

# miR-203a-3p.1 targets IL-24 to modulate hepatocellular carcinoma cell growth and metastasis

Wei Huo, Min Du, Xinyan Pan, Xiaomin Zhu, Yu Gao and Zhimin Li

Department of Medical Oncology, Dalian Municipal Central Hospital, China

#### Keywords

hepatocellular carcinoma; IL-24; metastasis; miR-203a-3p.1

#### Correspondence

Z. Li, Department of Medical Oncology, Dalian Municipal Central Hospital, Dalian, Liaoning 116033, China Tel: +86 0411 84412001 E-mail: yuzhimin17@163.com

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Hepatocellular carcinoma (HCC) is one of the most common causes of cancer-related death. Cytokines, including interleukin 24 (IL-24), play an important role in HCC. IL-24 inhibits HCC metastasis but the molecular mechanism by which this occurs is still unknown. MicroRNAs (miRNAs) are regulators of cancers including hepatocellular carcinoma (HCC). However, the role that miRNAs play in the regulation of IL-24 in HCC is unclear. The aim of this study was to investigate the effects of regulation of IL-24 by miR-203a-3p.1 on liver cancer cell proliferation and metastasis. IL-24 mRNA and miR-203a-3p.1 were detected by real-time RT-PCR, and IL-24 protein in the cell growth medium was measured by ELISA. A luciferase assay was used to verify that the IL-24 gene was the target of miR-203a-3p.1. Cell survival ability was detected by the MTT assay and colony formation. Cell metastasis was assayed by the Transwell system. The results showed that IL-24 could be down-regulated by miR-203a-3p.1 in HCC cells and that miR-203a-3p.1 acted as an onco-miRNA by targeting IL-24. Inhibition of miR-203a-3p.1 in cells could lead to the reversal of HCC cell proliferation and metastasis. The study highlights a novel molecular interaction between miR-203a-3p.1 and IL-24, which indicates that IL-24 and miR-203a-3p.1 may constitute potential therapeutic targets for HCC.

Hepatocellular carcinoma (HCC) is one of the most common causes of cancer-related death [1]. Cytokines play an important role in HCC including interleukin 24 (IL-24). IL-24 is a member of the IL-10 cytokine family [2]. It is released by both immune and nonimmune cells including peripheral blood mononuclear cells, monocytes, T and B cells, melanocytes and dermal keratinocytes [3]. IL-24 expression is lost in most cancer cells of human origin [2–4]. Studies have shown that loss of IL-24 expression is correlated with disease progression in prostate cancer [5], breast cancer [6], colon cancer [7], neuroblastoma [8], acute leukemia [9], oral squamous cell carcinoma [10,11], lung cancer [12] and other cancers [13–17]. Exogenous IL-24 expression has anti-tumor, anti-angiogenic and anti-metastatic properties and suppresses various signaling pathways, without harming normal cells [2–5].

IL-24 plays important roles in tumor suppression. miRNAs are non-coding RNAs of about 20 nucleotides in length, which are play important roles in gene regulation [18,19]. However, the regulation of IL-24 by miRNAs in HCC is unclear and the purpose of this study was to explore this question. Based on the predicted results from TARGETSCAN online, we found that miR-203a-3p.1 was one of the miRNAs that regulated IL-24 expression in HCC cells. In the present study, we show that IL-24 mRNA and protein were downregulated by miR-203a-3p.1 in HCC cells and miR-

#### Abbreviations

HCC, hepatocellular carcinoma; IL-24, interleukin 24; miRNA, microRNA; MTT, 3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-2-*H*-tetrazolium bromide; qRT-PCR, quantitative real-time PCR.

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203a-3p.1 acted as an onco-miRNA by targeting IL-24. Inhibition of miR-203a-3p.1 in cells could lead to inhibiting cell growth and metastasis. These data highlight a novel molecular interaction between miR-203a-3p.1 and IL-24, which indicates that IL-24 and miR-203a-3p.1 may constitute potential therapeutic targets for HCC.

### **Materials and methods**

#### **Cell culture**

HCC cell lines (SMMC-7721, HepG2, Huh7, Hep3B, 97L and 97H) and a normal liver cell line (LO2) were maintained in DMEM (Life Technologies, Carlsbad, CA, USA), supplemented with 10% fetal bovine serum (HyClone, Logan, UT, USA) and 1% penicillin/streptomycin (Life Technologies) at 37 °C, 5% CO<sub>2</sub>.

#### **HCC** tissues

Informed consent was provided by the patients, and the research procedures were approved by the Ethics Committee of Dalian Municipal Central Hospital. HCC tissues and their adjacent normal tissues as the controls were obtained from The First Affiliated Hospital (Luoyang, China).

#### **Quantitative real-time PCR**

Total RNA, including mature miRNA, was isolated from the cell lines and tissues using Trizol (Life Technologies), and the RNA was quantified by spectrophotometry (Nanodro<sup>®</sup> ND-1000) according to the manufacturer's instructions. Reverse transcription of the miRNA was performed using a miScript Reverse Transcription kit (Qiagen, Germantown, MD, USA) starting from 1 µg of total RNA. TaqMan miRNA assays kits (Life Technologies) were used to examine the specific miRNA expression by quantitative real-time PCR (qRT-PCR) according to the manufacturer's protocol. The qRT-PCR results, which were recorded as threshold cycle numbers ( $C_t$ ), were normalized against an internal control (U6 RNA), and the comparative threshold cycle method ( $\Delta\Delta C_t$ ) was used to determine the levels of miRNA expression.

#### miRNA mimics and vector construction

miR-203a-3p.1 (forward 5'-TGCTGCTAGTGGTCCTAA ACATTTCACGTTTTGGCCACTGACTGACGTGAAAT GTAGGACCACTAG-3'; reverse 5'-CCTGCTAGTGGT CCTACATTTCACGTCAGTCAGTGGCCAAAACGTG AAATGTTTAGGACCACTAGC-3') was synthesized. Lentiviral particles were obtained using the BLOCK-iT<sup>™</sup> PoIII miR RNAi Expression Vector kit with EmGFP (Thermo Fisher Scientific, Waltham, MA, USA). In brief, a small hairpin sequence corresponding to miR-203a-3p.1 was cloned into the pLenti6/V5-DEST vector, which was then packaged into replication-incompetent lentiviral particles in HEK293FT cells by co-transfecting pLenti6/V5 plasmid with the ViraPower Packaging Mix. Viral particles were collected 48 h post-transfection in the supernatant for transfection. Several clones were generated by limiting dilutions under blasticidin selection at  $10 \,\mu g \cdot m L^{-1}$  (Thermo Fisher Scientific). IL-24 lentivirus vector was the same for miR-203a-3p.1.

#### Luciferase assay

For measuring the effect of miR-203a-3p.1 on the 3'-UTR of IL-24, we generated a luciferase expression construct containing part of IL-24 3'-UTR. We amplified the wildtype fragment IL-24 mRNA that contained potential miR-203a-3p.1 binding sites. The PCR fragment was inserted into the pGL4 Basic Vector (Promega, Madison, WI, USA) using the KpnI/XhoI endonuclease restriction sites. Mutation of the IL-24 3'-UTR (Mut) was performed using a mutation kit (Stratagene, La Jolla, CA, USA). For luciferase activity assays, cells were co-transfected with 100 ng of wild-type or MUT1 and MUT2 IL-24 3'-UTR and 100 nm miR-203a-3p.1 or control mimics using Lipofectamine 2000. Luciferase activity was assayed using a luciferase assay kit from Promega referring to the manufacturer's protocol; after 48 h transfection, luciferase activity was measured and normalized to Renilla luciferase activity.

#### MTT assay

The proliferation of HCC cells was examined by 3-(4,5dimethyl-2-thiazolyl)-2,5-diphenyl-2-*H*-tetrazolium bromide (MTT) assay. A quantity of  $2 \times 10^3$  cells was seeded onto a 96-well plate, and at different time points 10 µL MTT solution (5 mg·mL<sup>-1</sup>, Sigma-Aldrich, St. Louis, MO, USA) was added to each well and the cells were cultured for 4 h. After the incubation, the supernatant was discarded and 150 µL dimethyl sulfoxide was added to each well until the crystals dissolved completely. The absorbance was measured using an ELISA reader.

#### **Statistical analysis**

All analyses were performed using the spss 16.0 statistical software package (SPSS Inc., Chicago, IL, USA) or Microsoft Excel. Groups of three were analyzed by one-way ANOVA. Student's *t* test was used to analyze data for two groups in the cell experiments. A non-parametric test was used to determine associations among clinicopathological variables. Differences between qualitative variables were compared with the chi-square test (Pearson's test) or

Fisher's exact test. Continuous parameters were presented as the mean  $\pm$  SD. Every experiment was completed independently at least three times. A *P* value <0.05 was considered significant.

## Results

# IL-24 is the target gene of miR-203a-3p.1 in HCC cells

Six human HCC cell lines were selected to detect IL-24 expression. We found that in two HCC cell lines, IL-24 expression was significantly lower than the normal cell line (Fig. 1A). To look for miRNAs targeting IL-24 in HCC cells, online prediction software (TARGETSCAN 7.0 and MIRBASE) was used to search for potential target genes. IL-24 expression is regulated by many miRNAs. The miRNAs with a high ability of IL-24 regulation are those such as miR-425-5p, miR-200b-3p, miR-200c-3p, miR-203a-3p.1 and others. We selected miR-203a-3p.1 because IL-24 mRNA expression was down-regulated significantly in Huh7 and Hep3B cells (Fig. 1B). The predicted targeting sequence of miR-203a-3p.1 is shown in Fig. 1C. IL-24

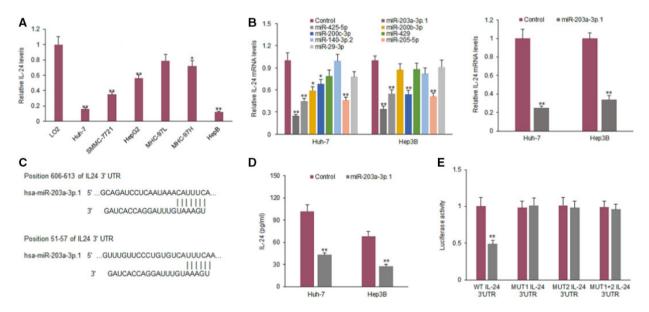
protein decreased in the Huh7 and Hep3B cell lines with miR-203a-3p.1 (Fig. 1D). Results from a luciferase assay showed that the luciferase activity of IL-24 3'-UTR (wild-type) in HCC cells was suppressed by miR-203a-3p.1, but was not changed for the mutated IL-24 3'-UTR (Fig. 1E).

#### miR-203a-3p.1 expression in clinical HCC samples

Firstly, the expression of miR-203a-3p.1 in HCC samples was examined by real-time RT-PCR. The levels of miR-203a-3p.1 were higher in HCC tissues than the normal samples (Fig. 2A). The average levels of miR-203a-3p.1 in HCC tissues were analyzed and the result showed that the levels of miR-203a-3p.1 in the tumor tissues were higher than in the normal tissues (Fig. 2B). These results suggested that miR-203a-3p.1 may promote HCC progression.

# High miR-203a-3p.1 expression in HCC cells increases cellular growth

To explore the role of miR-203a-3p.1 in cellular growth, Huh7 and Hep3B cells were transfected with miR-203a-3p.1 or the control for cell survival assays.



**Fig. 1.** The gene for IL-24 is the target of miR-203a-3p.1 in HCC cells. (A) IL-24 expression levels in HCC cell lines (SMMC-7721, HepG2, Huh7, Hep3B, 97L and 97H) and normal liver cell line (LO2) were examined by ELISA. (B) qRT-PCR was used to analyze IL-24 mRNA expression in CML cells. Huh7 and Hep3B cells were transfected with miR-203a-3p.1, miR-425-5p, miR-200b-3p, miR-200c-3p, miR-429, miR-140-3p.2, miR-205-5p or miR-29-3p mimics and IL-24 mRNA was assayed by qRT-PCR. (C) The IL-24 gene is predicted as a target of miR-203a-3p.1 using TARGETSCAN 7.0. (D) IL-24 protein in HCC cells. Huh7 and Hep3B cells were transfected with miR-203a-3p.1 mimics and IL-24 protein level was assayed by ELISA. (E) Luciferase activity of IL-24 3'-UTR and mutated IL-24 3'-UTR in HCC cells with miR-203a-3p.1 mimics transfection. Huh7 and Hep3B cells were transfected with wild-type or mutant type of IL-24 3'-UTR combined with miR-203a-3p.1 mimics and luciferase activity was assayed with a dual luciferase assay system. \*\**P* < 0.01; \**P* < 0.05.

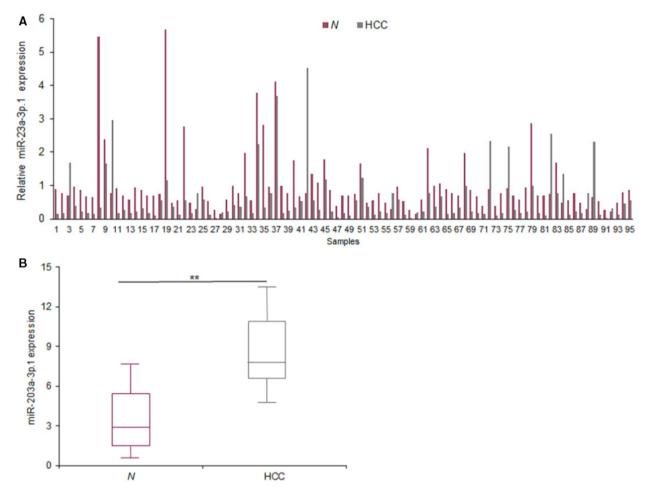


Fig. 2. miR-203a-3p.1 expression in clinical HCC samples. (A) The levels of miR-203a-3p.1 in 95 HCC samples. The levels of miR-203a-3p.1 were examined by real-time RT-PCR. (B) Data analysis from (A). N, normal samples. \*\*P < 0.01.

The MTT assay was used to examine cell proliferation, and the data showed that miR-203a-3p.1 could promote cell growth in Huh7 and Hep3B cells (Fig. 3A,B). We used a colony formation assay to evaluate cell proliferation, and the results showed that miR-203a-3p.1 could significantly increase colony formation rates in Huh7 and Hep3B cell lines (Fig. 3C,D).

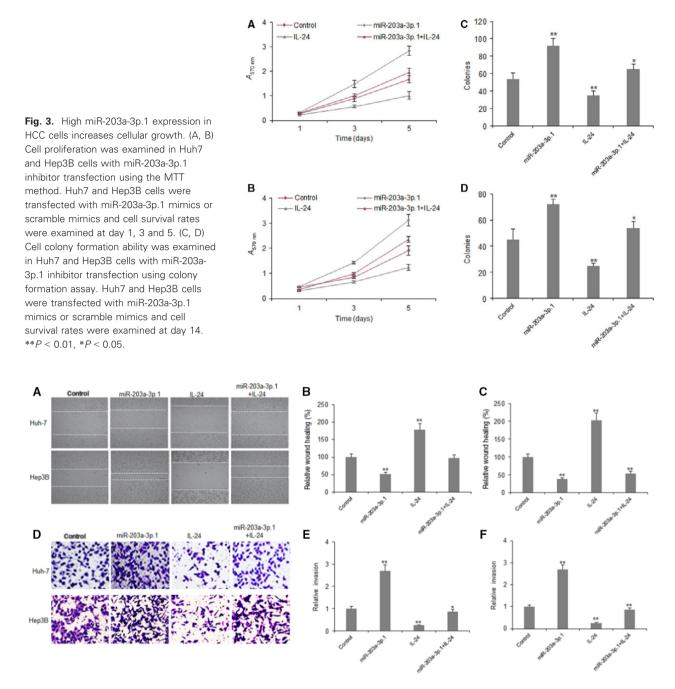
# miR-203a-3p.1 promotes HCC cell metastasis by targeting IL-24

In order to elucidate whether miR-203a-3p.1 is related to cancer metastasis by down-regulation of IL-24, HCC cells were transfected with miR-203a-3p.1 mimics or IL-24, and cell migration and metastasis were assayed with Transwell chambers. The results indicated that miR-203a-3p.1 increased migration of Huh7 and Hep3B cells (Fig. 4A–C). It was also demonstrated that miR-203a-3p.1 promoted invasion of Huh7 and Hep3B cells (Fig. 4D–F).

### Discussion

The gene for IL-24 has been reported as a tumor suppressor and IL-24 plays prominent roles in various cancers such as by inhibiting tumor growth, invasion and metastasis and promoting cell death. When miR-NAs bind to the target genes, the genes' expression is down-regulated at post-transcriptional levels. But IL-24 regulation by miRNAs in cancer has lacked investigation. The present research verified that IL-24 was down-regulated in HCC and its expression, cellular survival and metastasis were regulated by miR-203a-3p.1 in HCC cells.

It was shown that IL-24 is a selective anti-cancer agent regulating endoplasmic reticulum stress. IL-24 inhibits angiogenesis of cancer by suppressing vascular



**Fig. 4.** miR-203a-3p.1 promotes HCC cell metastasis by targeting IL-24. (A) Cell migration was examined in Huh7 and Hep3B cells using wound healing assay. Huh7 and Hep3B cells were transfected with miR-203a-3p.1 mimics or scramble mimics and then treated with IL-24; relative wound healing (%) were analyzed the next day. (B, C) Data from (A) were quantified. Left panel: Huh7 cell; right panel: Hep3B cells. (D) Cell invasion was examined in Huh7 and Hep3B cells using the Transwell system. Huh7 and Hep3B cells were transfected with miR-203a-3p.1 mimics or scramble mimics and then treated with IL-24, and cell invasion was analyzed the next day. (E, F) Data from (D) were quantified. Left panel: Huh7 cell; right panel: Hep3B cells. \*\*P < 0.01, \*P < 0.05.

endothelial growth factor, basic fibroblast growth factor, transforming growth factor, and so on. IL-24 could inhibit tumor metastasis by down-regulating genes for CD44 and matrix metallopeptidase 9 and other metastasis associated genes [13–17]. IL-24 is regulated by miRNAs such as miR-205 in oral cancer [11] and prostate cancer [20]. IL-24 was also found to be regulated by miR-203 but the type of miR-203 was not mentioned [21]. In another report, it was shown that IL-24 regulates miR-221 expression in cancer cells [22]. In this study, we identified that IL-24 was regulated by miR-203a-3p.1 in HCC cells.

The recent studies verify the important regulatory roles of miRNAs in HCC cell proliferation and metastasis. Some of the miRNAs in HCC are up-regulated and others are down-regulated. In our study, we identified that miR-203a-3p.1 was up-regulated in most HCC tissues used in this study and this was related to HCC metastasis. The cellular function analysis showed that miR-203a-3p.1 promoted HCC cell proliferation, migration and invasion by targeting IL-24, which indicates miR-203a-3p.1 is an onco-miRNA in HCC. There are no reports showing the role of miR-203a-3p.1 in HCC and other diseases. A report showed that miR-205 up-regulates IL-24 in KB oral cancer [10]. More studies such as ones using miRNA array need to be carried out to investigate IL-24 regulation by miRNAs.

In summary, the data presented in this study elucidate that miR-203a-3p.1 acts as on onco-miRNA by targeting IL-24. It is possible that miR-203a-3p.1 would be a therapeutic target for HCC. We hope that the combination of IL-24 and inhibiting miR-203a-3p.1 will be a good way to treat HCC in the future.

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# **Author contributions**

WH and ZL conceived and designed the project. WH, MD and XP acquired the data. WH and YG analyzed and interpreted the data. WH and ZL wrote the paper.

## References

- 1 Desai JR, Ochoa S, Prins PA and He AR (2017) Systemic therapy for advanced hepatocellular carcinoma: an update. *J Gastrointest Oncol* **8**, 243–255.
- 2 Menezes ME, Bhatia S, Bhoopathi P, Das SK, Emdad L, Dasgupta S, Dent P, Wang XY, Sarkar D and Fisher PB (2014) MDA-7/IL-24: multifunctional cancer killing cytokine. *Adv Exp Med Biol* **818**, 127–153.
- 3 Persaud L, De Jesus D, Brannigan O, Richiez-Paredes M, Huaman J, Alvarado G, Riker L, Mendez G, Dejoie J and Sauane M (2016) Mechanism of action and applications of interleukin 24 in immunotherapy. *Int J Mol Sci* 17, 869.
- 4 Panneerselvam J, Shanker M, Jin J, Branch CD, Muralidharan R, Zhao YD, Chada S, Munshi A and Ramesh R (2015) Phosphorylation of interleukin

- 5 Yu D, Zhong Y, Li X, Li Y, Li X, Cao J, Fan H, Yuan Y, Ji Z, Qiao B *et al.* (2015) ILs-3, 6 and 11 increase, but ILs-10 and 24 decrease stemness of human prostate cancer cells in vitro. *Oncotarget* **6**, 42687– 42703.
- 6 Menezes ME, Shen XN, Das SK, Emdad L, Guo C, Yuan F, Li YJ, Archer MC, Zacksenhaus E, Windle JJ et al. (2015) MDA-7/IL-24 functions as a tumor suppressor gene in vivo in transgenic mouse models of breast cancer. Oncotarget 6, 36928–36942.
- 7 Ma YF, Ren Y, Wu CJ, Zhao XH, Xu H, Wu DZ, Xu J, Zhang XL and Ji Y (2016) Interleukin (IL)-24 transforms the tumor microenvironment and induces anticancer immunity in a murine model of colon cancer. *Mol Immunol* **75**, 11–20.
- 8 Bhoopathi P, Lee N, Pradhan AK, Shen XN, Das SK, Sarkar D, Emdad L and Fisher PB (2016) mda-7/IL-24 Induces cell death in neuroblastoma through a novel mechanism involving AIF and ATM. *Cancer Res* 76, 3572–3582.
- 9 Cheng HR, Wu BQ, Chen L, Zhang ZX and Li B (2015) Expression and effect of serum interleukin-24 level on bone marrow mononuclear cells in children with acute leukemia. *Genet Mol Res* 14, 17281–17288.
- 10 Li J, Yang D, Wang W, Piao S, Zhou J, Saiyin W, Zheng C, Sun H and Li Y (2015) Inhibition of autophagy by 3-MA enhances IL-24-induced apoptosis in human oral squamous cell carcinoma cells. *J Exp Clin Cancer Res* 34, 97.
- Kim JS, Yu SK, Lee MH, Park MG, Park E, Kim SG, Lee SY, Kim CS, Kim HJ, Chun HS *et al.* (2013) MicroRNA-205 directly regulates the tumor suppressor, interleukin-24, in human KB oral cancer cells. *Mol Cells* 35, 17–24.
- 12 Panneerselvam J, Srivastava A, Muralidharan R, Wang Q, Zheng W, Zhao L, Chen A, Zhao YD, Munshi A and Ramesh R (2016) IL-24 modulates the high mobility group (HMG)A1/miR222/AKT signaling in lung cancer cells. *Oncotarget* 7, 70247–70263.
- 13 Tian H, Zhang D, Gao Z, Li H, Zhang B, Zhang Q, Li L, Cheng Q, Pei D and Zheng J (2014) MDA-7/IL-24 inhibits Nrf2-mediated antioxidant response through activation of p38 pathway and inhibition of ERK pathway involved in cancer cell apoptosis. *Cancer Gene Ther* 21, 416–426.
- 14 Shapiro BA, Vu NT, Shultz MD, Shultz JC, Mietla JA, Gouda MM, Yacoub A, Dent P, Fisher PB, Park MA *et al.* (2016) Melanoma differentiation-associated gene 7/IL-24 exerts cytotoxic effects by altering the alternative splicing of Bcl-x pre-mRNA via the SRC/PKCδ signaling axis. *J Biol Chem* **291**, 21669–21681.

- 15 Zhang J, Xu R, Tao X, Dong Y, Lv X, Sun A and Wei D (2016) TAT-IL-24-KDEL-induced apoptosis is inhibited by survivin but restored by the small molecular surviving inhibitor, YM155, in cancer cells. *Oncotarget* 7, 37030–37042.
- 16 Li G, Wu H, Cui L, Gao Y, Chen L, Li X, Liang T, Yang X, Cheng J and Luo J (2015) CD47-retargeted oncolytic adenovirus armed with melanoma differentiation-associated gene-7/interleukin-24 suppresses in vivo leukemia cell growth. *Oncotarget* 6, 43496–43507.
- 17 Panneerselvam J, Jin J, Shanker M, Lauderdale J, Bates J, Wang Q, Zhao YD, Archibald SJ, Hubin TJ and Ramesh R (2015) IL-24 inhibits lung cancer cell migration and invasion by disrupting the SDF-1/CXCR4 signaling axis. *PLoS ONE* 10, e0122439.
- 18 Yao M, Wang L, Yao Y, Gu HB and Yao DF (2014) Biomarker-based microRNA therapeutic strategies for

hepatocellular carcinoma. J Clin Transl Hepatol 2, 253–258.

- 19 Hayes CN and Chayama K (2016) MicroRNAs as biomarkers for liver disease and hepatocellular carcinoma. *Int J Mol Sci* 17, 280.
- 20 Majid S, Dar AA, Saini S, Yamamura S, Hirata H, Tanaka Y, Deng G and Dahiya R (2010) MicroRNA-205directed transcriptional activation of tumor suppressor genes in prostate cancer. *Cancer* 116, 5637–5649.
- 21 Primo MN, Bak RO, Schibler B and Mikkelsen JG (2012) Regulation of pro-inflammatory cytokines TNFα and IL24 by microRNA-203 in primary keratinocytes. *Cytokine* 60, 741–748.
- 22 Pradhan AK, Talukdar S, Bhoopathi P, Shen XN, Emdad L, Das SK, Sarkar D and Fisher PB (2017) mda-7/IL-24 mediates cancer cell-specific death via regulation of miR-221 and the Beclin-1 axis. *Cancer Res* 77, 949–959.