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## A Simple Paper-Based Colorimetric Device for Rapid Mercury(II) Assay

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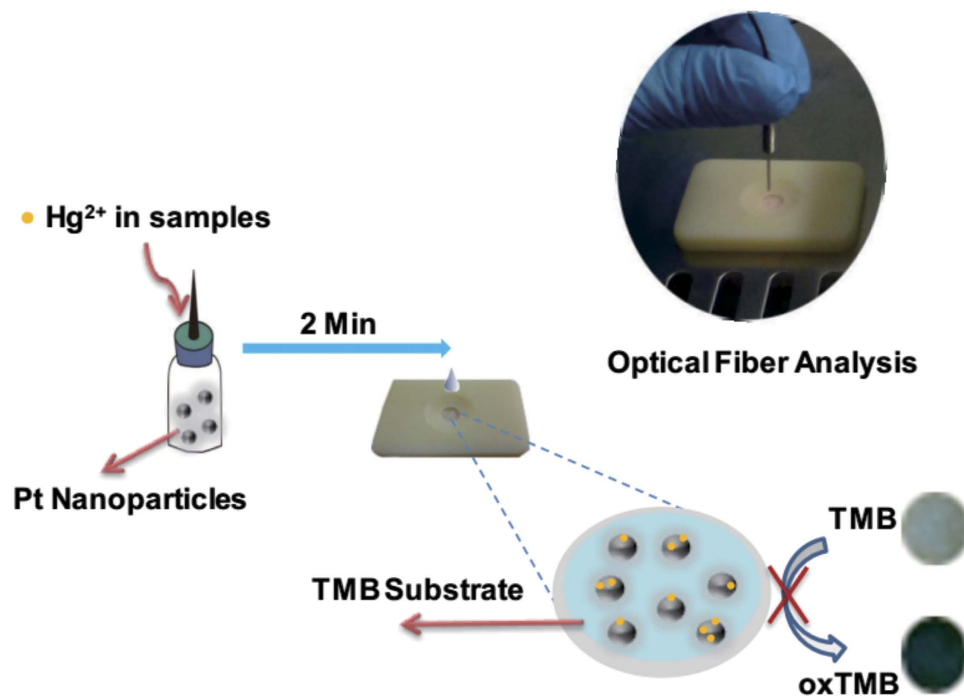
Contamination of the environment by mercury(II) ions ( $\text{Hg}^{2+}$ ) poses a serious threat to human health and ecosystems. Up to now, many reported  $\text{Hg}^{2+}$  sensors require complex procedures, long measurement times and sophisticated instrumentation. We have developed a simple, rapid, low cost and naked-eye quantitative method for  $\text{Hg}^{2+}$  environmental analysis using a paper-based colorimetric device (PCD). The sample solution to which platinum nanoparticles (PtNPs) have been added is dispensed to the detection zone on the PCD, where the 3,3',5,5'-tetramethylbenzidine (TMB) substrate has been pre-loaded. The PtNPs effect a rapid oxidation of TMB, inducing blue colorization on the PCD. However,  $\text{Hg}^{2+}$  in the solution rapidly interact with the PtNPs, suppressing the oxidation capacity and hence causing a decrease in blue intensity, which can be observed directly by the naked eye. Moreover,  $\text{Hg}^{2+}$  at concentrations as low as 0.01  $\mu\text{M}$ , can be successfully monitored using a fiber optic device, which gives a digital readout proportional to the intensity of the blue color change. This paper-based colorimetric device (PCD) shows great potential for field measurement of  $\text{Hg}^{2+}$ .

As a novel bioanalytical tool in recent years, paper-based analytical devices are widely used in environmental studies<sup>1,2</sup>, food safety analysis<sup>3-5</sup> and molecular diagnostics<sup>6-8</sup>. Paper-based devices have many advantages compared with conventional sensors, including ease-of-use, cost-effectiveness, simple fabrication and biocompatibility<sup>9-12</sup>. As paper-based analytical devices represent a relatively new analytical platform, many effective strategies have been established for multiplex analyte measurement, such as electrochemical<sup>13,14</sup> and chemiluminescent methods<sup>15</sup>, colorimetric methods<sup>16,17</sup>, FRET-based fluorescence methods<sup>18,19</sup>, and surface-enhanced Raman spectroscopy<sup>20,21</sup>. Among these strategies, colorimetric methods are well established because they are simple, quick to perform, and inexpensive. Various paper-based colorimetric device (PCDs) have been successfully developed in recent years. For example, Phillips *et al.*<sup>22</sup> described a paper-based microfluidic devices platform with naked eye detection, which required no complex or expensive instrumentation. Whiteside *et al.*<sup>23</sup> reported a paper-based ELISA platform with colorimetric detection that used a 96-microzone paper plate for the assay. In addition, there are many works focused on the use of smartphones as a new technology for the paper-based colorimetry<sup>24-26</sup>. By using smartphones, the colorimetric data can be rapidly converted to a digital image or a discrete number and provide an excellent solution for point-of-care (POC) test and real-time analysis.

One significant application in nanomaterials concerns the potential of enzyme mimetics<sup>27</sup>. Natural enzymes have many intrinsic drawbacks such as weak stability, high cost, and that they are easily influenced by environmental conditions<sup>28</sup>. So, naturally, some nanoparticles (NPs) with peroxidase-like or oxidase-like activities, such as graphene oxide (GO)<sup>29</sup>, single wall carbon nanotubes (SWNTs)<sup>30</sup>, carbon nanodots<sup>31</sup>,  $\text{MoS}_2$  nanosheets<sup>32</sup>,  $\text{Fe}_3\text{O}_4$  magnetic NPs<sup>33</sup>, cerium oxide nanoparticles<sup>34</sup>, platinum nanoparticles (PtNPs)<sup>35</sup> and gold nanoparticles (AuNPs)<sup>36</sup>, have been widely used as artificial enzymes for sensitive bioassay for their superior bioanalysis capacities over the past decade. Therefore, we have chosen PtNPs as a powerful nanozyme to catalyze the oxidation of TMB to generate signal amplification in colorimetric detection because of its excellent catalytic activity, good biocompatibility, easy synthesis and stability<sup>37</sup>.

As dangerous environmental pollutants, mercury(II) ions ( $\text{Hg}^{2+}$ ) are an ongoing environmental concern. In recent years, compact and portable instrumentation has become available for real-time and on-site trace  $\text{Hg}^{2+}$  screening of environmental and biological samples<sup>38-43</sup>. A novel lab-on-a-phone device with light-emitting diodes and dual wavelength illumination at 523 and 625 nm featured novel reaction chemistry based on  $\text{Hg}^{2+}$ -induced aptamer probe conformation changing and citrate-stabilized gold nanoparticles (AuNPs) aggregation<sup>44</sup>. Paresh *et al.*<sup>45</sup> reported a miniaturized and battery-operated device used to assay  $\text{Hg}^{2+}$  in water and in fish. The measurement principle was based on Hg(II) competing for binding with rhodamine B, whose fluorescence was quenched

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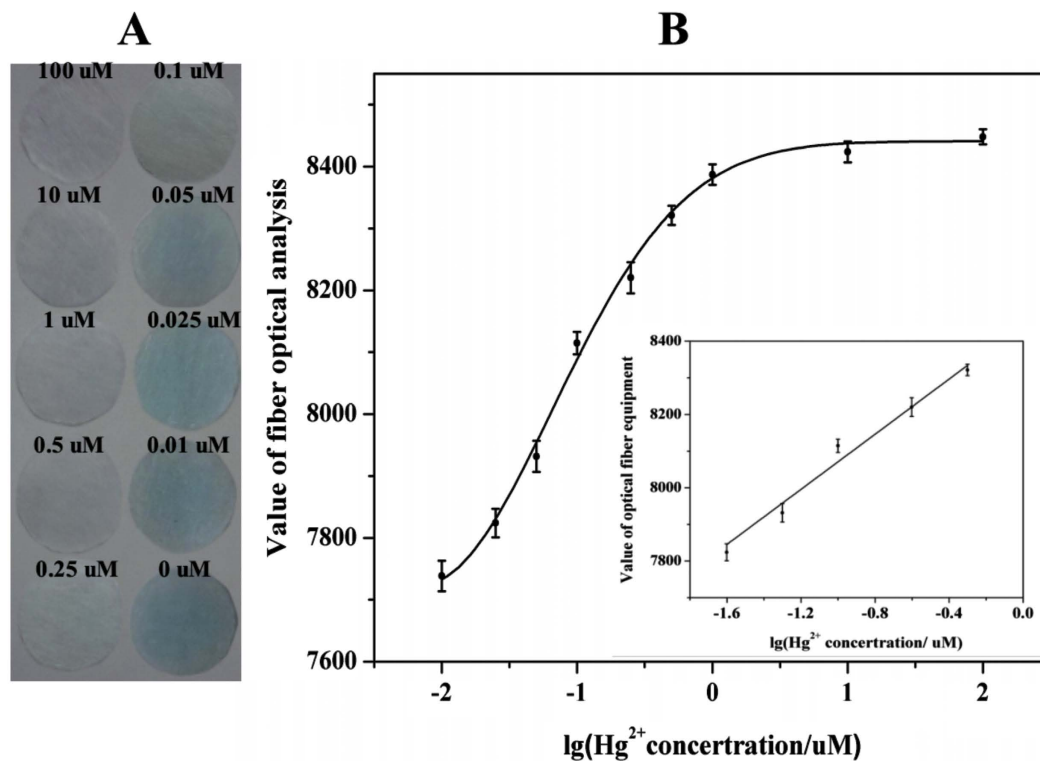
**Figure 1. Schematic Representation of PCD for Rapid Hg<sup>2+</sup> Assay.**

by AuNPs-based nanomaterial surface energy transfer (NSET). Meanwhile, as distinct from most schemes for Hg<sup>2+</sup> analysis, application of colorimetric approach for a single particle-based sensing of Hg<sup>2+</sup> have been developed recently which exhibited more attractive sensitivity<sup>46</sup>.

Recently, many works have discovered that mercury ions could interact with other metal nanoparticles to form bimetallic NPs<sup>47,48</sup>. In addition, it was also confirmed that the citrate around the surface of metal NPs was essential to promote the process of these combinations. In this work, we have introduced a novel design for a PCD that incorporated a Hg<sup>2+</sup>-induced color change and offered simple and real-time colorimetric detection of Hg<sup>2+</sup>. Our basic strategy of this PCD depended on the specific interaction between PtNPs and mercury ions and the Hg-Pt alloy can efficiently weaken its peroxidase-like catalytic activity. First, as showed in Fig. 1, the peroxidase-like activity of PtNPs is rapidly (2 minutes) inhibited through mixing and incubation with Hg<sup>2+</sup> solutions. As a result of the oxidation of TMB, which yields a blue color, the presence of Hg<sup>2+</sup> dramatically decreases the color intensity, which can be observed directly by the naked eye. Moreover, besides visual observation of the quantitative color change on the paper chip in response to the presence of Hg<sup>2+</sup>, a fiber optic laser device was also used to measure small changes in Hg<sup>2+</sup> concentration. The fiber optic emits at a red wavelength, which is well absorbed by the blue-colored complex and the decrease in the intensity of reflected light is recorded by the electrical readout system. In this way, the high intensity of the blue color, which develops on the paper-chip, corresponds to a low digital readout representing the assay of Hg<sup>2+</sup>. This novel measurement approach provides a rapid and robust sensor technology and has considerable potential for trace Hg<sup>2+</sup> analysis in diverse analytical scenarios.

## Results and Discussion

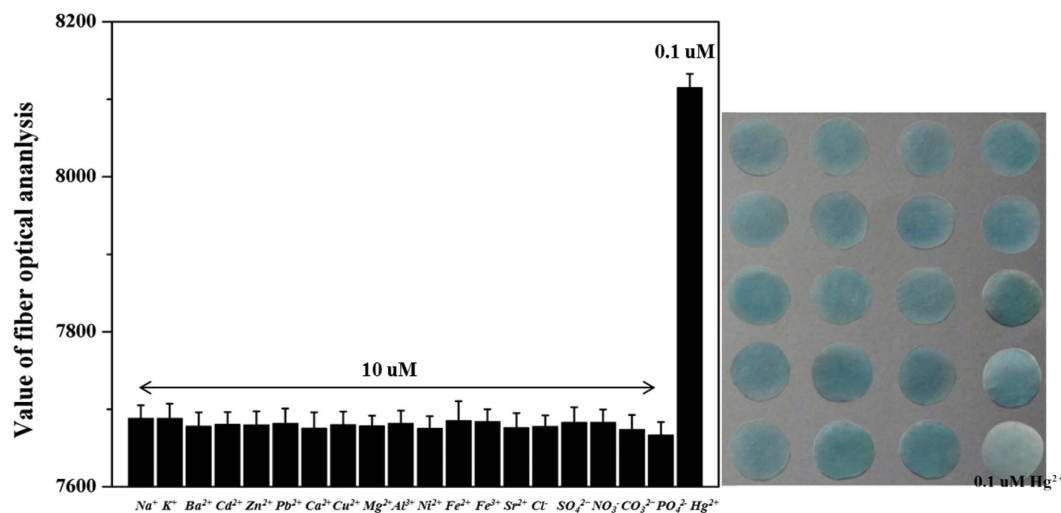
**Device design and colorimetric detection of Hg<sup>2+</sup>.** In this work, we designed a novel PCD which contained three components as shown in schematic 1: (1) the circular patterned paper can be treated as a miniature reactor for PtNPs catalyzing the oxidation of TMB<sup>49</sup>; (2) a plastic framework with central hole was used for positioning the paper chip; (3) a fiber optic analysis system was able to read-out the fading of the blue colored signal response as a result of the alteration in peroxidase catalytic activity because of reaction of the PtNPs with Hg<sup>2+</sup>. In the context of rapid and on-site analysis of real samples, the TMB/H<sub>2</sub>O<sub>2</sub> substrate solution was first dispensed on the paper-chip deposition zone. Then a small, widely-available plastic dropper bottle containing the sample solution with Hg<sup>2+</sup> and PtNPs was used to initiate the reaction. A few minutes later, one drop of the mixture was added to the detection zone where the PtNPs oxidized the TMB to induce the formation of a blue-colored sediment on the paper-chip. However, as the Hg<sup>2+</sup> presenting at water sample and the peroxidase-like activity of PtNPs would be rapidly inhibited after mixing and incubating with the solution, thus the blue oxidation of TMB decrease remarkably that could be viewed directly by the naked eye. The intensity of the blue color change for the PCD could readily be registered by digital camera or smart phones and the fiber optic system was used for signal quantitation by calculating the amount of light reflected from the detection zone by the paper surface. Because of key features such as simplicity of operation, convenience and rapid signal response, we believed this approach offers great potential for real-time, determination of Hg<sup>2+</sup> ions in diverse applications<sup>50</sup>.



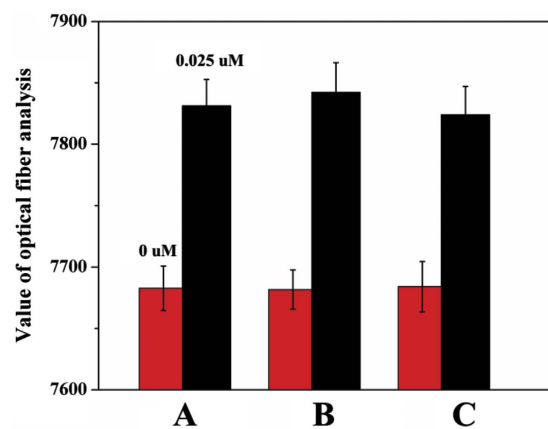
**Figure 2. Colorimetric results for Hg<sup>2+</sup> determination.** (A) Images for the PCD displayed the peroxidase-like inhibition of different Hg<sup>2+</sup> amounts. (B) The calibration curve indicated the correlation between value of optical fiber reader and the logarithm of Hg<sup>2+</sup> concentrations.

**Paper-based colorimetric assay.** For accurate and sensitive analysis, solutions with known concentrations of Hg<sup>2+</sup> ions were added to the storage-bottle where PtNPs had already been diluted to adequate volume. Following gentle shaking for 2 min, one drop was immediately dispensed onto the detection zone of the PCD which already had 10  $\mu$ L of TMB/H<sub>2</sub>O<sub>2</sub> solutions loaded. The detection zone was visually evaluated within 5 min to ensure oxidation of TMB. This simple operating procedure and short measurement time fulfilled key requirements for rapid, portable and field measurement of Hg<sup>2+</sup>. Samples with higher Hg<sup>2+</sup> concentrations gave higher intensity readings in the fiber optic analysis equipment. In Fig. 2B, the values increased as the target concentrations increased then values gradually reached steady-state at analyses concentration of ca. 0.5  $\mu$ M, which corresponded, by visual inspection, to images of patterned papers of low color intensity. Comparing the correlation between color change and the amount of Hg<sup>2+</sup>, the color of the paper-chip changed from dark blue to light blue when the concentration of Hg<sup>2+</sup> increased. Furthermore, it is worth noting that the colors of the patterned papers were almost colorless until the target concentration was below 0.1  $\mu$ M, which demonstrated that Hg<sup>2+</sup> ions can strongly inhibit the peroxidase-like activity of PtNPs. In addition, it also can be observed in the inset of Fig. 2B that the value of optical fiber reader exhibits a good linear correlation with the logarithm of Hg<sup>2+</sup> concentration over the range from 0.025  $\mu$ M to 0.5  $\mu$ M. The correlation equation was  $V = 8446.772 + 375.773 \lg C$ , where V was the response value of the fiber optical device and C was the concentration of Hg<sup>2+</sup>. The results demonstrated that this PCD could accurately discriminate Hg<sup>2+</sup> concentration at 0.01  $\mu$ M, which make it suitable for monitoring low content Hg<sup>2+</sup> in water samples. The reproducibility and stability of PCD was good since similar results were achieved for the three replicate analyses.

**Selectivity of PCD for Hg<sup>2+</sup> Determination.** The specificity of the method was investigated by viewing the response to other environmentally relevant metallic ions (Na<sup>+</sup>, K<sup>+</sup>, Ba<sup>2+</sup>, Cd<sup>2+</sup>, Zn<sup>2+</sup>, Pb<sup>2+</sup>, Ca<sup>2+</sup>, Cu<sup>2+</sup>, Mg<sup>2+</sup>, Al<sup>3+</sup>, Ni<sup>2+</sup>, Fe<sup>3+</sup>, Fe<sup>2+</sup>, Sr<sup>2+</sup>) and other common anions (Cl<sup>-</sup>, SO<sub>4</sub><sup>2-</sup>, NO<sub>3</sub><sup>-</sup>, CO<sub>3</sub><sup>2-</sup>, PO<sub>4</sub><sup>2-</sup>) under the same experimental conditions as used for Hg<sup>2+</sup> sensing. The final concentration of Hg<sup>2+</sup> ions and other ions was adjusted to 0.1  $\mu$ M and 10  $\mu$ M, respectively. By comparing the color intensities of the paper chips for different metallic ions and some anions, we found that all controlled ions did not exhibit inhibitory effects on the PtNPs, with the result that large blue precipitates formed within several min. as a result of catalysis of the TMB-H<sub>2</sub>O<sub>2</sub> reaction. Also, as shown in Fig. 3, the metal ions gave appropriate response signals when the fiber optic module was used. Clearly the presence of Hg<sup>2+</sup> in these samples would induce a large increase in readout value. These results indicated that the approach offers excellent selectivity for Hg<sup>2+</sup> detection and is well suited to field monitoring of environmental waters. However, probably due to lacking enough sensitivity, our PCD platform showed less positive response to S<sup>2-</sup> ions. We found that S<sup>2-</sup> ions performed slight inhibitory activity to PtNPs only its concentration was less than 0.1  $\mu$ M (This dates were not listed on the Fig. 3).



**Figure 3.** Selectivity of PCD for detection of  $\text{Hg}^{2+}$  (0.1  $\mu\text{M}$ ). Concentrations of metal ions and other anions, 10  $\mu\text{M}$ . The photograph shows PCD colorimetric response to various control ions and  $\text{Hg}^{2+}$  (indicated by the bold type words).



**Figure 4.** Real samples colorimetric assay by PCD with 0.025  $\mu\text{M}$   $\text{Hg}^{2+}$  (black) and no  $\text{Hg}^{2+}$  (red). (A) standard solution for  $\text{Hg}^{2+}$  analysis. (B,C)  $\text{Hg}^{2+}$  spiked pond and tap water samples analysis.

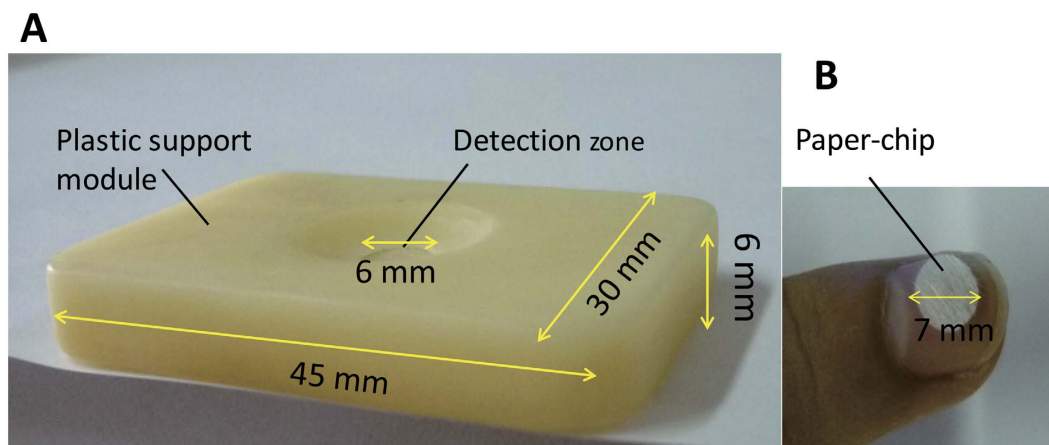
**Detection of  $\text{Hg}^{2+}$  In Real Samples.** On the basis of the sensitivity and selectivity studies, the practicality of deploying the PCD device for real water sensing was investigated. For this purpose, the signal intensities of  $\text{Hg}^{2+}$  spiked pond and tap water samples were compared with those achieved for standard solutions of  $\text{Hg}^{2+}$  (0  $\mu\text{M}$  or 0.025  $\mu\text{M}$ ), analyses being performed in triplicate. Figure 4 shows that the response values from pond and tap water samples were in good accord with the results of the standard solutions. Recovery ranged from 88% to 98.4% (Table S1) were analyzed by the addition and recovery experiments, which indicated that this new sensing platform would be suitable for the assay of trace  $\text{Hg}^{2+}$  in real samples.

## Conclusions

In summary, we have developed a portable PCD that utilizes PtNPs as a signal amplification probe for enzymatic oxidation of TMB whereby a colored product is produced on sensing paper. The device is also combined with a fiber optic module to give a digital readout of color intensity. The PCD permits determination of  $\text{Hg}^{2+}$  at concentrations as low as 0.01  $\mu\text{M}$  within several minutes. Additionally, the approach does not depend on expensive instrumentation and the simple operating procedures mean that non-specialists can perform the analysis. Given that this paper-based platform offers advantages of low-cost, convenience and near real-time analysis<sup>51</sup>, the approach has great potential as a new tool for field measurement of  $\text{Hg}^{2+}$  particularly in underdeveloped countries.

## Methods

**Materials and reagents.** Trisodium citrate ( $\text{Na}_3\text{C}_6\text{H}_5\text{O}_7 \cdot 2\text{H}_2\text{O}$ ) and 3,3',5,5'-tetramethylbenzidine (TMB) were purchased from Sigma-Aldrich. (St. Louis, MO). Hydrogen peroxide (30% w/w  $\text{H}_2\text{O}_2$ ) and  $\text{H}_2\text{PtCl}_6 \cdot (\text{H}_2\text{O})_6$  were obtained from Sinopharm Chemical Reagent Co. Ltd. (Shanghai, China). Glass fiber paper was purchased



**Figure 5.** Components of the portable paper-based colorimetric device (PCD). (A) Stereo structure of rectangular plastic module ( $45 \times 30 \times 6 \text{ mm}^3$ ) containing a round-hole (6 mm diameter). (B) Image of Hydrophilic glass fiber paper-chip with a diameter of 7 mm.

from Shanghai Kinbio Tech. Co. Ltd. Other reagents and chemicals were at least of analytical reagent grade and used without further purification. The TMB/ $\text{H}_2\text{O}_2$  colorimetric substrate solution consisted of 2% v/v TMB (30 mM) and  $\text{H}_2\text{O}_2$  (30% w/w), 140 mM NaAc in 20 mM Tris-HAc buffer (pH 4.0). All stock solutions were initially prepared by dissolving reagents in high purity water (Millipore water, 18.3 M $\Omega$ ).

**Synthesis of citrate-capped PtNPs.** The synthesis of citrate-PtNPs was based on a previously reported procedure that used a citrate reduction method<sup>37</sup>. First, all glassware and a magnetic stirrer bar were cleaned in aqua regia ( $\text{HCl}/\text{HNO}_3$  3:1 v/v), then rinsed with ultrapure water several times and oven-dried overnight. 7.78 mg of platinum chloride ( $\text{H}_2\text{PtCl}_6 \cdot 6\text{H}_2\text{O}$ ), dissolved in deionized water (40 mL), was vigorously mixed with trisodium citrate (0.4 mL, 100 mM). After stirring for 30 min at room temperature, freshly prepared aqueous sodium borohydride (200  $\mu\text{L}$ , 50 mM) was added dropwise to the mixture. Next, the mixture was stirred slowly until the color of the solution remained stable. Finally, the colloidal solution was stored at 4 °C in the dark. The stored solution of PtNPs was still color-uniformity, almost no aggregations and precipitates can be found during half a year period.

**Preparation and design of PCD.** The PCD design and features are displayed in Fig. 5, a previous report describing the printing technique used for device fabrication<sup>19</sup>. A circular-shaped paper chip of 7 mm diameter was designed using a drawing software package that was connected to a cutting plotter (Silhouette GAMEO, made in Vietnam). Subsequently, we designed a rectangular plastic module with a round-hole (6 mm diameter) as a detection zone which could support the paper-chips. As the paper was stacked into the printer, multiple 7 mm-diameter paper chips were fabricated at the same time to align with the hole in the plastic support module. Thus, the fabricated PCD, which combined the plastic support module with the paper-chip, was able to be used for  $\text{Hg}^{2+}$  assay.

**Digital Images and Dates collection.** We obtained the results of the PCD assays with a digital camera and a fiber optic device. The camera was used to clearly capture the color information of the paper-chip. Moreover, we selected a fiber optic device as a portable and off-site tool which could transmit the color change to a digital value for  $\text{Hg}(\text{II})$  analysis. Image of the fiber optic device with the PCD sensor was showed in Fig. S3, Support Information. For real-time result analysis, we vertically fixed a fiber optic probe above the detection zone of PCD at height of 0.3 mm. This made it suitable for accurately data acquisition. In principle, the probe emit the red laser, identify the reflective laser intensity and present it as a digital pattern simultaneously, thus light-colored paper-chips reflect most red laser and deep-colored chips would absorb more light which could induce a remarkable result discriminations.

**Colorimetric detection of  $\text{Hg}^{2+}$ .** For method development, several relatively simple procedures were followed. First, we used a plastic dropping bottle (5 ml) as a reactor to mix the samples containing  $\text{Hg}(\text{II})$  with the PtNPs solution. As the  $\text{Hg}(\text{II})$  was diluted in deionized water to specified concentrations, 2 ml of the  $\text{Hg}(\text{II})$  sample was added to the dropping bottle to effect rapid reaction (2 min.) with the PtNPs solution (1 ml). One drop of reagent (ca. 10  $\mu\text{L}$ ) from the dropper bottle was dispensed onto the detection zone where 10  $\mu\text{L}$  of TMB/ $\text{H}_2\text{O}_2$  colorimetric substrate solution had already been placed. The paper underwent complete reaction within 5 minutes and the colored images were captured by digital camera or smartphone and where the specific color intensity of the paper was measured by the fiber optic device.

**Real sample analysis.** Real samples of pond water and tap water were collected in 10 mL bottles and measured by the PCD (Support Information Fig. S3). The water samples were spiked to give a  $\text{Hg}(\text{II})$  concentration of 0.025  $\mu\text{M}$  and then mixed with PtNPs solution. After waiting 2 minutes, one drop of solution was placed on

the detection zone of the paper chip and allowed to react completely. The colored papers were analyzed and all analyses were performed at least three times.

## References

1. Meredith, N. A. *et al.* Paper-based analytical devices for environmental analysis. *Analyst* **141**, 1874–1887 (2016).
2. Sun, L.-J. *et al.* Paper-based electroanalytical devices for *in situ* determination of salicylic acid in living tomato leaves. *Biosens. Bioelectron.* **60**, 154–160 (2014).
3. Hossain, S. M. Z., Luckham, R. E., McFadden, M. J. & Brennan, J. D. Reagentless Bidirectional Lateral Flow Bioactive Paper Sensors for Detection of Pesticides in Beverage and Food Samples. *Anal. Chem.* **81**, 9055–9064 (2009).
4. Li, M. *et al.* “Periodic-Table-Style” Paper Device for Monitoring Heavy Metals in Water. *Anal. Chem.* **87**, 2555–2559 (2015).
5. Zhang, Y., Zuo, P. & Ye, B.-C. A low-cost and simple paper-based microfluidic device for simultaneous multiplex determination of different types of chemical contaminants in food. *Biosens. Bioelectron.* **68**, 14–19 (2015).
6. Connelly, J. T., Rolland, J. P. & Whitesides, G. M. “Paper Machine” for Molecular Diagnostics. *Anal. Chem.* **87**, 7595–7601 (2015).
7. Foudeh, A. M., Fatanat Didar, T., Veres, T. & Tabrizian, M. Microfluidic designs and techniques using lab-on-a-chip devices for pathogen detection for point-of-care diagnostics. *Lab Chip* **12**, 3249–3266 (2012).
8. Mu, X., Zhang, L., Chang, S., Cui, W. & Zheng, Z. Multiplex Microfluidic Paper-based Immunoassay for the Diagnosis of Hepatitis C Virus Infection. *Anal. Chem.* **86**, 5338–5344 (2014).
9. Hsu, M.-Y. *et al.* Monitoring the VEGF level in aqueous humor of patients with ophthalmologically relevant diseases via ultrahigh sensitive paper-based ELISA. *Biomaterials* **35**, 3729–3735 (2014).
10. Martinez, A. W., Phillips, S. T., Butte, M. J. & Whitesides, G. M. Patterned Paper as a Platform for Inexpensive, Low-Volume, Portable Bioassays. *Angew. Chem. Int. Ed.* **46**, 1318–1320 (2007).
11. Martinez, A. W., Phillips, S. T., Whitesides, G. M. & Carrilho, E. Diagnostics for the Developing World: Microfluidic Paper-Based Analytical Devices. *Anal. Chem.* **82**, 3–10 (2010).
12. Tao, H. *et al.* Metamaterials on Paper as a Sensing Platform. *Adv. Mater.* **23**, 3197–3201 (2011).
13. Liu, H. & Crooks, R. M. Paper-Based Electrochemical Sensing Platform with Integral Battery and Electrochromic Read-Out. *Anal. Chem.* **84**, 2528–2532 (2012).
14. Wu, Y., Xue, P., Kang, Y. & Hui, K. M. Paper-Based Microfluidic Electrochemical Immunodevice Integrated with Nanobioprobes onto Graphene Film for Ultrasensitive Multiplexed Detection of Cancer Biomarkers. *Anal. Chem.* **85**, 8661–8668 (2013).
15. Ge, L. *et al.* Three-dimensional paper-based electrochemiluminescence immunodevice for multiplexed measurement of biomarkers and point-of-care testing. *Biomaterials* **33**, 1024–1031 (2012).
16. Hong, J. I. & Chang, B.-Y. Development of the smartphone-based colorimetry for multi-analyte sensing arrays. *Lab Chip* **14**, 1725–1732 (2014).
17. Liang, P., Yu, H., Guntupalli, B. & Xiao, Y. Paper-Based Device for Rapid Visualization of NADH Based on Dissolution of Gold Nanoparticles. *ACS Appl. Mater. Inter.* **7**, 15023–15030 (2015).
18. He, M. & Liu, Z. Paper-Based Microfluidic Device with Upconversion Fluorescence Assay. *Anal. Chem.* **85**, 11691–11694 (2013).
19. Li, H., Fang, X., Cao, H. & Kong, J. Paper-based fluorescence resonance energy transfer assay for directly detecting nucleic acids and proteins. *Biosens. Bioelectron.* **80**, 79–83 (2016).
20. Chen, Y. *et al.* A paper-based surface-enhanced resonance Raman spectroscopic (SERRS) immunoassay using magnetic separation and enzyme-catalyzed reaction. *Analyst* **138**, 2624–2631 (2013).
21. Zhang, K. *et al.* Multifunctional Paper Strip Based on Self-Assembled Interfacial Plasmonic Nanoparticle Arrays for Sensitive SERS Detection. *ACS Appl. Mater. Inter.* **7**, 16767–16774 (2015).
22. Lewis, G. G., DiTucci, M. J. & Phillips, S. T. Quantifying Analytes in Paper-Based Microfluidic Devices Without Using External Electronic Readers. *Angew. Chem. Int. Ed.* **51**, 12707–12710 (2012).
23. Cheng, C. M. *et al.* Paper-based ELISA. *Angew. Chem. Int. Ed.* **49**, 4771–4774 (2010).
24. Lopez-Ruiz, N. *et al.* Smartphone-Based Simultaneous pH and Nitrite Colorimetric Determination for Paper Microfluidic Devices. *Anal. Chem.* **86**, 9554–9562 (2014).
25. Mei, Q. *et al.* Smartphone based visual and quantitative assays on upconversional paper sensor. *Biosens. Bioelectron.* **75**, 427–432 (2016).
26. Park, T. S., Li, W., McCracken, K. E. & Yoon, J.-Y. Smartphone quantifies Salmonella from paper microfluidics. *Lab Chip* **13**, 4832–4840 (2013).
27. Lin, Y., Ren, J. & Qu, X. Nano-Gold as Artificial Enzymes: Hidden Talents. *Adv. Mater.* **26**, 4200–4217 (2014).
28. Lin, Y., Ren, J. & Qu, X. Catalytically Active Nanomaterials: A Promising Candidate for Artificial Enzymes. *Accounts of Chemical Research* **47**, 1097–1105 (2014).
29. Song, Y., Qu, K., Zhao, C., Ren, J. & Qu, X. Graphene Oxide: Intrinsic Peroxidase Catalytic Activity and Its Application to Glucose Detection. *Adv. Mater.* **22**, 2206–2210 (2010).
30. Zhang, Y., Xu, C., Li, B. & Li, Y. *In situ* growth of positively-charged gold nanoparticles on single-walled carbon nanotubes as a highly active peroxidase mimetic and its application in biosensing. *Biosens. Bioelectron.* **43**, 205–210 (2013).
31. Shi, W. *et al.* Carbon nanodots as peroxidase mimetics and their applications to glucose detection. *Chem Commun* **47**, 6695–6697 (2011).
32. Wang, T. *et al.* Biosensor Based on Ultrasmall MoS<sub>2</sub> Nanoparticles for Electrochemical Detection of H<sub>2</sub>O<sub>2</sub> Released by Cells at the Nanomolar Level. *Anal. Chem.* **85**, 10289–10295 (2013).
33. Fan, J. *et al.* Direct evidence for catalase and peroxidase activities of ferritin–platinum nanoparticles. *Biomaterials* **32**, 1611–1618 (2011).
34. Asati, A., Santra, S., Kaittanis, C., Nath, S. & Perez, J. M. Oxidase-Like Activity of Polymer-Coated Cerium Oxide Nanoparticles. *Angew. Chem. Int. Ed.* **48**, 2308–2312 (2009).
35. Zhai, D. *et al.* Highly Sensitive Glucose Sensor Based on Pt Nanoparticle/Polyaniline Hydrogel Heterostructures. *ACS Nano* **7**, 3540–3546, doi: 10.1021/nn400482d (2013).
36. Lin, Y., Li, Z., Chen, Z., Ren, J. & Qu, X. Mesoporous silica-encapsulated gold nanoparticles as artificial enzymes for self-activated cascade catalysis. *Biomaterials* **34**, 2600–2610 (2013).
37. Wu, G.-W. *et al.* Citrate-Capped Platinum Nanoparticle as a Smart Probe for Ultrasensitive Mercury Sensing. *Anal. Chem.* **86**, 10955–10960 (2014).
38. Date, Y. *et al.* Trace-Level Mercury Ion (Hg<sup>2+</sup>) Analysis in Aqueous Sample Based on Solid-Phase Extraction Followed by Microfluidic Immunoassay. *Anal. Chem.* **85**, 434–440 (2013).
39. Huang, D. *et al.* Highly Sensitive Strategy for Hg<sup>2+</sup> Detection in Environmental Water Samples Using Long Lifetime Fluorescence Quantum Dots and Gold Nanoparticles. *Environ. Sci. Technol.* **47**, 4392–4398 (2013).
40. Lee, J.-S., Han, M. S. & Mirkin, C. A. Colorimetric Detection of Mercuric Ion (Hg<sup>2+</sup>) in Aqueous Media using DNA-Functionalized Gold Nanoparticles. *Angew. Chem. Int. Ed.* **46**, 4093–4096 (2007).
41. Li, J. *et al.* Engineering noble metal nanomaterials for environmental applications. *Nanoscale* **7**, 7502–7519 (2015).
42. Liu, D. *et al.* Highly Sensitive, Colorimetric Detection of Mercury(II) in Aqueous Media by Quaternary Ammonium Group-Capped Gold Nanoparticles at Room Temperature. *Anal. Chem.* **82**, 9606–9610 (2010).

43. Liu, D. *et al.* Highly Robust, Recyclable Displacement Assay for Mercuric Ions in Aqueous Solutions and Living Cells. *ACS Nano*. **6**, 10999–11008 (2012).
44. Wei, Q. *et al.* Detection and Spatial Mapping of Mercury Contamination in Water Samples Using a Smart-Phone. *ACS Nano*. **8**, 1121–1129 (2014).
45. Darbha, G. K., Ray, A. & Ray, P. C. Gold Nanoparticle-Based Miniaturized Nanomaterial Surface Energy Transfer Probe for Rapid and Ultrasensitive Detection of Mercury in Soil, Water, and Fish. *ACS Nano*. **1**, 208–214 (2007).
46. Liu, X. *et al.* Single Gold Nanoparticle-Based Colorimetric Detection of Picomolar Mercury Ion with Dark-Field Microscopy. *Anal. Chem.* **88**, 2119–2124 (2016).
47. Lien, C. W., Tseng, Y. T., Huang, C. C. & Chang, H. T. Logic control of enzyme-like gold nanoparticles for selective detection of lead and mercury ions. *Anal. Chem.* **86**, 2065–2072 (2014).
48. Chen, Z. *et al.* Colorimetric Signal Amplification Assay for Mercury Ions Based on the Catalysis of Gold Amalgam. *Anal. Chem.* **87**, 10963–10968 (2015).
49. Wei, H. & Wang, E. Nanomaterials with enzyme-like characteristics (nanozymes): next-generation artificial enzymes. *Chem. Soc. Rev.* **42**, 6060–6093 (2013).
50. Chen, G.-H. *et al.* Detection of Mercury(II) Ions Using Colorimetric Gold Nanoparticles on Paper-Based Analytical Devices. *Anal. Chem.* **86**, 6843–6849 (2014).
51. Cate, D. M., Adkins, J. A., Mettakoonpitak, J. & Henry, C. S. Recent Developments in Paper-Based Microfluidic Devices. *Anal. Chem.* **87**, 19–41 (2015).

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## Author Contributions

W.C. and X.F. conceived and designed the experiments. W.C., H.L. and H.C. performed the experiments. X.F. and H.C. analysed the data. J.K. and X.F. drafted the manuscript. All authors read and approved the final manuscript.

## Additional Information

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