FEBS openbio



C/EBPβ regulates the JAK/STAT signaling pathway in triple-negative breast cancer

Shu Wang¹, Dian Xia¹, Xianzhi Wang^{1,2}, Haowei Cao¹, Chaoshen Wu¹, Zhaoran Sun¹, Daoyong Zhang¹ (and Hao Liu²

1 Jiangsu Key Laboratory of Brain Disease and Bioinformation, Xuzhou Medical University, China

2 School of Pharmacy, Bengbu Medical College, China

Keywords

C/EBP β ; JAK; STAT; Triple-Negative Breast Cancer

Correspondence

D. Zhang, Jiangsu Key Laboratory of Brain Disease Bioinformation, Xuzhou Medical University, Xuzhou, Jiangsu, China E-mail: zhangdaoyong@tsinghua.org.cn H. Liu, School of Pharmacy, Bengbu Medical College, Bengbu, Anhui, China E-mail: liuhao6886@foxmail.com Shu Wang, Dian Xia and Xianzhi Wang contribute equally to this work

(Received 15 August 2020, revised 17 December 2020, accepted 3 March 2021)

doi:10.1002/2211-5463.13138

C/EBP β is a member of the CCAAT/enhancer-binding protein (C/EBP) family, which consists of a number of b-ZIP transcription factors. Although C/EBP β has been implicated in the development of certain cancers, including breast cancer, it remains unknown whether dysregulation of $C/EBP\beta$ in breast cancer is subtype-specific. Moreover, the underlying mechanisms by which C/EBPß regulates breast cancer carcinogenesis are not fully understood. Here, we present evidence that $C/EBP\beta$ is specifically overexpressed in human TNBC samples, but not in non-TNBC samples. $C/EBP\beta$ depletion dramatically suppressed TNBC cell growth, migration, invasion, and colony formation ability. A subsequent mechanistic study revealed that the JAK/STAT signaling pathway was upregulated in C/ EBPβ_high TNBC samples compared with C/EBPβ_low TNBC samples. C/EBP_β ChIP-seq and qPCR were performed to demonstrate that C/EBP_β directly binds to and regulates JAK/STAT signaling pathway genes in TNBC. Taken together, our data indicate the oncogenic role of C/EBP β in human TNBC and reveal a novel mechanism by which C/EBP^β promotes TNBC carcinogenesis.

C/EBPβ is a member of the CCAAT/enhancer-binding protein (C/EBP) family, whose members share a highly conserved leucine-zipper dimerization domain at the C-terminal and an adjacent basic DNA-binding region. The less conserved N-terminal activation domain interacts with various transcriptional coactivators and general transcriptional factors [1–4]. C/EBPs act as dimers to bind to DNA in a sequence-specific manner to regulate transcription of the target genes [5–7]. Increasing evidence shows that C/EBPs play important roles in cell growth, differentiation, apoptosis, inflammation, aging, and neuronal autophagy [8–13].

Emerging evidence has also implicated the crucial regulatory roles of C/EBP β in a few types of cancer, including breast cancer, pancreatic cancer, sarcoma,

and liver cancer [14–17]. Although C/EBP β is required for normal mammary gland growth and differentiation [18], its overexpression is capable of transforming normal human mammary epithelial cells, as evidenced by gaining anchorage independence, epithelial-to-mesenchymal transition marker gene expression, and the acquisition of an invasive phenotype [17]. These events are evidence that precisely regulated C/EBP β expression is critical in normal mammary gland development, and dysregulation leads to breast carcinogenesis. Moreover, C/EBP β is shown to be essential in maintaining mammary stem cells and luminal cell lineage commitment [19]. Although studies have indicated that C/EBP β has important roles in a few types of cancer, the regulatory role of C/EBP β has not been specifically

Abbreviations

C/EBPβ, CCAAT/enhancer-binding protein beta; TNBC, triple-negative breast cancer.

FEBS Open Bio **11** (2021) 1250–1258 © 2021 The Authors. *FEBS Open Bio* published by John Wiley & Sons Ltd on behalf of Federation of European Biochemical Societies This is an open access article under the terms of the Creative Commons Attribution License, which permits use,

and thoroughly studied in triple-negative breast cancer (TNBC), and the underlying mechanisms remain largely unknown.

In the present study, we attempted to dissect the potential role of C/EBP β specifically in TNBC and to understand the underlying mechanisms.

Materials and methods

Data source

mRNA expression data and clinical phenotype data of breast cancer from The Cancer Genome Atlas (TCGA) were downloaded from the UCSC Xena TCGA Hub. This included 1097 primary breast cancer samples and 113 normal breast tissue samples. According to the clinical phenotypes, 122 cases were TNBC and 617 cases were non-TNBC, whereas the remaining cases had at least one of the progesterone receptor (PR) status, ER status, and HER-2 status missing.

Cell culture and RNAi

The TNBC cell line BT549 was originally purchased from ATCC. BT549 cells were cultured in RPMI 1640 medium (Invitrogen, Invitrogen, CA, USA) supplemented with 10% FBS. The shRNAs targeting C/EBPβ and the control scrambled sequence were as follows: C/EBPβ-sh1, GCCGC CGCCTGCCTTTAAATC; C/EBPβ-sh2, GCCCTGAGTA ATCGCTTAAAG; shControl, TTCTCCGAACGTGTCA CGT. Lentivirus preparation was performed as previously described [20]. The lentivirus was then used to infect BT549 cells to knock down C/EBPβ expression followed by selection using puromycin.

Antibodies

The antibodies used in this study include anti-C/EBPβ (Cat. No. sc-7962; SANTA CRUZ, Santa Cruz, CA, USA) and anti-STAT5A (Cat. No. 13179-1-AP; ProteinTech, ProteinTech, IL, USA).

Quantitative real-time PCR

RNA extraction, reverse transcription, and quantitative real-time PCR were carried out as previously described [20].

EdU incorporation assay and clonogenicity formation assay

BT549 cells were seeded at a density of 3×10^3 cells per well in 96-well plates and were incubated overnight. The media was prepared using EdU (5-ethynyl-2'-deoxyuridine)

for a final concentration of 50 μ M, and the plates were incubated in a cell culture incubator for 2 h. The EdU incorporation assay was then performed using an EdU *In vitro* Imaging Kit (Ribo Bio, Guangdong, China) following the manufacturer's protocol. The tumor cell colony-forming ability was evaluated as previously described [20].

Wound-healing assay and Transwell cell invasion assay

Wound-healing and Transwell cell invasion assays were performed as previously prescribed [20].

Gene Set Enrichment Analysis

C/EBP β expression in TNBC was divided into C/EBP β -high and C/EBP β -low expression groups using the median value. For the Gene Set Enrichment Analysis (GSEA), upregulated genes in the C/EBP β -high group were used with the clusterProfiler R package.

Results

C/EBPβ expression was dysregulated in human triple-negative breast cancer

The study was designed to investigate the potential regulatory role of C/EBP β in TNBC. To this end, we first compared C/EBPB expression levels in TNBC samples (n = 122) versus non-TNBC samples (n = 617) using data obtained from the UCSC Xena data hub. We found that the C/EBP β was aberrantly overexpressed (more than twofold) in TNBC compared with non-TNBC samples (Fig. 1A). Moreover, the $C/EBP\beta$ mRNA level in TNBC was also significantly higher than that in normal breast tissue samples (Fig. 1B). We also assessed the C/EBPB expression in TNBC using another dataset (GSE58135) and observed a similar overexpression pattern in TNBC samples compared with the noncancerous adjacent samples (Fig. 1C). More importantly, we also observed a significant association between C/EBPB expression and poor survival probability among TNBC patients, indicating the promising prognostic value of C/EBPB in TNBC (Fig. 1D). Taken together, these results indicate that C/ EBP β may play a promoting role specifically in TNBC.

C/EBPβ regulates TNBC cell proliferation, colony formation, migration, and invasion

To directly assess the regulatory role of C/EBP β in TNBC, we knocked down C/EBP β expression in BT549 cells. Knockdown efficiency was evaluated by



Fig. 1. C/EBP β is overexpressed in human TNBC. (A, B) The mRNA expression data obtained from the TCGA project were used to examine C/EBP β expression in human TNBC (n = 122), non-TNBC (n = 617), and normal breast tissue (n = 113) samples. The phenotype data obtained from the UCSC xena TCGA data hub were used to define TNBC and non-TNBC samples. (C) RNA-seq data from GSE58135 was used to assess C/EBP β expression in human TNBC. (D) The Kaplan–Meier plotter (http://kmplot.com/analysis/) survival analysis for the relationship between the survival time of TNBC patients and C/EBP β expression.

real-time quantitative PCR and western blotting (Fig. 2A). On the other hand, the EdU proliferation assay was used to assess the regulatory role of C/EBP β in TNBC cell proliferation. As shown in Fig. 2B,C, C/EBP β knockdown significantly inhibited TNBC cell

proliferation. Next, we assessed the effect of C/EBP β knockdown on TNBC cell colony formation using a clonogenic assay. The results showed that C/EBP β knockdown dramatically impaired the colony formation ability of TNBC cells (Fig. 2D,E).



Fig. 2. Knockdown of C/EBP β inhibits TNBC cell proliferation and colony-forming ability. (A) The knockdown efficiency of C/EBP β was verified at the levels of transcription and translation. (B, C) The EdU proliferation assay was performed to determine the effect of C/EBP β knockdown on TNBC cell proliferation. Three independent experiments were carried out, and the results were represented as mean + SD Scale bar, 100 μ m. (D, E) The clonogenicity formation assay was performed to assess the effect of C/EBP β knockdown on the colony-forming ability of TNBC cells. Three independent experiments were carried out, and the results were represented as mean + SD. **P < 0.01, ***P < 0.01, uppaired *t*-test.

To further elucidate the role of C/EBP β in TNBC, we also investigated the regulatory role of C/EBP β in TNBC cell migration and invasion, which are both important processes in breast cancer metastasis. To this end, wound-healing and Transwell cell invasion assays were performed. As shown in Fig. 3A,B, we observed slower migration of TNBC cells upon C/EBP β knockdown. Furthermore, the Transwell cell invasion assay results demonstrated that C/EBP β knockdown dramatically impaired the invasiveness of TNBC cells (Fig. 3C,D). These results together support the conclusion that C/EBP β plays a role in promoting TNBC.

C/EBPβ regulates JAK/STAT signaling pathway genes in TNBC

We next explored the underlying mechanisms by which C/EBP β promotes TNBC carcinogenesis. To this end, we divided the TNBC cases into two groups, the C/EBP β _high group and the C/EBP β _low group, based on the median C/EBP β expression level in TNBC.

Then, we compared the two groups and identified the differentially expressed genes (DEGs) using the DEseq2 R package. The gene expression patterns between the C/EBP β _high group and the C/EBP β _low group were evidently distinct (Fig. 4A,B). The upregulated genes in the C/EBP β _high group were then used in the GSEA analysis, and the results indicate that the JAK/STAT signaling pathway genes were enriched and upregulated in the C/EBP β _high group compared with the C/EBP β low group (Fig. 4C).

As a transcriptional factor, C/EBP β directly binds to DNA in a sequence-specific manner to regulate the expression of target genes. We were interested in determining whether these JAK/STAT signaling pathway genes were directly bound by C/EBP β at the chromatin level. Since C/EBP β recognizes and binds to specific DNA sequences, we first tried to identify potential C/EBP β -binding sites at the JAK/STAT signaling pathway genes. We used the STAT5A gene as an example in this study. We first obtained a C/EBP β binding DNA sequence motif from the JASPAR database (Fig. 4D) and then loaded the bed file to the IGV



Fig. 3. Knockdown of C/EBP β inhibits TNBC cell migration and invasion. (A, B) The wound-healing assay was carried out to examine the effect of C/EBP β knockdown on TNBC cell migration. Three independent experiments were carried out, and the results were represented as mean + SD. Scale bar, 200 μ m. (C, D) The Transwell invasion assay was performed to examine the effect of C/EBP β knockdown on TNBC cell invasion. Three independent experiments were represented as mean + SD. Scale bar, 200 μ m. (C, D) The Transwell invasion assay was performed to examine the effect of C/EBP β knockdown on TNBC cell invasion. Three independent experiments were carried out, and the results were represented as mean + SD. Scale bar, 100 μ m. **P < 0.01, ***P < 0.01, unpaired *t*-test.

S. Wang et al.



Fig. 4. C/EBPβ regulates the JAK/STAT signaling pathway in human TNBC. (A) Hierarchical cluster analysis of DEGs (||ogFC| > 0.5 & P < 1E-4) between C/EBPβ_high and C/EBPβ_low TNBC samples. Each cell in the matrix represents the expression of a gene in an individual sample. (B) Volcano plot of differential gene expression between C/EBPβ_high and C/EBPβ_low TNBC samples. Each dot represents one gene that had detectable expression in the RNA-seq experiment. The horizontal dashed line marks the threshold (*P.adj* < 0.05) for defining a gene as significantly upregulated or downregulated in C/EBPβ_high samples compared with C/EBPβ_low samples. The vertical dashed lines represent twofold differences in expression. Significantly DEGs ($||og_2(FC)| > 1$ and *P.adj* < 0.05) are marked in red. (C) Gene set enrichment analysis (GSEA) score curves. The JAK/STAT signaling pathway enrichment plot is shown. Black bars represent the position of genes of the JAK/STAT signaling pathway in the ranked list together with the running enrichment score (plotted in green). (D) The C/EBPβ binding motif was obtained from the JASPAR database. The character and size of each logo represent the proportion of a base at a specific site. (E) The positions of potential C/EBPβ-binding sites at the STAT5A gene, defined by its binding motif, are shown. (F) C/EBPβ ChIP-seq data (GSM2330565) were analyzed to determine C/EBPβ-binding signals at the STAT5A gene. (G) qRT-PCR assays were performed to examine the effect of C/EBPβ knockdown on JAK/STAT signaling pathway genes. Three independent experiments were carried out, and the results were represented as mean + S.D. **P* < 0.05, ***P* < 0.01, ****P* < 0.001. Unpaired *t*-test. (H) Western blotting assay was carried out to confirm the change in STAT5A expression at the protein level upon C/EBPβ knockdown.

software to identify C/EBP β -binding motif sites at the STAT5A gene locus. As seen in Fig. 4E, we identified five potential C/EBP β -binding sites throughout the STAT5A gene body region.

However, the existence of a C/EBP β -binding motif sequence does not necessarily mean that C/EBP β is actually bound. The binding of C/EBP β to the motif sequence is regulated by many other factors, and its genome-binding profile varies from cell to cell. To verify whether these sites are actually bound by C/EBP β in TNBC, we analyzed the ChIP-seq data for C/EBP β binding in SUM159PT cells, a TNBC cell line. The results showed that there was a strong C/EBP β -binding signal at the 3' end of the STAT5A gene body

region, corresponding to the 5th C/EBPβ-binding motif locus identified above (Fig. 4F). Weak binding of C/ EBPB was also observed in the middle region, corresponding to the 2nd C/EBPβ-binding motif locus. To further demonstrate the regulatory role of C/EBP β in TNBC cells, we assessed the effect of C/EBPB depletion on the expression of genes involved in the JAK/ STAT pathway in TNBC cells. The results demonstrated that the majority of JAK/STAT signaling pathway genes examined here were downregulated upon C/EBP β knockdown, including STAT5A (Fig. 4G,H), suggesting a positive regulatory role of $C/EBP\beta$ in the expression of these genes. Taken together, our results support that C/EBPB directly binds to JAK/STAT signaling pathway genes and regulates their expression in TNBC.

Discussion

TNBC is an aggressive subtype of breast cancer in which the expression of estrogen receptor (ER), PR, and human epidermal growth factor receptor-2 is absent [21,22]. The survival time of TNBC patients is shorter, and the mortality rate is higher compared with that of non-TNBC patients. Moreover, TNBC is highly invasive, and nearly half of TNBC patients will have distant metastasis. Here, we examined the regulatory role of C/EBPB, specifically in TNBC. Our data showed that $C/EBP\beta$ is significantly upregulated in TNBC samples compared with non-TNBC and normal breast tissue samples. C/EBPB depletion in TNBC cells inhibited their proliferation rate and colony formation ability. Since TNBC is a highly invasive breast cancer subtype, we also investigated the role of C/EBP β in TNBC cell migration and invasion, which are both critical processes in cancer metastasis. Our data demonstrated that C/EBPß knockdown dramatically inhibited TNBC cell migration and invasion. A subsequent mechanistic study showed that C/EBP β binds to JAK/ STAT signaling pathway genes and regulates their expression. Given that the dysregulation of the JAK/ STAT signaling pathway plays an instrumental role in breast cancer development and metastasis as well as in other types of cancer [23-29], we concluded that C/ EBPβ promotes TNBC carcinogenesis probably via the regulation of the JAK/STAT signaling pathway.

C/EBP β is an intronless gene, and due to multiple translation start sites within the mRNA, three distinct isoforms of C/EBP β proteins (LAP*, LAP, and LIP) are expressed. The C/EBP β isoform LAP has been previously reported to have an impact on the tumor microenvironment in TNBC [30]. The involvement of LAP in tumor immunity indicates that C/EBP β plays

a much broader role in TNBC than we previously thought. Although we observed C/EBP β overexpression in TNBC samples, we do not know which specific isoform is eventually overexpressed at the protein level. Previous studies showed that LAP* was the only C/EBP β protein expressed in normal mammary tissue and was absent from breast cancer cells by contrast. Instead, the isoform LAP was reportedly upregulated in primary breast tumors [17]. Therefore, we hypothesize that LAP is the isoform that is overexpressed in TNBC as a result of upregulated C/EBP β mRNA levels. However, further experimental data are needed to confirm this hypothesis.

Conclusion

In the present study, we observed elevated C/EBP β expression specifically in TNBC samples compared with non-TNBC and normal breast tissue samples. Further studies showed that C/EBP β knockdown inhibited TNBC carcinogenesis. Moreover, a mechanistic study revealed that C/EBP β binds to and regulates JAK/STAT signaling pathway genes in TNBC cells, indicating that C/EBP β may promote TNBC carcinogenesis by regulating the JAK/STAT signaling pathway. In conclusion, our results demonstrate a promoting role of C/EBP β in human TNBC and reveal a novel underlying mechanism.

Acknowledgements

This work was supported by the National Natural Science Foundation of China (81972653) and the Natural Science Foundation of Jiangsu Province (BK20160224).

Conflict of interest

The authors declare no conflict of interest.

Data Accessibility

The data will be available from the corresponding author upon reasonable request.

Author contributions

SW, DX, XW, HC, CW, and ZS performed experimental work; SW, DX, XW, HC, DZ, and HL analyzed experimental data; DZ performed bioinformatic analyses; DZ and HL designed and supervised the study; DZ and HL wrote the manuscript. All authors read and approved the final manuscript.

References

- 1 Williams SC, Baer M, Dillner AJ and Johnson PF (1995) CRP2 (C/EBP beta) contains a bipartite regulatory domain that controls transcriptional activation, DNA binding and cell specificity. *EMBO J* **14**, 3170–3183.
- 2 Williamson EA, Xu HN, Gombart AF, Verbeek W, Chumakov AM, Friedman AD and Koeffler HP (1998) Identification of transcriptional activation and repression domains in human CCAAT/enhancer-binding protein epsilon. *J Biol Chem* **273**, 14796–14804.
- 3 Mink S, Haenig B and Klempnauer KH (1997) Interaction and functional collaboration of p300 and C/ EBPbeta. *Mol Cell Biol* **17**, 6609–6617.
- 4 Nerlov C and Ziff EB (1995) CCAAT/enhancer binding protein-alpha amino acid motifs with dual TBP and TFIIB binding ability co-operate to activate transcription in both yeast and mammalian cells. *EMBO J* 14, 4318–4328.
- 5 Agre P, Johnson PF and McKnight SL (1989) Cognate DNA binding specificity retained after leucine zipper exchange between GCN4 and C/EBP. *Science* 246, 922–926.
- 6 Vinson CR, Sigler PB and McKnight SL (1989) Scissors-grip model for DNA recognition by a family of leucine zipper proteins. *Science* 246, 911–916.
- 7 Moll JR, Acharya A, Gal J, Mir AA and Vinson C (2002) Magnesium is required for specific DNA binding of the CREB B-ZIP domain. *Nucleic Acids Res* 30, 1240–1246.
- 8 Niehrs C and Calkhoven CF (2020) Emerging role of C/EBPbeta and epigenetic DNA methylation in ageing. *Trends Genet* **36**, 71–80.
- 9 Cao Z, Umek RM and McKnight SL (1991) Regulated expression of three C/EBP isoforms during adipose conversion of 3T3-L1 cells. *Genes Dev* 5, 1538–1552.
- 10 Diehl AM (1998) Roles of CCAAT/enhancer-binding proteins in regulation of liver regenerative growth. J Biol Chem 273, 30843–30846.
- Poli V (1998) The role of C/EBP isoforms in the control of inflammatory and native immunity functions. *J Biol Chem* 273, 29279–29282.
- 12 Zahnow CA (2002) CCAAT/enhancer binding proteins in normal mammary development and breast cancer. *Breast Cancer Res* 4, 113–121.
- 13 Huang E, Huang H, Guan T, Liu C, Qu D, Xu Y, Yang J, Yan L, Xiong Y, Liang T *et al.* (2019) Involvement of C/EBPbeta-related signaling pathway in methamphetamine-induced neuronal autophagy and apoptosis. *Toxicol Lett* **312**, 11–21.
- 14 Cheng P, Chen Y, He TL, Wang C, Guo SW, Hu H, Ni CM, Jin G and Zhang YJ (2019) Menin coordinates C/EBPbeta-mediated TGF-beta signaling for epithelialmesenchymal transition and growth inhibition in pancreatic cancer. *Mol Ther Nucleic Acids* 18, 155–165.

- 15 Gardiner JD, Abegglen LM, Huang X, Carter BE, Schackmann EA, Stucki M, Paxton CN, Lor Randall R, Amatruda JF, Putnam AR *et al.* (2017) C/EBPbeta-1 promotes transformation and chemoresistance in Ewing sarcoma cells. *Oncotarget* 8, 26013–26026.
- 16 Maehara O, Ohnishi S, Asano A, Suda G, Natsuizaka M, Nakagawa K, Kobayashi M, Sakamoto N and Takeda H (2019) Metformin regulates the expression of CD133 through the AMPK-CEBPbeta pathway in hepatocellular carcinoma cell lines. *Neoplasia* 21, 545–556.
- 17 Bundy LM and Sealy L (2003) CCAAT/enhancer binding protein beta (C/EBPbeta)-2 transforms normal mammary epithelial cells and induces epithelial to mesenchymal transition in culture. *Oncogene* 22, 869–883.
- 18 Grimm SL and Rosen JM (2003) The role of C/ EBPbeta in mammary gland development and breast cancer. J Mammary Gland Biol Neoplasia 8, 191–204.
- 19 LaMarca HL, Visbal AP, Creighton CJ, Liu H, Zhang Y, Behbod F and Rosen JM (2010) CCAAT/enhancer binding protein beta regulates stem cell activity and specifies luminal cell fate in the mammary gland. *Stem Cells* 28, 535–544.
- 20 Shao S, Cao H, Wang Z, Zhou D, Wu C, Wang S, Xia D and Zhang D (2020) CHD4/NuRD complex regulates complement gene expression and correlates with CD8 T cell infiltration in human hepatocellular carcinoma. *Clin Epigenet* **12**, 31.
- 21 Wolff AC, Lazar AA, Bondarenko I, Garin AM, Brincat S, Chow L, Sun Y, Neskovic-Konstantinovic Z, Guimaraes RC, Fumoleau P *et al.* (2013) Randomized phase III placebo-controlled trial of letrozole plus oral temsirolimus as first-line endocrine therapy in postmenopausal women with locally advanced or metastatic breast cancer. *J Clin Oncol* **31**, 195–202.
- 22 Shao ZY, Chaudhri S, Guo M, Zhang LZ and Rea D (2016) Neoadjuvant chemotherapy in triple negative breast cancer: an observational study. *Oncol Res* 23, 291–302.
- 23 Banerjee K and Resat H (2016) Constitutive activation of STAT3 in breast cancer cells: a review. *Int J Cancer* 138, 2570–2578.
- 24 Wu S, Fu J, Dong Y, Yi Q, Lu D, Wang W, Qi Y, Yu R and Zhou X (2018) GOLPH3 promotes glioma progression via facilitating JAK2-STAT3 pathway activation. *J Neuro Oncol* 139, 269–279.
- 25 Wu QY, Ma MM, Fu L, Zhu YY, Liu Y, Cao J, Zhou P, Li ZY, Zeng LY, Li F *et al.* (2018) Roles of germline JAK2 activation mutation JAK2 V625F in the pathology of myeloproliferative neoplasms. *Int J Biol Macromol* **116**, 1064–1073.
- 26 Wu QY, Ma MM, Tong YX, Zhu YY, Liu Y, Cao J, Zhou P, Li ZY, Zeng LY, Wang XY *et al.* (2018) Effects of JAK2 V556F mutation on the JAK2's activity, structural stability and the transformation of Ba/F3 cells. *Int J Biol Macromol* **117**, 271–279.

- 27 Wu QY, Ma MM, Zhang S, Cao J, Yan ZL, Chen C, Li ZY, Zeng LY, Wang XY, Li F *et al.* (2019)
 Disruption of R867 and Y613 interaction plays key roles in JAK2 R867Q mutation caused acute leukemia. *Int J Biol Macromol* 136, 209–219.
- 28 Wu QY, Ma MM, Zhang S, Liu Y, Cao J, Yan ZL, Li ZY, Zeng LY, Wang XY, Li F *et al.* (2019) Loss of K607 and E877 interaction is a key reason for JAK2 K607N mutation caused acute myeloid leukemia. *Int J Biol Macromol* **124**, 1123–1131.
- 29 Chen C, Li F, Ma MM, Zhang S, Liu Y, Yan ZL, Chen W, Cao J, Zeng LY, Wang XY *et al.* (2019) Roles of T875N somatic mutation in the activity, structural stability of JAK2 and the transformation of OCI-AML3 cells. *Int J Biol Macromol* **137**, 1030–1040.
- 30 Li W, Tanikawa T, Kryczek I, Xia H, Li G, Wu K, Wei S, Zhao L, Vatan L, Wen B *et al.* (2018) Aerobic glycolysis controls myeloid-derived suppressor cells and tumor immunity via a specific CEBPB isoform in triplenegative breast cancer. *Cell Metab* 28, 87–103.e6