and proliferative lesions of the rat and mouse endocrine system. *J Toxicol Pathol.* **31**(Suppl): 1S–95S. 2018. [Medline] [CrossRef]
- Craigie EH. Bensley's Practical Anatomy of the Rabbit: An Elementary Laboratory Text-Book in Mammalian Anatomy. 8th ed. University of Toronto Press, Toronto. 1948.
- 44. La Perle KMD, and Dintzis SM. Endocrine system. In: Comparative Anatomy and Histology : A Mouse, Rat, and Human Atlas. 2nd ed. PM Treuting, SM Dintzis, and KS Montine (eds). Elsevier, London. 251–274. 2018.
- Caffe AR. Architecture of the mammalian pituitary cholinergic system with observations on a putative blood acetylcholine sensor. *Histol Histopathol.* 11: 537–551. 1996. [Medline]
- Kakita T, and Odell WD. Pituitary gland: one site of ultrashort-feedback regulation for control of thyrotropin. *Am J Physiol.* 250: E121– E124. 1986. [Medline]
- Reiter RJ. Comparative physiology: pineal gland. Annu Rev Physiol. 35: 305–328. 1973. [Medline] [CrossRef]
- Tan SQ, Thomas D, Jao W, Bourdeau JE, and Lau K. Surgical thyroparathyroidectomy of the rabbit. *Am J Physiol.* 252: F761–F767. 1987. [Medline]
- Rossouw DJ, and Chase CC. Ultrastructure of the capsule of the rabbit adrenal gland. *Acta Anat (Basel)*. 100: 538–544. 1978. [Medline] [CrossRef]
- Fazekas AG, and Sandor T. Unusual pathways of aldosterone biosynthesis in the rabbit adrenal. *Steroids.* 14: 161–177. 1969. [Medline] [CrossRef]
- Rosol TJ, Yarrington JT, Latendresse J, and Capen CC. Adrenal gland: structure, function, and mechanisms of toxicity. *Toxicol Pathol.* 29: 41–48. 2001. [Medline] [CrossRef]
- Henderson JR, and Daniel PM. A comparative study of the portal vessels connecting the endocrine and exocrine pancreas, with a discussion of some functional implications. *Q J Exp Physiol Cogn Med Sci.* 64: 267–275. 1979. [Medline]
- Lifson N, Kramlinger KG, Mayrand RR, and Lender EJ. Blood flow to the rabbit pancreas with special reference to the islets of Langerhans. *Gastroenterology*. 79: 466–473. 1980. [Medline] [CrossRef]
- King JE, and Ackerman GA. Erythropoiesis in the bone marrow of the fetal rabbit. *Anat Rec.* 157: 589–605. 1967. [Medline] [CrossRef]
- Kennedy DE, Witte PL, and Knight KL. Bone marrow fat and the decline of B lymphopoiesis in rabbits. *Dev Comp Immunol.* 58: 30–39. 2016. [Medline] [CrossRef]
- Bigelow CL, and Tavassoli M. Fatty involution of bone marrow in rabbits. Acta Anat (Basel). 118: 60–64. 1984. [Medline] [CrossRef]
- Elmore SA. Enhanced histopathology of the bone marrow. *Toxicol Pathol.* 34: 666–686. 2006. [Medline] [CrossRef]
- Cloyd GG, and Johnson GR. Lymphosarcoma with lymphoblastic leukemia in a New Zealand White rabbit. *Lab Anim Sci.* 28: 66–69. 1978. [Medline]
- Finnie JW, Bostock DE, and Walden NB. Lymphoblastic leukaemia in a rabbit: a case report. *Lab Anim.* 14: 49–51. 1980. [Medline] [CrossRef]
- Meier H, Fox RR, and Crary DD. Myeloid leukemia in the rabbit (Oryctolagus cuniculus). *Cancer Res.* 32: 1785–1787. 1972. [Medline]
- Toth LA, Olson GA, Wilson E, Rehg JE, and Claassen E. Lymphocytic leukemia and lymphosarcoma in a rabbit. J Am Vet Med Assoc. 197: 627–629. 1990. [Medline]
- Shibuya K, Tajima M, Kanai K, Ihara M, and Nunoya T. Spontaneous lymphoma in a Japanese White rabbit. *J Vet Med Sci.* 61: 1327–1329. 1999. [Medline] [CrossRef]

- Heatley JJ, and Smith AN. Spontaneous neoplasms of lagomorphs. Vet Clin North Am Exot Anim Pract. 7: 561–577, v. 2004. [Medline] [Cross-Ref]
- Gómez L, Gázquez A, Roncero V, Sánchez C, and Durán ME. Lymphoma in a rabbit: histopathological and immunohistochemical findings. J Small Anim Pract. 43: 224–226. 2002. [Medline] [CrossRef]
- White SD, Campbell T, Logan A, et al Lymphoma with cutaneous involvement in three domestic rabbits (Oryctolagus cuniculus). Vet Dermatol. 11: 61–67. 2000. [CrossRef]
- 66. Schummer A, Wilkens H, Vollmerhaus B, and Habermehl K-H. Lymphatic system. In: The Circulatory System, the Skin, and the Cutaneous Organs of the Domestic Mammals. A Schummer, H Wilkens, B Vollmerhaus, and K-H Habermehl (eds). Springer, Massachusetts. 269–440. 1981.
- Gaber W. Morphogenesis of the thymus in rabbit during prenatal and early postnatal periods. *J Electron (China)*. 2017; 10.15761/ HCE.1000118.
- Hassan AU, and Rasool Z. The Hassal of thymus: Hassals corpuscle histological and histopathological perspective. Sch J App Med Sci. 2(1B): 147–148. 2014.
- Bodey B, and Kaiser HE. Development of Hassall's bodies of the thymus in humans and other vertebrates (especially mammals) under physiological and pathological conditions: immunocytochemical, electronmicroscopic and in vitro observations. *In Vivo.* 11: 61–85. 1997. [Medline]
- Müllhaupt D, Wenger S, Kircher P, Pfammatter N, Hatt JM, and Ohlerth S. Computed tomography of the thorax in rabbits: a prospective study in ten clinically healthy New Zealand White rabbits. *Acta Vet Scand.* 59: 72. 2017. [Medline] [CrossRef]
- Maurer JK, Gibbons BA, and Bruce RD. Morphometric assessment of thymic size variation in laboratory rabbits. *Toxicol Pathol.* 18: 407– 411. 1990. [Medline] [CrossRef]
- Dimitrov RS. Comparative ultrasonographic, anatomotopographic and macromorphometric study of the spleen and pancreas in rabbit (Oryctolagus cuniculus). *Not Sci Biol.* 4: 14–20. 2012. [CrossRef]
- Marcato PS, and Rosmini R. Pathology of the rabbit and hare: a color atlas and compendium. Societa Editrice Esculapio, Bologna. 1986.
- Dunne AA, Plehn S, Schulz S, Levermann A, Ramaswamy A, Lippert BM, and Werner JA. Lymph node topography of the head and neck in New Zealand White rabbits. *Lab Anim.* 37: 37–43. 2003. [Medline] [CrossRef]
- Elmore SA. Histopathology of the lymph nodes. *Toxicol Pathol.* 34: 425–454. 2006. [Medline] [CrossRef]
- Soto-Miranda MA, Suami H, and Chang DW. Mapping superficial lymphatic territories in the rabbit. *Anat Rec (Hoboken)*. 296: 965–970. 2013. [Medline] [CrossRef]
- Willard-Mack CL. Normal structure, function, and histology of lymph nodes. *Toxicol Pathol.* 34: 409–424. 2006. [Medline] [CrossRef]
- Lanning D, Zhu X, Zhai S-K, and Knight KL. Development of the antibody repertoire in rabbit: gut-associated lymphoid tissue, microbes, and selection. *Immunol Rev.* 175: 214–228. 2000. [Medline] [CrossRef]
- Cesta MF. Normal structure, function, and histology of mucosa-associated lymphoid tissue. *Toxicol Pathol.* 34: 599–608. 2006. [Medline] [CrossRef]
- Stamatova-Yovcheva K, Dimitrov R, Kostov D, and Yovchev D. Anatomical macromorphological features of the liver in domestic rabbit (Oryctolagus cuniculus). *Trakia Journal of Sciences*. 10: 85–90. 2012.
- Bradley AE. New Zealand White Rabbit. In: Background lesions in laboratory animals: A color atlas, 1st ed. EF McInnes (ed). Saunders (Elsevier), Edinburgh. 87–91. 2012.
- Chandra SA, Stokes AH, Hailey R, Merrill CL, Melich DH, DeSmet K, Furst SM, Peterson RA, Mellon-Kusibab K, and Adler RR. Dermal toxicity studies: factors impacting study interpretation and outcome. *Toxicol Pathol.* 43: 474–481. 2015. [Medline] [CrossRef]
- Florizoone K, van der Luer R, and van den Ingh T. Symmetrical alopecia, scaling and hepatitis in a rabbit. *Vet Dermatol.* 18: 161–164. 2007. [Medline] [CrossRef]

- Ramirez CJ, Kim DY, Hanks BC, and Evans TJ. Copper toxicosis in New Zealand White rabbits (Oryctolagus cuniculus). *Vet Pathol.* 50: 1135–1138. 2013. [Medline] [CrossRef]
- Leipig M, Matiasek K, Rinder H, Janik D, Emrich D, Baiker K, and Hermanns W. Value of histopathology, immunohistochemistry, and real-time polymerase chain reaction in the confirmatory diagnosis of Encephalitozoon cuniculi infection in rabbits. *J Vet Diagn Invest.* 25: 16–26. 2013. [Medline] [CrossRef]
- Gupta BN. Duplication of the gall bladder in a rabbit. *Lab Anim Sci.* 25: 646. 1975. [Medline]
- Lee SP, and Scott AJ. Dihydrocholesterol-induced gallstones in the rabbit: evidence that bile acids cause gallbladder epithelial injury. *Br J Exp Pathol.* 60: 231–238. 1979. [Medline]
- Diribarne M, Mata X, Rivière J, Bouet S, Vaiman A, Chapuis J, Reine F, Fleurot R, Auvinet G, Deretz S, Allain D, Schibler L, Cribiu EP, and Guérin G. LIPH expression in skin and hair follicles of normal coat and Rex rabbits. *PLoS One*. 7: e30073. 2012. [Medline] [CrossRef]
- Jung EC, and Maibach HI. Animal models for percutaneous absorption. J Appl Toxicol. 35: 1–10. 2015. [Medline] [CrossRef]
- Nicoli S, Padula C, Aversa V, Vietti B, Wertz PW, Millet A, Falson F, Govoni P, and Santi P. Characterization of rabbit ear skin as a skin model for in vitro transdermal permeation experiments: histology, lipid composition and permeability. *Skin Pharmacol Physiol.* 21: 218– 226. 2008. [Medline] [CrossRef]
- Anderson JM, and Langone JJ. Issues and perspectives on the biocompatibility and immunotoxicity evaluation of implanted controlled release systems. J Control Release. 57: 107–113. 1999. [Medline] [CrossRef]
- Dincer Z, Jones S, and Haworth R. Preclinical safety assessment of a DNA vaccine using particle-mediated epidermal delivery in domestic pig, minipig and mouse. *Exp Toxicol Pathol.* 57: 351–357. 2006. [Medline] [CrossRef]
- Verdier F, Burnett R, Michelet-Habchi C, Moretto P, Fievet-Groyne F, and Sauzeat E. Aluminium assay and evaluation of the local reaction at several time points after intramuscular administration of aluminium containing vaccines in the Cynomolgus monkey. *Vaccine*. 23: 1359– 1367. 2005. [Medline] [CrossRef]
- Williams DF. Review. Tissue-biomaterial interactions. J Mater Sci. 22: 3421–3445. 1987. [CrossRef]
- Carlisle DB. On the relationship between mammary, sweat, and sebaceous glands. QJ Microsc Sci. 95: 79–83. 1954.
- 96. Diribarne M, Mata X, Chantry-Darmon C, Vaiman A, Auvinet G, Bouet S, Deretz S, Cribiu EP, de Rochambeau H, Allain D, and Guérin G. A deletion in exon 9 of the LIPH gene is responsible for the rex hair coat phenotype in rabbits (Oryctolagus cuniculus). *PLoS One.* 6: e19281. 2011. [Medline] [CrossRef]
- Moore DM, Zimmerman K, and Smith SA. Hematological assessment in pet rabbits: blood sample collection and blood cell identification. *Vet Clin North Am Exot Anim Pract.* 18: 9–19. 2015. [Medline] [CrossRef]
- Fayez I, Marai M, Alnaimy A, and Habeeb M. Thermoregulation in rabbits. In: Rabbit production in hot climates. M Baselga, IFM Marai (eds). Cahiers Options Méditerranéennes, Paris. 33–41. 1994.
- Ootsuka Y, and Tanaka M. Control of cutaneous blood flow by central nervous system. *Temperature*. 2: 392–405. 2015. [Medline] [CrossRef]
- 100. Sayed-Ahmed A, and Elnasharty MA. Pre and postnatal development of the rabbit thin skin. *Glob Vet.* **13**: 622–632. 2014.
- Melo AI, and González-Mariscal G. Communication by olfactory signals in rabbits: its role in reproduction. *Vitam Horm.* 83: 351–371. 2010. [Medline] [CrossRef]
- Mykytowycz R. Further observations on the territorial function and histology of the submandibular cutaneous (chin) glands in the rabbit, Oryctolagus cuniculus (L.). *Anim Behav.* 13: 400–412. 1965. [Medline] [CrossRef]
- Mykytowycz R, and Goodrich BS. Skin glands as organs of communication in mammals. *J Invest Dermatol.* 62: 124–131. 1974. [Medline] [CrossRef]
- Wales NA, and Ebling FJ. The control of the apocrine glands of the rabbit by steroid hormones. *J Endocrinol.* 51: 763–770. 1971. [Medline]

[CrossRef]

- 105. Heath E. Cytologic observations on the secretory cells of the chin (submandibular) gland and brown inguinal gland in the rabbit. *Cell Tissue Res.* 154: 399–408. 1974. [Medline] [CrossRef]
- Montagna W. The brown inguinal glands of the rabbit. Am J Anat. 87: 213–237. 1950. [Medline] [CrossRef]
- 107. Cerbón MA, Camacho-Arroyo I, Gamboa-Domínguez A, and González-Mariscal G. The rabbit submandibular gland: sexual dimorphism, effects of gonadectomy, and variations across the female reproductive cycle. J Comp Physiol A Neuroethol Sens Neural Behav Physiol. 178: 351–357. 1996. [Medline] [CrossRef]
- Fehr M, and Koestlinger S. Ectoparasites in small exotic mammals. Vet Clin North Am Exot Anim Pract. 16: 611–657. 2013. [Medline] [Cross-Ref]
- Hoppmann E, and Barron HW. Ferret and rabbit dermatology. J Exot Pet Med. 16: 225–237. 2007. [CrossRef]
- Snook TS, White SD, Hawkins MG, Tell LA, Wilson LS, Outerbridge CA, and Ihrke PJ. Skin diseases in pet rabbits: a retrospective study of 334 cases seen at the University of California at Davis, USA (1984– 2004). *Vet Dermatol.* 24: 613–617, e148. 2013. [Medline] [CrossRef]
- 111. Manjunatha DR, Mahesh V, and Ranganath L. Surgical management of aural hematoma in Russian Grey Giant rabbit - a case report. *International Journal of Agricultural Sciences and Veterinary Medicine*. 2: 37–38. 2014.
- d'Ovidio D, and Santoro D. Orodental diseases and dermatological disorders are highly associated in pet rabbits: a case-control study. *Vet Dermatol.* 24: 531–e125. 2013. [Medline] [CrossRef]
- 113. Harcourt-Brown F. Textbook of Rabbit Medicine. Reed Educational and Professional Publishing Ltd, Oxford. 2002.
- Kerr PJ, Liu J, Cattadori I, Ghedin E, Read AF, and Holmes EC. Myxoma virus and the Leporipoxviruses: an evolutionary paradigm. *Viruses*. 7: 1020–1061. 2015. [Medline] [CrossRef]
- 115. Meredith AL. Viral skin diseases of the rabbit. Vet Clin North Am Exot Anim Pract. 16: 705–714. 2013. [Medline] [CrossRef]
- Kanfer S, and Reavill DR. Cutaneous neoplasia in ferrets, rabbits, and guinea pigs. Vet Clin North Am Exot Anim Pract. 16: 579–598. 2013. [Medline] [CrossRef]
- 117. von Bomhard W, Goldschmidt MH, Shofer FS, Perl L, Rosenthal KL, and Mauldin EA. Cutaneous neoplasms in pet rabbits: a retrospective study. *Vet Pathol.* 44: 579–588. 2007. [Medline] [CrossRef]
- Florizoone K. Thymoma-associated exfoliative dermatitis in a rabbit. Vet Dermatol. 16: 281–284. 2005. [Medline] [CrossRef]
- Jassies-van der Lee A, van Zeeland Y, Kik M, and Schoemaker N. Successful treatment of sebaceous adenitis in a rabbit with ciclosporin and triglycerides. *Vet Dermatol.* 20: 67–71. 2009. [Medline] [CrossRef]
- 120. Rostaher Prélaud A, Jassies-van der Lee A, Mueller RS, van Zeeland YR, Bettenay S, Majzoub M, Zenker I, and Hein J. Presumptive paraneoplastic exfoliative dermatitis in four domestic rabbits. *Vet Rec.* 172: 155. 2013. [Medline]
- Chandra SA, Peterson RA, Melich D, Merrill CM, Bailey D, Mellon-Kusibab K, and Adler R. Dermal irritation of petrolatum in rabbits but not in mice, rats or minipigs. *J Appl Toxicol.* 34: 857–861. 2014. [Medline] [CrossRef]
- Rigdon RH. Local reaction to polyurethane--a comparative study in the mouse, rat, and rabbit. *J Biomed Mater Res.* 7: 79–93. 1973. [Medline] [CrossRef]
- 123. Buja LM, Kita T, Goldstein JL, Watanabe Y, and Brown MS. Cellular pathology of progressive atherosclerosis in the WHHL rabbit. An animal model of familial hypercholesterolemia. *Arteriosclerosis.* 3: 87–101. 1983. [Medline] [CrossRef]
- Nakano A, Kinoshita M, Okuda R, Yasuda T, Abe M, and Shiomi M. Pathogenesis of tendinous xanthoma: histopathological study of the extremities of Watanabe heritable hyperlipidemic rabbits. *J Orthop Sci.* 11: 75–80. 2006. [Medline] [CrossRef]
- Watanabe Y. Serial inbreeding of rabbits with hereditary hyperlipidemia (WHHL-rabbit). *Atherosclerosis*. 36: 261–268. 1980. [Medline] [CrossRef]

- Mitsuguchi Y, Ito T, and Ohwada K. Pathologic findings in rabbit models of hereditary hypertriglyceridemia and hereditary postprandial hypertriglyceridemia. *Comp Med.* 58: 465–480. 2008. [Medline]
- Quinton JF, Prelaud P, Poujade A, and Faivre NC. A case of actinic keratosis in a rabbit. J Exot Pet Med. 23: 283–286. 2014. [CrossRef]
- Calvert DT, Knight CH, and Peaker M. Milk accumulation and secretion in the rabbit. *Q J Exp Physiol.* **70**: 357–363. 1985. [Medline] [CrossRef]
- Propper AY, Howard BA, and Veltmaat JM. Prenatal morphogenesis of mammary glands in mouse and rabbit. *J Mammary Gland Biol Neopla*sia. 18: 93–104. 2013. [Medline] [CrossRef]
- Caba M, and González-Mariscal G. The rabbit pup, a natural model of nursing-anticipatory activity. *Eur J Neurosci.* 30: 1697–1706. 2009. [Medline] [CrossRef]
- Adlam C, Thorley CM, Ward PD, Collins M, Lucken RN, and Knight PA. Natural and experimental staphylococcal mastitis in rabbits. J Comp Pathol. 86: 581–593. 1976. [Medline] [CrossRef]
- Viana D, Selva L, Callanan JJ, Guerrero I, Ferrian S, and Corpa JM. Strains of Staphylococcus aureus and pathology associated with chronic suppurative mastitis in rabbits. *Vet J.* **190**: 403–407. 2011. [Medline] [CrossRef]
- Greene HS. Familial mammary tumors in the rabbit : li. Gross and microscopic pathology. *J Exp Med.* 70: 159–166. 1939. [Medline] [Cross-Ref]
- 134. Lipman NS, Zhao ZB, Andrutis KA, Hurley RJ, Fox JG, and White HJ. Prolactin-secreting pituitary adenomas with mammary dysplasia in New Zealand White rabbits. *Lab Anim Sci.* 44: 114–120. 1994. [Medline]
- 135. Sikoski P, Trybus J, Cline JM, Muhammad FS, Eckhoff A, Tan J, Lockard M, Jolley T, Britt S, and Kock ND. Cystic mammary adenocarcinoma associated with a prolactin-secreting pituitary adenoma in a New Zealand White rabbit (Oryctolagus cuniculus). *Comp Med.* 58: 297–300. 2008. [Medline]
- Petraitiene R, Petraitis V, Bacher J, Das SR, Parlow AF, and Walsh TJ. Cyclosporine A-induced mammary hyperplasia and hyperprolactinemia in New Zealand White rabbits. *Comp Med.* 51: 430–435. 2001. [Medline]
- 137. Krimer PM, Harvey SB, Blas-Machado U, Lauderdale JD, and Moore PA. Reversible fibroadenomatous mammary hyperplasia in male and female New Zealand White rabbits associated with cyclosporine A administration. *Vet Pathol.* 46: 1144–1148. 2009. [Medline] [CrossRef]
- Maratea KA, Ramos-Vara JA, Corriveau LA, and Miller MA. Testicular interstitial cell tumor and gynecomastia in a rabbit. *Vet Pathol.* 44: 513–517. 2007. [Medline] [CrossRef]
- 139. Shek JW, Wen GY, and Wisniewski HM. Atlas of the rabbit brain and spinal cord. Karger AG, Basel. 1986.
- 140. Pardo ID, Weber K, Cramer S, Krinke GJ, Butt MT, Sharma AK, and Bolon B. Atlas of normal microanatomy, procedural and processing artifacts, common background findings, and neurotoxic lesions in the peripheral nervous system of laboratory animals. *Toxicol Pathol.* 48: 105–131. 2020. [Medline] [CrossRef]
- Lindsey JR, and Fox RR. Inherited diseases and variations. In: Biology of the Laboratory Rabbit, 2nd ed. PJ Manning, DH Ringler, and CE Newcomer (eds). Academic Press, Orlando. 293–319. 1994.
- 142. Mooney MP, Aston CE, Siegel MI, Losken HW, Smith TD, Burrows AM, Wenger SL, Caruso K, Siegel B, and Ferrell RE. Craniosynostosis with autosomal dominant transmission in New Zealand White rabbits. *J Craniofac Genet Dev Biol.* 16: 52–63. 1996. [Medline]
- Rudmann D, and Colleton C. Interstitial adipocytes in the Beagle dog and New Zealand White rabbit choroid plexus. *Toxicol Pathol.* 47: 553– 555. 2019. [Medline] [CrossRef]
- 144. Ivens IA, Achanzar W, Baumann A, Brändli-Baiocco A, Cavagnaro J, Dempster M, Depelchin BO, Rovira AR, Dill-Morton L, Lane JH, Reipert BM, Salcedo T, Schweighardt B, Tsuruda LS, Turecek PL, and Sims J. PEGylated biopharmaceuticals: current experience and considerations for nonclinical development. *Toxicol Pathol.* 43: 959–983. 2015. [Medline] [CrossRef]

- Swank RL. The pyramidal tracts: an experimental study of the corticospinal and other components in the rabbit. *Arch Neurol Psychiatry*. 36: 530–541. 1936. [CrossRef]
- 146. Dixon D, Alison R, Bach U, Colman K, Foley GL, Harleman JH, Haworth R, Herbert R, Heuser A, Long G, Mirsky M, Regan K, Van Esch E, Westwood FR, Vidal J, and Yoshida M. Nonproliferative and proliferative lesions of the rat and mouse female reproductive system. J Toxicol Pathol. 27(Suppl): 1S–107S. 2014. [Medline] [CrossRef]
- 147. Limon M. Etude histologique et histogenetique de la glande interstitielle de ovaire. Arch d'Anat micr. 5: 155–190. 1902.
- 148. Mori H, and Matsumoto K. Development of the secondary interstitial gland in the rabbit ovary. *J Anat.* **116**: 417–430. 1973. [Medline]
- Nicosia SV, and Johnson JH. Surface morphology of ovarian mesothelium (surface epithelium) and of other pelvic and extrapelvic mesothelial sites in the rabbit. *Int J Gynecol Pathol.* 3: 249–260. 1984. [Medline] [CrossRef]
- Barberini F, Correr S, De Santis F, and Motta PM. The epithelium of the rabbit vagina: a microtopographical study by light, transmission and scanning electron microscopy. *Arch Histol Cytol.* 54: 365–378. 1991. [Medline] [CrossRef]
- 151. Barberini F, Sartori S, and Motta P. Changes in the surface morphology of the rabbit endometrium related to the estrous and progestational stages of the reproductive cycle a scanning and transmission electron microscopic study. *Cell Tissue Res.* **190**: 207–222. 1978. [Medline] [CrossRef]
- Bray MV, Weir EC, Brownstein DG, and Delano ML. Endometrial venous aneurysms in three New Zealand White rabbits. *Lab Anim Sci.* 42: 360–362. 1992. [Medline]
- Zook BC, Spiro I, and Hertz R. Malignant neoplasms of decidual origin (deciduosarcomas) induced by estrogen-progestin-releasing intravaginal devices in rabbits. *Am J Pathol.* **128**: 315–327. 1987. [Medline]
- 154. Zook BC, Jänne OA, Abraham AA, and Nash HA. The development and regression of deciduosarcomas and other lesions caused by estrogens and progestins in rabbits. *Toxicol Pathol.* 29: 411–416. 2001. [Medline] [CrossRef]
- Saito K, Nakanishi M, and Hasegawa A. Uterine disorders diagnosed by ventrotomy in 47 rabbits. *J Vet Med Sci.* 64: 495–497. 2002. [Medline] [CrossRef]
- 156. Jänne OA, Zook BC, Didolkar AK, Sundaram K, and Nash HA. The roles of estrogen and progestin in producing deciduosarcoma and other lesions in the rabbit. *Toxicol Pathol.* 29: 417–421. 2001. [Medline] [CrossRef]
- Cooper TK, Adelsohn D, and Gilbertson SR. Spontaneous deciduosarcoma in a domestic rabbit (Oryctolagus cuniculus). *Vet Pathol.* 43: 377–380. 2006. [Medline] [CrossRef]
- Morton D, Weisbrode SE, Wyder WE, Maurer JK, and Capen CC. Spermatid giant cells, tubular hypospermatogenesis, spermatogonial swelling, cytoplasmic vacuoles, and tubular dilatation in the testes of normal rabbits. *Vet Pathol.* 23: 176–183. 1986. [Medline] [CrossRef]
- Zwicker GM, and Killinger JM. Interstitial cell tumors in a young adult New Zealand White rabbit. *Toxicol Pathol.* 13: 232–235. 1985. [Medline] [CrossRef]
- Reineking W, Seehusen F, Lehmbecker A, and Wohlsein P. Predominance of granular cell tumours among testicular tumours of rabbits (Oryctolagus cuniculi f. dom.). *J Comp Pathol.* **173**: 24–29. 2019. [Medline] [CrossRef]
- Elchlepp JG. The urogenital organs of the Cottontail rabbit (Sylilagus floridanus). J Morphol. 91: 169–198. 1952. [CrossRef]
- 162. Holtz W, and Foote RH. The anatomy of the reproductive system in male Dutch rabbits (Oryctolagus cuniculus) with special emphasis on the accessory sex glands. *J Morphol.* 158: 1–20. 1978. [Medline] [CrossRef]
- 163. Skonieczna J, Madej JP, and Będziński R. Accessory genital glands in the New Zealand White rabbit: A morphometrical and histological study. J Vet Res (Pulawy). 63: 251–257. 2019. [Medline] [CrossRef]
- Zwicker GM, Killinger JM, and McConnel RF. Spontaneous vesicular and prostatic gland epithelial squamous metaplasia, hyperplasia, and

keratinized nodule formation in rabbits. *Toxicol Pathol.* **13**: 222–228. 1985. [Medline] [CrossRef]

- Carr EB. The vesicular seminialis of the rabbit. Proc Zool Soc Lond. 124: 675–683. 1954. [CrossRef]
- 166. Uthamanthil RK, Hachem RY, Gagea M, Reitzel RA, Borne AT, and Tinkey PT. Urinary catheterization of male rabbits: a new technique and a review of urogenital anatomy. J Am Assoc Lab Anim Sci. 52: 180–185. 2013. [Medline]
- Tsenova LHR, Ellison E, Manca C, and Kaplan G. Aerosol exposure system for rabbits: application to M. tuberculosis infection. *Appl Bio*saf. 11: 7–14. 2006. [CrossRef]
- Johnson-Delaney CA, and Orosz SE. Rabbit respiratory system: clinical anatomy, physiology and disease. *Vet Clin North Am Exot Anim Pract.* 14: 257–266, vi. 2011. [Medline] [CrossRef]
- 169. Pereira ME, Macri NP, and Creasy DM. Evaluation of the rabbit nasal cavity in inhalation studies and a comparison with other common laboratory species and man. *Toxicol Pathol.* **39**: 893–900. 2011. [Medline] [CrossRef]
- Phaneuf LR, Barker S, Groleau MA, and Turner PV. Tracheal injury after endotracheal intubation and anesthesia in rabbits. *J Am Assoc Lab Anim Sci.* 45: 67–72. 2006. [Medline]
- Choi HK, Finkbeiner WE, and Widdicombe JH. A comparative study of mammalian tracheal mucous glands. *J Anat.* 197: 361–372. 2000. [Medline] [CrossRef]
- Plopper CG, Halsebo JE, Berger WJ, Sonstegard KS, and Nettesheim P. Distribution of nonciliated bronchiolar epithelial (Clara) cells in intra- and extrapulmonary airways of the rabbit. *Exp Lung Res.* 5: 79–98. 1983. [Medline] [CrossRef]
- 173. Widdicombe JH, Chen LL, Sporer H, Choi HK, Pecson IS, and Bastacky SJ. Distribution of tracheal and laryngeal mucous glands in some rodents and the rabbit. *J Anat.* 198: 207–221. 2001. [Medline] [CrossRef]
- 174. Simons RS. Lung morphology of cursorial and non-cursorial mammals: lagomorphs as a case study for a pneumatic stabilization hypothesis. *J Morphol.* 230: 299–316. 1996. [Medline] [CrossRef]
- Bal HS, and Ghoshal NG. Morphology of the terminal bronchiolar region of common laboratory mammals. *Lab Anim.* 22: 76–82. 1988. [Medline] [CrossRef]
- Plopper CG, Mariassy AT, Wilson DW, Alley JL, Nishio SJ, and Nettesheim P. Comparison of nonciliated tracheal epithelial cells in six mammalian species: ultrastructure and population densities. *Exp Lung Res.* 5: 281–294. 1983. [Medline] [CrossRef]
- Cooper TK, Griffith JW, Chroneos ZC, Izer JM, Willing LB, and Peng X. Spontaneous lung lesions in aging laboratory rabbits (Oryctolagus cuniculus). *Vet Pathol.* 54: 178–187. 2017. [Medline] [CrossRef]
- Dale K. A method for inducing unilateral silicosis in rabbits by an injection technique with some observations on lung clearance and quantitative evaluation of experimental silicosis. *Scand J Respir Dis.* 54: 157–167. 1973. [Medline]
- Dale K. Early effects of quartz and titanium dioxide dust on pulmonary function and tissue. An experimental study on rabbits. *Scand J Respir Dis.* 54: 168–184. 1973. [Medline]
- Wallace WE, Gupta NC, Hubbs AF, Mazza SM, Bishop HA, Keane MJ, Battelli LA, Ma J, and Schleiff P. Cis-4-[(18)F]fluoro-L-proline PET imaging of pulmonary fibrosis in a rabbit model. *J Nucl Med.* 43: 413–420. 2002. [Medline]
- Fox RR, and Crary DD. Hereditary diaphragmatic hernia in the rabbit. J Hered. 64: 333–336. 1973. [Medline] [CrossRef]
- 182. Barros SS, Soares MP, and Gimeno EJ. Macrophages and giant cell proliferation associated with bone protein synthesis and calcification in the trachea and bronchi of rabbits intoxicated with Solanum glaucophyllum. *Vet Pathol.* **43**: 494–499. 2006. [Medline] [CrossRef]
- Wang X, Mabrey JD, and Agrawal CM. An interspecies comparison of bone fracture properties. *Biomed Mater Eng.* 8: 1–9. 1998. [Medline]
- 184. Muschler GF, Raut VP, Patterson TE, Wenke JC, and Hollinger JO. The design and use of animal models for translational research in bone tissue engineering and regenerative medicine. *Tissue Eng Part B Rev.*

16: 123-145. 2010. [Medline] [CrossRef]

- Pearce AI, Richards RG, Milz S, Schneider E, and Pearce SG. Animal models for implant biomaterial research in bone: a review. *Eur Cell Mater.* 13: 1–10. 2007. [Medline] [CrossRef]
- Haschek WM, Rousseaux CG, and Wallig MA. Fundamentals of Toxicologic Pathology. 2nd ed. Academic Press (Elsevier), Amsterdam. 2010.
- 187. Wancket LM. Animal models for evaluation of bone implants and devices: comparative bone structure and common model uses. *Vet Pathol.* 52: 842–850. 2015. [Medline] [CrossRef]
- Reinholz GG, Lu L, Saris DB, Yaszemski MJ, and O'Driscoll SW. Animal models for cartilage reconstruction. *Biomaterials*. 25: 1511–1521. 2004. [Medline] [CrossRef]
- Chu CR, Szczodry M, and Bruno S. Animal models for cartilage regeneration and repair. *Tissue Eng Part B Rev.* 16: 105–115. 2010. [Medline] [CrossRef]
- O'Driscoll SW, Saris DB, Ito Y, and Fitzimmons JS. The chondrogenic potential of periosteum decreases with age. *J Orthop Res.* 19: 95–103. 2001. [Medline] [CrossRef]
- 191. Shively MJ. Xeroradiographic anatomy of the domesticated rabbit (Oryctolagus cuniculus) part I: head, thorax, and thoracic limb. Swest Vet. 32: 219–233. 1979.
- Shively MJ. Xeroradiographic anatomy of the domesticated rabbit (Oryctolagus cuniculus) part II:abdomen, pelvis, and pelvic limb. *Swest Vet.* 33: 57–67. 1980.
- Greenaway JB, Partlow GD, Gonsholt NL, and Fisher KR. Anatomy of the lumbosacral spinal cord in rabbits. J Am Anim Hosp Assoc. 37: 27–34. 2001. [Medline] [CrossRef]
- Proffen BL, McElfresh M, Fleming BC, and Murray MM. A comparative anatomical study of the human knee and six animal species. *Knee*. 19: 493–499. 2012. [Medline] [CrossRef]
- 195. Brock K, Gallaugher L, Bergdall VK, and Dysko RC. Chapter 17: Mycoses and Non-Infectious Diseases. In: The Laboratory Rabbit, Guinea Pig, Hamster, and Other Rodents, 1st ed. MA Suckow, KA Stevens, and RP Wilson (eds). Academic Press (Elsevier), Amsterdam. 504–529. 2012:
- 196. Ikemura S, Yamamoto T, Nishida K, Motomura G, and Iwamoto Y. Gender difference in the development of steroid-induced osteonecrosis in rabbits. *Rheumatology (Oxford)*. 49: 1128–1132. 2010. [Medline] [CrossRef]
- 197. Kabata T, Kubo T, Matsumoto T, Hirata T, Fujioka M, Takahashi KA, Yagishita S, Kobayashi M, and Tomita K. Onset of steroid-induced osteonecrosis in rabbits and its relationship to hyperlipaemia and increased free fatty acids. *Rheumatology (Oxford)*. 44: 1233–1237. 2005. [Medline] [CrossRef]
- Nehrbass D, Arens D, and Zeiter S. Spontaneous bilateral avulsion fracture of the tuberositas tibiae in a New Zealand White rabbit a counterpart to Osgood-Schlatter disease in humans? *Exp Toxicol Pathol.* 67: 223–227. 2015. [Medline] [CrossRef]
- 199. Gough A, Johnson R, Campbell E, Hall L, Tylor J, Carpenter A, Black W, Basrur PK, Baragi VM, Sigler R, and Metz A. Quinolone arthropathy in immature rabbits treated with the fluoroquinolone, PD 117596. *Exp Toxicol Pathol.* 48: 225–232. 1996. [Medline] [CrossRef]
- 200. Mainil-Varlet P, Van Damme B, Nesic D, Knutsen G, Kandel R, and Roberts S. A new histology scoring system for the assessment of the quality of human cartilage repair: ICRS II. *Am J Sports Med.* 38: 880– 890. 2010. [Medline] [CrossRef]
- Laverty S, Girard CA, Williams JM, Hunziker EB, and Pritzker KP. The OARSI histopathology initiative - recommendations for histological assessments of osteoarthritis in the rabbit. *Osteoarthritis Cartilage*. 18(Suppl 3): S53–S65. 2010. [Medline] [CrossRef]
- Bendele AM. Animal models of osteoarthritis. J Musculoskelet Neuronal Interact. 1: 363–376. 2001. [Medline]
- Derry DM, Morrow E, Sadre N, and Flattery KV. Brown and white fat during the life of the rabbit. *Dev Biol.* 27: 204–216. 1972. [Medline] [CrossRef]
- 204. Baradi AF, and Hope J. Observations on ultrastructure of rabbit meso-

thelium. Exp Cell Res. 34: 33-44. 1964. [Medline] [CrossRef]

- Miao CY, and Li ZY. The role of perivascular adipose tissue in vascular smooth muscle cell growth. *Br J Pharmacol.* 165: 643–658. 2012. [Medline] [CrossRef]
- 206. Kempson RL, Fletcher CDM, Evans HL, Hendrickson MR, and Sibley RK. Atlas of Tumor Pathology, Tumors of the Soft Tissues. Armed Forces Institute of Pathology, Washington, D.C. 2001.
- 207. Calgüner E, Gözil R, Erdoğan D, Kurt I, Keskil S, Elmas C, and Sabuncuoğlu H. Atrophic and regenerative changes in rabbit mimic muscles after lidocaine and bupivacaine application. *Anat Histol Embryol.* 32: 54–59. 2003. [Medline] [CrossRef]
- Thuilliez C, Dorso L, Howroyd P, Gould S, Chanut F, and Burnett R. Histopathological lesions following intramuscular administration of saline in laboratory rodents and rabbits. *Exp Toxicol Pathol.* 61: 13–21. 2009. [Medline] [CrossRef]
- 209. Osheroff MR, Kobs DJ, Buccellato M, Croutch CR, Elcock LE, Burback BL, and Johnson JD. Comparative toxicology studies in Sprague-Dawley rats, Rhesus monkeys, and New Zealand White rabbits to determine a no observed adverse effect level for 1,1'-methylenebis[4-[(hydroxyimino)methyl]-pyridinium] dimethanesulfonate. Int J Toxicol. 32(Suppl): 59S-74S. 2013. [Medline] [CrossRef]
- Afifi AD, Al-Gailany AM, Salman JM, and Bahuth NB. Nerve and muscle in steroid-induced weakness in the rabbit. *Arch Phys Med Rehabil.* 58: 143–148. 1977. [Medline]
- Brucker M, Sati S, Spangenberger A, and Weinzweig J. Long-term fate of transplanted autologous fat in a novel rabbit facial model. *Plast Reconstr Surg.* 122: 749–754. 2008. [Medline] [CrossRef]
- Mendlowski B. Neuromuscular lesions in restrained rabbits. Vet Pathol. 12: 378–386. 1975. [Medline] [CrossRef]
- Serfilippi LM, Saladino BH, and Spainhour CB. Sarcocystis Infection in Laboratory Rabbits. Comp Med. 70: 300–301. 2020. [Medline] [CrossRef]
- 214. Abdo M, Haddad S, and Emam M. Development of the New Zealand White rabbit eye: I. Pre- and postnatal development of eye tunics. *Anat Histol Embryol.* 46: 423–430. 2017. [Medline] [CrossRef]
- Reichenbach A, Schnitzer J, Friedrich A, Knothe AK, and Henke A. Development of the rabbit retina: II. Müller cells. *J Comp Neurol.* 311: 33–44. 1991. [Medline] [CrossRef]
- Reichenbach A, Schnitzer J, Friedrich A, Ziegert W, Brückner G, and Schober W. Development of the rabbit retina. I. Size of eye and retina, and postnatal cell proliferation. *Anat Embryol (Berl).* 183: 287–297. 1991. [Medline]
- Sheppard LB. The anatomy and histology of the normal rabbit eye with special reference to the ciliary zone. *Arch Ophthalmol.* 66: 896–904.
 1961. [Medline] [CrossRef]
- Tauber H, Wachneldt TV, and Neuhoff V. Myelination in rabbit optic nerves is accelerated by artificial eye opening. *Neurosci Lett.* 16: 235–238. 1980. [Medline] [CrossRef]
- 219. Davis FA. The Anatomy and Histology of the Eye and Orbit of the Rabbit. *Trans Am Ophthalmol Soc.* **27**: 2–441, 402–441. 1929. [Medline]
- 220. Peiffer RL Jr, Pohm-Thorsen L, and Corcoran K. Models in Opthalmology and Vision Research. In: The Biology of the Laboratory Rabbit, 2nd ed. PJ Manning, DH Ringler, and CE Newcomer (eds). Academic Press, San Diego. 409–433. 1994.
- Prince JH. Cornea, Trabecular Region, and Sclera. In: The Rabbit in Eye Research. 1st ed. Prince JH (ed). Charles C Thomas Publisher, Illinois. 86–139. 1964.
- Williams D. The Rabbit. In: Veterinary Opthalmology: Vol II, 5th ed. KN Gelatt, BC Gilger, TJ Kern (eds). Wiley-Blackwell, Iowa. 1725– 1749. 2013.
- Hughes A. Topographical relationships between the anatomy and physiology of the rabbit visual system. *Doc Ophthalmol.* 30: 33–159. 1971. [Medline] [CrossRef]
- 224. Hughes A. A schematic eye for the rabbit. *Vision Res.* **12**: 123–138. 1972. [Medline] [CrossRef]
- 225. Greiner JV, Glonek T, Korb DR, Whalen AC, Hebert E, Hearn SL, Esway JE, and Leahy CD. Volume of the human and rabbit meibo-

mian gland system. *Adv Exp Med Biol.* **438**: 339–343. 1998. [Medline] [CrossRef]

- Jester JV, Nicolaides N, and Smith RE. Meibomian gland studies: histologic and ultrastructural investigations. *Invest Ophthalmol Vis Sci.* 20: 537–547. 1981. [Medline]
- Lambert RW, and Smith RE. Pathogenesis of blepharoconjunctivitis complicating 13-cis-retinoic acid (isotretinoin) therapy in a laboratory model. *Invest Ophthalmol Vis Sci.* 29: 1559–1564. 1988. [Medline]
- 228. Bergmanson JP, Doughty MJ, and Blocker Y. The acinar and ductal organisation of the tarsal accessory lacrimal gland of wolfring in rabbit eyelid. *Exp Eye Res.* **68**: 411–421. 1999. [Medline] [CrossRef]
- Chang M, Lee Y, and Baek S. The functional and histopathologic change in the levator palpebrae superioris and Müller muscle after subconjunctival injection of triamcinolone acetonide. *J Craniofac Surg.* 26: 285–289. 2015. [Medline] [CrossRef]
- Francis BA, Weiland JD, Sachs NA, and Chang EL. Benzalkonium chloride-induced denervation of orbicularis oculi muscle in rabbits. *Invest Ophthalmol Vis Sci.* 54: 1868–1872. 2013. [Medline] [CrossRef]
- Murphy EH, Garone M, Tashayyod D, and Baker RB. Innervation of extraocular muscles in the rabbit. *J Comp Neurol.* 254: 78–90. 1986. [Medline] [CrossRef]
- Prangen AD. A Study of the Comparative Anatomy of the Extra-ocular Muscles. *Trans Am Ophthalmol Soc.* 26: 353–380. 1928. [Medline]
- Sheppard LB. The anatomy and histology of the normal rabbit eye with special reference to the ciliary zone. *Arch Ophthalmol.* 67: 87–100.
 1962. [Medline] [CrossRef]
- 234. Prince JH, Diesem CD, Eglitis I, and Ruskell GL. The Rabbit. In: Anatomy and Histology of the Eye and Orbit in Domestic Animals, 1st ed. JH Prince, CD Diesem, I Eglitis, and GL Ruskell (eds). Charles C Thomas Publisher, Illinois. 260–297. 1960.
- 235. Prince JH, and Eglitis I. Uvea. In: The Rabbit in Eye Research. JH Prince (ed). Charles C Thomas Publisher, Illinois. 140–171. 1964.
- Butovich IA, Lu H, McMahon A, and Eule JC. Toward an animal model of the human tear film: biochemical comparison of the mouse, canine, rabbit, and human meibomian lipidomes. *Invest Ophthalmol Vis Sci.* 53: 6881–6896. 2012. [Medline] [CrossRef]
- 237. Korb DR, Greiner JV, Glonek T, Whalen A, Hearn SL, Esway JE, and Leahy CD. Human and rabbit lipid layer and interference pattern observations. *Adv Exp Med Biol.* 438: 305–308. 1998. [Medline] [Cross-Ref]
- Leonard BC, Yañez-Soto B, Raghunathan VK, Abbott NL, and Murphy CJ. Species variation and spatial differences in mucin expression from corneal epithelial cells. *Exp Eye Res.* 152: 43–48. 2016. [Medline] [CrossRef]
- Wei XE, Markoulli M, Zhao Z, and Willcox MD. Tear film break-up time in rabbits. *Clin Exp Optom.* 96: 70–75. 2013. [Medline] [Cross-Ref]
- Cain C, and Phillips TE. Developmental changes in conjunctiva-associated lymphoid tissue of the rabbit. *Invest Ophthalmol Vis Sci.* 49: 644–649. 2008. [Medline] [CrossRef]
- Chodosh J, Nordquist RE, and Kennedy RC. Comparative anatomy of mammalian conjunctival lymphoid tissue: a putative mucosal immune site. *Dev Comp Immunol.* 22: 621–630. 1998. [Medline] [CrossRef]
- Knop E, and Knop N. The role of eye-associated lymphoid tissue in corneal immune protection. J Anat. 206: 271–285. 2005. [Medline] [CrossRef]
- Andrew SE. Corneal diseases of rabbits. Vet Clin North Am Exot Anim Pract. 5: 341–356. 2002. [Medline] [CrossRef]
- Delaney KH. Diagnostic exercise: apparent corneal occlusion in a New Zealand White rabbit. *Contemp Top Lab Anim Sci.* 34: 76–77. 1995. [Medline]
- Katsuta O, Shinomiya K, Mochizuki T, Kikkawa C, Yoshimi M, and Ikuse T. Pseudopterygium: unique conjunctival stricture observed in Japanese White rabbit. *J Toxicol Pathol.* 21: 239–241. 2008. [CrossRef]
- 246. Kim JY, Williams DL, Rho KS, Kim KH, Lee YS, and Jeong SW. Surgical correction of aberrant conjunctival overgrowth in a rabbit: a case report. *Ir Vet J.* 66: 18. 2013. [Medline] [CrossRef]

- Roze M, Ridings B, and Lagadic M. Comparative morphology of epicorneal conjunctival membranes in rabbits and human pterygium. *Vet Ophthalmol.* 4: 171–174. 2001. [Medline] [CrossRef]
- Wagner F, and Fehr M. Common ophthalmic problems in pet rabbits. J Exot Pet Med. 6: 158–167. 2007. [CrossRef]
- Chan T, Payor S, and Holden BA. Corneal thickness profiles in rabbits using an ultrasonic pachometer. *Invest Ophthalmol Vis Sci.* 24: 1408– 1410. 1983. [Medline]
- Ichijima H, Petroll WM, Jester JV, Barry PA, Andrews PM, Dai M, and Cavanagh HD. In vivo confocal microscopic studies of endothelial wound healing in rabbit cornea. *Cornea*. 12: 369–378. 1993. [Medline] [CrossRef]
- 251. Labbé A, Liang H, Martin C, Brignole-Baudouin F, Warnet JM, and Baudouin C. Comparative anatomy of laboratory animal corneas with a new-generation high-resolution in vivo confocal microscope. *Curr Eye Res.* **31**: 501–509. 2006. [Medline] [CrossRef]
- Minkowski JS, Bartels SP, Delori FC, Lee SR, Kenyon KR, and Neufeld AH. Corneal endothelial function and structure following cryo-injury in the rabbit. *Invest Ophthalmol Vis Sci.* 25: 1416–1425. 1984. [Medline]
- Sailstad DM, and Peiffer RL Jr. Specular microscopic observations of the corneal endothelium in the normal rabbit. *Lab Anim.* 15: 393–395. 1981. [Medline] [CrossRef]
- 254. Wang X, Dong J, and Wu Q. Mean central corneal thickness and corneal power measurements in pigmented and white rabbits using Visante optical coherence tomography and ATLAS corneal topography. *Vet Ophthalmol.* 17: 87–90. 2014. [Medline] [CrossRef]
- Wang X, and Wu Q. Normal corneal thickness measurements in pigmented rabbits using spectral-domain anterior segment optical coherence tomography. *Vet Ophthalmol.* 16: 130–134. 2013. [Medline] [CrossRef]
- 256. Yüksel H, Türkcü FM, Ari Ş, Çinar Y, Cingü AK, Şahin M, Şahin A, Özkurt Z, and Çaça İ. Anterior segment parameters of rabbits with rotating Scheimpflug camera. *Vet Ophthalmol.* 18: 210–213. 2015. [Medline] [CrossRef]
- 257. Bechmann M, Thiel MJ, Neubauer AS, Ullrich S, Ludwig K, Kenyon KR, and Ulbig MW. Central corneal thickness measurement with a retinal optical coherence tomography device versus standard ultrasonic pachymetry. *Cornea.* 20: 50–54. 2001. [Medline] [CrossRef]
- Cintron C, Covington H, and Kublin CL. Morphogenesis of rabbit corneal stroma. *Invest Ophthalmol Vis Sci.* 24: 543–556. 1983. [Medline]
- Kaye GI, and Pappas GD. Studies on the cornea. I. The fine structure of the rabbit cornea and the uptake and transport of colloidal particles by the cornea in vivo. *J Cell Biol.* 12: 457–479. 1962. [Medline] [CrossRef]
- Ojeda JL, Ventosa JA, and Piedra S. The three-dimensional microanatomy of the rabbit and human cornea. A chemical and mechanical microdissection-SEM approach. *J Anat.* 199: 567–576. 2001. [Medline] [CrossRef]
- Wilson SE, and Hong JW. Bowman's layer structure and function: critical or dispensable to corneal function? A hypothesis. *Cornea*. 19: 417–420. 2000. [Medline] [CrossRef]
- Staatz WD, and Van Horn DL. The effects of aging and inflammation on corneal endothelial wound healing in rabbits. *Invest Ophthalmol Vis Sci.* 19: 983–986. 1980. [Medline]
- Van Horn DL, Sendele DD, Seideman S, and Buco PJ. Regenerative capacity of the corneal endothelium in rabbit and cat. *Invest Ophthalmol Vis Sci.* 16: 597–613. 1977. [Medline]
- 264. Wagner F, Brügmann M, Drommer W, and Fehr M. Corneal dermoid in a dwarf rabbit (Oryctolagus cuniculi). *Contemp Top Lab Anim Sci.* 39: 39–40. 2000. [Medline]
- Styer CM, Ferrier WT, Labelle P, Griffey SM, and Kendall LV. Limbic dermoid in a New Zealand White rabbit (Oryctolagus cuniculus). *Contemp Top Lab Anim Sci.* 44: 46–48. 2005. [Medline]
- Moore CP, Dubielzig R, and Glaza SM. Anterior corneal dystrophy of American Dutch belted rabbits: biomicroscopic and histopathologic findings. *Vet Pathol.* 24: 28–33. 1987. [Medline] [CrossRef]
- Boorman G, Crabbs TA, Kolenda-Roberts H, Latimer K, Miller AD, Muravnick KB, Nyska A, Ochoa R, Pardo ID, Ramot Y, Rao DB,

Schuh J, Suttie A, Travlos GS, Ward JM, Wolf JC, and Elmore SA. Proceedings of the 2011 National Toxicology Program Satellite Symposium. *Toxicol Pathol.* **40**: 321–344. 2012. [Medline] [CrossRef]

- Port CD, and Dodd DC. Two cases of corneal epithelial dystrophy in rabbits. *Lab Anim Sci.* 33: 587–588. 1983. [Medline]
- Durand-Cavagna G, Hubert MF, Gerin G, and Molon-Noblot S. Spontaneous pre-Descemet's membrane corneal opacities in rabbits. *Lab Anim Sci.* 48: 310–313. 1998. [Medline]
- Garibaldi BA, and Goad ME. Lipid keratopathy in the Watanabe (WHHL) rabbit. *Vet Pathol.* 25: 173–174. 1988. [Medline] [CrossRef]
- Janes RG. Changes in the Rabbit's Eye Caused by Cholesterol Feeding. *Am J Ophthalmol.* 58: 819–828. 1964. [Medline] [CrossRef]
- Roth SI, Stock EL, Siel JM, Mendelsohn A, Reddy C, Preskill DG, and Ghosh S. Pathogenesis of experimental lipid keratopathy. An ultrastructural study of an animal model system. *Invest Ophthalmol Vis Sci.* 29: 1544–1551. 1988. [Medline]
- 273. Sebesteny A, Sheraidah GA, Trevan DJ, Alexander RA, and Ahmed AI. Lipid keratopathy and atheromatosis in an SPF laboratory rabbit colony attributable to diet. *Lab Anim.* 19: 180–188. 1985. [Medline] [CrossRef]
- 274. Fernández-Navarro J, Aldea P, de Hoz R, Salazar JJ, Ramírez AI, Rojas B, Gallego BI, Triviño A, Tejerina T, and Ramírez JM. Neuroprotective effects of low-dose statins in the retinal ultrastructure of hypercholes-terolemic rabbits. *PLoS One.* 11: e0154800. 2016. [Medline] [Cross-Ref]
- 275. Rojas B, Ramírez AI, Salazar JJ, de Hoz R, Redondo A, Raposo R, Mendez T, Tejerina T, Triviño A, and Ramírez JM. Low-dosage statins reduce choroidal damage in hypercholesterolemic rabbits. *Acta Oph-thalmol.* 89: 660–669. 2011. [Medline] [CrossRef]
- 276. Salazar JJ, Ramírez AI, de Hoz R, Rojas B, Ruiz E, Tejerina T, Triviño A, and Ramírez JM. Alterations in the choroid in hypercholesterolemic rabbits: reversibility after normalization of cholesterol levels. *Exp Eye Res.* 84: 412–422. 2007. [Medline] [CrossRef]
- 277. Triviño A, Ramírez AI, Salazar JJ, de Hoz R, Rojas B, Padilla E, Tejerina T, and Ramírez JM. A cholesterol-enriched diet induces ultrastructural changes in retinal and macroglial rabbit cells. *Exp Eye Res.* 83: 357–366. 2006. [Medline] [CrossRef]
- 278. Yamagiwa Y, Takei Y, Koizumi H, Nemoto S, Kurata M, and Satoh H. Pathological features of corneal phospholipidosis in juvenile white rabbits induced by ocular instillation of chloroquine or amiodarone. *Toxicol Pathol.* 47: 26–34. 2019. [Medline] [CrossRef]
- Fine BS, Berkow JW, and Fine S. Corneal calcification. *Science*. 162: 129–130. 1968. [Medline] [CrossRef]
- McCulley JP. Ocular hydrofluoric acid burns: animal model, mechanism of injury and therapy. *Trans Am Ophthalmol Soc.* 88: 649–684. 1990. [Medline]
- Obenberger J, Ocumpaugh DE, and Cubberly MG. Experimental corneal calcification in animals treated with dihydrotachysterol. *Invest Ophthalmol.* 8: 467–474. 1969. [Medline]
- Doughman DJ, Olson GA, Nolan S, and Hajny RG. Experimental band keratopathy. *Arch Ophthalmol.* 81: 264–271. 1969. [Medline] [Cross-Ref]
- Economon JW, Silverstein AM, and Zimmerman LE. Band keratopathy in a rabbit colony. *Invest Ophthalmol.* 2: 361–368. 1963. [Medline]
- Short BG. Safety evaluation of ocular drug delivery formulations: techniques and practical considerations. *Toxicol Pathol.* 36: 49–62. 2008. [Medline] [CrossRef]
- Werner L, Chew J, and Mamalis N. Experimental evaluation of ophthalmic devices and solutions using rabbit models. *Vet Ophthalmol.* 9: 281–291. 2006. [Medline] [CrossRef]
- Bergmanson JP. The anatomy of the rabbit aqueous outflow pathway. Acta Ophthalmol (Copenh). 63: 493–501. 1985. [Medline] [CrossRef]
- 287. Grierson I, Nagasubramanian S, Edwards J, Millar LC, and Ennis K. The effects of various levels of intraocular pressure on the rabbit's outflow system. *Exp Eye Res.* 42: 383–397. 1986. [Medline] [CrossRef]
- Nishida S, Uchida H, Takeuchi M, Sui GQ, Mizutani S, and Iwaki M. Scanning electron microscope study of the rabbit anterior chamber

angle. Med Mol Morphol. 38: 54-62. 2005. [Medline] [CrossRef]

- 289. Samuelson DA, and Gelatt KN. Aqueous outflow in the beagle. II. Postnatal morphologic development of the iridocorneal angle: corneoscleral trabecular meshwork and angular aqueous plexus. *Curr Eye Res.* 3: 795–807. 1984. [Medline] [CrossRef]
- Sheppard LB. Intrascleral drainage channels of the normal rabbit eye. *Trans Am Ophthalmol Soc.* 57: 99–108. 1959. [Medline]
- Sheppard LB. The anatomy and histology of the normal rabbit eye with special reference to the ciliary zone. *Arch Ophthalmol.* 67: 87–100. 1962. [Medline] [CrossRef]
- 292. Bouhenni RA, Dunmire J, Sewell A, and Edward DP. Animal models of glaucoma. *J Biomed Biotechnol.* **2012**: 692609. 2012. [Medline] [CrossRef]
- Bunt-Milam AH, Dennis MB Jr, and Bensinger RE. Optic nerve head axonal transport in rabbits with hereditary glaucoma. *Exp Eye Res.* 44: 537–551. 1987. [Medline] [CrossRef]
- 294. Fox RR, Crary DD, Babino EJ Jr, and Sheppard LB. Buphthalmia in the rabbit. Pleiotropic effects of the (bu) gene and a possible explanation of mode of gene action. J Hered. 60: 206–212. 1969. [Medline] [CrossRef]
- Hanna BL, Sawin PB, and Sheppard LB. Recessive buphthalmos in the rabbit. *Genetics*. 47: 519–529. 1962. [Medline] [CrossRef]
- 296. Knepper PA, McLone DG, Goossens W, Vanden Hoek T, and Higbee RG. Ultrastructural alterations in the aqueous outflow pathway of adult buphthalmic rabbits. *Exp Eye Res.* 52: 525–533. 1991. [Medline] [CrossRef]
- 297. Tesluk GC, Peiffer RL, and Brown D. A clinical and pathological study of inherited glaucoma in New Zealand White rabbits. *Lab Anim.* 16: 234–239. 1982. [Medline] [CrossRef]
- Ueno A, Tawara A, Kubota T, Ohnishi Y, Inomata H, and Solomon AS. Histopathological changes in iridocorneal angle of inherited glaucoma in rabbits. *Graefes Arch Clin Exp Ophthalmol.* 237: 654–660. 1999. [Medline] [CrossRef]
- 299. Edward DP, and Bouhenni R. Anterior segment alterations and comparative aqueous humor proteomics in the buphthalmic rabbit (an American Ophthalmological Society thesis). *Trans Am Ophthalmol Soc.* **109**: 66–114. 2011. [Medline]
- 300. Zernii EY, Baksheeva VE, Iomdina EN, Averina OA, Permyakov SE, Philippov PP, Zamyatnin AA, and Senin II. Rabbit models of ocular diseases: new relevance for classical approaches. CNS Neurol Disord Drug Targets. 15: 267–291. 2016. [Medline] [CrossRef]
- Morrison JC, DeFrank MP, and Van Buskirk EM. Regional microvascular anatomy of the rabbit ciliary body. *Invest Ophthalmol Vis Sci.* 28: 1314–1324. 1987. [Medline]
- Pataky PE. Structure and innervation of the ciliary processes of the albino rabbit eye. *Anat Rec.* 168: 339–349. 1970. [Medline] [CrossRef]
- 303. Tserevelakis GJ, Avtzi S, Tsilimbaris MK, and Zacharakis G. Delineating the anatomy of the ciliary body using hybrid optical and photoacoustic imaging. *J Biomed Opt.* 22: 60501. 2017. [Medline] [CrossRef]
- Wegner K. Regional differences in ultrastructure of the rabbit ciliary processes: the effect of anesthetics and fixation procedures. *Invest Ophthalmol.* 6: 177–191. 1967. [Medline]
- 305. Funk RH. The vessel architecture of the pars plana in the cynomolgus monkey, rat and rabbit eye. A scanning electron microscopic study of plastic corrosion casts. *Ophthalmic Res.* 25: 337–348. 1993. [Medline] [CrossRef]
- 306. Langevin NE, Schafer KA, Turner OC, McPherson BJ, and Rose RE. Historical data: histopathology lesions observed in the eyes of control rabbits in topical ocular administration and contact lens studies. *Toxicol Pathol.* 46: 799–820. 2018. [Medline] [CrossRef]
- Griffith JW, Sassani JW, Bowman TA, and Lang CM. Osseous choristoma of the ciliary body in guinea pigs. *Vet Pathol.* 25: 100–102. 1988. [Medline] [CrossRef]
- Lynch GL, and Scagliotti RH. Osseous metaplasia in the eye of a dog. Vet Pathol. 44: 222–224. 2007. [Medline] [CrossRef]
- Williams D, and Sullivan A. Ocular disease in the guinea pig (Cavia porcellus): a survey of 1000 animals. *Vet Ophthalmol.* 13(Suppl): 54– 62. 2010. [Medline] [CrossRef]

- 310. Fernandez-Bueno I, Di Lauro S, Alvarez I, Lopez JC, Garcia-Gutierrez MT, Fernandez I, Larra E, and Pastor JC. Safety and biocompatibility of a new high-density polyethylene-based spherical integrated porous orbital implant: an experimental study in rabbits. *J Ophthalmol.* 2015: 904096. 2015. [Medline] [CrossRef]
- Kouchi M, Ueda Y, Horie H, and Tanaka K. Ocular lesions in Watanabe heritable hyperlipidemic rabbits. *Vet Ophthalmol.* 9: 145–148. 2006. [Medline] [CrossRef]
- 312. Kuszak JR. The ultrastructure of epithelial and fiber cells in the crystalline lens. *Int Rev Cytol.* **163**: 305–350. 1995. [Medline] [CrossRef]
- 313. Jeong MB, Kim NR, Yi NY, Park SA, Kim MS, Park JH, Jeong SM, Seo KD, Nam TC, Oh YS, Won MH, and Seo KM. Spontaneous ophthalmic diseases in 586 New Zealand White rabbits. *Exp Anim.* 54: 395–403. 2005. [Medline] [CrossRef]
- Munger RJ, Jensen VB, Bouldin TW, and Peiffer RL. Bilateral neuroepithelial choristomas of the optic disc in a cynomolgus monkey (Macaca fascicularis): a case report. *Vet Ophthalmol.* 5: 221–226. 2002. [Medline] [CrossRef]
- 315. Peng X, Roshwalb S, Cooper TK, Zimmerman H, and Christensen ND. High incidence of spontaneous cataracts in aging laboratory rabbits of an inbred strain. *Vet Ophthalmol.* 18: 186–190. 2015. [Medline] [Cross-Ref]
- Sanchez RF, Becker R, Dawson C, Escanilla N, and Lam R. Calculation of posterior chamber intraocular lens (IOL) size and dioptric power for use in pet rabbits undergoing phacoemulsification. *Vet Ophthalmol.* 20: 242–249. 2017. [Medline] [CrossRef]
- 317. Sanchez RF, Everson R, Hedley J, Dawson C, Lam R, Priestnall SL, Garcia de Carellan A, de Miguel C, and Seymour C. Rabbits with naturally occurring cataracts referred for phacoemulsification and intraocular lens implantation: a preliminary study of 12 cases. *Vet Ophthalmol.* 21: 399–412. 2018. [Medline] [CrossRef]
- Al-Khudari S, Donohue ST, Al-Ghoul WM, and Al-Ghoul KJ. Agerelated compaction of lens fibers affects the structure and optical properties of rabbit lenses. *BMC Ophthalmol.* 7: 19. 2007. [Medline] [CrossRef]
- Frenzel EM, Neely KA, Walsh AW, Cameron JD, and Gregerson DS. A new model of proliferative vitreoretinopathy. *Invest Ophthalmol Vis Sci.* 39: 2157–2164. 1998. [Medline]
- 320. Hsu YW, Yeh SM, Chen YY, Chen YC, Lin SL, and Tseng JK. Protective effects of taurine against alloxan-induced diabetic cataracts and refraction changes in New Zealand White rabbits. *Exp Eye Res.* 103: 71–77. 2012. [Medline] [CrossRef]
- 321. Kralinger MT, Kieselbach GF, Voigt M, Hayden B, Hernandez E, Fernandez V, and Parel JM. Experimental model for proliferative vitreoretinopathy by intravitreal dispase: limited by zonulolysis and cataract. *Ophthalmologica*. 220: 211–216. 2006. [Medline] [CrossRef]
- 322. Yuan L, Yao H, Xu Y, Chen M, Deng J, Song Y, Sui T, Wang Y, Huang Y, Li Z, and Lai L. CRISPR/Cas9-mediated mutation of αA-crystallin gene induces congenital cataracts in rabbits. *Invest Ophthalmol Vis Sci.* 58: BIO34–BIO41. 2017. [Medline] [CrossRef]
- 323. Gwon A, Mantras C, Gruber L, and Cunanan C. Concanavalin A-induced posterior subcapsular cataract: a new model of cataractogenesis. *Invest Ophthalmol Vis Sci.* 34: 3483–3488. 1993. [Medline]
- Hayasaka Y, Hayasaka S, and Nagaki Y. Ocular changes after intravitreal injection of methanol, formaldehyde, or formate in rabbits. *Pharmacol Toxicol.* 89: 74–78. 2001. [Medline] [CrossRef]
- Lubek BM, Avaria M, Basu PK, and Wells PG. Pharmacological studies on the in vivo cataractogenicity of acetaminophen in mice and rabbits. *Fundam Appl Toxicol.* 10: 596–606. 1988. [Medline] [CrossRef]
- 326. Morsy EA, Salem HM, Khattab MS, Hamza DA, and Abuowarda MM. Encephalitozoon cuniculi infection in farmed rabbits in Egypt. Acta Vet Scand. 62: 11. 2020. [Medline] [CrossRef]
- Özkan Ö, and Alcigir ME. Subacute stage of *encephalitozoon cuniculi* infection in eye lesions of rabbit in Turkey. *Iran J Parasitol.* 13: 301– 309. 2018. [Medline]
- Ashton N, Cook C, and Clegg F. Encephalitozoonosis (nosematosis) causing bilateral cataract in a rabbit. Br J Ophthalmol. 60: 618–631.
1976. [Medline] [CrossRef]

- Felchle LM, and Sigler RL. Phacoemulsification for the management of Encephalitozoon cuniculi-induced phacoclastic uveitis in a rabbit. *Vet Ophthalmol.* 5: 211–215. 2002. [Medline] [CrossRef]
- Giordano C, Weigt A, Vercelli A, Rondena M, Grilli G, and Giudice C. Immunohistochemical identification of Encephalitozoon cuniculi in phacoclastic uveitis in four rabbits. *Vet Ophthalmol.* 8: 271–275. 2005. [Medline] [CrossRef]
- 331. Latney LV, Bradley CW, and Wyre NR. *Encephalitozoon cuniculi* in pet rabbits: diagnosis and optimal management. *Vet Med (Auckl)*. **5**: 169–180. 2014. [Medline]
- Matsumoto B, Blanks JC, and Ryan SJ. Topographic variations in the rabbit and primate internal limiting membrane. *Invest Ophthalmol Vis Sci.* 25: 71–82. 1984. [Medline]
- 333. Los LI, van Luyn MJ, and Nieuwenhuis P. Organization of the rabbit vitreous body: lamellae, Cloquet's channel and a novel structure, the 'alae canalis Cloqueti'. *Exp Eye Res.* 69: 343–350. 1999. [Medline] [CrossRef]
- Ponsioen TL, Hooymans JM, and Los LI. Remodelling of the human vitreous and vitreoretinal interface--a dynamic process. *Prog Retin Eye Res.* 29: 580–595. 2010. [Medline] [CrossRef]
- 335. Lukáts A, Szabó A, Röhlich P, Vígh B, and Szél A. Photopigment coexpression in mammals: comparative and developmental aspects. *Histol Histopathol.* 20: 551–574. 2005. [Medline]
- Schnitzer J. The development of astrocytes and blood vessels in the postnatal rabbit retina. J Neurocytol. 17: 433–449. 1988. [Medline] [CrossRef]
- Jack RL. Regression of the hyaloid vascular system. An ultrastructural analysis. *Am J Ophthalmol.* 74: 261–272. 1972. [Medline] [CrossRef]
- Jack RL. Ultrastructure of the hyaloid vascular system. Arch Ophthalmol. 87: 555–567. 1972. [Medline] [CrossRef]
- Boillot T, Graille M, Williams D, and Rosolen SG. Unilateral persistent hyperplastic tunica vasculosa lentis and persistent hyperplastic primary vitreous in a rabbit. *Vet Ophthalmol.* 18: 510–514. 2015. [Medline] [CrossRef]
- Los LI, van Luyn MJ, Eggli PS, Dijk F, and Nieuwenhuis P. Vascular remnants in the rabbit vitreous body. II. Enzyme digestion and immunohistochemical studies. *Exp Eye Res.* 71: 153–165. 2000. [Medline] [CrossRef]
- Los LI, van Luyn MJ, and Nieuwenhuis P. Vascular remnants in the rabbit vitreous body. I. Morphological characteristics and relationship to vitreous embryonic development. *Exp Eye Res.* 71: 143–151. 2000. [Medline] [CrossRef]
- De Schaepdrijver L, Simoens P, Lauwers H, and De Geest JP. Retinal vascular patterns in domestic animals. *Res Vet Sci.* 47: 34–42. 1989. [Medline] [CrossRef]
- Morcos Y, and Chan-Ling T. Identification of oligodendrocyte precursors in the myelinated streak of the adult rabbit retina in vivo. *Glia.* 21: 163–182. 1997. [Medline] [CrossRef]
- Morcos Y, and Chan-Ling T. Concentration of astrocytic filaments at the retinal optic nerve junction is coincident with the absence of intraretinal myelination: comparative and developmental evidence. *J Neurocytol.* 29: 665–678. 2000. [Medline] [CrossRef]
- Schnitzer J. Distribution and immunoreactivity of glia in the retina of the rabbit. J Comp Neurol. 240: 128–142. 1985. [Medline] [CrossRef]
- Tripathi B, and Ashton N. Vaso-glial connections in the rabbit retina. Br J Ophthalmol. 55: 1–11. 1971. [Medline] [CrossRef]
- Uga S, and Smelser. Comparative study of the fine structure of retinal Müller cells in various vertebrates. *Invest Ophthalmol.* 12: 434–448. 1973. [Medline]
- Choudhury BP. Ganglion cell distribution in the albino rabbit's retina. Exp Neurol. 72: 638–644. 1981. [Medline] [CrossRef]
- Donatien P, and Jeffery G. Correlation between rod photoreceptor numbers and levels of ocular pigmentation. *Invest Ophthalmol Vis Sci.* 43: 1198–1203. 2002. [Medline]
- Oyster CW, Takahashi ES, and Hurst DC. Density, soma size, and regional distribution of rabbit retinal ganglion cells. J Neurosci. 1: 1331–

1346. 1981. [Medline] [CrossRef]

- Provis JM. The distribution and size of ganglion cells in the regina of the pigmented rabbit: a quantitative analysis. *J Comp Neurol.* 185: 121–137. 1979. [Medline] [CrossRef]
- Vaney DI. A quantitative comparison between the ganglion cell populations and axonal outflows of the visual streak and periphery of the rabbit retina. *J Comp Neurol.* 189: 215–233. 1980. [Medline] [CrossRef]
- 353. Steele-Russell I, Russell MI, Castiglioni JA, and Graham J. Differential retinal origins of separate anatomical channels for pattern and motion vision in rabbit. *Exp Brain Res.* 222: 99–111. 2012. [Medline] [Cross-Ref]
- Rapaport DH, and Stone J. The area centralis of the retina in the cat and other mammals: focal point for function and development of the visual system. *Neuroscience*. 11: 289–301. 1984. [Medline] [CrossRef]
- 355. Famiglietti EV, and Sharpe SJ. Regional topography of rod and immunocytochemically characterized "blue" and "green" cone photoreceptors in rabbit retina. *Vis Neurosci.* **12**: 1151–1175. 1995. [Medline] [CrossRef]
- 356. Juliusson B, Bergström A, Röhlich P, Ehinger B, van Veen T, and Szél A. Complementary cone fields of the rabbit retina. *Invest Ophthalmol Vis Sci.* 35: 811–818. 1994. [Medline]
- Röhlich P, van Veen T, and Szél A. Two different visual pigments in one retinal cone cell. *Neuron.* 13: 1159–1166. 1994. [Medline] [CrossRef]
- Lucas RJ. Mammalian inner retinal photoreception. Curr Biol. 23: R125–R133. 2013. [Medline] [CrossRef]
- Lira-Carrera MM, Gutiérrez-Amavizca BE, Álvarez-Araujo LJ, Aguirre-Ramírez M, and Pérez-León JA. Analysis of melanopsin gene expression in the rabbit retina at different ages. *Genet Mol Res.* 16. 2017; [CrossRef]. [Medline]
- Prince JH, and McConnell DG. Retina and optic nerve. In: The Rabbit in Eye Research, 1st ed. JH Prince (ed). Charles C Thomas Publisher, Illinois. 385–448. 1964.
- Robinson SR, and Dreher Z. Müller cells in adult rabbit retinae: morphology, distribution and implications for function and development. J Comp Neurol. 292: 178–192. 1990. [Medline] [CrossRef]
- Booij JC, Baas DC, Beisekeeva J, Gorgels TG, and Bergen AA. The dynamic nature of Bruch's membrane. *Prog Retin Eye Res.* 29: 1–18. 2010. [Medline] [CrossRef]
- 363. Ferrara D, Waheed NK, and Duker JS. Investigating the choriocapillaris and choroidal vasculature with new optical coherence tomography technologies. *Prog Retin Eye Res.* 52: 130–155. 2016. [Medline] [CrossRef]
- Nakaizumi Y. The Ultrastructure of Bruch's Membrane. I. Human, Monkey, Rabbit, Guinea Pig, and Rat Eyes. Arch Ophthalmol. 72: 380– 387. 1964. [Medline] [CrossRef]
- 365. Daniels AB, Froehler MT, Pierce JM, Nunnally AH, Calcutt MW, Bridges TM, LaNeve DC, Williams PE, Boyd KL, Reyzer ML, Lindsley CW, Friedman DL, and Richmond A. Pharmacokinetics, tissue localization, toxicity, and treatment efficacy in the first small animal (Rabbit) model of intra-arterial chemotherapy for retinoblastoma. *Invest Ophthalmol Vis Sci.* 59: 446–454. 2018. [Medline] [CrossRef]
- 366. Albrecht May C. Comparative anatomy of the optic nerve head and inner retina in non-primate animal models used for glaucoma research. *Open Ophthalmol J.* 2: 94–101. 2008. [Medline] [CrossRef]
- Ninomiya H, Inomata T, and Kanemaki N. Microvascular architecture of the rabbit eye: a scanning electron microscopic study of vascular corrosion casts. J Vet Med Sci. 70: 887–892. 2008. [Medline] [CrossRef]
- Sugiyama K, Bacon DR, Morrison JC, and Van Buskirk EM. Optic nerve head microvasculature of the rabbit eye. *Invest Ophthalmol Vis Sci.* 33: 2251–2261. 1992. [Medline]
- 369. Balaratnasingam C, Kang MH, Yu P, Chan G, Morgan WH, Cringle SJ, and Yu DY. Comparative quantitative study of astrocytes and capillary distribution in optic nerve laminar regions. *Exp Eye Res.* 121: 11–22. 2014. [Medline] [CrossRef]
- 370. Lockwood H, Reynaud J, Gardiner S, Grimm J, Libertiaux V, Downs JC, Yang H, and Burgoyne CF. Lamina cribrosa microarchitecture in normal monkey eyes part 1: methods and initial results. *Invest Oph-*

thalmol Vis Sci. 56: 1618–1637. 2015. [Medline] [CrossRef]

- Quigley HA. The contribution of the sclera and lamina cribrosa to the pathogenesis of glaucoma: Diagnostic and treatment implications. *Prog Brain Res.* 220: 59–86. 2015. [Medline] [CrossRef]
- 372. Ashton N, Tripathi B, and Knight G. Effect of oxygen on the developing retinal vessels of the rabbit. I. Anatomy and development of the retinal vessels of the rabbit. *Exp Eye Res.* 14: 214–220. 1972. [Medline] [CrossRef]
- 373. Schafer KA, and Render JA. Toxicologic Pathology of the Eye: Alterations of the Lens and Posterior Segment. In: Assessing Ocular Toxicology in Laboratory Animals. AB Wier, and M Collins (eds). Humana Press, New York. 219–257. 2013.
- 374. Kawasako K, Oshikata T, Kanno T, and Hamamura M. Neurofilament accumulation in rabbit retinas. *J Comp Pathol.* 153: 283–286. 2015. [Medline] [CrossRef]
- Ts'o MO, and Friedman E. The retinal pigment epithelium. I. Comparative histology. *Arch Ophthalmol.* 78: 641–649. 1967. [Medline] [Cross-Ref]
- Olsen TW, Aaberg SY, Geroski DH, and Edelhauser HF. Human sclera: thickness and surface area. *Am J Ophthalmol.* 125: 237–241. 1998. [Medline] [CrossRef]
- 377. Vurgese S, Panda-Jonas S, and Jonas JB. Scleral thickness in human eyes. *PLoS One*. 7: e29692. 2012. [Medline] [CrossRef]
- Ruskell GL. Blood Vessels of the Orbit and Globe. In: The Rabbit in Eye Research, 1st ed. JH Prince (ed). Charles C Thomas Publisher, Illinois. 514–553. 1964.
- 379. Sakai T. Comparative Anatomy of Mammalian Harderian Glands. In: Harderian Glands: Porphyrin Metabolism, Behavioral and Endocrine Effects. SM Webb, RD Hoffman, ML Puig-Domingo, and RJ Reiter (eds). Springer-Verlag, Berlin. 7–22. 1992.
- Gargiulo AM, Ceccarelli P, and Pedini V. The presence of granular excretory ducts in the rabbit zygomatic gland. *Anat Histol Embryol.* 25: 175–176. 1996. [Medline] [CrossRef]
- Koutavas H, Anderton PJ, and Millar TJ. Separation and characterisation of cyclic nucleotide phosphodiesterases from the lacrimal, harderian and zygomatic glands of the rabbit. *Curr Eye Res.* 15: 1191–1197. 1996. [Medline] [CrossRef]
- Eglitis I. Glands; Lacrimal, Harder, Nictitans. In: The Rabbit in Eye Research. JH Prince (ed). Charles C Thomas Publisher, Illinois. 38–56. 1964.
- Hittmair KM, Tichy A, and Nell B. Ultrasonography of the Harderian gland in the rabbit, guinea pig, and chinchilla. *Vet Ophthalmol.* 17: 175–183. 2014. [Medline] [CrossRef]
- Eltony SAM. A comparative study of the harderian gland in the female rat and female rabbit (a histological, histochemical, scanning electron microscopic and morphometric study). *Egypt J Histol.* 32: 46–65. 2009.
- 385. Shirama K, Ozawa S, Seyama Y, Kobayashi M, Sawamura S, and Yamada J. Postnatal development of the harderian gland in the rabbit: light and electron microscopic observations. *Microsc Res Tech.* 37: 572–582. 1997. [Medline] [CrossRef]
- Payne AP. The harderian gland: a tercentennial review. J Anat. 185: 1–49. 1994. [Medline]
- Rock CO, Fitzgerald V, Rainey WT Jr, and Snyder F. Mass spectral identification of 2-(O-acyl)hydroxy fatty acid esters in the white portion of the rabbit Harderian gland. *Chem Phys Lipids*. 17: 207–212. 1976. [Medline] [CrossRef]
- 388. Seyama T, Kasama T, Yasugi E, Park SH, and Kano K. Lipids in Harderian Glands and Their Significance. In: Harderian Glands: Porphyrin Metabolism, Behavioral and Endocrine Effects. SM Webb, RD Hoffman, ML Puig-Domingo, and RJ Reiter, (eds). Springer-Verlag, Berlin. 196–216. 1992.
- 389. Winterhager E, and Kühnel W. Membrane specializations of the cells of the Harderian gland of the rabbit with particular reference to the mechanism of exocytosis. *Cell Tissue Res.* 231: 623–636. 1983. [Medline] [CrossRef]
- 390. Wooding FB. Lipid droplet secretion by the rabbit harderian gland. J

Ultrastruct Res. 71: 68-78. 1980. [Medline] [CrossRef]

- Bayraktaroğlu AG, and Ergün E. Histomorphology of the Harderian gland in the Angora rabbit. *Anat Histol Embryol.* **39**: 494–502. 2010. [Medline] [CrossRef]
- Bjoerkman N, Nicander L, and Schantz B. On the histology and ultrastructure of the Harderian gland in rabbits. *Z Zellforsch Mikrosk Anat.* 52: 93–104. 1960. [Medline] [CrossRef]
- 393. Nadakavukaren MJ. The Mammalian Harderian Gland: Sexual Dimorphism, and its Regulation by Light and Steroids. In: Harderian Glands: Porphyrin Metabolism, Behavioral and Endocrine Effects. SM Webb, RD Hoffman, ML Puig-Domingo, and RJ Reiter (eds). Springer-Verlag, Berlin. 70–90. 1992.
- 394. Ding C, Parsa L, Nandoskar P, Zhao P, Wu K, and Wang Y. Duct system of the rabbit lacrimal gland: structural characteristics and role in lacrimal secretion. *Invest Ophthalmol Vis Sci.* 51: 2960–2967. 2010. [Medline] [CrossRef]
- 395. Schechter JE, Warren DW, and Mircheff AK. A lacrimal gland is a lacrimal gland, but rodent's and rabbit's are not human. *Ocul Surf.* 8: 111–134. 2010. [Medline] [CrossRef]
- Cornell-Bell AH, Sullivan DA, and Allansmith MR. Gender-related differences in the morphology of the lacrimal gland. *Invest Ophthalmol Vis Sci.* 26: 1170–1175. 1985. [Medline]
- 397. Stibbe EP. A comparative study of the nictitating membrane of birds and mammals. *J Anat.* **62**: 159–176. 1928. [Medline]
- 398. Schafer KA, and Render JA. Toxicologic Pathology of the Eye: Histologic Preparation and Alterations of the Anterior Segment. In: Assessing Ocular Toxicology in Laboratory Animals. AB Wier, and M Collins (eds). Humana Press, New York. 159–217. 2013:
- 399. Marini RP, Foltz CJ, Kersten D, Batchelder M, Kaser W, and Li X. Microbiologic, radiographic, and anatomic study of the nasolacrimal duct apparatus in the rabbit (Oryctolagus cuniculus). *Lab Anim Sci.* 46: 656–662. 1996. [Medline]
- 400. Frame NJ, and Burkat CN. Identifying an appropriate animal model for the nasolacrimal drainage system. *Ophthal Plast Reconstr Surg.* 25: 354–358. 2009. [Medline] [CrossRef]
- 401. Maeda S, Ishikawa M, Abe T, Konno S, and Sakuragi S. Lectin cytochemistry of the lacrimal sac epithelium in experimental dacryocystitis. *Jpn J Ophthalmol.* 43: 69–74. 1999. [Medline] [CrossRef]
- 402. Paulsen FP, Föge M, Thale AB, Tillmann BN, and Mentlein R. Animal model for the absorption of lipophilic substances from tear fluid by the epithelium of the nasolacrimal ducts. *Invest Ophthalmol Vis Sci.* 43: 3137–3143. 2002. [Medline]
- 403. Maurer KJ, Marini RP, Fox JG, and Rogers AB. Polycystic kidney syndrome in New Zealand White rabbits resembling human polycystic kidney disease. *Kidney Int.* 65: 482–489. 2004. [Medline] [CrossRef]
- Ogilvie RF, Sabour MS, and Horne NW. Light and electron microscopy of prednisolone-induced nephropathy in rabbits. *Diabetes.* 14: 595–605. 1965. [Medline] [CrossRef]
- 405. Rammer L, Nathorst-Windahl G, and Boberg J. Cortisone nephropathy in the rabbit. The effect of heparin on the morphology, plasma lipid levels and plasma lipoprotein lipase activity. *Acta Pathol Microbiol Scand.* **71**: 161–172. 1967. [Medline] [CrossRef]
- Rammer L. Cortisone nephropathy in the rabbit. The effect of warfarin administration. *Acta Pathol Microbiol Scand.* 74: 514–518. 1968. [Medline] [CrossRef]
- 407. Ritskes-Hoitinga J, Grooten HN, Wienk KJ, Peters M, Lemmens AG, and Beynen AC. Lowering dietary phosphorus concentrations reduces kidney calcification, but does not adversely affect growth, mineral metabolism, and bone development in growing rabbits. *Br J Nutr.* 91: 367–376. 2004. [Medline] [CrossRef]
- Burek JD, Duprat P, Owen R, Peter CP, and Van Zwieten MJ. Spontaneous renal disease in laboratory animals. *Int Rev Exp Pathol.* 30: 231–319. 1988. [Medline] [CrossRef]
- 409. Hinton M. Kidney disease in the rabbit: a histological survey. Lab Anim. 15: 263–265. 1981. [Medline] [CrossRef]