The antibacterial effect of topical ozone on the treatment of MRSA skin infection

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Abstract. Skin can be infected by many types of microorganisms, most commonly by gram-positive strains of Staphylococcus and Streptococcus spp. Treatment of Staphylococcus aureus (S. aureus) infections, particularly that of methicillin resistant Staphylococcus aureus (MRSA), is a challenge in clinical practice. Ozone therapy has proven to be one of the strongest antiseptics against the majority of microorganisms involved in skin infections. The purpose of the present study was to evaluate the microbicidal effects of topical ozone therapy on S. aureus and MRSA, and determine the clinical efficacy of ozone therapy on patients with MRSA skin infection. Microbicidal effects of ozonated oil and ozonated water were determined by plating and Kirby Bauer methods. Clinical efficacy and safety of topical ozone were evaluated in two cases with skin MRSA infection. The killing rates of ozonated oil for S. aureus and MRSA were greater when compared with the control oil group. Almost 100% of S. aureus were eliminated by ozonated oil following 5 min. Almost 100% MRSA were eliminated by ozonated oil following 15 min. In addition, 100% S. aureus and 100% MRSA were eliminated by ozonated water in 1 min. The inhibition zone diameters of ozonated oil for S. aureus and MRSA were 17 and 13 mm, respectively, which were significantly larger than the control group. Both cases of skin MRSA infection were completely healed with ozone therapy.

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Abbreviations: S. aureus, Staphylococcus aureus; MRSA, methicillin resistant Staphylococcus aureus; DMSO, Dimethylsulfoxide

Key words: ozone therapy, Staphylococcus aureus, methicillin resistant Staphylococcus aureus, skin infection

In conclusion, ozone therapy is a potential treatment for *S. aureus* and MRSA skin infection as it has great efficacy, few side effects and low-costs.

Introduction

Skin and soft tissue bacterial infections occur frequently in the general population. They are characterized by erythema, edema, and/or inflammation (1). and usually begin with an inflammatory process in epidermis, dermis, or subcutaneous tissues, they can spread to other parts of the body, leading to more serious symptoms (2).

Antibiotic therapy is the option of choice for the treatment. However, most of antibiotics could lose their potency over time due to the antimicrobial resistance (3). Microbial resistance to *Staphylococcus aureus* (*S. aureus*), especially methicillin resistant *Staphylococcus aureus* (MRSA) and vancomycin resistant-MRSA, is a grand challenge in clinical practices globally (4,5).

Ozone is naturally occurring gaseous molecule of triatomic allotrope of oxygen, formed recombination of oxygen atoms and represented as O₃ (6,7). The original application of Ozone was to sterilize microorganisms in drinking water (8). Now ozone has been used for treatment of open wounds, Herpes Zoster and Herpes Simplex (9,10), because of its anti-microorganism effectiveness (11). Additionally, ozone has other advantages like such as improving wound healing, enhancing immune, no side effect, no-toxic, environmental friendly and high efficacy (12). A few studies have shown that ozone therapy is efficient in killing many kinds of microorganisms, such as *S. aureus*, *Streptococci* spp, *Escherichia coli*, *Enterococcus faecalis*, and *P. aeruginosa* (13,14).

Studies also have shown that ozone therapy can disinfect against *S. aureus* and MRSA strain *in vitro* (14,15). *In vivo* studies have suggested that ozone therapy is safe and exhibits antibacterial effects for the treatment of peritonitis (16,17). Ozone has shown its efficacy on healing MRSA skin infections when combined with other drugs (18,19). However, the effect of ozone therapy alone in the treatment of MRSA skin infections is not to be determined.

This study aims to evaluate the microbicidal effects of topical ozone therapy on *S. aureus* and MRSA and determine its clinical efficacy on MRSA skin infections.

Materials and methods

Ethics approval and consent to participate. This study was approved by the Ethics Committee of The Third Xiangya Hospital, Central South University (Changsha, China) and was carried out in accordance with the approved guidelines. All patients provided written informed consent.

Material and equipment. The bacterial strains used in this study were *S. aureus* (ATCC 6538) and MRSA (ATCC 43300). Bacterial culture medium was purchased from Zhengzhou An Tu Biological Engineering Co., Ltd. (Zhengzhou, China). Dimethylsulfoxide (DMSO) was from Sigma-Aldrich (Merck KGaA, Darmstadt, Germany. Ozonated oil at the peroxide value of 2,000 to 2,200 mmol-equiv/Kg [provided by Hunan Health Care Technology Co., Ltd. (Hunan, China)] was obtained from the chemical reaction between ozone and camellia oil because of its high composition of unsaturated fatty acids. Ozonated water was created by Ozone Water Generating Instrument (from Hunan Health Care Technology Co., Ltd.) at the Dermatology Department of the Third Xiangya Hospital.

Plating method. The bacterial strains were diluted with PBS to get the concentration reaching at 10⁸ CFU/ml.

For ozonated oil: The test oil suspension was constituted by 400 μ l ozonized oil mixes, 50 μ l DMSO and 50 μ l microorganism suspension. Control oil suspension was constituted by 400 μ l basal oil mixes 50 μ l DMSO and 50 μ l microorganism suspension. Both groups were incubated in 37°C incubator for 1, 5, 10, 15, 20, and 30 min respectively. Then the suspension was plated into Petri dishes and grew at 37°C for 24 h. Then the number of colonies in each agar plate was calculated. The control suspension constituted by 400 μ l normal saline mixes 50 μ l DMSO and 50 μ l microorganism suspension was used as control to calculate the killing rate. Killing Rate (%)=(control colony number-oil colony number).

For ozonated water: 1.0 ml of bacteria liquid was mixed with 4.0 ml of ozonated water or PBS. 0.5 ml mixture was added into 4.5 ml neutralizer (Phosphate buffer solution including sodium thiosulfate) after oscillation for one minute. After that the sample was planted into Petri dishes and cultured in an incubator chamber at 37°C. The number of colonies in each agar plate was calculated after growing for 24 h. Killing Rate (%)=(control colony number-tested colony number)/control colony number).

Kirby bauer method. The bacteria samples prepared above were dropped on sterile cotton swabs. The surface was lightly and uniformly inoculated by cotton swab on agar plate. Then the scrips impregnated with ozonated oil or control oil were pasted into the agar plate, followed by incubation at 37°C for 16-18 h. The inhibitory ring test was performed. If the inhibition zone diameter was bigger than 7 mm, it was considered effective; otherwise, it was not.

Patients and ozone treatment. The Ethnic Committee of Third Xiangya Hospital approved the study, and the informed consents of all the participants were obtained. Two patients with skin MRSA infection were recruited in this study. In addition to skin infections, no other diseases were present in

the two patients. The skin lesions were washed or debrided by ozonated water in our therapy room once a day, followed by application of ozonated oil twice per day at home. Antibiotics and other drugs were not administrated during the ozone therapy.

Bacteria culture, drug sensitive test and PCR test of lesions. The lesions of patients were inoculated in the blood plate, then cultured in an incubator chamber with 5% CO₂ concentration at 35°C for 12 h. If there was colony formation of microorganism, the microorganism colony was stained by Gram staining. After confirming the gram-positive bacterial by the microscope examination, three bacterial colonies were added into physiological saline and were prepared with 0.5 turbidity ratio in VITEK2 system (VITEK2 gram-positive identification card) by electronic turbidimeter (DensiCHEK Plus, BioMérieux, Durham, NC, USA). Then bacterial species and drug sensitivity results of the samples were detected by Automatic Microorganisms Identification System (VITEK2-compact). The drug sensitivity results were further confirmed by VITEK2 AST-GP67 Test kit according to the manufacturer's protocol. Following the preliminary result of MRSA infection detected by Automatic Microorganisms Identification System, the microorganism colony was further confirmed as MRSA infection by the MRSA Quantitative Standards kit (no. Z-DD-0096-B; Liferiver Bio-Tech, San Diego, CA, USA) according to the manufacturer's protocol in CFX96™ Red-Time System.

Statistical analysis. The data were analyzed in SPSS 19.0 software (SPSS, Inc., Chicago, IL, USA). t-test was used to assess statistical differences between two groups. P<0.05 was considered to indicate a statistically significant difference.

Results

Killing rate of ozone on S. aureus and MRSA. The killing rate of ozonated oil on S. aureus was much higher than the control. Almost 100% S. aureus were killed in 5 min. For MRSA, the killing rate of ozonated oil was also much higher than the control oil. Almost 100% MRSA were killed within 15 min (Fig. 1). The killing rates over time between ozonated oil and control for S. aureus and MRSA were presented in Tables I and II. The ozonated water (1 mg/l) can sterilize 100% S. aureus and 100% MRSA in one minute (Fig. 2).

Bacterial inhibitory: Inhibition zone diameter. The inhibition zone diameters of ozonated oil for *S. aureus* and MRSA were 17 and 13 mm respectively, which were significantly much larger than the control (Fig. 3).

Practicing treatment

The first case. A 21-year-old male patient presented to the Dermatology Department of our hospital complaining of a painful abscess on his left calf muscle for approximately 20 days. Denied any systemic diseases or health issues. Skin examination revealed induration, bleeding and surrounding erythema in the left upper Achilles tendon (Fig. 4A). The overlying skin had become thin and felt fluctuant with spontaneous pus secretion. Previous treatments included systemic antibiotics,

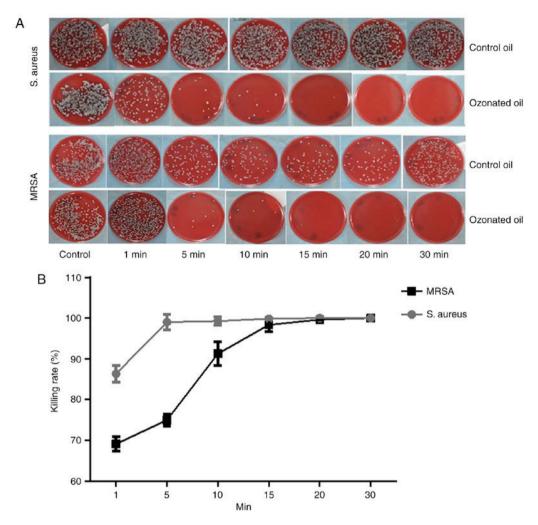


Figure 1. Killing rate of ozonated oil for *S. aureus* and MRSA. (A and B) Almost 100% *S. aureus* were killed in 5 min by ozonated oil. Almost 100% MRSA were killed within 15 min by ozonated oil. *S. aureus*, *Staphylococcus aureus*; MRSA, methicillin resistant *Staphylococcus aureus*.

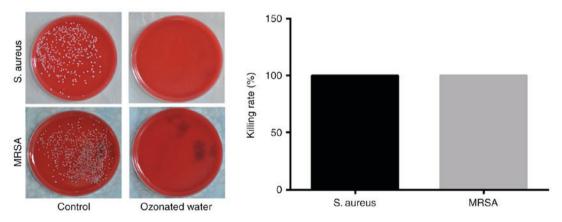


Figure 2. Killing rate of ozonated water for *S. aureus* and MRSA. The ozonated water can sterilize 100% *S. aureus* and 100% MRSA in one minute. *S. aureus*, *Staphylococcus aureus*; MRSA, methicillin resistant *Staphylococcus aureus*.

cleaning and dressing changes for more than 20 days without any visible effect. Tests for diabetes and immune-suppression syndrome were negative. Bacteria culture, drug sensitive test and PCR test confirmed skin MRSA infection (Fig. 5A and C). The strain was resistant to clindamycin, erythromycin, cefoxitin, oxacillin, penicillin-G, piperacillin, and azithromycin (Fig. 5A). After diagnosis of MRSA infection, the patient voluntarily

was put on topical application of ozone therapy. Ozonated water was used to wash the lesion for 10 min immediately, followed by soak the lesion for 20 min after debridement if necessary in our therapy room every day. The application of topical ozonated oil twice per day was carried out at home by the patient. Four days after treatment, the lesion was cleaned to remove necrotic tissue and pus secretion (Fig. 4B). Affected

Table I. Killing rate of ozonated oil on *S. aureus*.

	Killi		
Time (min)	Control oil (%)	Ozonated oil (%)	P-value
1	31.80±1.05	86.35±2.10	< 0.0001
5	40.81±8.27	99.01±1.90	< 0.0001
10	35.91±1.74	99.25±1.00	< 0.0001
15	36.04±3.37	99.84±0.24	< 0.0001
20	37.26±3.82	100±0.00	< 0.0001
30	37.16±1.44	100±0.00	< 0.0001

S. aureus, staphylococcus aureus.

Table II. Killing rate of ozonated oil on MRSA.

	Killi			
Time (min)	Control oil (%)	Ozonated oil (%)	P-value	
1	44.70±0.97	69.09±1.73	0.0037	
5	47.31±1.42	74.96±1.44	0.0004	
10	42.99±5.69	91.25±2.91	0.0001	
15	42.99±5.28	98.37±1.71	< 0.0001	
20	41.65±6.7	99.65±0.09	< 0.0001	
30	47.37±7.45	100±0.00	< 0.0001	

MRSA, methicillin resistant staphylococcus aureus.

areas were reduced by more than 70% during the first month (Fig. 4C) and was almost healed by the end of second month (Fig. 4D). Bacteria culture test revealed that tissues from the lesion did not develop bacteria one month after the topical ozone treatment.

The second case. An 8-year-old male patient presented to our department complaining of a big blister on the dorsal aspect of the left hand. Denied any systemic diseases and any treatment at the moment. On skin examination, a 2x2 cm ulcerated lesion with yellow crusts was seen with surrounding erythema and fluid exudation on the dorsal aspect of the left hand. Laboratory tests confirmed MRSA skin infection, resistant to clindamycin, cefotetan, erythromycin, oxacillin, penicillin-G, and trimethoprim/sulfa-methoxazole (Fig. 5B and D). A complete healing from the lesion was achieved after 12 days of treatment with a combination of ozonated water and ozonated oil using the same remedy as we described in the first case (Fig. 6). Bacteria culture test revealed that scales from the lesion did not develop bacteria 12 days after the topical ozone treatment.

Discussion

In this study, we demonstrated that ozonated oil and ozonated water have strong *in vitro* antibacterial effects against

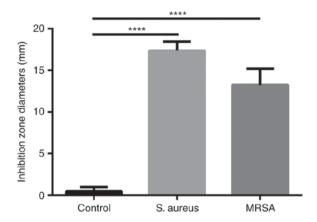


Figure 3. The inhibition zone diameters of ozoned oil for *S. aureus* and MRSA. The inhibition zone diameters of ozonated oil for *S. aureus* and MRSA were 17 and 13 mm, respectively. ****P<0.0001. *S. aureus, Staphylococcus aureus*; MRSA, methicillin resistant *Staphylococcus aureus*.

S. aureus and MRSA. This is the first study to prove that the topical application of ozone alone can be a powerful treatment option for MRSA skin infections.

Data in this study have shown that ozonated oil can sterilize up to 98% of S. aureus in 5 min and up to 98% of MRSA in 15 min while ozonated water (1 mg/l) can sterilize 100% of S. aureus and 100% of MRSA in one minute. This results indicate that ozone therapy has very powerful anti-microbial properties against gram positive microorganisms, which was confirmed by the bacterial inhibitory experiment of ozonated oil. Our results are consistent with reports regarding bacterial elimination in S. aureus or MRSA by ozone therapy (20-23). Notably ozone can sterilize both gram positive bacteria and gram negative bacteria (24). Ozone is an unstable molecule that rapidly decays to O₂ and releases a single oxygen atom. The single oxygen atom reacts with the cell membrane of the bacteria, attacks the cellular components, interrupts the normal cell activity and then destroys bacteria (23,25).

Ozone therapy has been used for infectious diseases such as conjunctivitis and keratitis (26), peritonitis (27), and surgical sepsis (28). In our clinical practice, topical application of ozone therapy is very effective for healing MRSA induced skin ulceration. The two cases got remarkable therapeutic effects after ozone treatment alone. Besides the high efficiency for sterilization, ozone therapy exhibits a potential effect in wound healing. The possible mechanisms include: i) Increasing oxygen levels, glucose and ATP transporter molecules in ischemic tissues; ii) increasing the activity of bone marrow stem cells, so as to promote angiogenesis and tissue regeneration; iii) the upregulation of the expression of antioxidant enzymes in blood; iv) Promoting the neuronal medium rise (29,30); and v) Inducing growth factors (31). Ozone therapy was also reported to reduce pain and swelling (32). Patients in this study also presented significant decrease in pain and swelling.

Diabetic foot ulcers are a challenging clinical problem, characterized by neuropathy, peripheral arterial diseases, foot deformities, and infection (33,34). *S. aureus* was the most common pathogen identified in Diabetic foot ulcers, representing 46% of culture-positive patients. And 15% were

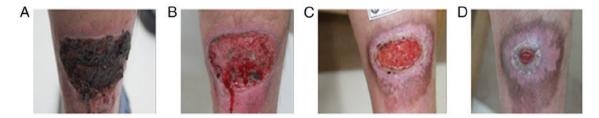


Figure 4. Topical application of ozone therapy on left calf muscle MRSA infection. (A) The lesion before ozone therapy. (B) The lesion after removing necrotic tissue and pus secretion. (C) The lesion after topical application of ozone therapy for one month. (D) The lesion after topical application of ozone therapy for two months. MRSA, methicillin resistant *Staphylococcus aureus*.

Α	Antibiotic name	KB (mm)	Drug sensitivity	В	Antibiotic name	KB (mm)	Drug sensitivity
	Clindamycin	, ,	Drug resistance		Clindamycin		Drug resistance
	Ciprofloxacin	1	Drug sensitivity	1	Ciprofloxacin	1	Drug sensitivity
	Erythromycin	>=8	Drug resistance	-	Cefotetan		Drug resistance
	Nitrofurantoin	>=o <=16	Drug resistance Drug sensitivity	-	Erythromycin	>=8	Drug resistance
		<=10	,	-	Nitrofurantoin	32	Drug sensitivity
	Cefoxitin		Drug resistance		Gentamicin	<=0.5	Drug sensitivity
	Gentamicin	<=0.5	Drug sensitivity		Levofloxacin	2	Drug sensitivity
	Levofloxacin	0.5	Drug sensitivity		Oxacillin	>=4	Drug resistance
	Oxacillin	>=4	Drug resistance		Penicillin-G	>=0.5	Drug resistance
	Penicillin-G	>=0.5	Drug resistance		Rifampin	<=0.5	Drug sensitivity
	Piperacillin		Drug resistance	1	Trimethoprim/sulfa-methoxazole	>=320	Drug resistance
	Rifampin	<=0.5	Drug sensitivity	1	Tetracycline	<=1	Drug sensitivity
	Trimethoprim/sulfa-methoxazole	<=10	Drug sensitivity	1	Vancomycin	2	Drug sensitivity
	Tetracycline	<=1	Drug sensitivity	1	Quinupristin-Dalfoprisdn	<=0.25	Drug sensitivity
	Vancomycin	1	Drug sensitivity		Linezolid	2	Drug sensitivity
	Quinupristin-Dalfoprisdn	0.5	Drug sensitivity		Moxifloxacin	<=0.25	Drug sensitivity
	Azithromycin		Drug resistance				
	Linezolid	2	Drug sensitivity	1			
	Moxifloxacin	<=0.25	Drug sensitivity	1			
	Tigecycline	<=0.12	Drug sensitivity	1			
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Figure 5. Drug sensitive test and PCR test confirmed skin MRSA infection. (A) The results of drug sensitive test of the first case. (B) The results of drug sensitive test of the second case. (C) The melting curve of PCR test confirmed the MRSA infection of the first case. (D) The melting curve of PCR test confirmed the MRSA infection of the second case. PCR, polymerase chain reaction; MRSA, methicillin resistant *Staphylococcus aureus*.

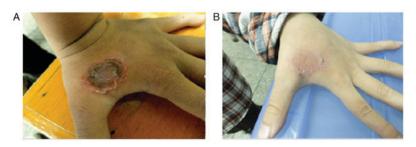


Figure 6. Topical application of ozone therapy on the left-hand MRSA infection. (A) The lesion before ozone therapy. (B) The lesion after topical application of ozone therapy for 12 days. MRSA, methicillin resistant *Staphylococcus aureus*.

classified as MRSA (35). Ozone application was not only to kill *S. aureus* or MRSA, but also reported to significantly reduce the lesion area in patient with diabetic foot (31,36,37). Ozone also can improve glycemic control by controlling hyperglycemia and insulin sensitivity and preventing oxidative stress associated with diabetes mellitus and its complications (38). Because of the low-cost, ozone therapy can also reduce the medical bills for patients with diabetes mellitus and its complications.

Ozonated water can keep the bactericidal effect for approximately 30 min, while ozonated oil can maintain its sterilization ability persistently (39-42). This is why ozonated oil was applied to our patients after washing with ozonated water to increase the effective time.

Although ozone therapy can kill microbes, improve wound healing, reduce pain and swelling at minimal cost with almost no side effect, it is contraindicated in several diseases such as Blood Coagulation Failure, Bleeding Organs, Thrombocytopenia, Ozone Alergia, Hemorrhagic or Apoplectic Stroke, Ozone Intolerance (43).

In summary, ozone therapy is potential treatment for *S. aureus* and MRSA skin infections with great efficacy, low side effects, and low-cost.

Acknowledgements

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References

- Dréno B, Araviiskaia E, Berardesca E, Gontijo G, Sanchez Viera M, Xiang LF, Martin R and Bieber T: Microbiome in healthy skin, update for dermatologists. J Eur Acad Dermatol Venereol 30: 2038-2047, 2016.
- Diaz JH and Lopez FA: Skin, soft tissue and systemic bacterial infections following aquatic injuries and exposures. Am J Med Sci 349: 269-275, 2015.
- 3. Sully EK and Geller BL: Antisense antimicrobial therapeutics. Curr Opin Microbiol 33: 47-55, 2016.
- 4. Yang X, Zhang J, Yu S, Wu Q, Guo W, Huang J and Cai S: Prevalence of *Staphylococcus aureus* and methicillin-resistant *Staphylococcus aureus* in retail ready-to-eat foods in China. Front Microbiol 7: 816, 2016.
- Miller WR, Bayer AS and Arias CA: Mechanism of action and resistance to daptomycin in *Staphylococcus aureus* and enterococci. Cold Spring Harb Perspect Med 6: pii: a026997, 2016.
- 6. Greene AK, Few BK and Serafini JC: A comparison of ozonation and chlorination for the disinfection of stainless steel surfaces. J Dairy Sci 76: 3617-3620, 1993.
- Fiessinger F, Richard Y, Montiel A and Musquere P: Advantages and disadvantages of chemical oxidation and disinfection by ozone and chlorine dioxide. Sci Total Environ 18: 245-261, 1981.
- 8. Stalder K and Klosterkötter W: Studies on the reappearing of a bacterial flora in drinking water after ozonization (author's transl). Zentralbl Bakteriol Orig B 161: 474-481, 1976 (In German).
- Liu J, Zhang P, Tian J, Li L, Li J, Tian JH and Yang K: Ozone therapy for treating foot ulcers in people with diabetes. Cochrane Database Syst Rev: Cd008474, 2015.

- Bassi P, Sbrascini S, Mattassi R, D'Angelo F and Franchina A: Ozone in the treatment of herpes zoster. Riv Neurobiol 28: 328-333, 1982 (In Italian).
- 11. Moureu S, Violleau F, Ali Haimoud-Lekhal D and Calmon A: Ozonation of sunflower oils: Impact of experimental conditions on the composition and the antibacterial activity of ozonized oils. Chem Phys Lipids 186: 79-85, 2015.
- 12. Zhong J, Allen K, Rao X, Ying Z, Braunstein Z, Kankanala SR, Xia C, Wang X, Bramble LA, Wagner JG, et al: Repeated ozone exposure exacerbates insulin resistance and activates innate immune response in genetically susceptible mice. Inhal Toxicol 28: 383-392, 2016.
- Farac RV, Pizzolitto AC, Tanomaru JM, Morgental RD, Lima RK and Bonetti-Filho I: Ex-vivo effect of intracanal medications based on ozone and calcium hydroxide in root canals contaminated with *Enterococcus faecalis*. Braz Dent J 24: 103-106, 2013.
- Heß S and Gallert C: Šensitivity of antibiotic resistant and antibiotic susceptible Escherichia coli, Enterococcus and Staphylococcus strains against ozone. J Water Health 13: 1020-1028, 2015.
- Sharma M and Hudson JB: Ozone gas is an effective and practical antibacterial agent. Am J Infect Control 36: 559-563, 2008.
- 16. Gadzhiev ND, Nasirov Mla, Sushkov SV and Klimova EM: Effect of combined and local cytokine- and ozone therapy on the indices of lipid peroxidation, endogenous intoxication and ferroproteins in diffuse peritonitis. Vestn Khir Im I I Grek 173: 38-41, 2014 (In Russian).
- 17. Kolesova OE, Vasil'ev IT, Volkhovskaia NB, Mumladze RB, Tkachenko SB and Savina GD: Correction of the antioxidative system during ozone therapy in peritonitis. Vestn Ross Akad Med Nauk: 34-39, 2010 (In Russian).
- 18. Tamai M, Matsushita S, Miyanohara H, Imuta N, Ikeda R, Kawai K, Nishi J, Sakamoto A, Shigihara T and Kanekura T: Antimicrobial effect of an ultrasonic levitation washer disinfector with silver electrolysis and ozone oxidation on methicillin-resistant Staphylococcus aureus. J Dermatol 40: 1020-1026, 2013.
- Solovăstru LG, Stîncanu A, De Ascentii A, Capparé G, Mattana P and Vâţă D: Randomized, controlled study of innovative spray formulation containing ozonated oil and α-bisabolol in the topical treatment of chronic venous leg ulcers. Adv Skin Wound Care 28: 406-409, 2015.
- 20. Gulmen S, Kurtoglu T, Meteoglu I, Kaya S and Okutan H: Ozone therapy as an adjunct to vancomycin enhances bacterial elimination in methicillin resistant *Staphylococcus aureus* mediastinitis. J Surg Res 185: 64-69, 2013.
- 21. Al-Saadi H, Potapova I, Rochford ET, Moriarty TF and Messmer P: Ozonated saline shows activity against planktonic and biofilm growing *Staphylococcus aureus* in vitro: A potential irrigant for infected wounds. Int Wound J 13: 936-942, 2016
- 22. Wilczyńska-Borawska M, Leszczyńska K, Nowosielski C and Stokowska W: Ozone in dentistry: Microbiological effects of gas action depending on the method and the time of application using the ozonytron device. Experimental study. Ann Acad Med Stetin 57: 99-103, 2011.
- 23. Yamayoshi T and Tatsumi N: Microbicidal effects of ozone solution on methicillin-resistant *Staphylococcus aureus*. Drugs Exp Clin Res 19: 59-64, 1993.
- 24. Almaz ME and Sönmez IŞ: Ozone therapy in the management and prevention of caries. J Formos Med Assoc 114: 3-11, 2015.
- Komanapalli IR and Lau BH: Inactivation of bacteriophage lambda, *Escherichia coli*, and *Candida albicans* by ozone. Appl Microbiol Biotechnol 49: 766-769, 1998.
- 26. Gierek-Lapińska A, Antoszewski Z, Myga B and Skowron J: Preliminary report on using therapeutic ozone in infectious conjunctivitis and keratitis and in corneal degeneration. Klin Oczna 94: 137-138, 1992 (In Polish).
- 27. Erginel B, Erginel T, Aksoy B and Dokucu AI: Effect of Ozone Therapy (OT) on healing of colonic anastomosis in a rat model of peritonitis. Balkan Med J 31: 249-253, 2014.
- 28. Parkhisenko IuA and Glukhov AA: Use of ozone therapy and hydro-pressure technologies in complex intensive therapy of surgical sepsis. Khirurgiia (Mosk): 55-58, 2001 (In Russian).
- Verrazzo G, Coppola L, Luongo C, Sammartino A, Giunta R, Grassia A, Ragone R and Tirelli A: Hyperbaric oxygen, oxygen-ozone therapy, and rheologic parameters of blood in patients with peripheral occlusive arterial disease. Undersea Hyperb Med 22: 17-22, 1995.
- 30. Bocci VA: Scientific and medical aspects of ozone therapy. State of the art. Arch Med Res 37: 425-435, 2006.

- 31. Zhang J, Guan M, Xie C, Luo X, Zhang Q and Xue Y: Increased growth factors play a role in wound healing promoted by noninvasive oxygen-ozone therapy in diabetic patients with foot ulcers. Oxid Med Cell Longev 2014: 273475, 2014.
- 32. Kazancioglu HO, Kurklu E and Ezirganli S: Effects of ozone therapy on pain, swelling, and trismus following third molar surgery. Int J Oral Maxillofac Surg 43: 644-648, 2014.
- 33. Noor S, Khan RU and Ahmad J: Understanding diabetic foot infection and its management. Diabetes Metab Syndr 11: 149-156,
- 34. Kateel R, Adhikari P, Augustine AJ and Ullal S: Topical honey for the treatment of diabetic foot ulcer: A systematic review. Complement Ther Clin Pract 24: 130-133, 2016.
- 35. Reveles KR, Duhon BM, Moore RJ, Hand EO and Howell CK: Epidemiology of methicillin-resistant Staphylococcus aureus diabetic foot infections in a large academic hospital: Implications for antimicrobial stewardship. PLoS One 11: e0161658, 2016.
- 36. Gazin IK: Pathophysiological aspects of endotoxicosis complicated with purulent infection of the foot and correction of endotoxicosis with conventional treatment and with application of ozonized physiological solution in patients suffering from diabetes mellitus. Patol Fiziol Eksp Ter: 23-25, 2008 (In Russian).
- 37. Martínez-Sánchez G, Al-Dalain SM, Menéndez S, Re L, Giuliani A, Candelario-Jalil E, Alvarez H, Fernández-Montequín JI and León OS: Therapeutic efficacy of ozone in patients with diabetic foot. Eur J Pharmacol 523: 151-161, 2005.

- 38. Al-Dalain SM, Martínez G, Candelario-Jalil E, Menéndez S, Re L, Giuliani A and León OS: Ozone treatment reduces markers of oxidative and endothelial damage in an experimental diabetes model in rats. Pharmacol Res 44: 391-396, 2001.
- 39. Bialoszewski D, Pietruczuk-Padzik A, Kalicinska A, Bocian E, Czajkowska M, Bukowska B and Tyski S: Activity of ozonated water and ozone against Staphylococcus aureus and Pseudomonas aeruginosa biofilms. Med Sci Monit 17: BR339-BR344, 2011.
- 40. Burke FJ: Ozone and caries: A review of the literature. Dent Update 39: 271-272, 275-278, 2012.
- 41. Valacchi G, Zanardi I, Lim Y, Belmonte G, Miracco C, Sticozzi C, Bocci V and Travagli V: Ozonated oils as functional dermatological matrices: Effects on the wound healing process using SKH1 mice. Int J Pharm 458: 65-73, 2013.
- 42. Pai ŠA, Gagangras SA, Kulkarni SS and Majumdar AS: Potential of ozonated sesame oil to augment wound healing in rats. Indian
- J Pharm Sci 76: 87-92, 2014. 43. Zhang YB, Xiang YP, Huang JH, Gao L, Chen M, Kathy W, Li M, Chen J, Yang S and Lu J: Combined ozone hydrotherapy for atopic dermatitis: Evaluation of efficacy and detection of interleukin-4 and nerve growth factor levels in peripheral blood from patients before and after treatment. Chin J Dermatol 49: 736-738, 2016 (In Chinese).



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