

ORIGINAL RESEARCH

ZNF277 regulates ovarian cancer cell proliferation and invasion through inhibition of PTEN

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Department of Gynaecology, Linyi Central Hospital, Linyi, Shandong, 276400, People's Republic of China **Background:** ZNF277 is a transcription factor that is overexpressed in several cancers. However, its clinical role in ovarian cancer (OC) has not been reported yet. The present study aims to investigate the expression of ZNF277 in patients with OC, and to reveal the effects of ZNF277 on the proliferation, migration, and invasion of OC cells.

Methods: Using The Cancer Genome Atlas database, we found that higher expression of ZNF277 was correlated with poorer survival times of OC patients. This study used functional experiments, such as Cell Counting Kit-8 assay, colony formation assay, wound healing assay, and transwell invasion assay. Mechanistically, using quantitative chromatin immunoprecipitation assay, luciferase reporter assay, quantitative reverse-transcription PCR, and Western blot we identified the potential mechanism.

Results: We confirmed for the first time that the expression of ZNF277 is significantly increased in OC tissues and cell lines and that it is closely associated with the adverse clinical features of OC patients. We demonstrated that overexpression of ZNF277 potentiated the proliferation, migration, and invasion of SKOV3 and OVCAR3 loss-of-function experiments showed that the silencing of ZNF277 reduced the proliferation, migration, and invasion of OC cells. Mechanistically, using quantitative chromatin immunoprecipitation assay, luciferase reporter assay, quantitative reverse-transcription PCR, and Western blot we identified that PTEN was a direct downstream target for ZNF277. PTEN expression antagonized the tumor-promoting function of ZNF277.

Conclusion: Taken together, the results of the current study demonstrated that ZNF277 exerted a promoting role in the progression of OC and might act as a promising biomarker and therapeutic target for OC patients.

Keywords: ZNF277, ovarian cancer, proliferation, migration, invasion, PTEN

Introduction

Ovarian cancer (OC) is a common malignant tumor with a serious threat to the health of females all over the world. Despite numerous advances in integrated therapeutic approach, including surgery, chemotherapy, radiotherapy, and immune therapy, the overall survival rate remains unsatisfactory with a mean 5-year survival rate of ~40%. Since frequent relapse and metastasis result in poor prognosis, the examination of the underlying mechanisms involved in OC remains to be elucidated. 5

PTEN, a dual lipid/protein phosphatase,⁶ works as a well-known tumor suppressor by negatively regulating the AKT signaling pathway,⁷ and has been found to play important roles in the progression of various human cancers including OC.^{8,9} Accumulating studies have confirmed that several miRNAs such as MiR-200a,¹⁰ MiR-19b,¹¹ and MiR-216a¹² could promote the invasion and migration of OC by targeting the 3'UTR of PTEN, thus suppressing the PTEN/AKT pathway. However, the suppression of PTEN by the binding of transcription factors to the promoter region in OC remains unclear.

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ZNF277 contains five repeats of a C₂H₂-type zinc finger motif, with RNA polymerase II transcription factor activity, sequence-specific DNA binding activity, and is localized to human chromosome 7q31.1.13 With evolutionary conservation, ZNF277 has been reported to be a critical regulator in differentiation and plays an important role in cellular senescence and cancer. 14,15 Till now, the function of ZNF277 in OC remains to be investigated. The objective of the study is to offer insights into ZNF277 function and suggest ZNF277 as a potential target for OC.

Materials and methods

Tissue samples and cell lines

All tissue samples were obtained from the Department of Gynaecology, Linyi Central Hospital (Shandong, China) between February 2011 and December 2013. Eighty-six tissue specimens and their corresponding adjacent non-tumor tissues were obtained from female patients, with age ranging from 40 to 55 years, who underwent tumor surgical resections. The tissues were stored at -80°C for later use. All patients had not undergone chemo- or radiotherapy before surgery. The study was approved by the Clinical Research Ethics Committee of Linyi Central Hospital (No 2013-111). The informed consent and written approvals were obtained from all the participants.

The human OC cell lines SKOV3, OVCAR3, A2780, and CAOV3 were purchased from the American Type Culture Collection (ATCC, Manassas, VA, USA). Normal human ovarian surface epithelial cells (HOSEPiCs) were purchased from the Cell Bank of Type Culture Collection of Chinese Academy of Sciences (Shanghai, China). The relative cells were cultured in DMEM or RPMI-1640, respectively (Thermo Fisher Scientific, Waltham, MA, USA), containing 10% FBS, 100 U/mL penicillin, and 100 mg/mL streptomycin (Thermo Fisher Scientific), and were cultured at 37°C in a humidified atmosphere with 5% CO₂.

Cell transfection

Lipofectamine® 2000 was obtained from Thermo Fisher Scientific. Transfection was performed according to the protocol recommended by the manufacturer. Forty-eight hours after transfection, the cells were collected and used for further experiments. The sequences of the ZNF277 short hairpin RNA (shZNF277) were purchased from GenePharma Co., Ltd. (Shanghai, China).

RNA extraction and quantitative reversetranscription PCR (RT-qPCR)

Total RNA was extracted using 1 mL of TRIzol® reagent (Thermo Fisher Scientific) from 100 µg of human OC tissues for each specimen or OC cell lines according to the manufacturer's protocol. UV spectrophotometer (Thermo Fisher Scientific) was used to monitor the total RNA concentration and purity. A PrimeScript RT reagent kit (Thermo Fisher Scientific) was used to reverse-transcribe RNA into cDNAs. RT-qPCR system (ABI 7500; Applied Biosystems, Foster City, CA, USA) with SYBR Green Master Mix (Toyobo, Tokyo, Japan) was used for qPCR. GAPDH was used as a reference gene. The thermocycler conditions were as follows: 95°C for 45 seconds and then 40 cycles at 90°C for 15 seconds, 60°C for 10 seconds, and 72°C for 45 seconds. The primers used were as follows: ZNF277: 5'-GAACATGCTTTCAACA TTGG-3' (forward) and 5'-ATCAGACCAGTCATCT TCCT-3' (reverse); GAPDH: 5'-AGCCACATCGCTCAG ACAC-3' (forward) and 5'-GCCCAATACGAC-CAAATCC-3' (reverse). The 2–ΔΔCT method was performed to demonstrate the relative expression of all the target genes. ¹⁶

Western blot

Equal amount of protein (30 µg) in each group was separated by 10% SDS-PAGE and then transferred to nitrocellulose membranes. Next, the membranes were blocked with 5% non-fat milk in Tris-buffered saline with 0.1% Tween (Thermo Fisher Scientific) for 1 hour at room temperature. The membrane was incubated with primary antibodies at 37°C for 2 hours, and then incubated with horseradish peroxidaseconjugated anti-rabbit secondary antibodies (Sigma-Aldrich Co., St Louis, MO, USA) at 37°C (dilution, 1:5,000) for 2 hours. The protein bands were detected using Pierce Electrochemiluminescence Plus Substrate (Thermo Fisher Scientific) on Kodak film developer (Fujifilm, Tokyo, Japan). The primary antibodies used were as follows: PTEN (ab32199; Abcam, Cambridge, UK) and ZNF277 (ab96299; Abcam).

Luciferase reporter assay

Luciferase reporter assay was performed using the Dual-Luciferase Reporter Assay System (Promega Corporation, Madison, WI, USA) according to the manufacturer's protocol. The relative cells were cultured in 24-well plates at 37°C overnight, and then co-transfected with luciferase reporter plasmids with wild-type or mutant PTEN reporter constructs together with the ZNF277 overexpression or knockdown construct which were purchased from Genepharma (Shanghai, China). Forty-eight hours after transfection, cells were harvested and the luciferase activity was measured.

Quantitative chromatin immunoprecipitation assay

Briefly, cells were cross-linked with 1% formaldehyde, sonicated, and pre-cleared. After incubation with 5–10 μg

of the appropriate antibody, protein A/G Sepharose CL-4B beads were added into the immune complex. The beads were then washed in buffers and DNA was eluted for quantitative chromatin immunoprecipitation assay.

Proliferation assay

The 100 μ L OV cells (5×10³ cells/well) were seeded in 96-well culture plates (Corning Incorporated, New York, NY, USA), and after culturing for 24, 48, 72, and 96 hours at 37°C, 20 μ L of MTT reagent (Dojindo Molecular Technologies, Inc., Kumamoto, Japan) was added and plates were incubated for another 4 hours. The supernatant was removed, and 150 μ L of dimethyl sulfoxide was added into each well. The absorbance was measured using an ELX-800 spectrometer reader (Omega Bio-Tek, Inc., Norcross, GA, USA) at 490 nm. The OD values represent the relative proliferation level of the cells. The experiments were performed in triplicate.

Colony formation assay

The transfected cells were seeded into six-well plates with 1,000 cells/well, and cultured at 37°C with 5% CO₂ for 14 days. Then, colonies were fixed with 100% methanol for 30 minutes and stained with 0.2% crystal violet for 15 minutes (Thermo Fisher Scientific) at room temperature. The total number of colonies (>50 cells) was counted using light microscopy (magnification, ×40; Olympus, Tokyo, Japan). The experiment was performed in triplicate.

Wound healing assay

In order to detect the migration ability, transfected cells were trypsinized and seeded into a 12-well plate at the density of 2×10^5 cells per well. After overnight incubation, we scratched the confluent monolayer across the plate using a 10 μL pipette tip. The wells were washed with PBS for three times to make sure no suspending cells existed in the wound areas. The wounded monolayer was photographed immediately. Subsequently, the cells were incubated in low-serum medium for 24 hours and the wound images were taken again. The migration ability was calculated by the ratio of the healing width at 24 hours to the wound width at 0 hours.

Transwell invasion assay

Invasion assays were performed using transwell inserts (8 μ M pore size; Costar, Cambridge, MA, USA). The transwell inserts were coated with 100 μ L of Matrigel (BD Biosciences) and 500 μ L of FBS-free DMEM was added into 24-well plates. The OC cells were added into the top chamber, while 1,000 μ L of DMEM with 20% FBS

was added into the bottom chamber. After incubation for 48 hours, 4% paraformaldehyde was used to fix the invasive cells for 20 minutes, and then 0.5% crystal violet (Beyotime) was used for staining. The cells were counted under an inverted microscope (magnification, ×40, TS100; Nikon, Tokyo, Japan) in five random fields of view. The experiment was repeated three times.

Statistical analysis

All statistical analyses were performed using SPSS 17.0 software (SPSS, Inc., Chicago, IL, USA). Data were presented as the mean ± SD. The log-rank test was used for the Kaplan–Meier curve to evaluate the association between the expression of ZNF277 and the survival rate. Parametric statistics were carried out, Student's *t*-test was used to compare differences between the two groups, and two-way ANOVA was used for comparison amongst three groups or between two groups with two factors, followed by a least significant difference post hoc comparison test. *P*<0.05 was considered to indicate a statistically significant difference. All experiments were repeated at least three times.

Results

ZNF277 expression was upregulated in ovarian cancer tissues and cell lines

As we know, the expression profile of ZNF277 in OC has not been elucidated yet. In the present study, we first use the online database of the Human Protein Atlas. The expression pattern of ZNF277 in OC tissues and normal ovary was examined by using IHC data from https://www. proteinatlas.org/, as Uhlen et al reported.¹⁷ As shown in Figure 1A, ZNF277 was upregulated in OC tissues. Further, we used the The Cancer Genome Atlas (TCGA) database to analyze the correlation between the expression of ZNF217 and the prognosis. Survival analysis showed that patients with ZNF277 overexpression had a significantly shorter overall survival (OS) time compared with those with lower ZNF277 expression (Figure 1B). RT-qPCR was used to detect the expression of ZNF277 mRNA in the primary OC tissues and tumor-adjacent normal tissues obtained from patients treated in Linyi Central Hospital. The OC tissues had significantly increased expression of ZNF277 mRNA compared with the adjacent normal tissues, and the difference was statistically significant (Figure 1C). As shown in Table 1, higher expression of ZNF277 was correlated with the lymph node metastasis, tumor size, and Federation Internationale of Gynecologie and Obstetrigue stage. Further, the expression of ZNF217 mRNA was assessed in SKOV3, Liu et al Dovepress

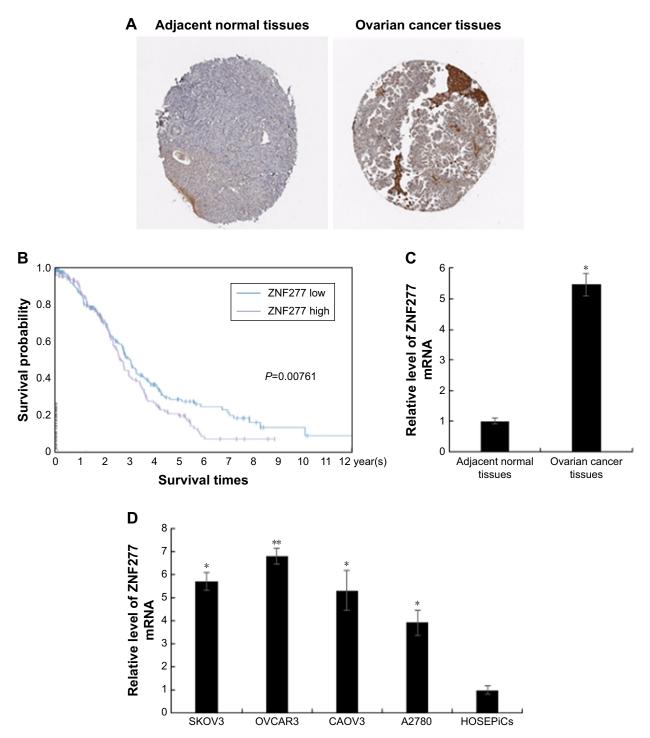


Figure 1 ZNF277 expression was upregulated in ovarian cancer (OC) tissues and cell lines. (A) The Human Protein Atlas database was used to detect the expression of ZNF277 in OC tissues and normal ovary tissues. (B) TCGA database was used to detect the survival times of patients with high ZNF277 expression and those with low ZNF277 expression, and Kaplan–Meier curves were drawn. (C) RT-qPCR was used to reveal the expression of ZNF277 in OC tissues and tumor-adjacent normal tissues, and the reference protein GAPDH was used as a negative control (n=3). Data are presented as the mean ± SD of three independent experiments (n=3; *P<0.05 vs tumor-adjacent normal tissues). (D) RT-qPCR was used to reveal the expression of ZNF277 in the OC cell lines SKOV3, OVCAR3, CAOV3, and A2780 compared to the human ovarian surface epithelial cells (HOSEPiCs). Results are presented as the mean ± standard error of three independent experiments (*P<0.05, **P<0.01 vs HOSEPiCs). Abbreviations: TCGA, The Cancer Genome Atlas; RT-qPCR, quantitative reverse-transcription PCR.

OVCAR3, A2780, CAOV3, and HOSEPiCs cells. As shown in Figure 1D, ZNF277 expression was upregulated in OC cell lines compared with HOSEPiCs. These results indicated that ZNF277 might have an important role in OC progression.

ZNF277 facilitated the proliferation abilities of OC cells

To further explore the function of ZNF277 in the proliferation and invasion of epithelium ovarian cancer, we transfected

Table 1 The clinicopathologic variables identified in 86 ovarian cancer patients

Variables	No (n=86)	ZNF277 protein expression		P-value
		Low (n=32)	High (n=54)	
Age (years)				0.158
<50	38	11	27	
≥50	48	21	27	
Ascites (mL)				0.729
≤100	49	19	30	
>100	37	13	24	
Serum CA-125 (U/mL)				0.468
≤35	44	18	26	
>35	42	14	28	
Tumor size				0.014
≤3 cm	39	20	19	
>3 cm	47	12	35	
Lymph node metastasis				0.012
Negative	44	22	22	
Positive	42	10	32	
FIGO stage				0.044
I–II	39	19	20	
III–IV	47	13	34	

Note: The expression of ZNF277 is based on the mRNA level, and the mean expression is used as cutoff to define the high or low expression of ZNF277. **Abbreviations:** CA, cancer antigen; FIGO, Federation Internationale of Gynecologie and Obstetrigue.

ZNF277 plasmid or ZNF277 siRNA into SKOV3 and OVCAR3 cells. The transfection efficiency was measured by RT-qPCR (Figure 2A and B). Cell Counting Kit-8 (CCK-8) assays were performed to assess the function of ZNF277 on OC cell proliferation. Compared with the control group, SKOV3 and OVCAR3 cells transfected with ZNF277 exhibited significantly increased proliferation ability, while knockdown of ZNF277 decreased the proliferation in a time-dependent manner (Figure 2C and D). Similarly, colony formation assays revealed that the number of cell colonies generated was obviously decreased in the siZNF277 transfected groups, while in the ZNF277 overexpression groups, the colonies were significantly increased (Figure 2E and F).

ZNF277 promoted migration and invasion in OC cells

To fully explore the putative biological functions of ZNF277 using wound healing assay, the cells were treated with ZNF277 and migration pattern was studied for 24 hours. The results showed that the migration capacity of SKOV3 and OVCAR3 cells increased, while knockdown of ZNF277

decreased cell migration (Figures 3A and B and S1). Transwell assays were then performed to monitor the invasive function of ZNF277 on OC cells. As shown in Figure 3C and D, in response to ZNF277 knockdown, the invasive ability of SKOV3 and OVCAR3 cells in vitro was specifically suppressed.

PTEN was a direct downstream target of ZNF277 in OC cells

To elucidate the molecular mechanisms underlying the promotion effect of ZNF277 on OC progression, we further identified the key target genes of ZNF277 in SKOV3 and OVCAR3 cells using qChIP assay; among them, the binding of PTEN is the most remarkable one (Figure 4A). To further confirm that ZNF277 could directly target the promoter of PTEN, luciferase reporter assay was performed in SKOV3 and OVCAR3 cells. As shown in Figure 4B, forced expression of ZNF277 in the two cell lines significantly suppressed the luciferase activity of wild-type promoter of PTEN. On the other hand, inhibition of ZNF277 obviously enhanced the luciferase activity of promoter (Figure 4C), while the overexpression or knockdown of ZNF277 had no significant influence on the mutant promoter of PTEN (Figure 4D). So, we can hypothesize that ZNF277 could regulate PTEN at transcription level. The mRNA and protein expression of PTEN was significantly downregulated in response to ZNF277 overexpression (Figure 4E), while the mRNA and protein level of PTEN increased following the silencing of ZNF277 (Figure 4F).

ZNF277 promoted OC cell proliferation by targeting PTEN

After confirming PTEN was a direct target of ZNF277 in OC, we further investigated whether PTEN has mediator effects on the function of ZNF277 in OC cells. Firstly, PTEN overexpression construct was transfected into SKOV3 and OVCAR3 cells, and RT-qPCR analysis was used to confirm the transfection efficiency of PTEN (Figure 5A). Then CCK-8 assay showed that overexpression of PTEN could reduce the proliferation ability and partly abrogate the promoting effects of ZNF277 of the two cells (Figure 5B and C). Consistent with the proliferation assay, overexpression of PTEN in cells transfected with ZNF277 also resulted in retarded colony formation (Figure 5D and E).

ZNF277 regulated OC cell migration and invasion through PTEN

Previous reports have indicated that PTEN has an important role in cell migration and invasion. Therefore, we probed Liu et al Dovepress

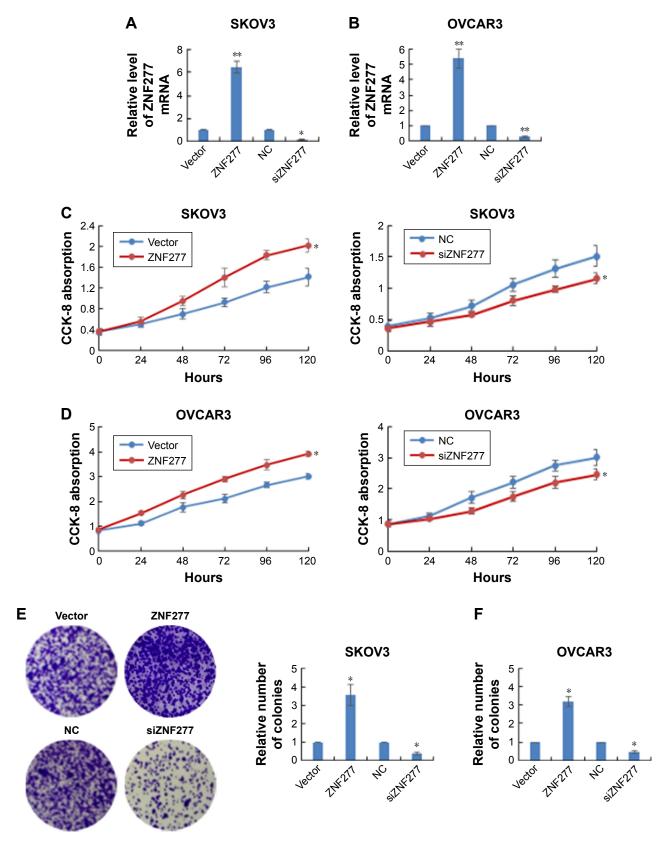


Figure 2 ZNF277 facilitated the proliferation ability of OC cells. (**A**) RT-qPCR was used to analyze the mRNA expression levels of ZNF277 after exposure to the ZNF277 plasmid or ZNF277-specific siRNA in SKOV3 and OVCAR3 cells (**B**). (**C**) ZNF277 affected the growth rate of OC cells. Cell growth viability was assayed in SKOV3 and OVCAR3 cells (**D**) transfected with vector, ZNF277 plasmid, NC, and siZNF277 using CCK-8 at 0, 24, 48, 72, and 96 hours time points. (**E**) ZNF277 facilitated the colony formation of OC cells. Colony formation was performed in stably overexpressing ZNF277 and in ZNF277 knockdown SKOV3 and OVCAR3 cells (**F**). Results are presented as mean ± SD, and error bars represent the mean ± SD of three independent experiments. *P<0.05, **P<0.01 as compared with the control group. **Abbreviations:** OC, ovarian cancer; RT-qPCR, quantitative reverse-transcription PCR; NC, negative control; CCK-8, Cell Counting Kit-8.

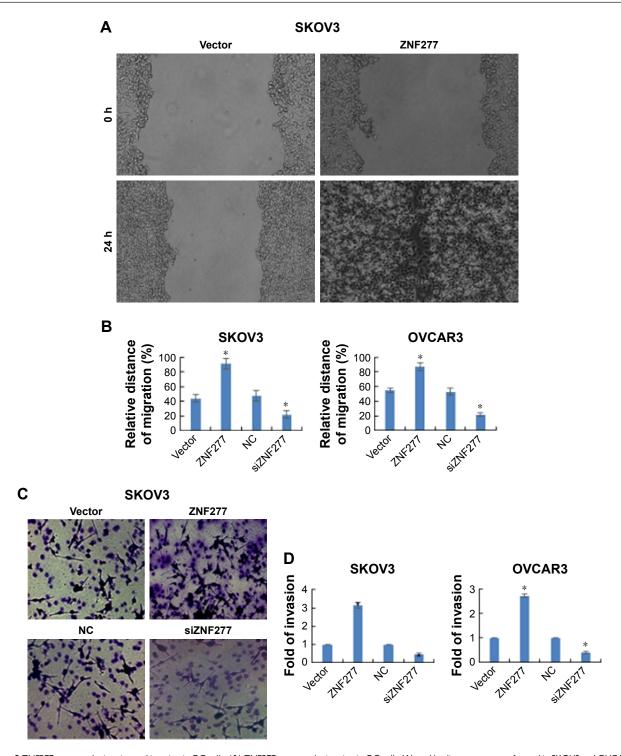


Figure 3 ZNF277 promoted migration and invasion in OC cells. (A) ZNF277 promoted migration in OC cells. Wound healing assay was performed in SKOV3 and OVCAR3 cells (B) following transfection with vector, ZNF277 plasmid, NC, and siZNF277. Analysis of the wound closure rate is shown, and data are presented as the mean ± SD of at least three independent experiments. (C) ZNF277 promoted invasion in OC cells. Transwell assays were used to measure the effect of ZNF277 on cell invasion in SKOV3 and OVCAR3 cells (D) transfected with vector, ZNF277 plasmid, NC, and siZNF277. Representative images of the invasion assays are shown. The data are presented as mean ± SD of three independent experiments. *P<0.05 as compared with the control group.

Abbreviations: OC, ovarian cancer; NC, negative control.

whether the promotion of OC cell migration and invasion by ZNF277 was mediated by targeting PTEN. As shown in Figure 6A and B, when SKOV3 and OVCAR3 were transfected with PTEN, the migratory ability was significantly decreased compared with that of the control group, whereas PTEN restoration counteracted the promotion of migrated SKOV3 and OVCAR3 cells by ZNF277. The effect of PTEN on OC cell invasion was also evaluated. Transwell

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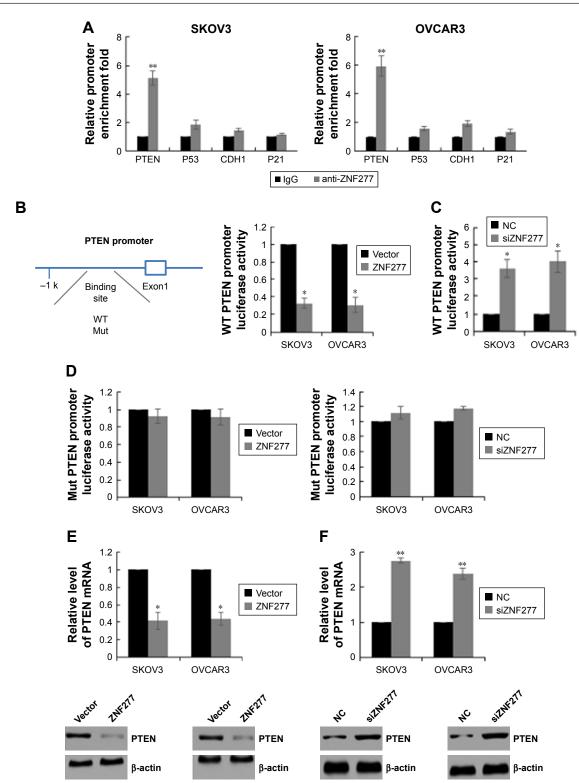


Figure 4 PTEN was a direct downstream target of ZNF277 in OC cells. (A) The binding of ZNF277 to the promoter of PTEN. qChIP was performed in SKOV3 and OVCAR3 cells to identify key target genes for ZNF277, such as P53, PTEN, CDH1, and P21. Normal IgG was used as a negative control (**P<0.01 as compared with the normal IgG). (B) Luciferase reporter activities driven by wild-type PTEN promoter were examined in SKOV3 or OVCAR3 cells transfected with Vector or ZNF277. (C) Luciferase reporter activities driven by wild-type PTEN promoter were examined in SKOV3 or OVCAR3 cells transfected with NC or siZNF277. Error bars indicate mean ± SD of three independent experiments (*P<0.05 vs Vector). (D) Luciferase reporter activities driven by mutant (Mut)-type PTEN promoter were examined in SKOV3 or OVCAR3 cells transfected with vector or ZNF277, NC or siZNF277. (E) Vector or ZNF277 was transfected into SKOV3 and OVCAR3 cells, and then the expression levels of PTEN were monitored using RT-qPCR and Western blot. The experiment was performed in triplicate and subsequently subjected to statistical analysis. (F) NC or siZNF277 was transfected into SKOV3 and OVCAR3 cells, and then the expression levels of PTEN were monitored using RT-qPCR and Western blot. Error bars indicate mean ± SD of three independent experiments (*P<0.05, **P<0.01 vs NC).

Abbreviations: OC, ovarian cancer; RT-qPCR, quantitative reverse-transcription PCR; qChIP, quantitative chromatin immunoprecipitation; NC, negative control; WT, wild type; Mut, mutant type.

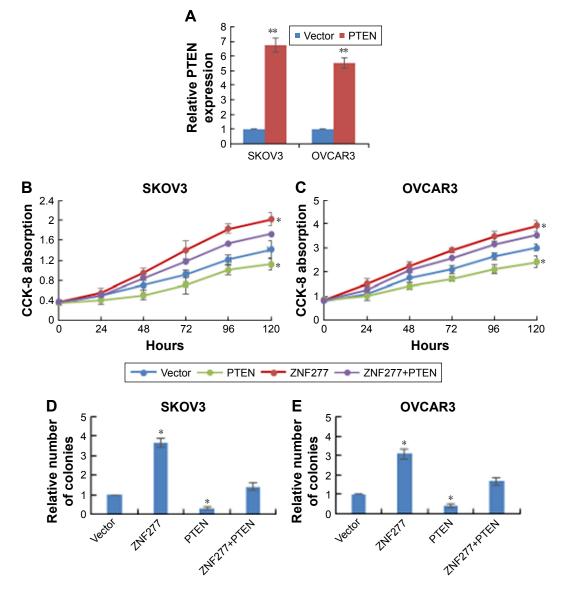


Figure 5 ZNF277 promoted OC cell proliferation by targeting PTEN. (**A**) The mRNA expression of PTEN was determined by RT-qPCR in SKOV3 and OVCAR3 cells transfected with PTEN overexpression plasmid. Error bars indicate mean ± SD of three independent experiments (**P<0.05 vs vector). (**B**) Cell viability of SKOV3 cells and (**C**) OVCAR3 cells transfected with vector, PTEN, ZNF217, and ZNF217+PTEN was determined by CCK-8 assay. Error bars represent mean ± SD of three independent experiments (*P<0.05 vs vector). (**D**) Cell growth of SKOV3 cells and (**E**) OVCAR3 cells transfected with vector, PTEN, ZNF217, and ZNF217+PTEN was determined by colony formation assay. Error bars represent mean ± SD of three independent experiments. *P<0.05 vs vector. **Abbreviations:** OC, ovarian cancer; RT-qPCR, quantitative reverse-transcription PCR; CCK-8, Cell Counting Kit-8.

assay showed the number of invasive cells was significantly decreased by PTEN overexpression. Analogously, the increased invasion of ZNF277 cells was reduced by PTEN overexpression (Figure 6C and D).

Discussion

Thus far, not only literature but also gene bank searches have revealed little regarding the function of ZNF277. Although ZNF277 overexpression is reported in chronic lymphocytic leukemia, colon neoplasia, and endocrine tumors, ¹⁵ the role of ZNF277 in OC still remains elusive. Herein, we demonstrate the important role of ZNF277 in promotion of OC. OC tissues

and cell lines exhibited increased expression of ZNF277 compared with the normal tissues and cell lines. Furthermore, using TCGA database, survival analysis showed that higher level of ZNF277 was closely associated with poorer overall survival of OC patients. Therefore, ZNF277 might play an oncogenic role in OC. The cells transfected with ZNF277 exhibited significantly increased viability and proliferation, while knockdown of ZNF277 markedly decreased the growth rate. We also found that ZNF277 could regulate the migration and invasion of OC cells.

The C₂H₂-type zinc finger motif of ZNF277 is found in numerous transcription factors, ¹⁸ and the capacity of the

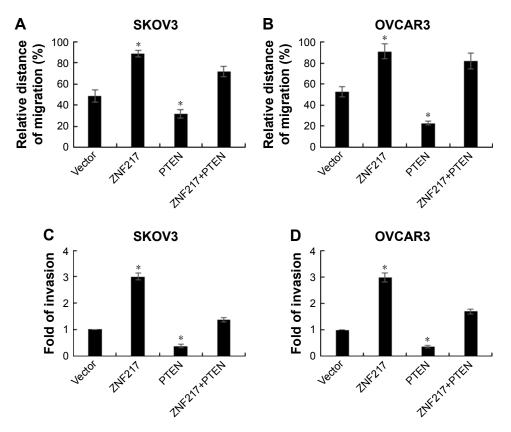


Figure 6 ZNF277 regulated OC cell migration and invasion through PTEN. (A) ZNF277 regulated OC cell (SKOV3) migration through PTEN (* * P<0.05 vs vector). (B) OVCAR3 cells were transfected with vector, PTEN, ZNF217, and ZNF217+PTEN, respectively, and wound healing assay was performed; error bars represent the mean \pm SD of three independent experiments (* * P<0.05 vs vector). (C) ZNF277 regulated OC cell (SKOV3) invasion through PTEN (* * P<0.05 vs vector). (D) OVCAR3 cells were transfected with vector, PTEN, ZNF217, and ZNF217+PTEN, respectively, and transwell assay was performed accordingly. Error bars represent mean \pm SD of three independent experiments (* * P<0.05 vs vector). Abbreviation: OC, ovarian cancer.

zinc finger motif is to bind specific DNA elements. Here, we reported ZNF277 could directly bind to the promoter of PTEN. We identified PTEN as the downstream target gene of ZNF277. The evidence was as follows: overexpression or suppression of ZNF277 could significantly influence the luciferase activity of wild-type promoter of PTEN, while little effect was observed on the mutant promoter of PTEN. Overexpression of ZNF277 resulted in significantly inhibited mRNA and protein level of PTEN, while knockdown of ZNF277 led to obviously increased PTEN expression. Although a panel of putative tumor suppressors were also measured upon ZNF277 overexpression or knockdown, only PTEN expression was shown to be significantly altered. By downregulating the targeted PTEN, ZNF277 may activate the PI3K/Akt signaling pathway, resulting in the OC cell proliferation and invasion. Additionally, the link between ZNF277 and PTEN has rarely been established before. To our knowledge, our study represents the first evidence that ZNF277 suppresses PTEN in OC. PTEN is a wellknown tumor suppressor which plays an important role in the development of various human cancers. 6,9 It is widely known that loss of PTEN could enhance cancer cell proliferation, migration, and invasion, thereby leading to poor prognosis of patients with OC. ^{19,20} Given the significant roles of PTEN, we conclude that ZNF277 is an important molecule in OC. These data also indicate that PTEN is not only the downstream target of ZNF277 but also the functional mediator of ZNF277 in OC, and might provide a potential targeted therapy for the treatment of OC. In the future, further investigations are warranted to explain the clinical utility of ZNF277.

Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Disclosure

The authors report no conflicts of interest in this work.

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Supplementary material

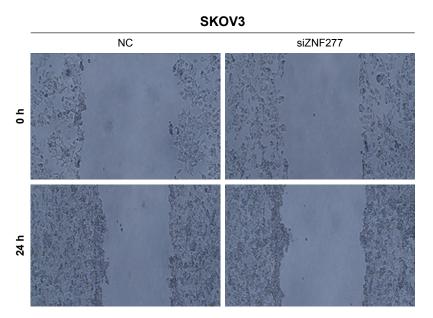


Figure \$1 ZNF277 promoted migration in OC cells. Wound healing assay was performed in SKOV3 cells following transfection with NC and siZNF277. Pictures of the

Abbreviations: OC, ovarian cancer; NC, negative control.

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