



# Draft Genome Sequence of Freshwater-Derived *Streptomyces* sp. Strain BPSDS2, Isolated from Damte Stream, Northeast India

Zothanpuia,<sup>a</sup> Ajit Kumar Passari,<sup>a</sup> Purbajyoti Deka,<sup>a</sup> Vinay Rajput,<sup>b</sup> Lakshmi P. M. Priya,<sup>c</sup>  Mahesh Dharne,<sup>b</sup> Syed Dastager,<sup>b</sup> Oommen K. Mathew,<sup>c</sup> Abeer Hashem,<sup>d,e</sup> Elsayed Fathi Abd\_ Allah,<sup>f</sup>  Bhim Pratap Singh<sup>a</sup>

<sup>a</sup>Department of Biotechnology, Mizoram University, Aizawl, Mizoram, India

<sup>b</sup>NCIM Resource Centre, CSIR-National Chemical Laboratory, Pune, India

<sup>c</sup>AgriGenome Labs Pvt Ltd., SmartCity Kochi, Kerala, India

<sup>d</sup>Botany and Microbiology Department, College of Science, King Saud University, Riyadh, Saudi Arabia

<sup>e</sup>Mycology and Plant Disease Survey Department, Plant Pathology Research Institute, Agriculture Research Center, Giza, Egypt

<sup>f</sup>Plant Production Department, College of Food and Agriculture Science, King Saud University, Riyadh, Saudi Arabia

**ABSTRACT** We report the draft genome sequence of *Streptomyces* sp. strain BPSDS2, isolated from freshwater sediments in Northeast India. The draft genome has a size of 8.27 Mb and 7,559 protein-coding sequences.

*Streptomyces* spp. are Gram-positive filamentous spore-forming bacteria, belong to the most dominant genus of the *Actinobacteria*, have high G+C content in their DNA (1), and are remarkably rich sources of bioactive compounds, accounting for the production of over two-thirds of the commercially available antibiotics in current use (2). *Streptomyces* spp. account for the production of 75% of antibiotics in use from the total of 70% of antibiotics produced by the phylum *Actinobacteria* (3). In the course of our screening program for new bioactive compounds from freshwater actinobacteria, *Streptomyces* sp. strain BPSDS2 was isolated from a freshwater sediment sample from Damte Stream (23°73'N, 92°80'E) in Mizoram, Northeast India.

The sediment sample was diluted in distilled water, and the dilutions were spread over the surface of starch casein agar (SCA) solid agar medium. The plates were incubated at 28°C for 7 days, and pure colonies were obtained after repeated subculturing on fresh isolation medium (4). The pure colonies were then transferred to tryptone-yeast extract broth (ISP1 broth) liquid medium, and axenicity was confirmed by Gram staining. Cells were harvested, and genomic DNA was extracted using the PureLink genomic DNA isolation kit (catalog number K182002; Thermo Scientific Invitrogen). The 16S rRNA gene was amplified using universal bacterial primers. The primers, reactions, and conditions of the PCR were exactly as reported in our previous studies (5). The obtained sequences were compared with the reference strains of actinobacteria from the NCBI genomic database using a BLASTn search to determine similarity percentages. Based on 16S rRNA gene sequence analysis, strain BPSDS2 (GenBank accession number [MG711553](https://doi.org/10.1128/MRA.00874-19)) was found to be closely similar to *Streptomyces* sp. strain QLS87 (GenBank accession number [JQ838120](https://doi.org/10.1128/MRA.00874-19)), with 99% similarity.

For genome sequencing, 500 ng of the isolated good-quality genomic DNA was fragmented using a Covaris M220 sonicator. The fragmented genomic DNA was end repaired and further processed for ligation of Illumina adaptors with the NEBNext Ultra DNA library preparation kit (catalog number E7370L; NEB), as per the recommendation of the manufacturer. An adaptor-ligated enriched library was purified using AMPure XP beads. Library size distribution was checked on an Agilent TapeStation D1000 DNA chip (product number 5067-5583). *Streptomyces* sp. strain BPSDS2 genomic DNA was se-

**Citation** Zothanpuia, Passari AK, Deka P, Rajput V, Priya LPM, Dharne M, Dastager S, Mathew OK, Hashem A, Abd\_ Allah EF, Singh BP. 2019. Draft genome sequence of freshwater-derived *Streptomyces* sp. strain BPSDS2, isolated from Damte Stream, Northeast India. *Microbiol Resour Announc* 8:e00874-19. <https://doi.org/10.1128/MRA.00874-19>.

**Editor** Julia A. Maresca, University of Delaware

**Copyright** © 2019 Zothanpuia et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Bhim Pratap Singh, [bhimpratap@gmail.com](mailto:bhimpratap@gmail.com).

**Received** 21 July 2019

**Accepted** 30 September 2019

**Published** 24 October 2019

quenced with the  $2 \times 150$ -bp paired-end read length sequencing protocol of the Illumina MiSeq platform. The quality check of the reads was done using FastQC (6), and the generated sequencing reads were filtered to remove low-quality reads using Trim Galore v0.5.0 (7), with set default parameters. Unicycler v0.4.8 (8) was used for *de novo* assembly. The *Streptomyces* sp. strain BPSDS2 draft genome sequence contains 54 contigs, with an  $N_{50}$  value of 349,850 bp. The genome coverage is  $\sim 234.0\times$ , and the estimated genome length is 8,272,875 bp, with an average G+C content of 71.82%. The *Streptomyces* sp. strain BPSDS2 genome was annotated on the PATRIC Web server (9) using the RAST tool kit (RASTtk) (10), and it contains 7,546 protein-coding sequences (CDS), 65 tRNA genes, and 3 rRNA genes. The annotation of this strain consists of 2,498 hypothetical proteins and 5,048 proteins with functional assignments, including 1,233 proteins with Enzyme Commission (EC) numbers, 1,070 with Gene Ontology (GO) assignments, and 968 proteins that were mapped to the KEGG pathway.

**Data availability.** This whole-genome shotgun project has been deposited at the NCBI database under GenBank accession numbers [STGN01000001](#) to [STGN01000054](#). The BioSample accession number is [SAMN11159182](#). The BioProject identifier is [PRJNA527763](#). The short-read data have been submitted to the SRA under run accession number [SRR8742575](#).

## ACKNOWLEDGMENTS

This work was supported by DBT's Unit of Excellence program for NE (grant 102/IFD/SAN/4290-4291/2016-2017). We also thank the Deanship of Scientific Research at King Saud University for funding this research group (RG-1435-014).

## REFERENCES

- Goodfellow M, Fiedler HP. 2010. A guide to successful bioprospecting: informed by actinobacterial systematics. *Antonie Van Leeuwenhoek* 98:119–142. <https://doi.org/10.1007/s10482-010-9460-2>.
- Bentley SD, Chater KF, Cerdeño-Tárraga A-M, Challis GL, Thomson NR, James KD, Harris DE, Quail MA, Kieser H, Harper D, Bateman A, Brown S, Chandra G, Chen CW, Collins M, Cronin A, Fraser A, Goble A, Hidalgo J, Hornsby T, Howarth S, Huang C-H, Kieser T, Larke L, Murphy L, Oliver K, O'Neil S, Rabinowitsch E, Rajandream M-A, Rutherford K, Rutter S, Seeger K, Saunders D, Sharp S, Squares R, Squares S, Taylor K, Warren T, Wietzorrek A, Woodward J, Barrell BG, Parkhill J, Hopwood DA. 2002. Complete genome sequence of the model actinomycete *Streptomyces coelicolor* A3(2). *Nature* 417:141–147. <https://doi.org/10.1038/417141a>.
- Ser H, Mutalib AB, Yin N, Chan K, Goh B, Lee L. 2015. Evaluation of antioxidative and cytotoxic activities of *Streptomyces pluripotens* MUSC 137 isolated from mangrove soil in Malaysia. *Front Microbiol* 6:1398. <https://doi.org/10.3389/fmicb.2015.01398>.
- Zothanpuia, Passari AK. 2018. Bioprospection of actinobacteria derived from freshwater sediments for their potential to produce antimicrobial compounds. *Microb Cell Fact* 17:68. <https://doi.org/10.1186/s12934-018-0912-0>.
- Zothanpuia, Passari AK, Gupta VK, Singh BP. 2016. Detection of antibiotic-resistant bacteria endowed with antimicrobial activity from a freshwater lake and their phylogenetic affiliation. *PeerJ* 4:e2103. <https://doi.org/10.7717/peerj.2103>.
- Andrews S. 2010. FastQC: a quality control tool for high throughput sequence data. <http://www.bioinformatics.babraham.ac.uk/projects/fastqc/>.
- Krueger F. 2015. Trim Galore: a wrapper tool around Cutadapt and FastQC to consistently apply quality and adapter trimming to FastQ files. [https://www.bioinformatics.babraham.ac.uk/projects/trim\\_galore/](https://www.bioinformatics.babraham.ac.uk/projects/trim_galore/).
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. *PLoS Comput Biol* 13:e1005595. <https://doi.org/10.1371/journal.pcbi.1005595>.
- Wattam AR, Abraham D, Dalay O, Disz TL, Driscoll T, Gabbard JL, Gillespie JJ, Gough R, Hix D, Kenyon R, Machi D, Mao C, Nordberg EK, Olson R, Overbeek R, Pusch GD, Shukla M, Schulman J, Stevens RL, Sullivan DE, Vonstein V, Warren A, Will R, Wilson MJ, Yoo HS, Zhang C, Zhang Y, Sobral BW. 2014. PATRIC, the bacterial bioinformatics database and analysis resource. *Nucleic Acids Res* 42:D581–D591. <https://doi.org/10.1093/nar/gkt1099>.
- Brettin T, Davis JJ, Disz T, Edwards RA, Gerdes S, Olsen GJ, Olson R, Overbeek R, Parrello B, Pusch GD, Shukla M, Thomason JA, III, Stevens R, Vonstein V, Wattam AR, Xia F. 2015. RASTtk: a modular and extensible implementation of the RAST algorithm for building custom annotation pipelines and annotating batches of genomes. *Sci Rep* 5:8365. <https://doi.org/10.1038/srep08365>.